

Cardioprotective Effect Afforded by Transient Exposure to Phosphodiesterase III Inhibitors

The Role of Protein Kinase A and p38 Mitogen-Activated Protein Kinase

Shoji Sanada, MD; Masafumi Kitakaze, MD, PhD; Philip J. Papst, BS; Hiroshi Asanuma, MD; Koichi Node, MD; Seiji Takashima, MD; Masanori Asakura, MD; Hisakazu Ogita, MD; Yulin Liao, MD; Yasuhiko Sakata, MD; Akiko Ogai, PhD; Tomi Fukushima, BS; Junko Yamada, BS; Yoshiro Shinozaki, MD; Tsunehiko Kuzuya, MD; Hidezo Mori, MD, PhD; Naohiro Terada, MD, PhD; Masatsugu Hori, MD, PhD

Background—Phosphodiesterase III inhibitors (PDEIII-Is) improve the hemodynamic status of heart failure via inotropic/vasodilatory effects attributable to the increase in intracellular cAMP level. Direct cardioprotection by PDEIII-Is and its underlying mechanisms, however, have not been identified. We tested the infarct size-limiting effect of PDEIII-Is and the roles of cAMP, protein kinase (PK) A, PKC, and mitogen-activated protein kinase (MAPK) families in open-chest dogs.

Methods and Results—Milrinone, olprinone (PDEIII-Is), or dibutyryl-cAMP (db-cAMP) was injected intravenously 30 minutes before 90-minute ischemia, followed by 6 hours of reperfusion. Olprinone was also examined with an intracoronary cotreatment with a PKA inhibitor (H89), a PKC inhibitor (GF109203X), an extracellular signal-regulated kinase kinase (MEK) inhibitor (PD98059), or a p38 MAPK inhibitor (SB203580) throughout the preischemic period. Either PDEIII-Is or db-cAMP caused substantial hemodynamic changes, which returned to control levels in 30 minutes. Collateral flow and percent risk area were identical for all groups. Both PDEIII-Is and db-cAMP increased myocardial p38 MAPK activity during the preischemic period, which was blocked by H89, but not by GF109203X. Both PDEIII-Is and db-cAMP reduced infarct size ($19.1 \pm 4.1\%$, $17.5 \pm 3.3\%$, and $20.3 \pm 4.8\%$, respectively, versus $36.1 \pm 6.2\%$ control, $P < 0.05$ each). Furthermore, the effect of olprinone was blunted by either H89 ($35.5 \pm 6.4\%$) or SB203580 ($32.6 \pm 5.9\%$), but not by GF109203X or PD98059. H89, GF109203X, PD98059, or SB203580 alone did not influence infarct size.

Conclusions—Pretreatment with PDEIII-Is has cardioprotective effects via cAMP-, PKA-, and p38 MAPK-dependent but PKC-independent mechanisms in canine hearts. (*Circulation*. 2001;104:705-710.)

Key Words: phosphodiesterase ■ infarction ■ kinases

Phosphodiesterase type III inhibitors (PDEIII-Is) used for the treatment of severe heart failure have 2 major cardiovascular effects,¹ ie, vasodilatory and inotropic effects, via the elevation of intracellular cAMP levels in both vascular smooth muscle cells² and cardiomyocytes.³ The former effect reduces vascular resistance, and the latter increases myocardial contractility to improve hemodynamic status,⁴ with less cardiac oxygen demand than catecholamines.⁵ It has been shown,⁶ however, that long-time exposure to either inotropic or cardiotoxic agents fails to provide any better outcomes. Conversely, Adamopoulos et al⁷ reported that intermittent short-time exposure to dobutamine, a potent β -adrenoceptor agonist, for 3 weeks causes better functional recovery and exercise tolerance for several months in patients with chronic

heart failure. Furthermore, Lochner et al⁸ showed that transient β -adrenergic stimulation before sustained ischemia mimics the cardioprotection of ischemic preconditioning.⁹ Thus, we hypothesized that there is a novel cardioprotective mechanism afforded by transient but not persistent use of inotropic/cardiotoxic agents that overcome or mask the demerits of long-term β -adrenergic stimulation, such as β -adrenoceptor downregulation¹⁰ or proarrhythmic effects.⁶

Treatment with PDEIII-Is or catecholamines increases intracellular calcium levels ($[Ca^{2+}]_i$) in cardiomyocytes.³ We have reported that a transient increase in myocardial $[Ca^{2+}]_i$ mimics the infarct size limitation by ischemic preconditioning via protein kinase C (PKC) activation.¹¹ Therefore, transient pretreatment with PDEIII-Is may reduce infarct size,

Received January 26, 2001; revision received April 10, 2001; accepted April 11, 2001.

From the Department of Internal Medicine and Therapeutics, Osaka University Graduate School of Medicine, Suita, and the Department of Physiological Science, Tokai University School of Medicine, Isehara (Y. Shinozaki, H.M.), Japan; and the Department of Pediatrics, National Jewish Medical Research Center, Denver, Colo (P.J.P., N.T.).

Correspondence to Masafumi Kitakaze, MD, PhD, Department of Internal Medicine and Therapeutics, Osaka University Graduate School of Medicine, 2-2 Yamadaoka, Suita, 565-0871, Japan. E-mail kitakaze@medone.med.osaka-u.ac.jp

© 2001 American Heart Association, Inc.

Circulation is available at <http://www.circulationaha.org>

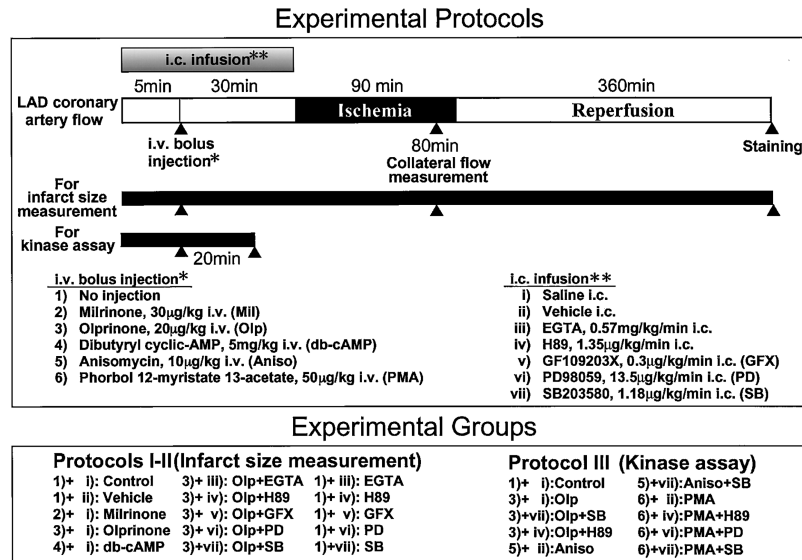


Figure 1. All experimental protocols (top) and experimental groups (bottom) in this study. Abbreviations are in parentheses.

which may be mediated through the transient increase in intracellular cAMP or $[Ca^{2+}]_i$ and PKC activation. Other reports showed that PKC activates extracellular signal-regulated kinase (ERK) 1/2, c-Jun N-terminal protein kinase (JNK), and 38-kDa mitogen-activated protein kinase (p38 MAPK),^{12,13} which are also reported to merge into the cascade in the early-phase cardioprotection in ischemic preconditioning.^{14,15} Lochner et al,⁸ however, showed that the cardioprotection afforded by β -adrenergic stimulation does not involve PKC. Taking them together, the cardioprotection afforded by short-time treatment with PDEIII-Is may involve the cellular signal(s) activated by protein kinase A (PKA) independently of the transient increase in myocardial $[Ca^{2+}]_i$ or PKC activation. No reports, however, have addressed the direct cardioprotection by short-time exposure to PDEIII-Is and the underlying mechanisms. Therefore, we tested whether transient exposure to the potent PDEIII-Is olprinone¹⁶ or milrinone¹⁷ limits infarct size and whether PKA, PKC, or MAPK activation is involved in the underlying mechanisms.

Methods

All procedures were performed in careful conformance with the *Guide for the Care and Use of Laboratory Animals* published by the National Institutes of Health (NIH publication No. 85-23, revised 1985).

Instrumentation

Beagle dogs weighing 9 to 14 kg were anesthetized with sodium pentobarbital (30 mg/kg IV) and prepared as described previously.¹⁸ We cannulated and perfused the left anterior descending coronary artery (LAD) with blood from the left carotid artery through an extracorporeal bypass tube, and aortic blood pressure (ABP) was monitored at this tube. In all experiments, mean ABP, heart rate (HR), and PO_2 in the systemic arterial blood in control conditions averaged 108 ± 2.1 mm Hg, 129 ± 2.2 bpm, and 112 ± 3.5 mm Hg, respectively. ABP and HR were measured continuously during the experiment.

Experimental Protocols

Protocol 1: Effects of PDEIII-Is or cAMP on Infarct Size

We used 34 dogs in this protocol. After hemodynamic stabilization, we administered the PDEIII-Is milrinone (Yamanouchi, 30 μ g/kg) or

olprinone (Eisai, 20 μ g/kg), dibutyryl cAMP (db-cAMP, Sigma, 5 mg/kg), or saline intravenously (Mil group, n=8; Olp group, n=8; db-cAMP group, n=9; and control group, n=9, respectively) and observed hemodynamic changes for 30 minutes. The agents were dissolved in saline. Then, the bypass tube was occluded for 90 minutes, followed by 6 hours of reperfusion. The doses of these agents, determined according to our preliminary experiments (data not shown), provide significant hemodynamic effects and return to the control level in 30 minutes.

Protocol 2: Effects of Inhibition of Calcium, PKA, PKC, and p38 MAPK on the Infarct Size-Limiting Effect of the PDEIII-Is

In another 40 dogs, procedures identical to those in protocol 1 were performed with olprinone, with an intracoronary infusion into the LAD of the calcium chelator EGTA (Sigma; $0.57 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$: 34 mg/mL at an infusion rate of $0.0167 \text{ mL} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$), the selective PKA inhibitor H89 (Alexis; $1.35 \text{ } \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$: 80 μ g/mL at the same rate), the selective PKC inhibitor GF109203X (GFX, Sigma; $0.3 \text{ } \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$: 18 μ g/mL at the same rate), the selective MEK inhibitor PD98059 (PD, Calbiochem; $13.5 \text{ } \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$: 0.8 mg/mL at the same rate), or the selective p38 MAPK inhibitor SB203580 (SB, Calbiochem; $1.18 \text{ } \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$: 70 μ g/mL at the same rate), between 5 minutes before intravenous injection and the onset of bypass occlusion (Olp+EGTA group, n=10; Olp+H89 group, n=7; Olp+GFX group, n=7; Olp+PD group, n=7; and Olp+SB group, n=9, respectively). The drugs were dissolved in saline with polyethylene glycol and ethanol (vehicle; final dose of <1%). The dose of EGTA was confirmed in our previous report¹¹ to block the calcium-induced preconditioning effect without any systemic hemodynamic changes. The doses of other drugs were determined according to previous reports^{11,15} and were also evaluated to inhibit the individual kinases specifically in this system in the present study. In 46 other dogs, we administered EGTA, H89, GF109203X, PD98059, SB203580, or vehicle alone identically for 35 minutes before bypass occlusion to check their effects on infarct size (EGTA group, n=6; H89 group, n=8; GFX group, n=7; PD group, n=7; SB group, n=9; and vehicle group, n=9, respectively). In protocols 1 and 2, we measured myocardial collateral blood flow at 80 minutes of ischemia by injecting nonradioactive microspheres into the left atrium and comparing the content of accumulated spheres in the ischemic myocardium with those in the arterial blood, as described previously.¹⁸ Infarct size was evaluated as described previously.¹⁸ After 6 hours of reperfusion, the LAD was reoccluded, and Evans blue dye was injected intravenously to determine the risk/nonischemic area. The heart was then removed and sliced 6 to 7 mm in width. The nonischemic area was identified

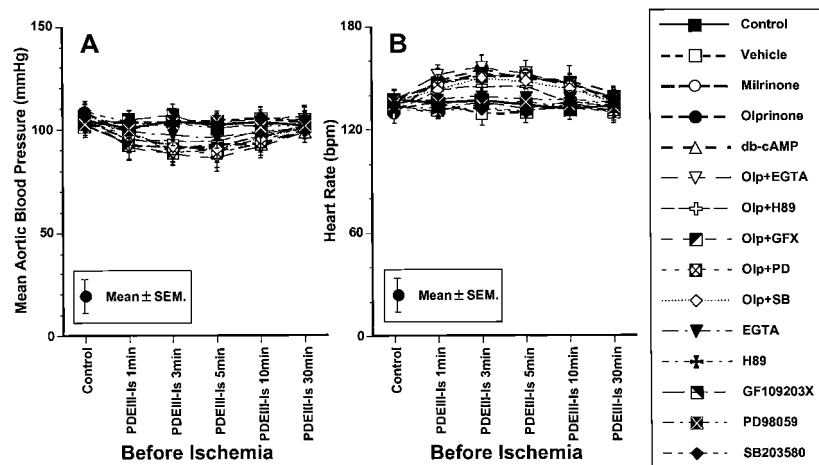


Figure 2. Sequential changes in ABP (A) and HR (B) before sustained ischemia in all 15 groups in protocols 1 and 2.

by blue stain, and the ischemic region was incubated for triphenyl-tetrazolium chloride staining to determine the infarct area.

Protocol 3: Specificity of the Individual Kinase Inhibitors and Effect of PDEIII-Is on the Kinase Activity in the In Vivo Canine Model

We evaluated the specificity of the individual kinase inhibitors used in this study on inhibiting PKA, PKC, or MAPKs (ERK, JNK, and p38 MAPK) in the in vivo canine system. Thirty other dogs received intravenous bolus injection of either db-cAMP (5 mg/kg), phorbol 12-myristate 13-acetate (PMA; 50 μ g/kg), or anisomycin (10 μ g/kg), with intracoronary infusion of saline, H89, PD98059, or SB203580 into the LAD (see Figure 1 for details) ($n=3$ each). After 20 minutes of each injection, we quickly placed samples of myocardial tissue supplied by the LAD into liquid nitrogen and stored them at -80°C . In this protocol, the kinase activities were assayed as follows: for PKC, each myocardial sample was homogenized, separated into membrane and cytosolic fractions by centrifugation, immunoprecipitated, and subjected to Western blotting with specific antibodies against PKC- α , - β , and - γ (Amersham) as described previously.¹⁹ For PKA, JNK, and ERK, each myocardial sample was homogenized, immunoprecipitated, and subjected to Western blotting as described previously^{20,21} with specific antibodies against phospho-CREB (Upstate Biotechnology) for PKA, specific antibodies against phospho-Jun (Upstate Biotechnology) for JNK, and specific antibody against phospho ERK-1/2 (Upstate Biotechnology), respectively. Other dogs, after hemodynamic stabilization, received intravenous bolus injection of either saline, db-cAMP, milrinone, or olprinone (control, db-cAMP, Mil, or the Olp groups, respectively; $n=4$ each), along with an intracoronary infusion of H89 or GF109203X into the LAD from 5 minutes before the intravenous injection and for 25 minutes (Mil+H89, Olp+H89, Mil+GF109203X, or Olp+GF109203X group, respectively; $n=4$ each; see Figure 1 for details). Thereafter, we quickly sampled myocardial tissue into liquid nitrogen and stored it at -80°C . One gram of each myocardial tissue sample in protocol 3 was homogenized, immunoprecipitated, and subjected to in vitro p38 MAPK activity assay as reported previously.²² Then, homogenates in extraction buffer were incubated with rabbit antiserum raised against the COOH-terminal peptide sequence of p38 MAPK (New England Biolabs). After recombinant protein G-Sepharose 4B (Zymed Laboratories) was added, they were incubated and washed with extraction buffer and PAN buffer. For kinase assay, the immunoprecipitates were suspended in assay buffer containing a recombinant amino-terminal fragment of ATF-2 (20 to 50 ng) (Upstate Biotechnology) as a substrate and $[\gamma\text{-}^{32}\text{P}]\text{ATP}$. After the reaction was terminated, the mixture was boiled, separated by electrophoresis on an SDS gel, and subjected to autoradiography. The kinase activity was quantified with a PhosphorImager (Molecular Dynamics).

All details of the above protocols are given in Figure 1.

Criteria for Exclusion

To ensure that all of the animals included in the data analysis of infarct size were healthy and exposed to similar extents of ischemia, the previously described standards¹⁸ were used.

Statistical Analysis

Each value was expressed as mean \pm SEM. Statistical analyses were performed by use of ANOVA with Fisher's post hoc test to determine significance at the $P<0.05$ level for group pairs that exhibited statistically significant differences.

Results

Mortality and Exclusions

One hundred twenty dogs were randomly divided into 15 groups for assessment of infarct size and ventricular fibrillation. Eight and 13 dogs were matched to the exclusion criteria of ventricular fibrillation during sustained ischemia and during reperfusion, respectively. Eleven were also excluded because of myocardial collateral blood flow $>15 \text{ mL} \cdot 100 \text{ g}^{-1} \cdot \text{min}^{-1}$. Therefore, 88 dogs were used for data analysis.

Changes in Hemodynamic Parameters, Risk Area, and Collateral Blood Flow

The current changes of mean ABP and HR were compared in the 15 groups in protocols 1 and 2 before the onset of

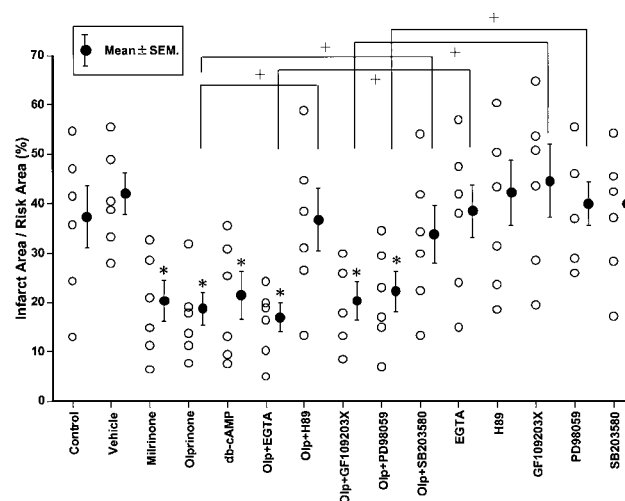


Figure 3. Infarct size in all 15 groups in protocols 1 and 2. + $P<0.05$, * $P<0.05$ vs control.

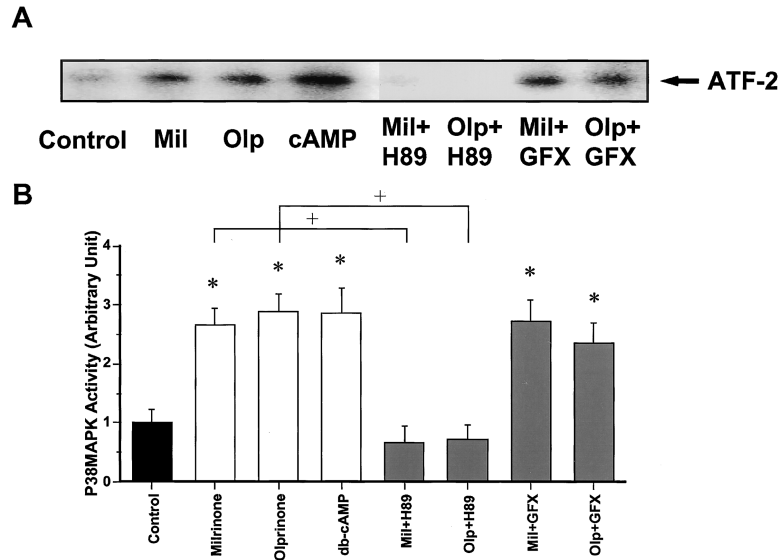


Figure 4. p38 MAPK activity estimated by amount of phosphorylated substrate of activated p38 MAPK. Immunoblotting of representative specimens (A) and changes in mean value (B) (mean±SEM). Each bar indicates mean of n=4 each. Abbreviations as in Figure 1. +*P*<0.05, **P*<0.05 vs control.

coronary occlusion (Figure 2). PDEIII-Is and db-cAMP caused an ≈15% decrease in mean ABP (2A) and ≈15% increase in HR (2B), which was not significantly influenced by the intracoronary administration of EGTA, H89, GF109203X, PD98059, or SB203580 and returned to the control level in 30 minutes. The mean ABP or HR during ischemia and reperfusion and either risk area or collateral blood flow were comparable in all of the 15 groups in protocols 1 and 2 (data not shown).

Infarct Size

Figure 3 shows infarct size in the 15 groups of protocols 1 and 2. Preischemic single treatment with either olprinone, milrinone, or db-cAMP significantly attenuated infarct size (19.1±4.1%, 17.5±3.3%, and 20.3±4.8%, respectively, *P*<0.05 each versus control). This cardioprotective effect of olprinone was blunted by cotreatment with either H89 or SB203580 (35.5±6.4% and 32.6±5.9%, respectively), but not by EGTA, GF109203X, or PD98059 (15.8±2.9%, 19.1±3.9%, and 21.0±4.1%, respectively). Treatment with

EGTA, H89, GF109203X, PD98059, or SB203580 alone did not affect infarct size (37.3±5.3%, 41.0±6.6%, 43.3±7.4%, 38.7±4.5%, and 38.6±6.1%, respectively).

Specificity of the Individual Kinase Inhibitors in This Model and Effect of PDEIII-Is on the Kinase Activity

Figure 4 shows the results for the measurement of p38 MAPK activity, with representative cases (4A) and mean values (4B). Milrinone, olprinone, and db-cAMP equally activated p38 MAPK (2.6- to 2.9-fold), which was completely blunted by H89 and unaffected by GFX.

Figure 5 shows the measurement of PKA (5A), PKC (5B), ERK (5C), or JNK (5D) activity, with the representative cases (top of each panel) and the mean values (bottom of each panel) for each group. Olprinone increased PKA activity in myocardium after 20 minutes of treatment (4.6-fold), which was completely blocked by H89 but was unaffected by SB203580. PMA significantly increased the PKC translocation and the ERK activity (2.8-fold), but olprinone did not activate either of them.

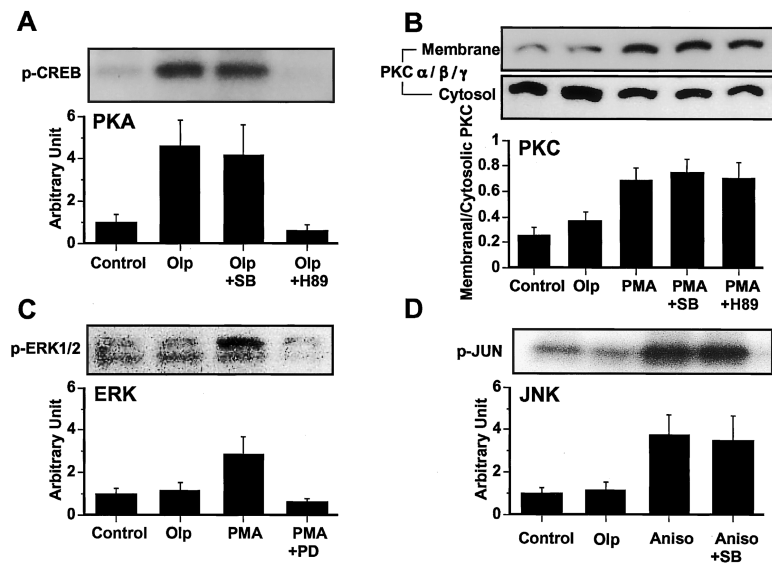


Figure 5. Measurement of PKA (A), PKC (B), ERK (C), and JNK (D) activity, with representative cases (top panels) and mean values (bottom panels) for each group (mean±SEM). See text for details. Each bar indicates mean of n=3 each. Abbreviations as in Figure 1.

Neither H89 nor SB203580 affected the translocation of PKC by PMA. PD98059 blunted the activation of ERK, indicating that the dose of PD98059 in this study can inhibit ERK in this system. Anisomycin activated JNK (3.8-fold), whereas olprinone did not. In addition, the dose of SB203580 in this study did not affect JNK by anisomycin.

Discussion

We have shown that the transient pretreatment with PDEIII-Is can limit infarct size independently of systemic hemodynamic changes and collateral blood flow. Furthermore, either the augmentation of PKA activity or subsequent p38 MAPK activation mediates this effect. This observation may explain the preconditioning effect by transient β -adrenergic stimulation.⁸

Specificity of the Individual Kinase Inhibitors in the In Vivo Canine System

First, olprinone activated both PKA and p38 MAPK, but not PKC, ERK, or JNK in this system. Of course, in the present canine model, we confirmed that PMA activated both PKC and ERK and anisomycin activated JNK.

Second, SB203580 in this protocol blocked only p38 MAPK activation, not PKA, PKC, or JNK activation, indicating that the dose of SB203580 in this system is specific to p38 MAPK. Although we did not check the direct effect of SB on ERK, both the inability of olprinone to activate ERK and the observation that PD98059, which blocked ERK activation in this system, failed to blunt the infarct size limitation by olprinone confirmed that the effect of SB203580 to blunt the cardioprotective effect by olprinone is afforded by blocking p38 MAPK.

Furthermore, H89, which blocked cardioprotection by olprinone, did block PKA, whereas it did not block PKC, indicating that (1) PKC is not related to the olprinone-induced cardioprotection and (2) the dose of H89 in this study is selective for PKA and does not affect PKC in this system.

Taken together, we can conclude that the pharmacological interventions in this study work properly to activate or block the individual kinases selectively in this canine model.

Direct Link Between PKA and p38 MAPK

The present results suggest that p38 MAPK activation plays a crucial role in mediating cardioprotection afforded by short-time exposure to PDEIII-Is, after a transient increase in intracellular cAMP level and PKA activity. PDEIII-Is cause a prompt increase in cAMP in cardiomyocytes, which is maximized within 10 minutes,³ followed by a rapid activation of PKA. Furthermore, PKA activation is reported to cause sufficient augmentation of p38 MAPK activity through a protein tyrosine phosphatase, completely independently of PKC activation.^{23,24} Therefore, PDEIII-Is can rapidly activate p38 MAPK. We understand, however, that this interpretation of the results is perfectly confirmed when we observe that H89 has no effect on p38 MAPK activity when added directly to the *in vitro* p38 MAPK activity assay.

Timing for the Activation of p38 MAPK to Provide Cardioprotection

The next issue is the time window in which activation of p38 MAPK can mediate cardioprotection. The transient activation

of p38 MAPK in the preischemic phase may cause cardioprotection, because we showed here that the concomitant administration of either H89 or SB203580 alone before ischemia abolishes either the enhancement of p38 MAPK activity in the preischemic period or the infarct size-limiting effect. This may be further supported by a recent report.²⁵

Conversely, we do not have direct data on whether the transient use of PDEIII-Is during ischemia/reperfusion is cardioprotective. Nakano et al¹⁵ reported that the activation of p38 MAPK is beneficial, because ischemic preconditioning elicits the activation of p38 MAPK during ischemia and protects the myocardium, which does not occur in control ischemia. There are contradictory reports, however, to elucidate the role of p38 MAPK activation during ischemia; the activation of p38 MAPK is unfavorable,^{25–27} because sustained ischemia and reperfusion activate p38 MAPK in either the ischemic or reperfused phase, which enhances myocardial apoptosis or myocardial death, which is prevented by p38 MAPK inhibition during ischemia or in the early phase of reperfusion. Furthermore, persistent p38 MAPK activation has been established by recent reports²⁸ to be deleterious to the heart, because it can induce myocardial hypertrophy, apoptosis, and fibrosis, leading to cardiac maladaptation or myocardial dysfunction. Thus, we should further consider the role of p38 MAPK activation during ischemia or in early reperfusion in the cardioprotection in this study.

Further Downstream Effectors of p38 MAPK

There are some candidates for the downstream effectors of p38 MAPK, which may also be the final effectors of cardioprotection. Because persistent p38 MAPK activation fails to protect myocardium, identification and stimulation of the downstream effectors that are tolerable for persistent stimulation may be essential for beneficial clinical use.

The translocation of HSP27 occurs after ischemic preconditioning through the transient activation of p38 MAPK²⁵ and protects myocardium from ischemic injuries. The previous study reported that the translocated HSP27 accumulates in the Z bands of myofibril²⁹ and prevents conformational changes or fragmentation in myofibril and cytoskeleton.^{30,31} This may also explain the cardioprotection against contraction failure, because a recent report showed that the structural components of myofibrils, especially in Z bands, are disorganized in failing human heart.³²

In addition, it has also been reported that both the opening of ATP-sensitive potassium channels on mitochondria³³ and nitric oxide³⁴ are activated or induced by p38 MAPK and also mediate the cardioprotection. Further investigations concerning this issue may reveal the complete cellular mechanisms of cardioprotection afforded by β -adrenergic preconditioning or the mechanism of cardioprotection against ischemia/reperfusion injury.

Finally, taking these arguments together, there may be at least 2 possible ways to extend our present observations to the clinical situation. The direct short-time activation of p38 MAPK instead of transient administration of PDEIII-Is may be useful, because the transient, but not the persistent, activation of p38 MAPK is cardioprotective. Another way may include gene transfer or direct activation of HSP27 to elicit maximal cardioprotection.

Acknowledgments

This study was supported by Grants-in-Aid for Scientific Research 12470153 and 12877107 from the Ministry of Education, Culture, Sports, Science, and Technology of Japan and by grants from the Smoking Research Foundation of Japan. We thank Jun Fujiki and Hideki Koyama for technical assistance and advice.

References

- Ludmer PL, Wright RF, Arnold JM, et al. Separation of the direct myocardial and vasodilator actions of milrinone administered by an intracoronary infusion technique. *Circulation*. 1986;73:130–137.
- Tajimi M, Ozaki H, Sato K, et al. Effect of a novel inhibitor of cyclic AMP phosphodiesterase, E-1020, on cytosolic Ca⁺⁺ level and contraction in vascular smooth muscle. *Naunyn Schmiedebergs Arch Pharmacol*. 1991;344:602–610.
- Satoh H, Endoh M. Effects of a new cardiotoxic agent 1,2-dihydro-6-methyl-2-oxo-5-[imidazo (1,2-a) pyridin-6-yl]-3-pyridine carbonitrile hydrochloride monohydrate (E-1020) on contractile force and cyclic AMP metabolism in canine ventricular muscle. *Jpn J Pharmacol*. 1990;52:215–224.
- Jaski BE, Fifer MA, Wright RF, et al. Positive inotropic and vasodilator actions of milrinone in patients with severe congestive heart failure: dose-response relationships and comparison to nitroprusside. *J Clin Invest*. 1985;75:643–649.
- Monrad ES, Baim DS, Smith HS, et al. Milrinone, dobutamine, and nitroprusside: comparative effects on hemodynamics and myocardial energetics in patients with severe congestive heart failure. *Circulation*. 1986;73:III-168–III-174.
- Packer M, Carver JR, Rodeheffer RJ, et al. Effect of oral milrinone on mortality in severe chronic heart failure. The PROMISE Study Research Group. *N Engl J Med*. 1991;325:1468–1475.
- Adamopoulos S, Piepoli M, Qiang F, et al. Effects of pulsed beta-stimulant therapy on beta-adrenoceptors and chronotropic responsiveness in chronic heart failure. *Lancet*. 1995;345:344–349.
- Lochner A, Genade S, Tromp E, et al. Ischemic preconditioning and the β -adrenergic signal transduction pathway. *Circulation*. 1999;100:958–966.
- Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation*. 1986;74:1124–1136.
- Tohmeh JF, Cryer PE. Biphasic adrenergic modulation of beta-adrenergic receptors in man: agonist-induced early increment and late decrement in beta-adrenergic receptor number. *J Clin Invest*. 1980;65:836–840.
- Node K, Kitakaze M, Sato H, et al. Role of intracellular Ca²⁺ in activation of protein kinase C during ischemic preconditioning. *Circulation*. 1997;96:1257–1265.
- Ping P, Zhang J, Huang S, et al. PKC-dependent activation of p46/p54 JNKs during ischemic preconditioning in conscious rabbits. *Am J Physiol*. 1999;277:H1771–H1785.
- Kim SO, Baines CP, Critz SD, et al. Ischemia induced activation of heat shock protein 27 kinases and casein kinase 2 in the preconditioned rabbit heart. *Biochem Cell Biol*. 1999;77:559–567.
- Ping P, Zhang J, Cao X, et al. PKC-dependent activation of p44/p42 MAPKs during myocardial ischemia-reperfusion in conscious rabbits. *Am J Physiol*. 1999;276:H1468–H1481.
- Nakano A, Baines CP, Kim SO, et al. Ischemic preconditioning activates MAPK2 in the isolated rabbit heart: evidence for involvement of p38 MAPK. *Circ Res*. 2000;86:144–151.
- Sugioka M, Ito M, Masuoka H, et al. Identification and characterization of isoenzymes of cyclic nucleotide phosphodiesterase in human kidney and heart, and the effects of new cardiotoxic agents on these isoenzymes. *Naunyn Schmiedebergs Arch Pharmacol*. 1994;350:284–293.
- Colucci WS. Positive inotropic/vasodilator agents. *Cardiol Clin*. 1989;7:131–144.
- Kitakaze M, Node K, Minamoto T, et al. Role of activation of protein kinase C in the infarct size-limiting effect of ischemic preconditioning through activation of ecto-5'-nucleotidase. *Circulation*. 1996;93:781–791.
- Kitakaze M, Funaya H, Minamoto T, et al. Role of protein kinase C- α in activation of ecto-5'-nucleotidase in the preconditioned canine myocardium. *Biochem Biophys Res Commun*. 1997;239:171–175.
- Houglum K, Lee KS, Chojkier M. Proliferation of hepatic stellate cells is inhibited by phosphorylation of CREB on serine 133. *J Clin Invest*. 1997;99:1322–1328.
- Sanchez I, Hughes RT, Mayer BJ, et al. Role of SAPK/ERK kinase-1 in the stress-activated pathway regulating transcription factor c-Jun. *Nature*. 1994;372:794–798.
- Ishizuka T, Terada N, Gerwins P, et al. Mast cell tumor necrosis factor α production is regulated by MEK kinases. *Proc Natl Acad Sci U S A*. 1997;94:6358–6363.
- Saxena M, Williams S, Tasken K, et al. Crosstalk between cAMP-dependent kinase and MAP kinase through a protein tyrosine phosphatase. *Nat Cell Biol*. 1999;1:305–311.
- Blanco-Aparicio C, Torres J, Pulido R. A novel regulatory mechanism of MAP kinases activation and nuclear translocation mediated by PKA and the PTP-SL tyrosine phosphatase. *J Cell Biol*. 1999;147:1129–1136.
- Sanada S, Kitakaze M, Papst PJ, et al. Role of phasic dynamism of p38 mitogen-activated protein kinase activation in ischemic preconditioning of the canine heart. *Circ Res*. 2001;88:175–180.
- Ma XL, Kumar S, Gao F, et al. Inhibition of p38 mitogen-activated protein kinase decreases cardiomyocyte apoptosis and improves cardiac function after myocardial ischemia and reperfusion. *Circulation*. 1999;99:1685–1691.
- Mackay K, Mochly-Rosen D. An inhibitor of p38 mitogen-activated protein kinase protects neonatal cardiac myocytes from ischemia. *J Biol Chem*. 1999;274:6272–6279.
- Zhang D, Gaussin V, Taffet GE, et al. TAK1 is activated in the myocardium after pressure overload and is sufficient to provoke heart failure in transgenic mice. *Nat Med*. 2000;6:556–563.
- Yoshida K, Aki T, Harada K, et al. Translocation of HSP27 and MKBP in ischemic heart. *Cell Struct Funct*. 1999;24:181–185.
- Bluhm WF, Martin JL, Mestral R, et al. Specific heat shock proteins protect microtubules during simulated ischemia in cardiac myocytes. *Am J Physiol*. 1998;275:H2243–H2249.
- Huot J, Houle F, Spitz DR, et al. HSP27 phosphorylation-mediated resistance against actin fragmentation and cell death induced by oxidative stress. *Cancer Res*. 1996;56:273–279.
- Heling A, Zimmermann R, Kostin S, et al. Increased expression of cytoskeletal, linkage, and extracellular proteins in failing human myocardium. *Circ Res*. 2000;86:846–853.
- Baines CP, Liu GS, Birincioglu M, et al. Ischemic preconditioning depends on interaction between mitochondrial KATP channels and actin cytoskeleton. *Am J Physiol*. 1999;276:H1361–H1368.
- Bellmann K, Burkart V, Bruckhoff J, et al. p38-dependent enhancement of cytokine-induced nitric-oxide synthase gene expression by heat shock protein 70. *J Biol Chem*. 2000;275:18172–18179.

Cardioprotective Effect Afforded by Transient Exposure to Phosphodiesterase III Inhibitors: The Role of Protein Kinase A and p38 Mitogen-Activated Protein Kinase

Shoji Sanada, Masafumi Kitakaze, Philip J. Papst, Hiroshi Asanuma, Koichi Node, Seiji Takashima, Masanori Asakura, Hisakazu Ogita, Yulin Liao, Yasuhiko Sakata, Akiko Ogai, Tomi Fukushima, Junko Yamada, Yoshiro Shinozaki, Tsunehiko Kuzuya, Hidezo Mori, Naohiro Terada and Masatsugu Hori

Circulation. 2001;104:705-710

doi: 10.1161/hc3201.092216

Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

Copyright © 2001 American Heart Association, Inc. All rights reserved.

Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://circ.ahajournals.org/content/104/6/705>

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

Reprints: Information about reprints can be found online at:
<http://www.lww.com/reprints>

Subscriptions: Information about subscribing to *Circulation* is online at:
<http://circ.ahajournals.org/subscriptions/>