

Characterization of extraintestinal pathogenic *Escherichia coli* isolated from captive wild felids with bacteremia

Vania M. Carvalho,¹ Lika Osugui, Ariela P. Setzer, Rodrigo P. G. Lopez, Antonio F. Pestana de Castro, Kinue Irino, José L. Catão-Dias

Abstract. Diseases caused by extraintestinal pathogenic *Escherichia coli* (ExPEC) in wild felids are rarely reported. Although urinary tract infections are infrequently reported in domestic cats, such infections when present are commonly caused by ExPEC. The present work characterized ExPEC strains isolated from 2 adult felines, a snow leopard (*Panthera uncia*) and a black leopard (*Panthera pardus melas*), that died from secondary bacteremia associated with urinary tract infections. Isolates from both animals were classified into the B2 phylogenetic group and expressed virulence genotypes that allowed them to cause severe disease. In addition, strains from the black leopard showed multidrug resistance.

Key words: Bacteremia; extraintestinal pathogenic *Escherichia coli*; multidrug resistance; urinary tract infections; wild felids.

Extraintestinal pathogenic *Escherichia coli* (ExPEC) strains are characterized by specific virulence factors (VFs) and are related to a heterogeneous group of human disorders. Among extraintestinal diseases caused by *E. coli*, urinary tract infections (UTI) are the most frequently reported, along with neonatal meningitis and septicemia.¹⁵ ExPEC strains have also been associated with animal diseases, especially in poultry and pets.^{12,15} There is also evidence that ExPEC strains isolated from animals present zoonotic potential.^{3,13} There is little published information associating this group of pathogens with diseases in wild animals. The purpose of the current study was to characterize *E. coli* isolates obtained from a snow leopard (*Panthera uncia*) and a black leopard (*Panthera pardus melas*) housed in a Brazilian zoological park that died due to UTIs associated with secondary bacteremia.

The first animal, an adult male snow leopard, was brought to the veterinary hospital and presented with hematuria. Urinalysis and the determination of the urea and creatinine levels in blood indicated a primary renal disease. Despite supportive therapy, death occurred within 24 hr. Necropsy revealed suppurative hemorrhagic cystitis (Fig. 1), nephritis, and suppurative hemorrhagic pyelonephritis (Fig. 2). Histopathology showed multifocal interstitial nephritis, multifocal suppurative pyelonephritis, severe necrotizing cystitis, splenic lymphocytosis, and multisystemic petechiae.

The second case was a 17-year-old female black leopard that had a prior history of recurring UTIs, without etiological diagnosis, that started 9 months before death. On the last UTI manifestation, 7 days of enrofloxacin treatment was adopted with apparent success, according to pre- and post-treatment urinalysis. The animal was brought to the hospital exhibiting lethargy and pale mucous membranes.



Figure 1. Urinary bladder; snow leopard (*Panthera uncia*). The urinary bladder is distended and hemorrhagic and contains a large amount of bloody urine.

Complementary exams revealed azotemia and moderate neutrophilia, compatible with infection. The animal died 48 hr after presentation. Chronic pyelonephritis, multifocal

From Laboratório de Biologia Molecular e Celular, Universidade Paulista, São Paulo, SP, Brazil (Carvalho, Osugui), Autonomus Veterinarian, São Paulo, SP, Brazil (Setzer), Fundação Parque Zoológico de São Paulo, São Paulo, SP, Brazil (Lopez), Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, SP, Brazil (Pestana de Castro), Instituto Adolfo Lutz, Centro de Bacteriologia, São Paulo, SP, Brazil (Irino), Laboratório de Patologia Comparada de Animais Selvagens, FMVZ, Universidade de São Paulo, São Paulo, SP, Brazil (Catão-Dias).

¹Corresponding Author: Vania M. Carvalho, Laboratório de Biologia Molecular e Celular, Universidade Paulista, Av. José Maria Whitaker, 290, CEP: 04057-000 São Paulo, SP, Brazil. vaniamc@uol.com.br



Figure 2. Kidney; snow leopard (*Panthera uncia*). The medulla shows congestion and diffuse hemorrhages associated with cortical-medullary white-tan striations and hemorrhagic pyelonephritis.

cardiac petechiae, and severe pulmonary edema were the pathological lesions.

The animals were necropsied immediately after death. Heart blood samples from both animals and a vesical urine sample from the snow leopard were aseptically collected and cultivated in brain–heart infusion broth,^a blood agar,^a and MacConkey agar.^a Pure *E. coli* strains from the 3 samples were isolated and identified using a commercial system.^b

Serotyping was performed at a reference center for *E. coli* serotyping (Instituto Adolfo Lutz, São Paulo, SP, Brazil), according to standard international procedures of tube agglutination test¹⁰ and using currently available O (O1–O181) and H (H1–H56) antisera. The molecular characterization of the strains was performed by polymerase chain reaction (PCR), following protocols¹² previously reported for putative genes encoding VFs, including adhesins (*papA*, *papEF*, *sfa*, *afaI*, *fimH*, *iha*), siderophores (*iucD*, *fyuA*), toxins (*cnf-1*, *hlyA*), protectins (*traT*, *cvaC*), invasive factor (*ibeA*), and a pathogenicity island (*malX*). The phylogenetic group of isolates, as well as hemolysin expression, were determined according to methods previously described.^{4,6} The susceptibility to antibiotics was determined according to Clinical and Laboratory Standards Institute guidelines^{8,9} using the *E. coli* strain ATCC 25922 as a control and the following antibiotic disks^c: ampicillin (10 µg), amoxicillin (30 µg), cephalixin (30 µg), cefoxitin (30 µg), ceftiofur (30 µg), gentamicin (10 µg), streptomycin (10 µg), enrofloxacin (5 µg), ciprofloxacin (5 µg), chloramphenicol (30 µg), tetracycline (30 µg), nitrofurantoin (300 µg), and trimethoprim–sulfamethoxazole (25 µg).

Diseases caused by ExPEC in wild felids are rarely described. Although UTIs are infrequently reported in domestic cats, such infections when present are commonly caused by ExPEC.² In previous reports,^{5,16} cases of necrotizing pneumonia have been associated with ExPEC in both domestic

Table 1. Strain origin, serotype, phylogenetic group, virulence-associated genes, and antibiotic susceptibility of extraintestinal pathogenic *Escherichia coli* isolated from captive wild felids with bacteremia of urinary tract origin.*

	Snow leopard†	Black leopard
	OR:H-	ONT:H9
Virulence-associated genes‡	B2	B2
Adhesins		
<i>sfa</i>	+	–
<i>fimH</i>	+	+
Toxins		
<i>hlyA</i>	+	–
<i>cnf-1</i>	+	–
Siderophores		
<i>iucD</i>	–	+
<i>fyuA</i>	+	–
Protectin		
<i>traT</i>	–	+
Pathogenicity island		
<i>malX</i>	+	–
Hemolysin expression	+	–
Antibiotic resistance	No	AMP, AMOX, CEF, CTX, CIP, ENO, TET, CL

* R = rough; NT = nontypeable; + = positive result; – = negative result; No = no resistance observed; AMP = ampicillin; AMOX = amoxicillin; CEF = ceftiofur; CTX = cefotaxime; CIP = ciprofloxacin; ENO = enrofloxacin; TET = tetracycline; CL = chloramphenicol.

† Both strains from urine and blood had the same profile.

‡ The genes *papC*, *papEF*, *afaI*, *iha*, *cvaC*, and *ibeA* were not found in the strains.

cats and tiger cubs. Regarding nondomestic cats, a report of pyometra involving *E. coli* was described in a lioness,¹ and colisepticemia was reported in a tiger.¹⁴ However, VFs were not characterized in either of those cases. The present report describes the occurrence of bacteremia in 2 captive wild cats that were associated with ExPEC infections of urinary tract origin. The results obtained regarding the investigated VFs, serotyping, and phylogenetic classification of the isolates are presented in Table 1.

In both cases, although the isolates could not be associated with the classical ExPEC serogroups, the presence of virulence markers linked with ExPEC¹⁵ was demonstrated. There were, however, interesting differences between the isolates. The strains isolated from blood and urine of the snow leopard presented the same phylogenetic group and virulence gene profile and were susceptible to all of the antibiotics tested, showing a great range of VFs, which included those classically related to ExPEC, such as S fimbriae (*sfa*), the yersiniabactin receptor (*fyuA*), and genes for CNF-1 (*cnf-1*) and hemolysin (*hlyA*) toxins; the expression of the latter was demonstrated in a phenotype test. These strains also had the gene *malX*, which was linked to the pathogenicity island

of urosepticemic human *E. coli* (CFT073). Some reviews of the literature indicate a close relationship between this island and high virulence potential.^{11,15}

In contrast, the strain isolated from the black leopard showed multidrug resistance to 8 different active compounds (5 different classes) but a narrower range of virulence markers, including *fimH*, *iucD*, and *traT*. Data from a published report¹¹ has mentioned the negative association involving antimicrobial resistance and VFs and/or phylogenetic background, ascribing this phenomenon to the balance between host compromise and bacterial fitness advantage. However, hybrid plasmids codifying multiple VFs along with multidrug resistance has already been found in avian pathogenic *E. coli* strains with zoonotic potential.¹³ The finding of multidrug resistance and VFs in strains isolated from captive animals is worrisome because the selective pressure exerted by extensive antibiotic use may lead to the selection of more pathogenic strains, which may threaten the animals' survival.

Although in lower number, all 3 virulence markers found in the black leopard isolate indicate potential for the establishment of systemic infections. Type 1 fimbriae (*fimH*), in addition to having an adhesion function, also has a role in invasion. The siderophore encoded by the *iucD* gene, although not very prevalent in isolates from domestic animals, has been detected in high percentages of human isolates from patients with pyelonephritis.¹¹ Finally, the strains in the current study were classified into the phylogenetic group B2, which is known to include the ExPEC strains responsible for the most severe and invasive syndromes in animals and human beings.^{7,11} Considering the zoonotic potential of ExPEC, as well as the severity of the reported cases, it is believed that the characterization of such conditions is important to improve clinical approaches for wild animals kept in captivity.

Acknowledgements

The authors thank LAB&VET Diagnostics and Veterinarian Consultation Ltd. for providing bacterial strains, Dr. James R. Johnson for kindly providing the controls strains, and Glória Jafet for the assistance with the picture.

Sources and manufacturers

- a. Difco Laboratories Inc., Sparks, MD.
- b. API, bioMérieux, Marcy l'Etoile, France.
- c. Cefar, Jurubatuba, SP, Brazil.

Declaration of conflicting interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: Financial support was made by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP; grant #06/54343-2).

References

1. Baker R, Henderson R: 1983, Pyometra in an African lioness. *J Am Vet Med Assoc* 183:1314.
2. Barsanti JA: 2006, Genitourinary infections. *In: Infectious diseases of the dog and cat*, ed. Greene CE, 3rd ed., pp. 935–961. Saunders Elsevier, Philadelphia, PA.
3. Bélanger L, Garenaux A, Harel J, et al.: 2011, *Escherichia coli* from animal reservoirs as a potential source of human extraintestinal pathogenic *E. coli*. *FEMS Immunol Med Microbiol* 62:1–10.
4. Beutin L, Montenegro M, Zimmermann S, Stephan R: 1986, Characterization of hemolytic strains of *Escherichia coli* belonging to classical enteropathogenic O-serogroups. *Zentralbl Bakteriell Mikrobiol Hyg A* 261:266–279.
5. Carvallo FR, Debroy C, Baeza E, et al.: 2010, Necrotizing pneumonia and pleuritis associated with extraintestinal pathogenic *Escherichia coli* in a tiger (*Panthera tigris*) cub. *J Vet Diagn Invest* 22:136–140.
6. Clermont O, Bonacorsi S, Bingen E: 2000, Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Appl Environ Microbiol* 66:4555–4558.
7. Clermont O, Olier M, Hoede C, et al.: 2011, Animal and human pathogenic *Escherichia coli* strains share common genetic backgrounds. *Infect Genet Evol* 11:654–662.
8. Clinical and Laboratory Standards Institute (CLSI): 2008, Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals; approved standard—third edition. CLSI document M31-A3. CLSI, Wayne, PA.
9. Clinical and Laboratory Standards Institute (CLSI): 2008, Performance standards for antimicrobial susceptibility testing; approved standard—eighteenth international supplement. CLSI document M100-S18. CLSI, Wayne, PA.
10. Ewing WH: 1986, The genus *Escherichia*. *In: Edwards and Ewing's identification of Enterobacteriaceae*, 4th ed., pp. 93–134. Elsevier Science, New York, NY.
11. Johnson JR, Russo TA: 2005, Molecular epidemiology of extraintestinal pathogenic (uropathogenic) *Escherichia coli*. *Int J Med Microbiol* 295:383–404.
12. Johnson JR, Stell AL: 2000, Extended virulence genotypes of *Escherichia coli* strains from patients with urosepsis in relation to phylogeny and host compromise. *J Infect Dis* 181:261–272. Erratum in *J Infect Dis* 2000, 181:2122.
13. Johnson TJ, Jordan D, Kariyawasam S, et al.: 2010, Sequence analysis and characterization of a transferable hybrid plasmid encoding multidrug resistance and enabling zoonotic potential for extraintestinal *Escherichia coli*. *Infect Immun* 78:1931–1942.
14. Singh KR, Rajeswari KR, Char L: 1994, Colisepticaemia in a tiger. *Indian Vet J* 71:406.
15. Smith JL, Fratamico PM, Gunther NW: 2007, Extraintestinal pathogenic *Escherichia coli*. *Foodborne Pathog Dis* 4:134–163.
16. Sura R, Van Kruiningen HJ, DeRoy C, et al.: 2007, Extraintestinal pathogenic *Escherichia coli*-induced acute necrotizing pneumonia in cats. *Zoonoses Public Health* 54:307–313.