Cite this: Nat. Prod. Rep., 2011, 28, 290

www.rsc.org/npr



Secondary metabolites of fungi from marine habitats†

Mostafa E. Rateb^{ab} and Rainer Ebel^{*a}

Received 1st November 2010 DOI: 10.1039/c0np00061b

Covering: 2006 to mid-2010

Marine-derived fungi continue to produce structurally unique secondary metabolites, a considerable number of which display promising biological and pharmacological properties. This review gives an overview of new natural products from marine-derived fungi and their biological activities, focusing on the period from 2006 until mid-2010. Overall, 690 structures and 478 references are presented.

- 1 Introduction
- 2 Definition of marine fungi
- 3 Biology of marine fungi
- 3.1 Taxonomy and biological diversity
- 3.2 Ecology of fungal communities
- 4 Chemistry and biological activity
- 4.1 Sources of fungi for chemical studies
- 4.2 General aspects of secondary metabolism
- 4.3 Polyketides
- 4.4 Prenylated polyketides/meroterpenoids
- 4.5 Terpenoids
- 4.6 Peptides including diketopiperazines
- 4.7 Alkaloids and other nitrogen-containing metabolites
- 4.8 Shikimate-derived metabolites
- 4.9 Lipids
- 5 Conclusion
- 6 References

1 Introduction

In recent years, secondary metabolites obtained from marinederived fungi have gained considerable attention, as many of them are structurally unique and possess interesting biological and pharmacological properties. From the time of the more-or-less accidental discovery of cephalosporin C in 1949, which was produced by a culture of a *Cephalosporium* species obtained off the Sardinian coast, it took another thirty years for marine-derived fungi to be more systematically evaluated for their chemical potential. A sharp increase in the number of published structures

from this source occurred at the beginning of the 1990s, and this trend continues today. It is the aim of this review to give an overview on secondary metabolites from marine-derived fungi and their biological activities, focusing on the period from 2006 until mid-2010. For earlier years, review papers with a similar scope have been published in NPR, including those by Bugni & Ireland in 2004,1 and Saleem et al. in 2007.2 We also mention briefly important developments that have occurred in terms of characterisation of microbial (and most importantly fungal) communities by molecular biological methods, first from algae, but recently also from sponges. Sponges and algae are the top-ranking sources of marine fungal diversity, as expressed in the number of new structures reported. On the other hand, the true origin of these fungal strains has been the subject of controversy, with the extreme positions being that they are true symbionts to that they are mere contaminants washed into the sea from terrestrial habitats. Very recent reports, however, provide increasing evidence that, similar to the case of marine bacteria, fungal diversity assessed by molecular methods considerably exceeds culturable diversity, and also that specifically adapted fungal associates might exist.

2 Definition of marine fungi

Marine fungi are an ecologically rather than physiologically or taxonomically defined group of organisms. According to the "classic" definition that appears to be universally accepted in the scientific community, marine fungi are divided into two groups, obligate and facultative marine fungi – *obligate* marine fungi are those that grow and sporulate exclusively in a marine or estuarine habitat, while *facultative* marine fungi are those from freshwater or terrestrial milieus able to grow (and possibly also to sporulate) in the marine environment.³ It has been suggested that indigenous marine species (obligate and facultative) should be separated from non-indigenous species (sometimes referred to as "contaminants" or "transients", *i.e.* terrestrial or freshwater species that are dormant in marine habitats) based on their germination ability,³ but in practical terms this is difficult to achieve. Moreover, there is

^aMarine Biodiscovery Centre, Department of Chemistry, University of Aberdeen, Meston Walk, Aberdeen, AB24 3UE, Scotland, UK

^bPharmacognosy Department, Faculty of Pharmacy, Beni-Suef University, Salah Salem St., 62111 Beni-Suef, Egypt

 $[\]dagger$ This paper is part of an NPR themed issue on Marine Natural Products.

an increasing tendency to identify marine fungi by molecular biological methods that do not require sexually reproducing life stages, for example sequencing of rDNA, instead of traditional approaches based on morphological characteristics.

3 Biology of marine fungi

3.1 Taxonomy and biological diversity

Based on the distinction between obligate and facultative marine fungi, it can clearly be stated that among the former there are taxa which are exclusively found in the marine environment, while by definition, this is not the case for the latter. A recent monographic treatment of filamentous marine fungi listed at total of 530 species in 321 genera, among them 424 species within Ascomycota (in 251 genera), 94 species of anamorphic fungi (in 61 genera) and only 12 species within Basidiomycota (in 9 genera).⁴ These figures apply to those marine taxa that can be isolated or cultivated by classical microbial techniques, and that have been studied in detail and taxonomically described by marine mycologists, while the overall figures are expected to be much higher. For example, analysis of the mangrove plant Kandelia candel, sampled at a single nature reserve in Hong Kong, resulted in more than 50 taxonomically distinct isolates for this one host species.⁵ It has also been suggested that the fungal diversity in individual habitats so far has been underestimated considerably, exemplified by the situation of sedimentborne marine fungi which avoid microscopic detection due to their tendency to form aggregates.⁶

On the other hand, there is a pronounced difference regarding the biological diversity accessible by cultivation-based methods, and the diversity evident from molecular biological studies which rely on gene sequences present in environmental samples. This phenomenon has been demonstrated when analysing methane hydrate bearing deep-sea marine sediments for fungal gene sequences,⁷ but is by no means restricted to marine fungi. On the contrary, similar observations have been made in virtually all microbiological disciplines, with molecular studies giving evidence for the presence of taxonomic groups which have no known closely related cultivated representatives.

In terms of global fungal diversity, a commonly used estimate is that while 74,000 fungal species have been described, the overall expected diversity amounts to 1.5 million species.⁸ While critics have questioned the validity of this figure, a more recent "rigorous, minimum estimate of global fungal diversity" came to the conclusion that there are at least 712,000 extant fungal species worldwide.⁹

3.2 Ecology of fungal communities

Marine fungal strains have been obtained from virtually every possible marine habitat, including inorganic matter (soil, sediments, sandy habitats, artificial substrates, and the water column), marine microbial communities, marine plants (algae, sea grasses, driftwood and other higher plants, especially mangrove plants), marine invertebrates (most notably sponges, but also corals, ascidians, holothurians, bivalves, crustaceans) and vertebrates (mainly fishes). However, it is worth mentioning that the fraction of culturable isolates is very low, *i.e.* in the range of 1% or less, with regard to the overall estimated biodiversity, similar to the situation with bacteria. As described above, marine plants, especially from mangrove habitats, harbor an enormous diversity of fungal associates. Since algae are the most prevalent source of marine fungi for chemical studies (see below), it is not surprising that they have been the subject of meticulous studies regarding their fungal communities. Using molecular techniques such as denaturing gradient gel-electrophoresis (DGGE), it has been demonstrated that the brown alga Fucus serratus harbours a distinct pattern of fungi which based on their rDNA sequences belonged to the Lindra, Lulworthia, Engyodontium, Sigmoideal Corollospora complex, and Emericellopsis/Acremonium-like ribotypes. In a parallel culture-based approach, 336 isolates representing 35 genera of the Ascomycota and Zygomycota were obtained, including Sigmoidea marina and Dendryphiella salina, together with members of the genera Acremonium (most of them



Mostafa Rateb

Mostafa Rateb received his BPharm from the Faculty of Pharmacy, Cairo University, in 2000. He obtained his MPharm in plant natural products isolation and plant tissue culture in 2005 from the Department of Pharmacognosy, Faculty of Pharmacy, Cairo University under the supervision of Prof. Ali El-Shamy and Prof. Seham El-Hawary. He is now in his final year of his PhD, studying the isolation and structure elucidation of natural products from marine and under-explored

habitats at the Marine Biodiscovery Centre, University of Aberdeen under the supervision of Prof. Marcel Jaspars and Dr Rainer Ebel.



Rainer Ebel

Rainer Ebel received his PhD at University of Würzburg, Germany, where he studied the chemical ecology of Mediterranean sponges under the guidance of Professor Peter Proksch. He undertook postdoctoral research with Professor Phil Crews at University of California at Santa Cruz, working on the natural products chemistry of sponge-derived fungi. In 2000, he joined the Institute for Pharmaceutical Biology and Biotechnology at University of Düsseldorf, Germany, and in

2007, he transferred to the University of Aberdeen, where he currently holds the position of lecturer in Organic Chemistry at the recently established Marine Biodiscovery Centre.

representing the new endophytic species Acremonium fuci), Cladosporium and Fusarium.¹⁰⁻¹²

On the other hand, marine animals, especially sponges, have also been demonstrated to represent a rich source of fungal diversity. The ecological role of fungal associates within sponges has been controversial from the very beginning, since due to their enormous filtering capacities, sponges are to be expected to effectively concentrate microorganisms from the surrounding water column. However, for sponge-associated bacteria, it has been shown that most of the bacterial strains present within sponges (where they can contribute to up to 40% of the volume) are highly adapted and specialized symbionts, with spongederived taxa collected from various geographic locations and different sponge hosts being more closely related to each other than to any other free-living taxa.¹³⁻¹⁵ Numerous cultivationbased studies with marine sponges have shown enormous biological diversity for the isolates obtained, with at least some of them displaying close affiliation with fungi from marine habitats, although in most cases genera already described from terrestrial habitats were dominating.¹⁶⁻²¹ However, a recent DGGE-based study provided evidence for sponge-specific patterns of distribution, and possibly even for the existence of sponge-specific fungal clades.22

4 Chemistry and biological activity

4.1 Sources of fungi for chemical studies

Fig. 1 gives an overview of new chemical structures from marinederived fungi reported in the literature until mid-2010. It should be noted that it is not always possible to assign a given report to a given source, since there is a deplorable general trend to disclose little to virtually no information at all, especially for fungi from mangrove habitats.²³ Thus, while it was possible to assign publications until 2002 to categories such as plants or sediments,¹ this is no longer possible in recent years, as reflected by the introduction of an additional category "mangrove habitats" in the key report on marine natural products published annually in this journal.²³ Based on these considerations, it can be estimated that roughly two-thirds of all new compounds reported from marine fungi are derived from isolates from living matter, with an approximately even split between plant and



Fig. 1 New compounds from marine-derived fungi until mid-2010, divided by sources of the fungal strains.

animal sources in the broadest term, while the remaining compounds are due to fungi from non-living sources, most notably sediments. Within the individual groups, algae are the predominant sources for fungal diversity, closely followed by sponges and mangrove habitats. An interesting newly emerging source is the deep sea, and only a handful of reports exist describing new secondary metabolites from fungi derived from this habitat, all of them actually obtained from deep sea sediments, which consequently have been assigned to the sediment category. It is interesting to compare the present chart to the one provided 8 years ago by Bugni and Ireland.¹ The most dramatic change that has occurred in the meantime is certainly the dramatic increase in reports from Chinese scientists, with most of them studying fungal isolates from mangrove areas on the subtropical island of Hainan.

4.2 General aspects of secondary metabolism

From the number of new compounds published each year (Fig. 2), it is evident that natural products chemistry of marine-derived fungi is rapidly developing, and still has not reached its climax. While in the period covered by the first review of this series, *i.e.* from the beginning until 2002, 272 new structures had been reported, in 2009 alone more than two-thirds of this figure was reached, and it can be projected that in 2010, approximately 200 new structures will be reported. The total number of new natural products from marine-derived fungi currently exceeds 1000.

Fig. 3 gives an overview of new chemical structures from marine-derived fungi reported in the literature until mid-2010, based on their putative biogenetic origin. In most cases, assignments of a given metabolite to a certain category is only based on structural considerations, and only represents the authors' personal judgement. It is also clear that any approach to classify the enormous structural diversity of marine fungal-derived metabolites according to biogenetic categories is somewhat arbitrary, since there are numerous examples of mixed biogenesis, such as PKS–NRPS (polyketide synthase–non-ribosomal peptide synthase) hybrids, *e.g.* responsible for the generation of pseurotin congeners,²⁴ or the numerous cases of prenylated alkaloids and prenylated polyketides. Obviously, for some groups of compounds it was necessary to compromise in order to avoid too many different categories, as exemplified by the very



Fig. 2 New compounds from marine-derived fungi, according to year of publication.



Fig. 3 New compounds from marine-derived fungi until mid-2010, divided by structural/biogenetic types.

common diketopiperazines, which have been listed as (di-)peptides, rather than alkaloids.

As evident from Fig. 3, polyketides play a dominant role, and if prenylated polyketides and nitrogen-containing polyketides (grouped as alkaloids in this overview) are taken into account, their total share will exceed 50% of all new natural products from marine-derived fungi, which is similar to the situation of terrestrial fungi. A clearly emerging picture based on molecular sequence data is that fungal polyketide synthases (PKS) can be grouped into three subtypes, *i.e.* non-reducing, partially-reducing and highly-reducing, and for the limited number of fungal secondary metabolites for which biogenetic gene clusters have been identified and characterised, this division is also reflected in the relevant structures of the natural products.²⁵ However, no attempt was undertaken in this review to further divide the large group of polyketides along these lines, since the vast number of reported structures would have required extensive speculation.

4.3 Polyketides

Shimalactones A (1) and B (2) are unusual polyketides with a novel carbon framework containing a bicyclo[4.2.0]octadiene and an oxabicyclo[2.2.1]heptane unit.^{26,27} They are produced by an isolate of *Emericella variecolor* which was obtained from a sediment sample collected in Japan. 1 and 2 induced neuritogenesis in neuroblastoma Neuro2A cells at a concentration of 10 μ g mL⁻¹, but were cytotoxic at higher concentrations.



Chemical analysis of the fungus *Penicillium rugulosum*, isolated from the Mediterranean sponge *Chondrosia reniformis*, yielded a series of structurally unusual tricyclic and bicyclic pentaenes, prugosenes A1 (3), A2 (4), A3 (5), B1 (6), B2 (7), C1 (8), and C2 (9).²⁸ Through feeding studies labelled precursors, **3** was established as an undecaketide, with all branching methyl groups being derived from *S*-adenosyl-methionine. The oxabicyclo[2.2.1]heptane core present in **3–5** had previously only been reported for

shimalactones A (1) and B (2) from an marine-derived *Emericella* variecolor,^{26,27} and more recently for coccidiostatin A, a metabolite from a terrestrial isolate of *Penicillium rugulosum*.²⁹ Upon treatment of **3** with diluted alkali, **6** was obtained as a major and **8** as a minor product, suggesting that **6** is formed by hydrolytic cleavage of the C-2–C-3 bond, while the formation of the cyclopentenone system of **8** and **9** should include decarboxylation of the bicyclic system of the A-type prugosenes (**3–5**). None of the compounds were active when tested for antimicrobial activity against several bacterial and one fungal strain.



The fungus *Rhizopus* sp. was isolated from the Chinese bryozoan *Bugula* sp., and was found to produce aspericins A–C

(10–12), along with the related asperic acid.³⁰ Asperic acid, so far the only compound reported from nature with the same carbon skeleton as present in 10–12, had previously been obtained from a saltwater culture of sponge-derived *Aspergillus niger*.³¹ When tested for cytotoxic properties against four different cell lines, only 12 displayed moderate activity.



Bisorbicillinoids have frequently been reported from marinederived fungi, and are thought to arise biosynthetically via a Diels-Alder cycloaddition of two sorbicillinol or oxosorbicillinol moieties. Oxosorbiquinol (13) and its 10,11-dihydro derivative (14) were discovered when investigating a deep-sea isolate of *Phialocephala* sp., collected from sediments at a depth of 5059 m.³² Compounds 13 and 14 displayed weak cytotoxic activity against five different cancer cell lines. Apart from sorbiquinol,³³ previously reported form a terrestrial Trichoderma longibrachiatum, 13 and 14 are the only bisorbicillinoids connected through the six-membered ring of one monomer and the unsaturated side chain of the other one. The same strain also yielded the first sorbicillin trimer, trisorbicillinone A (15), which displayed moderate cytotoxicity against P388 and HL60 cells.34 A recent re-examination of the fermentation broth yielded three further congeners, trisorbicillinones B-D (16-18), all of which exhibited only very weak cytotoxicity.35 The sorbicillinoid 6demethylsorbicillin (19) and the bisorbicillinoid 10,11-dihydrobisvertinolone (20) were isolated together with nine known sorbicillinoids and bisorbicillinoids from the fungus Trichoderma sp. which was obtained from a Chinese sediment sample.³⁶ All compounds exhibited mild cytotoxicity towards the HL-60 cell line, and also led to an increase in the percentage of cells in the sub- G_1 fraction, compared to the control.





Trichodermanones A–D (21–24) are new sorbicillinoid polyketide derivatives with an unprecedented tricyclic ring system produced by the fungus *Trichoderma* sp. isolated from the Caribbean sponge *Agelas dispar*.³⁷ Compounds 21–23 exhibited moderate radical scavenging activity in the DPPH assay, but were devoid of antimicrobial, antiparasitic and cytotoxic properties, or inhibitory properties toward phosphatase, acetylcholine esterase and trypsin. Based on structural considerations, 21–24 are thought to be biosynthesised by Diels–Alder cycloaddition of the hexaketide sorbicillinol with a triketide unit. An isolate of *Penicillium citrinum*, obtained from an unidentified

Japanese demosponge, yielded JBIR-59 (**25**), along with a series of known sorbicillinoids.³⁸ Based on structural considerations, **25** is assumed to be formed by reaction of sorbicillin and the known trihydroxyquinol USF 406A,³⁹ a metabolite of the soilborne fungus *Mortierella* sp.



The fungus *Penicillium* sp. which was isolated from the surface of the drifting cotton clothing collected off an island in Korea, was found to produce two new resorcylic acid-containing 14membered lactones, 8'-hydroxyzearalanone (**26**) and 2'-hydroxyzearalanol (**27**).⁴⁰ Zearalanone, originally described from a terrestrial *Gibberella zeae*,⁴¹ and three further known congeners were also detected in the culture broth. Compounds **26** and **27** did not possess radical scavenging activity, antibacterial activity, or inhibitory properties toward tyrosinase, even though similar biological activities had previously been reported for zearalanones. Compound **26**, zearalenone and zearalenol were likewise identified as metabolites of a *Fusarium* sp. derived from an undisclosed Japanese sponge, together with the new derivative 5'hydroxyzearalenol (**28**),⁴² which was devoid of antifungal or antimitotic activity.



A strain of the lignicolous mangrove ascomycete *Aigialus parvus* was reinvestigated and yielded six new nonaketide metabolites, aigialomycins F (29) and G (30a/30b), 7',8'-dihydroaigialospirol (31), 4'-deoxy-7',8'-dihydroaigialospirol (32), and rearranged macrolides 33 and 34,⁴³ along with the known compounds hypothemycin, aigialomycins A and B, aigialospirol, 4-*O*-demethylhypothemycin, and aigialone, previously reported from the same fungus.^{44,45} None of the new metabolites displayed antimalarial or cytotoxic properties. Hypothemycin is a resorcylic macrolide that was first reported from a terrestrial specimen

of *Hypomyces trichothecoides*,⁴⁶ and has attracted considerable attention due to its protein kinase inhibitory profile.⁴⁷



Two new 12-membered lactones of the lasiodiplodin-type, **35** and **36**, in addition to known lasiodiplodins, were identified as metabolites of an unidentified endophytic fungus, which was isolated from the Chinese brown alga *Sargassum* sp.⁴⁸ Only the known congeners displayed various degrees of antimicrobial activity.



Chemical analysis of the marine fungus *Curvularia* sp., isolated from the red alga *Acanthophora spicifera* collected in Guam, resulted in the discovery of the novel macrolide apralactone A (37), together with the antipodes of known curvularin macrolides (38–43).⁴⁹ The dimeric curvularin (44) was recognised as an artefact. Compound 37 is a 14-membered phenyl acetic acid macrolactone, and the first such compound with a 4-chromanone substructure. Compounds 37, 38, and 40–42 displayed varying Published on 12 January 2011. Downloaded by Pennsylvania State University on 13/05/2016 04:15:05.

degrees of cytotoxicity towards a panel of 36 human tumour cell lines, with mean IC_{50} values in the range 1.25 to 30 $\mu M.$



An unidentified endophytic fungus, isolated from the Chinese mangrove plant *Sonneratia apetala*, produced two new 10-membered lactones, (3R,5R)-sonnerlactone (**45**) and its (3R,5S) diastereoisomer (**46**).⁵⁰ Both **45** and **46** displayed weak inhibitory effects towards the multi-drug resistant cancer cell line KV/MDR.



Three pentaketides, aspinotriols A (47) and B (48) and aspinonediol (49), together with the known aspinonene⁵¹ and dihydroaspyrone,52 metabolites from a terrestrial isolate of Aspergillus ochraceus, were obtained from the culture of an Aspergillus ostianus strain isolated from an unidentified marine sponge at Pohnpei, Micronesia.53 Since the same strain previously had produced new chlorinated pentaketides when fermented in natural seawater,⁵⁴ a culture medium containing large amounts of bromine was used. Even though no brominated compounds were detected, the metabolite profile of the fungus changed considerably compared to the previous study. When screened for activity towards MRSA and cytotoxic properties, only aspinonene displayed mild inhibitory properties towards mouse lymphocytic leukemia cells.53 A further chemical investigation of the same fungus under identical culture conditions led to the discovery of three new 14-membered macrolides, aspergillides A-C (50-52).55 Even though 14-membered macrolides in

general are frequently reported from fungi, **50–52** are unusual since they represent the first congeners with an additional tetrahydropyran or dihydropyran ring, respectively. All three proved inactive when tested for antimicrobial activity toward MRSA, however, the compounds exhibited moderate cytotoxicity toward mouse lymphocytic leukemia cells (L1210).



The fungus *Curvularia* sp., obtained from the red alga *Acanthophora spicifera* collected in Guam, produced four tenmembered lactones, curvulide A (53), a mixture of curvulides B₁ (54) and B₂ (55) and another congener 56.⁵⁶ Compounds 53–56 are related to other lactones including modiolides⁵⁷ or decarestrictines,⁵⁸ previously reported from a marine-derived *Paraphaeosphaeria* sp. and from terrestrial *Penicillium* spp., respectively, but differ with regard to their oxygenation pattern. None of the new congeners was active when tested for antibacterial, antifungal, or antialgal properties.



The strain *Phomopsis* sp. of undisclosed marine origin produced two very closely related new 10-membered macrolides, phomolides A (**57**) and B (**58**), and the new 3-(hydroxymethyl)-6-methyl-6,7-dihydrobenzofuran-4(5*H*)-one (**59**).⁵⁹ All compounds were devoid of cytotoxic activity, but displayed antimicrobial activity against *Escherichia coli*, *Candida albicans*, and *Saccharomyces cerevisiae*, while **59** also showed strong activity against *Bacillus subtilis*.



The fungus *Chaetomium globosum*, originally isolated from the Japanese fish *Mugil cephalus*, is the source of a complex series of

azaphilones, chaetomugilins. Initially, chaetomugilins A-C (60-62) were reported, of which 60 and 62 exhibited significant cvtotoxic activity against P388 and HL-60 cell lines.⁶⁰ A subsequent report disclosed the structures of chaetomugilins D-F (63-65), and all compounds were tested for cytotoxic activity profiles against a panel of 39 human cancer cell lines.⁶¹ Compound 65 exhibited remarkable growth inhibition in combination with selective cytotoxic activity, and evaluation of the activity profiles with the COMPARE program suggested that the mode of action for 60, 62 and 65 might be different from any other anticancer drug developed to date. Further chemical analysis revealed chaetomugilins G (66) and H (67), with 66 exhibiting significant cytotoxicity against the P388 and HL-60 cell lines, while 67 was moderately active against KB cell line.⁶² Furthermore, secochaetomugilins A (68) and D (69) were discovered, of which 69 exhibited significant cytotoxic activity against the cell lines P388, HL-60, L1210 and KB.63 Investigations into the stability and the chemical reactivity indicated that neither of compounds 66-69 was an artefact resulting from degradation of 60 or 61. The next additions to this series of azaphilones were chaetomugilins I-O (70-76), and all compounds apart from 74 exhibited significant cytotoxic activity against various cancer cell lines.⁶⁴ Compound 70 was subjected to detailed activity profiling in a panel of 39 human cancer cell lines, and the COMPARE program suggested that it possessed a novel mechanism of action. Recently, the two stereoisomers of 60, 11-epi-chaetomugilin A (77) and 4'-epichaetomugilin A (78), were discovered upon reinvestigation of the same fungus.⁶⁵ Compound 77 exhibited moderate activity against P388 and HL-60 cell lines, but marginal activity against L1210 and KB cell lines, whereas 78 was only marginally active against all cell lines.





Trichodermatides A–D (79–82) have been characterised from the fungus *Trichoderma reesei*, obtained from a Chinese sediment sample.⁶⁶ Compound 79 is the first example of a pentacyclic polyketide with a ketal moiety, and similar to the remaining compounds, displayed mild cytotoxic activity against the A375-S2 (human melanoma) cell line. Octaketide derivatives with an α , β -unsaturated cyclohexenone fused to a pyran ring such as 79– 82 have so far been reported from the terrestrial fungus *Trichoderma harzianum*,^{67,68} but do not appear to occur elsewhere in Nature.



The fungus *Trichoderma koningii*, isolated from marine mud of the South China Sea, produced 7-*O*-methylkoninginin D (83) and trichodermaketones A–D (84–87), together with four koninginin derivatives.⁶⁹ Koninginin D had previously been reported form a terrestrial strain of *Trichoderma koningii*.⁷⁰

Compounds **83–87** did not display antimicrobial activity against methicillin-resistant *Staphylococcus aureus* or *Candida albicans*, but **84** showed synergistic antifungal activity against *C. albicans* with ketoconazole, albeit at rather high concentrations.

An isolate of the fungus *Fusarium* sp. (section *Liseola*), which was obtained from driftwood collected in Japan, yielded neofusapyrone (**88**), fusapyrone (**89**), and deoxyfusapyrone (**90**).⁷¹ Compounds **89** and **90** had initially been described from *Fusarium semitectum* and were thought to be α -pyrones,⁷² while their structures were revised to γ -pyrones in the present investigation. All three displayed weak antifungal activity against *Aspergillus clavatus*.



The endophytic fungus *Ascochyta salicorniae* was obtained from the green alga *Ulva* sp., collected from the German North Sea. A reinvestigation of this strain, which previously had the tetramic acids ascosalipyrrolidinone A and B as well as the polyketide ascosalipyrone,⁷³ resulted in the discovery of biogenetically unique cycloethers, ascospiroketals A (91) and B (92),⁷⁴ composed of a methylated diketide attached to a highly modified octaketide *via* an ester link. From a biosynthetic point of view, the presence of two additional carbon atoms attached to C-2 of the octaketide part is striking, since extender units other than malonyl-CoA are not known from fungal polyketides, and these two carbons thus most probably arise from geminal biomethylation *via S*-adenosyl-methionine, a very rare process that seems to be unprecedented in the context of fungal polyketides. Two closely related metabolites, cephalosporolides H (93) and I (94), were obtained from a *Penicillium* sp. isolated from the Chinese mangrove plant *Kandelia candel*,⁷⁵ and shortly after, the same group of authors also reported penisporolides A (95) and B (96) from a *Penicillium* sp. isolated from the same source,⁷⁶ although it is not clear whether or not both strains were identical. Compounds 95 and 96 showed mild inhibitory activity toward xanthine oxidase and 3α -hydroxysteroid dehydrogenase, while 93 and 94 were inactive.



The fungus *Nodulisporium* sp., obtained from a soft coral from Thailand, produced a tetronic acid, nodulisporacid A (97).⁷⁷ The same communication also reported the presence of vermelhotin,⁷⁸ previously described from an unidentified terrestrial fungal strain, in the culture broth of an unidentified sponge-derived fungal strain. Both compounds occurred as equilibrium E/Zmixtures. Compound 97 is structurally related to lowdenic acid, a metabolite from a terrestrial *Verticillium* sp.⁷⁹ When tested for cytotoxic properties, 97 was inactive, but its methyl and benzyl ester displayed a pronounced increase in activity, similar in potency to vermelhotin. Both 97 and vermelhotin also exhibited moderate antiplasmodial activity.



Dehydroxychlorofusarielin B (98) is a decalin derivative of Aspergillus sp., obtained from the surface of the Korean marine brown alga Sargassum horneri.⁸⁰ The structure of 98 was established by X-ray crystallography, and it is structurally closely related to fusarielins A and B, previously described from a soilborne Fusarium sp.81 that were also detected in the culture broth. All three compounds displayed moderate antibacterial activity against Staphylococcus aureus, methicillin-resistant S. aureus, and multidrug-resistant S. aureus. Shortly after this report, fusarielin E (99) was described from the culture broth of a marine-derived Fusarium sp. of undisclosed origin by a Chinese research group.⁸² Even though 99 is claimed to differ from 98 with regard to the configuration at C-11, C-15 and C-16, their NMR data are virtually identical, and it is thus very likely that both compounds are actually identical. At high concentrations, 99 inhibited the conidia growth of Pyricularia oryzae by a swelling effect and induced curling deformation of the mycelia.



Nigrosporapyrones A–D (100–103) were reported, together with a series of known metabolites, from the fungus *Nigrospora* sp. isolated from the sea fan *Annella* sp. in Thailand.⁸³ Compound 100 exhibited mild antibacterial activity against *Staphylococcus aureus* and methicillin-resistant *S. aureus*.



The fungus Aspergillus glaucus, which was cultured from sediment around mangrove roots in China, yielded aspergiolide A (104), a novel anthraguinone derivative with an unprecedented naphtho[1,2,3-de]chromene-2,7-dione skeleton, the structure of which was secured by X-ray diffraction analysis.⁸⁴ Compound 104 displayed pronounced cytotoxicity at submicromolar concentrations towards A-549 and HL-60 cells, while it was less active against BEL-7402 and P388 cell lines by approximately two orders of magnitude. More recently, through feeding experiments with labelled acetate units, the biosynthesis of 104 was demonstrated to follow the classical acetate-malonate pathway of aromatic polyketides, and to proceed via coupling of an octaketide and a pentaketide precursor.85 A subsequent study yielded a further congener, aspergiolide B (105), together with seven new polyketides.⁸⁶ Compounds 106–108 are unusual naphthyl furanosides, isoasperflavin (109) is a constitutional isomer of the known asperflavin from a terrestrial-derived Aspergillus flavus,⁸⁷ 110 was identified as the (+)-enantiomer of the recently described variecolorquinone A from the halotolerant Chinese fungus Aspergillus variecolor,88 while 111 and 112 are new physicon-emodin bisanthrones. When tested against the most susceptible cell lines A-549 and HL-60 of the previous study, the activity of 105 was comparable to 104, whereas 111 and 112 were only moderately active.



Secondary metabolites produced by the endophytic fungus *Monodictys putredinis*, obtained from an undisclosed green alga,

collected in Tenerife, Spain, were subjected to an in-depth evaluation for potential cancer chemopreventive effects.89,90 Initially, four new monomeric xanthones, monodictysins A-C (113-115) and monodictyxanthone (116), and the benzophenone monodictyphenone (117) were obtained, all of them thought to be biogenetically derived by oxidative cleavage of a common anthraquinone precursor either between C-4a or C-10a and carbonyl C-10.89 A more recent reinvestigation resulted in the two dimeric chromanones monodictyochromes A (118) and B (119), consisting of two unusual naphthoquinone units, which are probably also derived from an anthraquinone precursor via a xanthone intermediate, and which are connected by phenol oxidative coupling in a regioselective manner. This coupling should also occur under strict stereoselectivity, since 118 was identified as the (P)-atropisomer, while 119 represents the (M)stereoisomer as was deduced by comparison of the pertaining CD spectra with the model compounds, (P)- and (M)-orsellinic acid camphanate.⁹⁰ The cancer chemopreventive potential of **113–119**, expressed as their ability to either inhibit or induce certain enzymes involved in biotransformation of potential carcinogenic agents, was studied using a series of assay systems. At concentrations in the lower micromolar range 114, 118 and 119 were found to inhibit cytochrome P450 1A, an enzyme that is involved in the metabolic conversion of procarcinogens into carcinogens. Similarly, 114, 115, 118 and 119 also moderately induced NAD(P)H:quinone reductase, a carcinogen-detoxifying enzyme. Furthermore, 115, 118 and 119 were weak inhibitors of aromatase activity, which is essential for the biosynthesis of estrogens.





The endophytic fungus *Phomopsis* sp., isolated from the Chinese mangrove tree *Excoecaria agallocha*, produced

phomopsis-H76 A (120), a dimeric chromanone closely related to monodictyochrome B (119), as well as phomopsis-H76 B (121), and C (122), structurally unique dimeric pyrano[4,3-b]pyran-5(2H)-ones.⁹¹ In zebra fish embryos, 113 led to a significant acceleration of sub-intestinal vessel plexus (SIV) branching, and all three compounds lacked cytotoxic or antibacterial properties.



Chaetocyclinones A-C (123-125) were obtained from the culture broth of Chaetomium sp., isolated from an undisclosed marine alga.⁹² Compound 125 was active towards selected phytopathogenic fungi, but was not found to be cytotoxic. Compound 123 is closely related to anhydrofulvic acid, a metabolite of Carpenteles brefeldianum (now named Eupenicillium brefeldianum),93 while 125 differs from vinaxanthone, previously described from Penicillium vinaceum94 only with regard to the oxygenation pattern. Using feeding experiments with ¹³C-labelled acetate, the biosynthesis of 123 was shown to proceed via a linear heptaketide intermediate, which would then undergo oxidative cleavage and recyclisation. Accordingly, the biosynthesis of 125 should involve an unusual two-fold aldol condensation of two highly reactive heptaketide intermediates, but it could not be ruled out that this reaction could occur spontaneously, and that 125 thus represents an artefact.92



An endophytic *Aspergillus niger*, isolated from the Chinese brown alga *Colpomenia sinuosa*, yielded three new naphtho- γ -pyrones,

nigerasperone A–C (**126–128**).⁹⁵ Compound **126** is a linear naphtho- γ -pyrone, while **127** and **128** are dimers of two angular and two linear naphtho- γ -pyrones, respectively. Compound **128** showed weak antifungal activity against *Candida albicans* and moderate antioxidant activity in the DPPH assay, but none of the compounds exhibited cytotoxic activity.



8'-O-Demethylnigerone (129) and 8'-O-demethylisonigerone (130) are two new structurally related dimeric naphtho- γ pyrones which were detected in the culture broth of the fungus Aspergillus carbonarius, isolated from a Chinese sediment sample.96 Both compounds exhibited mild inhibitory activity against Mycobacterium tuberculosis. In a screening effort directed at discovering tyrosine kinase inhibitors using human umbilical vein endothelial cell lysate, out of a total of 200 fungal strains tested, only the fungus Hypocrea vinosa isolated from beach sand collected in Japan displayed activity. Chemical analysis revealed the presence of two new bisnaphtho- γ -pyrones, hypochromins A (131) and B (132), besides the closely related known SC2051, the diketocongener of 131 which so far has only been reported in the patent literature.97 All three compounds inhibited tyrosine kinase activity in HUVEC lysate, and in addition also inhibited HUVEC proliferation, migration, and tubule formation, thus displaying antiangiogenic potential.

A new naphtho- γ -pyrone, 2-benzyl-5-hydroxy-6,8-dimethoxy-4*H*-benzo[*g*]chromen-4-one (133), was reported from the endophytic fungus *Phomopsis* sp., isolated from the Chinese mangrove tree *Excoecaria agallocha*.⁹⁸ Compound 133 exhibited moderate cytotoxicity against HEp-2 and HepG2 cell lines. In a screening program aimed at discovering inhibitors of the bacterial two-component regulatory system (TCS), an isolate of







Ascochyta sp. from a floating scrap of festering rope in a Japanese fishing port yielded the spirodioxynaphthalene ascochytatin (134).⁹⁹ 134 which is structurally related to palmarumycins,^{100,101} exhibited stronger activity against the temperature-sensitive mutant *Bacillus subtilis* CNM2000 than against wild-type strain 168, suggesting that the compounds inhibited the function of the YycG/YycF-TCS in the bacterium. Furthermore, 134 displayed strong antimicrobial activity against Gram-positive bacteria and *Candida albicans*, and displayed cytotoxicity toward two mammalian cancer cell lines with IC₅₀ values in the lower micromolar range.



7-*epi*-8-Hydroxyaltertoxin I (135) and 6-*epi*-stemphytriol (136), together with the known perylenequinones altertoxin I and stemphyperylenol, were discovered during chemical analysis of the fungus *Alternaria alternata*, which was cultivated from the inner tissue of the Chinese red alga *Laurencia* sp.¹⁰² Altertoxin I is a known mycotoxin from the plant pathogenic *Alternaria alternata*,¹⁰³ while stemphyperylenol was initially reported from *Stemphylium botryosum* var. *lactucum*.¹⁰⁴ None of the compounds displayed any significant antibacterial or antifungal activity.



The fungus *Alternaria* sp. was isolated from the Chinese mangrove plant *Sonneratia alba*, and identified as a source xanalteric acids I (137) and II (138), besides a series of known compounds including perylenequinones such as altertoxin I.¹⁰⁵

Compounds **137** and **138** appear to represent the first natural products with a 10*H*-phenaleno[1,2,3-*de*]chromene skeleton, the only other report of which being that of xanalteric acid, a degradation product of the fungal toxin cercosporin which was obtained following addition of the latter to cultures of the bacterium *Xanthomonas campestris* pv. *zinnia*.^{106,107} Compounds **137** and **138** exhibited weak antibiotic activity against multidrug-resistant *Staphylococcus aureus* and only marginal cytotoxicity against L5178Y cells.



The endophytic fungus *Sporothrix* sp., isolated from the Chinese inshore mangrove tree *Kandelia candel*, produced three metabolites with unique ring systems, sporothrins A–C (**139–141**), besides the known 1-hydroxy-8-methoxynaphthalene and 1,8-dimethoxynaphthalene.¹⁰⁸ Biogenetically, **139** and **140** are assumed to arise *via* multiple oxidative coupling reactions of three 1,8-dihydroxynaphthalene units. This assumption was supported by molecular biological investigations revealing the presence of a non-reducing polyketide synthase gene in the genomic DNA of *Sporothrix* sp., which based on BLAST homology search is very likely to be involved in 1,3,6,8-tetrahydroxynaphthalene (T4HN) biosynthesis. Compounds **139** and **140** displayed modest cytotoxic activity against the hepG2 cell line, whereas **139** additionally strongly inhibited acetylcholine esterase (AChE) activity.



The fungal strain *Chaetomium* sp., isolated from an undisclosed Greek alga, was found to produce three structurally unusual xanthones, chaetoxanthones A–C (142–144).¹⁰⁹ The dioxane/tetrahydropyran moieties present in 142 and 143 are reminiscent of intermediates of aflatoxin biosynthesis, while 144 is a chlorinated xanthone substituted with a tetrahydropyran ring. Compound 143 displayed selective activity against *Plasmodium falciparum*, with an IC₅₀ value of 0.5 µg mL⁻¹ without being cytotoxic, whereas 144 was moderately active against *Trypanosoma cruzi*, with an IC₅₀ value of 1.5 µg mL⁻¹.



An Aspergillus sp. strain that was isolated from sea water in China, yielded a bifuran-substituted xanthone, asperxanthone (145) and a biphenyl, asperbiphenyl (146), together with a series of known fatty acid monoglycerides.¹¹⁰ Compounds 145 and 146 moderately inhibited the multiplication of tobacco mosaic virus (TMV). Two new xanthones, 8-(methoxycarbonyl)-1-hydroxy-9oxo-9H-xanthene-3-carboxylic acid (147) and dimethyl 8methoxy-9-oxo-9H-xanthene-1,6-dicarboxylate (148)were obtained, together with one known congener, methyl 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylate (149), from the culture broth of the fungus *Penicillium* sp. that was isolated from the bark of the Chinese mangrove plant, Acanthus ilicifolius.111 The spectral data of 149 indicated that it is in fact identical to janthinone, reported initially as a lactone from a terrestrial endophytic Penicillium janthinellum,112 but in the current study, its structure was revised to 149, which was secured by X-ray crystallographic analysis. Compound 148 exhibited modest antifungal activity against Fusarium oxysporum f. sp. cubense. More recently, 7-hydroxyjanthinone (150) was isolated from Penicillium sp., an endophyte from Melia azedarach growing in a Chinese mangrove habitat, and proved to be devoid of cytotoxic activity.113



Griseusin C (151) is a new, structurally unusual naphthoquinone derivative which was produced by a strain of *Penicillium* sp., an endophyte from the Chinese mangrove plant *Kandelia candel*. Compound 151 displayed very mild inhibitory activity towards 3α -hydroxysteroid dehydrogenase, but was inactive against xanthin oxidase and horseradish peroxidase.¹¹⁴

An endophytic fungal strain *Eurotium rubrum*, isolated from the Chinese mangrove plant *Hibiscus tiliaceus*, yielded four new anthraquinone derivatives including the bisdihydroanthracenone eurorubrin (**152**), two seco-anthraquinones 2-*O*-methyl-9-dehydroxyeurotinone (**153**) and 2-*O*-methyl-4-O-(α -D-ribofuranosyl)-9-dehydroxyeurotinone (**154**) and the anthraquinone glycoside 3-O-(α -D-ribofuranosyl)questin (**155**), together with three known compounds.¹¹⁵ Compound **152** displayed strong radical scavenging activity in the DPPH assay, exceeding that of the positive control, butylated hydroxytoluene (BHT), whereas the remaining compounds were moderately active.



The fungus *Microsporum* sp. was isolated from the surface of the Korean red alga *Lomentaria catenata*, and was shown to produce the anthracene glycoside, asperflavin ribofuranoside (**156**).¹¹⁶ This compound exhibited considerable radical scavenging activity in the DPPH assay, and also showed moderate antibacterial activity against methicillin-resistant and multidrugresistant *Staphylococcus aureus*.



156 (R = α-D-ribofuranosyl)

Monodictyquinone A (157) is an anthraquinone that, together with three known congeners was isolated from the marine-derived *Monodictys* sp., obtained from the Japanese sea urchin, *Anthocidaris crassispina*.¹¹⁷ Compound 157 showed antimicrobial activity against *Bacillus subtilis, Escherichia coli* and *Candida albicans*, but was not cytotoxic toward HeLa cells even at high concentrations. The endophytic fungus *Fusarium* sp. from an undisclosed Chinese mangrove plant was reported as a source of the new anthraquinone, 5-acetyl-2-methoxy-1,4,6-trihydroxy-anthraquinone (158), besides a series of known naphthoquinones.¹¹⁸



6,8,1'-Tri-*O*-methylaverantin (159) was isolated, besides a series of known anthraquinones, from an unidentified fungus

This journal is $\ensuremath{\textcircled{}}$ The Royal Society of Chemistry 2011

obtained from the Chinese mangrove plant *Acanthus ilicifolius*.¹¹⁹ Two new hexahydroanthrones, tetrahydrobostrycin (160) and its 1-deoxy congener (161), were isolated from the fungus *Aspergillus* sp. obtained from an unidentified Indonesian alga, together with the known pigment bostrycin and the plant hormone abscisic acid.¹²⁰ Compound 160 displayed weak antibacterial activity against *Staphylococcus aureus* and *Escherichia coli*, while 161 was active against *S. aureus*. Both compounds displayed no antifungal activity against yeasts.



The fungus *Halorosellinia* sp., isolated from decayed wood in Hong Kong and a salt lake in the Bahamas, yielded two new anthraquinones **162** and **163**, besides three known derivatives.¹²¹ Three new hehahydroanthraquinones, fusaquinones A–C (**164– 166**) were isolated from the fermentation medium of the fungus *Fusarium* sp., obtained from mangrove sediment in China.¹²² Compound **164** is characterized by an unusual 7,9-epoxide bridge. Compounds **164–166** were inactive when tested for cytotoxic properties.



An unidentified endophytic fungal strain isolated from the Chinese mangrove *Bruguiera gymnorrhiza* produced two new aromatic lactones, 1,10-dihydroxy-8-methyldibenzo[*b,e*]oxepine-6,11-dione (**167**) and 3-(hydroxymethyl)-6,8-dimethoxy-2*H*-chromen-2-one (**168**), along with a known coumarin derivative and the 8-demethyl congener of **168**, which so far had only been obtained synthetically, but not as a natural product.¹²³

A new lactone, 1,8-dihydroxy-10-methoxy-3-methyldibenzo[*b,e*]oxepine-6,11-dione (**169**), and two new xanthones, 1-hydroxy-8-(hydroxymethyl)-6-methoxy-3-methyl-9*H*-xanthen-9-one (**170**) and 1-hydroxy-8-(hydroxymethyl)-3-methoxy-6methyl-9*H*-xanthen-9-one (**171**) were reported from the endophytic fungus *Phoma* sp., isolated from the roots of the Chinese mangrove plant *Avicennia marina*.¹²⁴ Compounds **169–171** were inactive when tested for cytotoxic properties.



The known depsidone unguinol (**172**), a metabolite of *Aspergillus unguis*¹²⁵ and *A. nidulans*,¹²⁶ was identified as an inhibitor of the C₄ plant enzyme pyruvate phosphate dikinase (PPDK), a potential herbicide target, in the course of a systematic screening of more than 2000 extracts generated from approximately 450 marine-derived fungal strains from the collection of the Australian Institute of Marine Sciences (AIMS).¹²⁷ The producing strain was originally obtained from the Australian sponge *Ianthella reticulata*, and by molecular methods was identified as being related to both *A. unguis* and *A. nidulans*, with the analysis of 18S rDNA sequences giving a slightly different result from that of ITS sequences.

(-)-Byssochlamic acid (173), previously only known as a synthetic product, was described as a metabolite of an unidentified mangrove fungus obtained from an undisclosed Chinese location.¹²⁸ The same strain also yielded the hydroxylated congener, 1-hydroxybyssochlamic acid (174).¹²⁹ Compound 173 displayed mild cytotoxic effects towards HEp-2 and HepG2 cells, while 174 was inactive.



2240B (175) is a new alternariol derivative that was isolated, together with alternariol and two further known congeners, from the culture of an unidentified fungus obtained from an undisclosed Chinese mangrove habitat.¹³⁰ An endophytic *Guignardia* sp. was isolated from the bark of the Chinese mangrove plant *Kandelia candel.*¹³¹ Besides the known vermistatin (176), a metabolite of *Penicillium vermiculatum*,¹³² the two new oxygenated congeners 177 and 178 were detected. Compound

vestigation additionally yielded 6-demethylvermistatin (179).¹³³

176 exhibited weak and 177 moderate cytotoxic activity, but both

compounds were devoid of antimicrobial activity. A later rein-



(3S,3'R)-3-(3'-Hydroxybutyl)-7-methoxyphthalide (180) and (*S*)-3-butyl-7-methoxyphthalide (181) were isolated from the culture broth of the fungal strain CRIF2 belonging to the order Pleosporales, obtained from an unidentified sponge in Thailand.¹³⁴ 181 which was obtained synthetically before, but was isolated for the first time as a natural product, exhibited mild cytotoxicity towards several cell lines. 180 and 181 are closely related to known phthalides such as (–)-3-butyl-7-hydroxyphthalide, a cytotoxic metabolite of *Penicillium vulpinum*.^{135,136}



Xyloketal H (182), previously only known as a synthetic intermediate, is a metabolite of the fungus *Xylaria* sp., obtained from the seeds of an undisclosed angiosperm mangrove tree from Hong Kong.¹³⁷ Reinvestigation of the same fungus yielded the new congener 183, to which unfortunately the same name was assigned.¹³⁸ Subsequently, the new analogues xyloketal J (184), xyloester A (185) and xyloallenolide B (186) were discovered, together with (2-(prop-1-en-2-yl)-2,3-dihydrobenzofuran-5-yl)methanol, the benzofuran moiety present in 185 and 186,





which had previously only been obtained synthetically,¹³⁹ but apparently was not known as a natural product.¹⁴⁰ Compounds **184–186** were inactive when tested for antibacterial activity.

The fungus *Penicillium* sp. was isolated from the sea fan *Annella* sp., collected in Thailand, and found to produce the ketal penicipyrone (**187**) and the γ -lactone, penicilactone (**188**), together with three known brefeldin-type macrolides.¹⁴¹ Only one of the known compounds displayed mild antifungal activity.



An endophytic strain of Ascochyta salicornia was cultivated from the inner tissue of the green alga Ulva sp., collected from the German North Sea coast. Upon fermentation on a solid seawater-based medium, two new lactones ascolactones A (189) and B (190),¹⁴² were detected besides two previously described polyketides - ascochital, a metabolite previously reported from the marine fungus Kirschsteiniothelia maritima,143 and the wellknown phytotoxin ascochitine, produced by terrestrial Ascochyta spp.¹⁴⁴ The absolute configurations of **189** and **190** were assigned based on a combination of quantum-chemical CD calculations and chemical degradation.^{142,145} When tested for inhibitory properties against a series of protein phosphates, which were predicted by in silico screening, the known ascochitine was moderately active against MPtpB (mycobacterial protein tyrosine phosphatase B), while 189 and 190 were inactive. The fungus Sporothrix sp., isolated from an undisclosed mangrove at the



South China Sea coast, yielded the new griseofulvin derivative **191**.¹⁴⁶

1-Deoxyrubralactone (**192**) was obtained, besides the known talaroflavone, from an unidentified fungal strain isolated from undisclosed Japanese algae.¹⁴⁷ Compound **192** is a deoxy derivative of the known rubralactone, a metabolite of the terrestrial fungus *Penicillium rubrum*,¹⁴⁸ while talaroflavone had previously been reported from the terrestrial *Talaromyces flavus*.¹⁴⁹ Compound **192** and talaroflavone selectively inhibited eukaryotic DNA polymerases of the X and Y families, but were inactive against other eukaryotic or prokaryotic polymerases, or other DNA metabolic enzymes.

Stoloniferols A (193) and B (194) are two new isocoumarin derivatives, produced by the fungus *Penicillium stoloniferum* which was isolated from an unidentified Chinese sea squirt.¹⁵⁰ In the same study, the halophilic fungus *Penicillium notatum*, obtained from sediments of Qinghai Lake, a saline lake in China, yielded the known dihydrocitrinone, previously described from *Aspergillus carneus*.¹⁵¹ None of the compounds exhibited cytotoxicity against various cell lines. The closely related 6,8-dihydroxy-3,4,7-trimethylisochroman-1-one (195) was reported from *Penicillium* sp., an endophyte of the Chinese mangrove *Bruguiera sexangula*, and demonstrated to possess moderate cytotoxic activity towards the K562 cell line.¹⁵²



Methyl 8-hydroxy-6-methoxy-3,4,5-trimethyl-1-oxoisochroman-7-carboxylate (**196**) was obtained from an unidentified endophytic fungus obtained from an undisclosed Chinese mangrove.¹⁵³ From a culture of a sterile mycelium, isolated from the Japanese green alga *Codium fragile*, (3R,4aR,5S,6R)-6-hydroxy-5-methylramulosin (**197**) and three known mellein congeners were obtained.¹⁵⁴ **197** displayed weak cytotoxic activity toward HeLa cells, whereas the remaining compounds were inactive.



Two endophytic fungi of algae, *Acremonium* sp. from the German red alga *Plocamium* sp. and *Nodulisporium* sp. from a Greek algal species of unknown taxonomy, yielded

This journal is © The Royal Society of Chemistry 2011

acremonisol A (198) and (3*R*)-7-hydroxy-5-methylmellein (199), respectively. Compounds 198 and 199 were inactive in several bioassays, including tests for antibacterial, antifungal, antialgal, or cytotoxic properties.¹⁵⁵ An unidentified endophytic strain, obtained from the dropper of *Kandelia candel* growing in an estuarine mangrove in China, was found to produce the new isochroman 6-hydroxy-3-methylisochroman-5-carboxylic acid (200), besides a series of known natural products.¹⁵⁶



The endophytic fungus *Talaromyces* sp., isolated from the Chinese mangrove plant *Kandelia candel*, yielded two new isochromenones 7-epiaustdiol (**201**) and 8-*O*-methylepiaustdiol (**202**), besides a diverse series of known polyketides.¹⁵⁷ The fungus *Penicillium* sp., obtained from the surface of the Korean green alga *Ulva pertusa*, yielded the new redoxcitrinin (**203**)¹⁵⁸ together with the structurally related known phenol A¹⁵⁹ and citrinin H2,¹⁶⁰ all of them being derivatives of the well-known citrinin.^{161,162} All compounds were found to possess moderate radical scavenging activity in the DPPH assay.



In the course of a screening for antimicrobial substances targeting FtsZ, which is a structural homolog of eukaryotic tubulin, the new sulfoalkylresorcinol (**204**), was identified in the culture extract of the fungus *Zygosporium* sp., isolated from an undisclosed hard coral in Palau.¹⁶³ At very high concentrations, **204** inhibited the GTPase activity of FtsZ and also inhibited FtsZ polymerization, in addition to exhibiting antimicrobial activity against various bacterial strains.



Pichiafurans A–C (205–207) and pichiacins A (208) and B (209) have been characterised from the yeast *Pichia membranifaciens* which was obtained from the Korean marine sponge *Petrosia* sp.¹⁶⁴ Compounds 205–207 are furyl ethers with 2-phenylethanol (likely to be derived from the shikimate pathway), while 208 and 209 are esters consisting of 2-phenylethanol and short-chain ω -hydroxy acids. The fungus *Massarina* sp., isolated from a marine mud sample collected at low depth in the Palau Islands, yielded spiromassaritone (210) and massariphenone (211), which did not exhibit significant antimicrobial or cytotoxic

activity.¹⁶⁵ Compound **210** is a stereoisomer of V214w, previously reported from an unidentified fungus.¹⁶⁶ For the related metabolite arthropsolide A, produced by *Arthropsis truncata*, it has been proposed that its biogenesis includes the condensation of a pentaketide precursor and malic acid.¹⁶⁷



The unusual tricyclic compound 2106 A (**212**) was obtained from the culture broth of an unidentified fungus from the mangrove *Avicennia marina* in Hong Kong, and its structure was established by X-ray diffraction analysis.¹⁶⁸ Protulactones A (**213**) and B (**214**) were reported from the fungus *Aspergillus* sp., isolated from an inter-tidal sediment sample collected in Korea.¹⁶⁹ Both the unusual furo[3,2-*b*] furan ring system present in **213** and the dioxabicyclo[3.3.1]nonane ring system as found in **214** appear to be unprecedented in fungi, at least in the genus *Aspergillus*, but are present in plant-derived metabolites such as goniofufurone from *Goniothalamus giganteus*,¹⁷⁰ or a tetraketide from *Euscaphis japonica*.¹⁷¹



A sterile mycelium obtained from an undisclosed sponge of unknown geographic origin produced three new metabolites, 1-((1R,2R,6S,8S,8aS)-2,8-dihydroxy-1,2,6-trimethyl-1,2,6,7,8,8a-hexahydronaphthalen-1-yl)-3-methoxypropan-1-one (**215**), 4,8dihydroxy-7-(2-hydroxyethyl)-6-methoxy-3,4-dihydronaphthalen-1(2*H*)-one (**216**) and 1-methylnaphthalene-2,6-dicarboxylic acid (**217**).¹⁷² A fungal strain, identified as belonging to the Pleosporales (Ascomycota), was isolated from driftwood from the Baltic coast in Germany, and subsequent chemical analysis led to the discovery of six naphthalenone derivatives, balticols A–F (**218–223**),¹⁷³ besides the known altechromone A, previously described from *Alternaria* sp.¹⁷⁴ At non-cytotoxic concentrations, all compounds displayed antiviral activity against influenza A virus or Herpes simplex virus (HSV) type I, most notably **221** and **222**, with IC₅₀ values ranging between 0.01 and 0.1 µg ml⁻¹.



An isolate of Periconia byssoides, obtained from the sea hare Aplysia kurodai, and which is the well-established source of peribvsins with cell-adhesion inhibitory properties.^{175,176} was subjected to further chemical study, resulting in the discovery of pericosines A-E (224-228).¹⁷⁷ A preliminary report¹⁷⁸ of the structures for 224 and 225 had been revised with regard to the relative stereochemistry based on the synthesis of the enantiomer of 224.179 The absolute configuration of pericosine D (227) was later established by total synthesis.¹⁸⁰ Compounds 224, 225 and 227 exhibited significant growth inhibition against the murine P388 cell line, whereas 226 and 228 were inactive. In a diseaseoriented panel of human cancer cell lines (HCC panel) in the Japanese Foundation for Cancer Research, 224 and 228 displayed moderate activity in general, but 224 was selective for HBC-5 and SNB-75 cell lines, and also showed significant in vivo tumour inhibitory activity. Likewise, 224 was found to inhibit EGFR protein kinase and topoisomerase II.177





The fungus *Phaeosphaeria spartinae*, an endophyte of the German alga *Ceramium* sp., produced spartinols A–D (229–232).¹⁸¹ Compound 231 exhibited weak inhibitory activity towards human leukocyte elastase (HLE). Nigrosporanees A (233) and B (234) were isolated from fungus *Nigrospora* sp, which was obtained from the sea fan *Annella* sp. collected Thailand.¹⁸² Compound 233 displayed moderate cytotoxicity towards MCF-7 and Vero cells, while 233 and 234 also showed weak radical scavenging activity.



The fungal strain *Exophiala* sp., isolated from the surface of the Korean sponge *Halichondria panicea*, was the source of chlorohydroaspyrones A (235) and B (236),¹⁸³ besides three known polyketides including aspyrone, an epoxide which yielded 235 and 236 upon hydrolysis with HCl. Compounds 235 and 236 exhibited mild antibacterial activity against *Staphylococcus aureus*, methicillin-resistant *S. aureus*, and multidrug-resistant *S. aureus*.



Three new epoxydon esters, nigrospoxydons A–C (237–239), and the new pyrone, nigrosporapyrone (240), were reported from the fungus *Nigrospora* sp. which was isolated from the sea fan *Annella* sp. in Thailand.¹⁸⁴ Compound 237 is an ester of abscisic

acid and epoxydon, a well-known metabolite from originally described from a terrestrial *Phoma* sp.¹⁸⁵ Both abscisic acid and epoxydon were also detected in the culture medium, and the latter as well as **237** displayed mild antibiotic activity against *Staphylococcus aureus* and methicillin-resistant *S. aureus*.



The endophytic fungus *Penicillium* sp., isolated from the Chinese mangrove plant *Aegiceras corniculatum*, was found to produce a diverse suite of metabolites including the new leptosphaerone C (**241**), penicillenone (**242**) and 9-demethyl FR-901235 (**243**), besides the new prenylated polyketide arugosin I (**311**). Compounds **241** and **242** displayed marked cytotoxicity towards P388 cells.¹⁸⁶ A strain of *Aspergillus flavus*, obtained from the Chinese marine alga *Enteromorpha tubulosa*, yielded two new 5-hydroxy-2-pyrones, **244** and **245**.¹⁸⁷ Compound **244** induced the production of cAMP on GPR12 transfected CHO and HEK293 cells in a dose-dependent manner, indicating that the compound might be a possible ligand for GPR12.



A strain of *Penicillium terrestre*, obtained from a sediment sample in China, yielded a series of new gentisyl alcohol polymers, including the trimeric terrestriol A (**246**), the dimeric terrestrols B– H (**247–253**) and the monomer **254**.¹⁸⁸ All compounds displayed moderate levels of cytotoxicity against four different cancer cell lines, and were also moderately active as radical scavengers. In addition, **252** displayed moderate inhibitory activity against protein tyrosine kinases Src and KDR. Bromochlorogentisylquinones A (**255**) and B (**256**) are metabolites of the



The fungus *Aspergillus* sp., isolated from a Chinese sediment sample, was found to produce the diphenyl ether dimethyl 2,3'-dimethylosoate (**257**), together with three known compounds.¹⁹⁰ Compound **257** displayed weak cytotoxic properties towards K562 cells, which might be due to arresting the cell cycle in the S phase, as demonstrated by flow cytometry, and also a marked increase in apoptotic cells. Two new diaryl ethers, phomopsides A (**258**) and B (**259**), were reported from the endophytic fungus *Phomopsis* sp., isolated from an undisclosed Chinese mangrove plant.¹⁹¹ In addition, the known excelsione, a depsidone previously described from an endophytic fungus of the New Zealand endemic tree *Knightia excelsa*, was also detected.¹⁹²



An unidentified fungus collected from the rhizosphere in a Chinese mangrove habitat yielded the new biphenyl derivative methyl 3,5'-dihydroxy-4',5-dimethoxy-2'-methylbiphenyl-2-carboxylate (260).¹⁹³ The endophytic fungus *Penicillium commune*, which was isolated from the Chinese semi-mangrove plant *Hibiscus tiliaceus*, afforded a new ester of orsellinic acid with glycerol (261), besides a suite of mostly structurally simple known metabolites.¹⁹⁴



Ethyl 2-(3-hydroxy-2-(7-hydroxyoctanoyl)-5-methoxyphenyl)acetate (**262**) was produced by the endophytic fungus *Phomopsis* sp., isolated from the Chinese mangrove tree *Excoecaria agallocha*.¹⁹⁵ In addition, the closely related dothiorelones A–C, previously described from the mangrove endophytic fungus *Dothiorella* sp.,¹⁹⁶ were also detected. Compound **262** displayed weak cytotoxicity against HEp-2 and HepG2 cells.



The fungus *Pestalotiopsis* sp., isolated from the Chinese mangrove plant *Rhizophora mucronata*, proved to be a prolific source of new secondary metabolites. At first, six new chromones, pestalotiopsones A–F (**263–268**) were characterized, together with a closely related known congener.¹⁹⁷ Compound **268** displayed moderate cytotoxicity towards the murine cancer



This journal is © The Royal Society of Chemistry 2011



cell line L5178Y, while the remaining compounds were inactive. A later reinvestigation yielded new cytosporone derivatives, cytosporones J–N (**269–273**) and five new coumarins, pestalasins A–E (**274–278**), together with the known cytosporone C, dothiorelone B and the new alkaloid pestalotiopsoid A (**647**).¹⁹⁸ Cytosporone C was initially described from an endophytic *Cytospora* sp.,¹⁹⁹ while dothiorelone B was reported from the mangrove endophytic fungus *Dothiorella* sp.¹⁹⁶ None of the new compounds exhibited significant cytotoxic effects.

Chromanone A (279) was produced by an endophytic *Penicillium* sp, isolated from the Egyptian green alga *Ulva* sp.²⁰⁰ Besides a potent scavenging activity against hydroxyl radicals, 279 was found to inhibit cytochrome P450 1A, and to induce mEH (epoxide hydrolase) as well as glutathione *S*-transferase, enzymes involved in carcinogen metabolism. The fungal strain *Botrytis* sp., isolated from the surface of the Korean red alga *Hyalosiphonia caespitosa*, gave a new α -pyrone, (*E*)-6-(hept-1-enyl)-2*H*-pyran-2-one (280),²⁰¹ together with (*E*)-6-(pent-1-enyl)-2*H*-pyran-2-one, which had so far only been obtained synthetically.²⁰² Compound 280 exhibited tyrosinase inhibitory activity with an IC₅₀ value of 4.5 μ M, and was thus more active than the positive control, kojic acid.



Three new derivatives (**281–283**) of the known α -pyrone infectopyrone were isolated from the culture broth of the fungus *Petriella* sp., obtained from the Mediterranean sponge *Suberites domuncula*.²⁰³ Their parent compound infectopyrone had previously been described from the plant pathogenic fungus



Alternaria infectoria.²⁰⁴ Its 2,3-dihydro congener **281** exhibited pronounced cytotoxicity towards the L5178Y cell line.²⁰³

The fungus *Trichoderma viride*, isolated from the Caribbean sponge *Agelas dispar*, was identified as the source of a new α -pyrone, trichopyrone (**284**), together with ten known compounds, most of them sorbicillinoids and bisorbicillinoids.²⁰⁵ Compound **284**, which is closely related to the previously described xylarone from *Xylaria hypoxylon*,²⁰⁶ did not display significant biological activity when tested for antiviral, antimicrobial, estrogenic, antioxidative or cytotoxic properties. The fungus *Paecilomyces lilacinus*, derived from the Korean sponge *Petrosia* sp., produced two new α -pyrones, paecilopyrones A (**285**) and B (**286**), and two cyclohexenone hydroperoxides (**287** and **288**).²⁰⁷



5-(Ethynyloxy)-3-hydroxy-3,6-dihydro-2*H*-pyran-2-one (**289**) and 4-(butoxymethyl)benzene-1,2-diol (**290**), the latter due to its C_6-C_1 motif and its hydroxylation patterns probably rather representing a phenylpropanoid instead of a polyketide, were reported together with a suite of structurally simple known aromatic compounds from an unidentified fungal strain obtained from the tree *Clerodendrum inerme* growing in the intertidal zone of a Chinese coastal region.²⁰⁸ The methoxylated (instead of butoxylated) analogue of **290** was also among the isolated compounds, and is a compound which up to that point was only known synthetically.



The fungus *Penicillium* sp., isolated from the tree *Clerodendrum inerme* growing in the inter-tidal zone of the South China Sea, yielded three new esters of desoxypatulinic acid **291–293**.²⁰⁹ Desoxypatulinic acid, which was also detected in the culture broth, is a common mycotoxin initially reported from *Penicillium patulum*.²¹⁰ The fungal strain *Trichoderma atroviride*, isolated from sediment near the Chinese mangrove *Ceriops tagal*, produced the new compound 3-hydroxy-5-(4-hydroxybenzyl)dihydrofuran-2(3*H*)-one (**294**).²¹¹

Two new benzaldehyde derivatives, 6-ethyl-2-hydroxy-4methoxy-3-methylbenzaldehyde (295) and 6-ethyl-2,4-dihydroxy-3-methylbenzaldehyde (296), were obtained from an



undisclosed fungus, supposedly endophytic to an undisclosed Chinese mangrove plant.²¹² Based on the strain designation, it could actually be a previously studied *Penicillium* sp. isolated from *Acanthus ilicifolius*, although this was not specifically stated in this communication.



The known aquastatin A (297), produced by the Korean sediment-borne fungus *Cosmospora* sp., was isolated as a potent protein tyrosine phosphatase 1B (PTP1B) inhibitory component.²¹³ 297 is an orcinol depside that had previously been reported from the fungus *Fusarium aquaeductuum*, but the absolute configuration of its galactose moiety had not been determined.²¹⁴ The fungus *Trichoderma* sp. was isolated from marine sediment in the South China Sea and found to produce a new cyclopentenone, trichoderone (298), besides a known steroid.²¹⁵ Compound 298 displayed selective cytotoxicity towards six cancer cell lines and was moderately active, probably by inducing apoptosis.



In the culture extract of the fungus *Botrytis* sp., obtained from the surface of the Korean green alga *Enteromorpha compressa*, the new cyclopentenone botrytinone (**299**) was detected, besides the new nitrogen-containing cyclopentenone bromomyrothenone B (**661**).²¹⁶ Compound **299** did not display significant activity during a screening for radical scavenging, tyrosinase inhibitory and antimicrobial properties. Subsequently, **299** and **661** were also detected in the culture medium of *Rhizopus stolonifer*, isolated from the surface of the Korean brown alga, *Sargassum horneri*.²¹⁷



4.4 Prenylated polyketides/meroterpenoids

An isolate of *Myrothecium roridum*, obtained from a wood sample collected in Palau, was reported to produce three new macrocyclic trichothecenes, 12'-hydroxyroridin E (**300**), roridin Q (**301**), and 2',3'-deoxyroritoxin D (**302**), while one new compound, roridin R (2',3'-dihydro-2'-hydroxyroridin H (**303**), was isolated from *Myrothecium* sp. which was obtained from an unidentified Indonesian sponge.²¹⁸ Compound **301** is characterized by a unique ether moiety at position C-13' and thus contains a third hydroxyacid moiety not present in other trichothecenes. Compounds **300**, **301** and **303** displayed cytotoxic activity towards the murine leukemia cell line L1210 with IC₅₀ values of 0.19, 31.2, and 0.45 μ M, respectively, while **303** showed antifungal activity against *S. cerevisiae* at 1 μ g per disc.



Chemical analysis of the culture broth of a fungus of the genus *Aspergillus*, isolated from an unidentified Hawaiian sponge, resulted in the characterisation of four new meroterpenoids, tropolactones A–D (**304–307**).²¹⁹ **304–306** contain a substituted 2,4,6-cycloheptatriene-2-one (tropone) ring, which is presumed to arise from an oxidative ring expansion of the 2,5-cyclohexadiene-2-one ring system of **307**. Tropolactones A–C (**304–306**) showed weak cytotoxicity against HCT-116 human colon adenocarcinoma cells.



The fungus *Aspergillus insuetus*, isolated from the Mediterranean sponge *Petrosia ficiformis*, yielded two new meroterpenoids terretonins E (**308**) and F (**309**), together with the known diketopiperazine aurantiamine.²²⁰ Compounds **308** and **309** are derivatives of the previously reported terretonin²²¹ and terretonins A–D²²² isolated from *A. terreus*. Compounds **308** and **309** showed midrange potency as mammalian mitochondrial respiratory chain inhibitors, interacting with NADH oxidase activity.



Breviones F–H (**310–312**) are new spiroditerpenoid-type meroterpenoids which were produced by the deep-sea fungus *Penicillium* sp., obtained from a sediment sample which was collected at a depth of 5115 m.²²³ They are structurally related to breviones A–E, allelopathic agents from a terrestrial isolate of *Penicillium brevicompactum*.^{224,225} Compounds **310–312** displayed moderate cytotoxicity towards HeLa cells, while **310** also inhibited replication of HIV-1 in C8166 cells.



The fungus *Phoma* sp. was obtained from the Caribbean sponge *Ectyplasia perox*, and identified as the producer of two unusual cyclohexenone sesquiterpenoids, epoxyphomalins A (**313**) and B (**314**).²²⁶ Based on structural considerations, their biosynthesis is assumed to proceed *via* a farnesylated epoxydon

derivative, yielding an analogue of the known 22-deactylyanuthone A, reported from a marine-derived *Aspergillus niger*,²²⁷ which upon cyclisation and hydroxylation would yield **313**. Cytotoxic properties were investigated using a panel of 36 human tumour cell lines, and mean IC₅₀ values of **313** and **314** were 0.11 and 1.25 μ g mL⁻¹, respectively. The observed cytotoxic selectivity pattern of **313** did not correlate with those of reference anticancer agents with known mechanisms of action in the COMPARE analysis.



Arugosins G (**315**) and H (**316**) are prenylated benzophenones produced by the fungus *Emericella nidulans* var. *acristata*, which was isolated from an undisclosed Mediterranean green alga.²²⁸ Compounds **315** and **316** were tested in a variety of assays for antibacterial, antifungal and antialgal activities, but only **316** displayed mild activity towards *Mycotypha microspora* and *Chlorella fusca*. In addition, **315** and **316** were devoid of cytotoxic or immunostimulating activity. The endophytic fungus *Penicillium* sp., isolated from the Chinese mangrove plant *Aegiceras corniculatum*, was found to produce arugosin I (**317**), besides further new polyketides leptosphaerone C (**241**), penicillenone (**242**) and 9-demethyl FR-901235 (**243**).¹⁸⁶

The fungus *Paecilomyces* sp. was isolated from the bark of an undisclosed Taiwanese mangrove tree, and was found to produce the prenylated xanthone, paeciloxanthone (**318**).²²⁹ **318** displayed pronounced cytotoxicity against hepG2 (human liver cancer) cells, was active against the fungi *Curvularia lunata* and *Candida albicans*, and also inhibited acetyl choline esterase (AChE) activity. 1,7-Dihydroxy-2-methoxy-3-(3-methylbut-2-enyl)-9*H*-xanthen-9-one (**319**) and 1-hydroxy-4,7-dimethoxy-6-(3-oxobutyl)-9*H*-xanthen-9-one (**320**) were reported from an unidentified endophytic fungus, isolated from the Chinese mangrove tree *Avicennia marina*.²³⁰ Compounds **319** and **320** displayed mild cytotoxicity towards the KB and KB_v200 cell lines.





3,5,8-Trihydroxy-2,2-dimethyl-3,4-dihydropyrano[3,2b]xanthen-6(2H)-one (**321**) was reported from an unidentified endophytic fungus, obtained from a Chinese mangrove tree of the genus *Avicennia*.²³¹ Cultivation of the endophytic fungus *Chaetomium globosum*, isolated from the Chinese red alga *Polysiphonia urceolata*, resulted in the isolation of the new benzaldehyde derivative chaetopyranin (**322**), together with ten known compounds.²³² Compound **322** displayed moderate radical-scavenging properties, and also exhibited moderate to weak cytotoxic activity toward several tumour cell lines.



Cytosporins D (**323**) and cytosporin E (**324**) are metabolites of the fungus *Eutypella scoparia*, isolated from the marine pulmonate mollusc *Onchidium* sp.²³³ Compounds **323** and **324** are derivatives of cytosporins A–C, angiotensin II binding inhibitors previously described from an endophytic *Cytospora* sp.²³⁴ When tested for antimicrobial properties, **323** and **324** proved inactive towards *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*.



Awajanoran (325) is a new dihydrobenzofuran derivative, produced by the fungus *Acremonium* sp. which was obtained

from a sea mud sample in Japan.²³⁵ Compound **325** displayed moderate cytotoxic activity towards the A549 (human lung adenocarcinoma) cell line, and was also moderately active against five different bacterial strains as well as *Candida albicans*.



An isolate of *Aspergillus ustus*, obtained from the rhizosphere soil of the mangrove plant *Bruguiera gymnorrhiza* in China, produced ustusoranes A–F (**326–331**) together with the closely related known pergillin and eight new drimane sesquiterpenes.²³⁶ Pergillin had initially been reported as a plant growth inhibitor from a terrestrial isolate of *Aspergillus ustus*.²³⁷ Compound **330** displayed significant cytotoxicity against HL-60 cells, with an IC₅₀ value of 0.13 μ M.



The endophytic fungus *Phaeosphaeria spartinae*, isolated from a German red alga of the genus *Ceramium*, was the source of spartinoxide (**332**), which is the enantiomer of the known compound A82775C.²³⁸ The latter was previously reported from an unknown fungus of the order Sphaeropsidales.²³⁹ Compound **332** showed potent inhibition of human leukocyte elastase (HLE), but was inactive against bovine trypsin acetyl cholinesterase from *Electrophorus electricus*, and porcine cholesterol esterase.



JBIR-37 (333) and JBIR-38 (334) are glucosylated benzenediols that were obtained from the culture broth the fungus

Acremonium sp., isolated from an undisclosed Japanese sponge of the class Demospongiae.²⁴⁰ **333** and **334** were inactive when tested for cytotoxicity towards two cell lines. The endophytic fungus *Talaromyces* sp., isolated from the mangrove *Kandelia candel* in Hong Kong, yielded tenelate A (**335**) and B (**336**), together with the known compound, tenellic acid.²⁴¹ The latter had previously been reported from the aquatic fungus *Dendrospora tenella*.²⁴²





An unidentified fungus from an undisclosed Chinese mangrove habitat was reported to produce methyl 4-hydroxy-3-(3-methylbut-2-enyloxy)benzoate (**337**), exhibiting moderate antibacterial, antifungal and cytotoxic properties.²⁴³ 8-Hydroxy-3-methyl-5-(3-methylbut-2-enyloxy)isochroman-1-one (**338**) was detected in the culture broth of the fungus *Cephalosporium* sp., isolated from an undisclosed Chinese mangrove plant.²⁴⁴



The fungus Acremonium sp., isolated from the Korean sponge Stelletta sp., produced three new meroterpenoids,



chlorocylindrocarpol (339), acremofuranone A (340) and acremofuranone B (341), besides the new sesquiterpenoid dihydroxybergamotene (414) and nine known sesquiterpenoids.²⁴⁵ Compound 339 is a chlorinated analogue of cylindrocarpol, previously described from *Cylindrocarpon lucidum*,²⁴⁶ while the cyclic skeleton of 340 and 341 is unprecedented in Nature. Although some of the known metabolites exhibited various anti-inflammatory properties, 339–341 were inactive in the respective bioassays.

4.5 Terpenoids

The fungus *Phomopsis* sp., isolated from the Chinese mangrove plant, *Hibiscus tiliaceus*, produced four new unusual ring A-seco-oleanes, namely 3,4-seco-olean-11,13-dien-4,15 α ,22 β ,24-tetraol-3-oic acid (**342**), 3,4-seco-olean-11,13-dien-4,7 β ,22 β ,24-tetraol-3-oic acid (**343**), 3,4-seco-olean-13-en-4,7 α ,15 α ,22 α ,24-pentaol-3-oic acid (**344**), and 3,4-seco-olean-13-en-4,15 α ,22 α ,24-tetraol-3-oic acid (**345**).²⁴⁷ This is the first report of A-seco-oleanes in fungi, while oleane-type triterpenes, frequently found in terrestrial plants, have only rarely been reported from microbial including fungal sources. Interestingly, it has been demonstrated that some fungi are able to convert oleananes into A-seco-oleanes, which might be relevant with regard to the biosynthetic origin of **342–345**.^{248,249}



The rearranged triterpene, 6β,16β-diacetoxy-25-hydroxy-3,7dioxy-29-nordammara-1,17(20)-dien-21-oic acid (**346**) was isolated as well as a new pseurotin congener and three new diketopiperazines from a culture of the fungus *Aspergillus sydowi*, obtained from a driftwood sample collected in China.²⁵⁰ Compound **346** displayed significant antibiotic activity towards *Escherichia coli*, *Bacillus subtilis* and *Micrococcus lysoleikticus*. The new friedelane triterpene, 3β-hydroxyfriedelan-17β-carboxylic acid (**347**) was isolated from an unidentified mangrove endophytic fungus.²⁵¹



The fungus Gymnascella dankaliensis was isolated from the Japanese sponge Halichondria japonica, and yielded a series of structurally unusual steroid-type compounds, the pattern of which varied depending on media composition.²⁵² Dankasterones A (348)²⁵³ and B (349) were obtained when glucose in the original medium was replaced by soluble starch, while gymnasterones A-D (350-353) were isolated from malt-glucose-yeast media.²⁵² 348 and 349 are most unusual steroids possessing a $13(14 \rightarrow 8)abeo-8$ -ergostane skeleton, which so far only has been described once from Nature, resulting from a photochemical reaction of the insect molting hormone, (20R)-hydroxyecdysone.²⁵⁴ On the other hand, **350** is structurally intriguing since it represents an unprecedented steroid alkaloid with an additional ring and an amide-linked side chain derived from gymnastatins, a group of polyketides likewise described from Gymnascella dankaliensis. Compounds 348, 349 and 351-353 exhibited significant growth inhibition against the murine P388 cancer cell line, whereas 348 also exhibited potent growth inhibition against human cancer cell lines.



353 (R¹, R² = O)

The marine-derived fungus *Rhizopus* sp., isolated from the Chinese bryozoan *Bugula* sp., yielded six new ergosterols,

3 β -hydroxy-(22*E*,24*R*)-ergosta-5,8,22-trien-7,15-dione (**354**), 3 β -hydroxy-(22*E*,24*R*)-ergosta-5,8,14,22-tetraen-7-one (**355**), 3 β ,15 β -dihydroxy-(22*E*,24*R*)-ergosta-5,8(14),22-trien-7-one (**356**), 3 β ,15 β -dihydroxy-(22*E*,24*R*)-ergosta-5,8(14),22-trien-7-one (**357**), 3 β -hydroxyl-(22*E*,24*R*)-ergosta-5,8(14),22-trien-7,15-dione (**358**), and 5 α ,8 α -epidioxy-23,24(*R*)-dimethylcholesta-6,9(11),22-trien-3 β -ol (**359**).²⁵⁵ All compounds showed cytotoxic activity to varying degrees against four different cancer cell lines.



Six new fatty acid esters of the known steroids (22*E*)-ergosta-7,22-diene-3 β ,5 α ,6 α -triol (**360–363**) and (22*E*)-ergosta-7,22-diene-3 β ,5 α ,6 β -triol (**364**, **365**) were isolated from the fungus *Aspergillus awamori* isolated from soil around the mangrove plant *Acrostichum speciosum* in Hainan, China, besides various known steroids and their esters.²⁵⁶ All compounds exhibited mild cytotoxic activity against B16 and SMMC-7721 cell lines. Further analysis yielded the two new oxidized sterols, (22*E*)-7 α -



This journal is © The Royal Society of Chemistry 2011

methoxy- 5α , 6α -epoxyergosta-8(14),22-dien- 3β -ol (**366**) and (22*E*)- 3β -hydroxy- 5α , 6α , 8α , 14α -diepoxyergosta-22-en-7-one (**367**), both of which displayed weak cytotoxic activity towards the lung cancer cell line A549.²⁵⁷ Conformational analysis on the basis of the observed NOEs in the ROESY spectrum indicated that the cyclohexene oxide system in ring B of **366** adopted an *endo*-boat rather than a half-chair conformation.

The fungus *Penicillium* sp. was obtained from an undisclosed moss collected from the South Pole. Chemical analysis revealed the presence of a new sterol, ergosta-8(14),22-diene-3 β ,5 α ,6 β ,7 α -tetraol (**368**), together with four known sterols. **368** exhibited pronounced cytotoxicity against the human liver cancer cell line Hep G with an IC₅₀ value of 10.4 µg mL⁻¹.²⁵⁸



An unidentified fungus that was isolated from the surface of the Japanese brown alga *Ishige okamurae*, and based on DNA sequence analysis was grouped into the order Dothideales was identified as the producer of phomactin I (**369**), 13-*epi*-phomactin I (**370**) and phomactin J (**371**).²⁵⁹ Phomactins are an intriguing class of fungal diterpenes which have raised considerable attention due to their activity as platelet-activating factor (PAF) antagonists,²⁶⁰ and were initially discovered in cultures of the fungus *Phoma* sp. obtained from the shell of the crab *Chionoecetes opilio*,^{261,262} and it is therefore remarkable that the unidentified fungus in this study was taxonomically not closely related to the genus *Phoma*.



The fungus *Acremonium striatisporum* was isolated from superficial mycobiota of the sea cucumber *Eupentacta fraudatrix* collected from the Sea of Japan. Over the course of eight years, it was repeatedly studied and proved to be an exceptionally rich source of new isopimaradiene diterpene glycosides, virescenosides. The latest addition to this series were virescenosides V–X (**372–374**), which like most of the remaining virescenosides are substituted with an unusual β -D-altropyranose moiety.²⁶³

The marine fungus *Cryptosphaeria eunomia* var. *eunomia*, isolated from an unidentified Micronesian sponge, was found to produce the new pimarane-type diterpene 11-deoxydiaporthein A (**375**),²⁶⁴ together with known compounds from terrestrial fungi, diaporthein A and B, metabolites of *Diaporthe* sp.²⁶⁵ and scopararane A, previously described from *Eutypella scoparia*.²⁶⁶







A deep sea sediment-borne *Penicillium* sp., obtained from a depth of 5080 m, was found to produce a series of new diterpenes, conidiogenones B–G (**376–381**), besides four new diketopiperazine-type alkaloids (**501–504**).²⁶⁷ **376–381** are structurally related to conidiogenol and conidiogenone, metabolites from a terrestrial isolate of *Penicillium cyclopium* which were reported to possess potent conidiation-inducing activity.²⁶⁸ When tested for cytotoxic properties against four different cell lines, **376–381** displayed various degrees of activity, with **377** exhibiting potent activity towards HL-60 and BEL-7402 cells.



Peribysins H²⁶⁹ (**382**), I (**383**)²⁶⁹ and J (**384**)²⁷⁰ are eremophilane-type sesquiterpenoids produced by the fungus *Periconia byssoides* which was originally isolated from the Japanese sea hare, *Aplysia kurodai*. Similar to most other peribysins A–G^{175,176} which have been established as cell-adhesion inhibitors, **382** and **383** inhibited the adhesion of HL-60 cells to HUVEC (human umbilical vein endothelial cells) more potently than the standard control in this assay system, herbimycin A.



The fungus *Penicillium* sp., isolated from sea mud in the Bering Sea, yielded two new eremophilane sesquiterpenes, 3-acetyl-9,7(11)-dien-7 α -hydroxy-8-oxoeremophilane (**385**) and 3-acetyl-13-deoxyphomenone (**386**).²⁷¹ Compound **386** had been synthesized in the course of the structure elucidation of sporogen A0 I from a mycophilic *Hansfordia* sp.,²⁷² but had not been previously reported as a natural product. The epoxide **386** exhibited pronounced cytotoxic activity in the nanomolar range against three different cell lines, while the ring-opened alcohol **385** was less active by several orders of magnitude.



JBIR-27 (**387**) and JBIR-28 (**388**) were reported from the fungus *Penicillium* sp. which was obtained from the Japanese tunicate *Didemnum molle*.²⁷³ In addition, the closely related eremophilanes phomenone²⁷⁴ and its 13-deoxy congener,²⁷⁵ also known as sporogen-A01, were also obtained, which previously had been described from *Phoma exigua* var. *non-oxydabilis* and *Hansfordia pulvinata*, respectively. Compound **388** and the two known compounds showed moderate cytotoxicity towards the HeLa cell line, whereas **387** was inactive.



A marine-derived ascomycete related to the genus *Cryptosphaeria*, isolated from an unidentified ascidian in the Bahamas, produced cryptosphaerolide (**389**), an eremophilane type sesquiterpenoid bearing both an *exo*-methylene and an ester function.²⁷⁶ **389** is related to berkleasmin A, a metabolite of the terrestrial saprobic fungus, *Berkleasmium nigroapicale*.²⁷⁷ In the Mcl-1/Bak fluorescence resonance energy transfer (FRET) assay, **389** displayed inhibitory activity towards the Mcl-1 protein, a cancer drug target involved in apoptosis. In addition, **389** also showed significant cytotoxicity against the HCT-116 human colon carcinoma cell line, with IC₅₀ values in the lower μ M range.

An undisclosed mangrove endophytic fungal strain yielded the sesquiterpene microsphaeropsisin A (**390**),²⁷⁸ closely related to microsphaeropsisin described previously from a sponge-derived *Microsphaeropsis* sp.²⁷⁹ Due to its instability neither the relative

stereochemistry nor the biological activity of **390** could be investigated.



Seven new drimane sesquiterpenoids, hydroxylated derivatives of drim-7-en-6-one (391-393) and esters of 68.9a-dihydroxy-5adrim-7-en-11,12-olide with polyunsaturated acid substituents at C-6 (394-397), together with the related known compounds deoxyuvidin B, strobilactone B and RES-1149-2, were obtained from cultures of the fungus Aspergillus ustus, which was isolated from the Mediterranean sponge Suberites domuncula.^{203,280} Compounds 394, 395, and RES-1149-2 showed cytotoxic activity against a panel of tumour cell lines, and 395 was the most active, with an EC₅₀ value of 0.6 μ g mL⁻¹ against the L5178Y cell line. In a study which was published almost simultaneously, another isolate of Aspergillus ustus, obtained from the rhizosphere soil of the mangrove plant Bruguiera gymnorrhiza in China, produced eight drimane sesquiterpenes, termed ustusols A-C (391, 398, 399) and ustusolates A-E (400-403, 395), out of which two proved to be identical to the ones reported from the spongederived fungal strain mentioned above, besides another occurrence of RES-1149-2, and six new isochromane derivatives.²³⁶ In



This journal is © The Royal Society of Chemistry 2011



this latter report, the absolute configuration of **391** was established based on its CD spectrum and the octant rule for cyclohexenones, and **395** and **402** were found to exhibit moderate cytotoxicity, while **400** was weakly active. It is noteworthy that compounds **392** and **398**, reported from the two research groups as C-2 epimers, displayed virtually identical ¹³C NMR data, and are thus very likely to be identical. RES-1149-2 had previously been described as a metabolite from *A. ustus* var. *pseudoreflectus* isolated from a soil sample,^{281,282} and was found to act as endothelin type B receptor antagonist.²⁸³

Four new phenolic bisabolane-type sesquiterpenoids, (+)-sydowic acid (404), (+)-methyl sydowate (405), 7-deoxy-7,14didehydrosydonic acid (406) and 7-deoxy-7,8-didehydrosydonic acid (407), together with the known (+)-sydonic acid were isolated from a marine-derived *Aspergillus* sp., obtained from the Chinese gorgonian *Dichotella gemmacea*.²⁸⁴ The enantiomer of 404 ((-)-sydowic acid), as well as the corresponding racemate and sydonic acid, had previously been obtained from terrestrial strains of *Aspergillus sydowi*.^{285–287} Compounds 404, 405 and sydonic acid exhibited weak antibacterial activity against *Staphylococcus aureus*, but were inactive against methicillin-resistant *S. aureus*.







bisabolane sesquiterpenoids substituted with a diphenyl ether unit, and two new phenolic bisabolane sesquiterpenoids, (S)-(+)-11-dehydrosydonic acid (**410**) and (7*S*,11*S*)-(+)-12-acetoxysydonic acid (**411**).²⁸⁸ Compound **408** exhibited moderate cytotoxicity against the HL-60 cell line, while **409** inhibited the proliferation of A549 and HL-60 cells, while **410** and **411** were inactive.

Verticinols A (**412**) and B (**413**) are two new hydroxylated bisabolane-type sesquiterpenes which were reported from the fungus *Verticillium tenerum*, isolated from an unidentified marine alga.²⁸⁹ Although **412** and B **413** were tested for a variety of effects, *i.e.* antibacterial, antifungal, antialgal, antiplasmodial, antiviral, and cytotoxic activity as well as protein kinase inhibition or fat-accumulation inhibitory activity against 3T3-L1 murine adipocytes, they did not display significant activity in any of these test systems.

The fungus *Acremonium* sp., isolated from the Korean sponge *Stelletta* sp., produced the new sesquiterpenoid dihydroxybergamotene (**414**) and nine known sesquiterpenoids, together with three new meroterpenoids (**339–341**).²⁴⁵ **414** is a dihydroxylated congener of β -*trans*-bergamotene, which was reported from a terrestrial strain of *Aspergillus fumigatus*,²⁹⁰ but otherwise has widespread occurrence in essential oils of higher plants. Although some of the known metabolites exhibited various anti-inflammatory properties, **414** was inactive in the respective bioassays.



Four new hydroxylated sclerosporin derivatives 15-hydroxysclerosporin (**415**), 12-hydroxysclerosporin (**416**), 11-hydroxysclerosporin (**417**) and 8-hydroxysclerosporin (**418**), besides the known (+)-sclerosporin were produced by the fungus *Cadophora malorum*, isolated from the green alga *Enteromorpha* sp. upon long-term fermentation in a medium supplemented with artificial sea salt.²⁹¹ Sclerosporin was initially characterised as a sporogenic metabolite of a terrestrial isolate of *Sclerotinia fruticula*, and is a rare example of a fungal-derived cadinane-type sesquiterpene.^{292,293} Compounds **415–418** were subjected to a variety of assays, but were found devoid of significant biological activity, apart from **418** which showed a weak fat-accumulation inhibitory activity against 3T3-L1 murine adipocytes.

A large-scale fermentation of the fungus *Tryblidiopycnis* sp., an endophyte from woody tissue of a *Kandelia* sample from Hong Kong, yielded the new chlorinated monoterpene, (1S, 2S, 3S, 4R)-3-chloro-4-(2-hydroxypropan-2-yl)-1-methylcyclohexane-1,2-diol (**419**).²⁹⁴

4.6 Peptides including diketopiperazines

RHM1 (420) and RHM2 (421) are highly *N*-methylated linear octapeptides produced by the fungus *Acremonium* sp., which was



isolated from the sponge Teichaxinella sp. collected in Papua New Guinea.²⁹⁵ The fungus was considered by the authors "a notably atypical Acremonium sp.", since its unusual morphological characteristics were in disagreement with the results of the molecular biological identification which suggested a close similarity to Leucosphaerina indica. Compounds 420 and 421 exhibited mild cytotoxicity against murine L1210 cells, while 420 additionally possessed antibiotic properties against Staphylococcus epidermidis. Reinvestigation of the same fungal strain revealed the presence of additional RHM congeners, RHM3 (422) and RHM4 (423).²⁹⁶ In addition, two new efrapeptin derivatives, efrapeptin $E\alpha$ (424) and H (425), were characterised, besides the known efrapeptides E, F and G, so far only known from terrestrial sources. Efrapeptins are linear pentadecapeptides produced by the soil hyphomycete Tolypocladium niveum that have attracted considerable attention due to their ability to inhibit the soluble (F_1) part of the mitochondrial ATPase.²⁹⁷ The structure elucidation of 424 and 425, most importantly establishing their amino acids sequence, heavily relied on MS and MSⁿ analysis, while NMR spectroscopy was mainly used to establish the presence or absence of individual amino acids by individual characteristic signals.

The fungus *Tolypocladium* sp., obtained from sea mud in Japan, was recently identified as a source of the new efrapeptin analogue, efrapeptin J (**426**), together with the known efrapeptins F and G.²⁹⁸ In a reporter gene assay, all three compounds were shown to inhibit the expression of luciferase controlled by the GRP78 promoter in a dose-dependent manner. Furthermore, **426** also inhibited the protein expression of GRP78 in HT1080 cells and MKN-74 human gastric cancer cells, and induced cell death in HT1080 cells under endoplasmic reticulum stress.

The fungal strain *Trichoderma* sp., isolated from an unidentified marine sponge of undisclosed geographic origin, produced three new aminolipopeptides, trichoderins A (**427**), A1 (**428**) and B (**429**).²⁹⁹ Compounds **427–429** are characterized by the presence of an 2-amino-6-hydroxy-4-methyl-8-oxodecanoic acid (AHMOD) moiety, which they share with previously described aminolipopeptides such as the leucinostatins or trichopolyns, metabolites of *Penicillium lilacinum*³⁰⁰ and *Trichoderma polysporum*,³⁰¹ respectively. Compounds **427–429** showed potent anti-mycobacterial activity against *Mycobacterium smegmatis*, *M. bovis* and *M. tuberculosis*, with MIC values in the range 0.02– 2.0 µg mL⁻¹, and were effective against both actively growing and dormant states.

Six new peptaibols, asperelines A–F (**430–435**), characterised by an uncommon prolinol residue at the C-terminus, were produced by the psychrophilic fungus *Trichoderma asperellum*,



collected from a sediment sample in the Antarctic.³⁰² The absolute configurations of some of the amino acid residues were determined using a new method of ¹H NMR spectroscopic comparison of complexes formed with the chiral shift reagent Ru(D₄-Por*)CO. **430–435** exhibited weak inhibitory activity towards the phytopathogenic fungi *Alternaria solani* and *Pyricularia oryzae*, as well as against the bacteria *Staphylococcus aureus* and *Escherichia coli*.

A strain of *Trichoderma longibrachiatum* was isolated from blue mussel, *Mytilus edulis*, collected in France. Using various mass spectrometric techniques, eight new short peptaibols were identified, each of them comprising 11 amino acids, trichobrachins A I–IV (**436–439**) and trichobrachins B I–IV (**440– 443**).³⁰³ Reinvestigation of the same strain led to the partial characterisation by mass spectrometry of a further microheterogenous mixture of 30 peptaibols, out of which 21 were new.³⁰⁴ All shared the common motif Ac-Aib-xxx-xxx-Aib-Pro-xxx-Aib-Pro-xxol, and were assigned the names trichobrachins A when the residue in position 2 was an asparagine, and trichobrachin C when it was a glutamine.

The fungus *Clonostachys* sp., isolated from an unidentified Japanese sponge, yielded the symmetric cyclodepsipeptide IB-01212 (444).³⁰⁵ When its cytotoxic activity was evaluated against a panel of 14 different human tumour cell lines, 444 was most active against LN-caP (prostate cancer), SK-BR3 (breast cancer), HT29 (colon cancer), and HELA (cervix cancer) cell lines, with GI₅₀ values ranging in the order of 10⁻⁸ M.



Sclerotides A (445) and B (446), cyclic hexapeptides containing anthranilic acid and a dehydrotryptophan unit, were isolated in a nutrient-limited hypersaline medium from the marine-derived halotolerant *Aspergillus sclerotiorum*, isolated from a Chinese salt field.³⁰⁶ Compounds 445 and 446 exhibited moderate antifungal activity against *Candida albicans*, while 434 also showed weak cytotoxic activity against the HL-60 cell line and antibacterial activity against *Pseudomonas aeruginosa*. Compounds 445 and 446 were photointerconvertible *via* a radical mechanism.

From the brown alga *Durvillaea antarctica*, collected off the coast of New Zealand, an endophytic strain of *Gliocladium* sp. was obtained. Bioactivity profiling using a coupled HPLC/ microtiter assay system revealed that both the antimicrobial and the cytotoxic activity of the fungal crude extract were due to the presence of the known compound, 4-keto-clonostachydiol, a 14-membered macrodiolide previously reported in the patent literature. In addition, the new cyclodepsipeptide gliotide (**447**) was characterised, which proved inactive in the cytotoxicity and antimicrobial assays.³⁰⁷



An unidentifiable fungus was isolated from the sponge *Ian-thella* sp. collected in Papua New Guinea. In a seawater-based medium, it was found to produce two new guangomides A (448) and B (449), together with a new destruxin derivative, homo-destcardin (450).³⁰⁸ Compounds 448 and 449 are unusual due to the presence of two lactone moieties, while 450 is the first example of a destruxin analogue obtained from marine-derived fungi. Both 448 and 449 were devoid of cytotoxic activity, but displayed weak antibacterial activity against *Staphylococcus epidermidis* and *Enterococcus durans*.



Emericellamides A (451) and B (452) are cyclic lipodepsipeptides that were found to be produced by competing co-culture of the fungus *Emericella* sp., isolated from the surface of the green alga *Halimeda*. sp. collected in Papua New Guinea, in the presence of the marine actinomycete *Salinispora areni-cola*.³⁰⁹ Compounds **451** and **452** were produced in low yields by *Emericella* sp. alone, but upon co-culture their levels were enhanced by 100-fold. Both compounds showed modest antibacterial activities against methicillin-resistant *Staphylococcus aureus*. More recently, the biogenetic gene cluster for the production of **451** comprising a PKS–NRPS hybrid synthase has been identified through genomic data mining in the fully sequenced genome of *Aspergillus nidulans*.³¹⁰



The fungus *Scopulariopsis brevicaulis* was isolated from the Mediterranean sponge *Tethya aurantium*, and was shown to produce the two cyclodepsipeptides, scopularides A (**453**) and B (**454**).³¹¹ Their 3-hydroxy-4-methylalkanoic acid moiety is similar to the corresponding moiety in emericellamides A (**451**) and B (**452**), and both compounds displayed only weak antibacterial activity, but significantly inhibited growth of several tumour cell lines.



The fungus *Beauveria felina* was isolated from the green alga *Caulerpa* sp. collected off the coast of Brazil. Chemical investigation of its cytotoxic and anti-tuberculosis active butanone extract led to the discovery of two new destruxins, [β -Me-Pro] destruxin E chlorohydrin (**455**) and pseudodestruxin C (**456**), along with five known cyclic depsipeptides.³¹² The absolute configuration of the (2*R*,4*S*)-5-chloro-2,4-dihydroxypentanoic acid residue in **455** was established by the *J*-based configuration method in combination with chiral derivatisation. In a subsequent communication, the multi-screening approach leading to

the discovery of destruxins including **455** and **456** was explained in more detail.³¹³



The fungal strain *Spicellum roseum*, isolated from the Caribbean sponge *Ectyplasia perox*, yielded two new cyclohexadepsipeptides, spicellamides A (**457**) and B (**458**).³¹⁴ Both compounds displayed moderate cytotoxicity against neuroblastoma cells, but were inactive when assayed for antimicrobial effects against various test bacteria, fungi and algae. Since hydrolysis using Marfey's method revealed the presence of both D- and L-alanine in **457** and **458**, the position of this amino acid was determined by using a NOESY data in combination with molecular modelling.

Two new cyclic depsipeptides, 1962A (**459**) and 1962B (**460**), were produced by an unidentified fungal endophyte, obtained from the mangrove plant *Kandelia candel* collected in Hong Kong.³¹⁵ **459** and **460** were both found to contain one p-amino acid each. **459** exhibited mild cytotoxic activity towards human breast cancer MCF-7 cells.



Zygosporamide (461) is a new cyclic pentadepsipeptide which was obtained upon seawater-based fermentation of the fungus *Zygosporium masonii*.³¹⁶ Interestingly, the fungal strain was isolated from a cyanobacterium collected off Maui, Hawaii. So far, cyanobacteria have largely been neglected as a potential source of fungal diversity. 461 proved inactive against various drug-resistant bacteria, but displayed highly selective cytotoxicity in the National Cancer Institute cell line assay, with GI_{50} values ranging in the nanomolar range.

The cyclic pentadepsipeptide alternaramide (**462**) was detected in the culture broth of the fungus *Alternaria* sp. which was isolated from Korean sediment sample.³¹⁷ Compound **462** displayed weak antimicrobial activity against *Bacillus subtilis* and *Staphylococcus aureus*, and also inhibited protein tyrosine phosphatase 1B (PTP1B).

An Australian marine-derived isolate of *Aspergillus versicolor*, recovered from beach sand, was grown on a cellophane raft over



nutrient-rich media, and by this treatment was induced to produce two new cyclopentapeptides, cotteslosins A (463) and B (464), besides a new quinazoline-type alkaloid.³¹⁸ Compound 463 and 464 display structural similarities to the cyclic pentadepsipeptide sansalvamide A, previously reported from a seagrassderived *Fusarium* sp.,³¹⁹ and 464 was found to exhibit weak cytotoxicity against several cancer cell lines.



The fungus Microsporum cf. gypseum, obtained from a sample of the bryozoan Bugula sp. collected in the U.S. Virgin Islands, was found to produce two new cyclic tetrapeptides, microsporins A (465) and B (466).³²⁰ Compound 465 was synthesised by solidphase synthesis using a sulfonamide linker resin. 465 and 466 displayed marked inhibitory properties towards histone deacetylase, exhibited cytotoxic activity against HCT-116 cells (human colon adenocarcinoma), and were active in the National Cancer Institute's 60-cancer-cell panel. 465 contains the unusual amino acid, (2S)-2-amino-8-hydroxydecanoic acid, while 466 contains its oxidised derivative, (S)-2-amino-8-oxodecanoic acid (Aoda), respectively. The latter is present in other fungal cyclic tetrapeptides including the potent protozoan HDAC inhibitors apicidin³²¹ and 9,10-deepoxy-chlamydocin,³²² described from Fusarium pallidoroseum and Peniophora sp., respectively.

A strain of *Trichoderma reesei* separated from sea mud in China was found to produce the cyclotetrapeptide trichoderide A (**467**) which exhibited moderate cytotoxicity towards the A375-S2 melanoma cell line.³²³



The new JBIR-15 (**468**), together with the known aspochracin, were obtained from the culture broth of the fungus *Aspergillus sclerotiorum* which was isolated from the Japanese sponge *Mycale* sp.³²⁴ Aspochracin, an insecticidal cyclic tripeptide which differs from **468** by an *N*-methylated alanine residue, was originally reported from *A. ochraceus*.³²⁵ Both **468** and asprochacin were inactive when tested for cytotoxicity towards two cell lines.



The Australian sediment-derived fungus *Acremonium* sp. produced a novel family of lipodepsipeptides, acremolides A–D (469–472), together with known chaetoglobosins.³²⁶ Compounds 469–472 were devoid of antibacterial, antifungal or cytotoxic properties, nor did they synergize the cytotoxicity of the chaetoglobosins. The absolute stereochemistry of amino acid residues in 469–472 was established using a new C₃ Marfey's method, capable of discriminating between diastereomeric amino acid pairs such as isoleucine and *allo*-isoleucine, which are otherwise hard to resolve.



A new lipopeptide, fellutamide C (473), was isolated from a strain of *Aspergillus versicolor* which was obtained from the Korean sponge *Petrosia* sp.³²⁷ Compound 473 displayed mid-range cytotoxicity against several cancer cell lines, and is a derivative of the previously described fellutamides A and B, metabolites of a marine fish-derived *Penicillium fellutanum*,³²⁸ which however were not detected in the extract of *Aspergillus versicolor*.



Two new indolic enamides, terpeptins A (474) and B (475), were characterised from the fungus *Aspergillus* sp., an endophyte of the Chinese mangrove plant *Acanthus ilicifolius*, together with the known terpeptin.³²⁹ The latter is a mammalian cell cycle inhibitor initially described from *Aspergillus terreus*.³³⁰ All three compounds exhibited moderate cytotoxicity towards the A-549 cell line. Two further derivatives, JBIR-81 (476) and JBIR-82 (477), in addition to terpeptin, were reported recently from another *Aspergillus* sp., isolated from the Japanese brown alga *Sargassum* sp.³³¹ All three terpeptins displayed free radical scavenging activities as expressed by their ability to protect N18-RE-105 cells against L-glutamate toxicity, and were considerably more potent than the positive control, α -tocopherol.



The fungus *Aspergillus* sp., which was isolated from the common mussel, *Mytilus edulis*, yielded a series of complex prenylated diketopiperazines, notoamides A–D (**478–481**)³³² and F–K (**486–491**),³³³ with diverse modifications in their backbones. Compounds **478**, **479** and **488** contain the same intriguing spiroindolinone skeleton as encountered in sclerotiamide, previously reported as a metabolite of a terrestrial *Aspergillus sclerotiorum*,³³⁴ or in paraherquamide, a toxin produced by *Penicillium paraherquei*.³³⁵ Compounds **486**, **487** and **489** are related to stephacidin A (**492**), earlier described from an Indian isolate of *Aspergillus ochraceus*.³³⁶ The remaining congeners could be considered biosynthetic intermediates, since all of them are characterised by the presence of a rearranged isoprenyl moiety either at C-2 or C-3 of the indole, but lack additional cyclisation steps which link this substituent to the pyrrolo[1,2-*a*]pyrazine

This journal is © The Royal Society of Chemistry 2011

system in 478, 479 and 486-489, possibly by means of an intramolecular Diels-Alder reaction.337 Compounds 478-480 and 489 showed moderate cytotoxicity against HeLa cells, while the remaining congeners proved inactive. Moreover, 480 was found to induce G2/M-cell cycle arrest. Reinvestigation of the same fungus vielded notoamide E (482) which represents a short-lived biogenetic intermediate, and feeding studies with synthetic doubly ¹³C-labeled **482** resulted in the discovery of notoamides E2 (483), E3 (484) and E4 (485).³³⁸ A subsequent report³³⁹ disclosed the structures of notoamides L-N (493-495) and (-)-versicolamide B (496), the antipode of (+)-versicolamide B which was previously reported from a terrestrial fungicolous isolate of A. versicolor.³⁴⁰ Interestingly, a marine isolate of A. ostianus, obtained from an unidentified Japanese sponge, likewise yielded related congeners, including the new 21-hydroxystephacidin A (497), besides notoamide F (486).³⁴¹ Very recently, further notoamide derivatives, notoamides O-O (498-500) and notoamide R were reported from the original producer Aspergillus sp., but the latter compound appears to be identical to 497.342 Compound 498 possesses a hemiacetal/hemiaminal ether function which is unprecedented in the notoamide class of compounds.

A deep sea sediment-borne *Penicillium* sp., obtained from a depth of 5080 m, was found to produce the new diketopiperazine-type alkaloids meleagrins B (501) and C (502), roquefortins F (503) and G (504), besides a series of new diterpenes, conidiogenones B–G (376–381).²⁶⁷ Compounds 501 and 502 are related to meleagrin (a compound initially described from *Penicillium meleagrinum*,³⁴³ and since then rather frequently detected in various terrestrial and marine isolates of the genus *Penicillium*), but 501 is structurally most unusual since it also contains a diterpene-derived moiety. 503 and 504 are derivatives of roquefortin C, a very common mycotoxin from various species of *Penicillium* subgenus *Penicillium*.³⁴⁴ When tested for cytotoxic properties against four different cell lines, 501 displayed moderate activity against all cells, while the remaining compounds were less active.

The fungus Aspergillus fumigatus, isolated from the Chinese holothurian Stichopus japonicus, also produced a diverse suite of seven new prenylated indole diketopiperazine alkaloids.345 Compound 505 was identified as a structurally unique spiro-3indolinone derivative, while spirotryprostatins C-E (506-508) are spiro-2-indolinones related to spirotryprostatins A and B, previously described as mammalian cell cycle inhibitors from a terrestrial isolate of Aspergillus fumigatus.^{346,347} Compounds 509 and 510 which were obtained as an inseparable mixture of diastereomers, display the same skeleton as present in fumitremorgin, another well-known tremorgenic toxin from A. fumigatus.³⁴⁸ 13-Oxoverruculogen (511) contains a 1,2,4-dioxazocane ring formed from two isoprene moieties cross-linked by a peroxide bridge, and is a derivative of the known verruculogen, a metabolite from Penicillium verruculosum.³⁴⁹ Compounds 506-511 displayed moderate cytotoxic activity against four different cell lines.

Brevicompanines D–H (**513–517**) along with the known *allo*brevicompanine B (**512**) and fructigenine, were produced by the fungus *Penicillium* sp., obtained from a deep ocean sediment sample collected at a depth of 5080 m.³⁵⁰ Compound **512** was up to that point known as a synthetic product, and had not been reported as a natural product before. Brevicompanine alkaloids



were first reported as plant growth regulators from *Penicillium brevicompactum*.^{351,352} Compounds **514** and **517** were found to inhibit lipopolysaccharide (LPS)-induced nitric oxide production in BV2 microglial cells at concentrations at which they did not exhibit cytotoxic effects.

Eurotium rubrum, an endophytic fungus from the Chinese mangrove plant *Hibiscus tiliaceus*, was found to produce two new

threefold prenylated diketopiperazine derivatives, dehydrovariecolorin L (**518**) and dehydroechinulin (**519**).³⁵³ Both compounds displayed neither cytotoxicity nor radical scavenging activity. Compound **518** is the 8,9-dehydro derivative of variecolorin L, recently described from a halotolerant strain of *Aspergillus variecolor*,³⁵⁴ while **519** is the 8,9-dehydro congener of echinulin, described 50 years ago from *Aspergillus glaucus*.³⁵⁵

A series of prenylated diketopiperazines, 6-methoxyspirotryprostatin B (520), 18-oxotryprostatin A (521), and



14-hydroxyterezine (**522**), together with a new pseurotin congener and a new rearranged dammarane triterpene, were isolated from a culture of the fungus *Aspergillus sydowi*, obtained from a driftwood sample collected in China.²⁵⁰ Spirotryprostatin B and tryprostatin A have originally been described from *Aspergillus fumigatus* BM939, isolated from a sea sediment sample in Japan, and were identified as mammalian cell cycle inhibitors,^{347,356} while terezine D is a metabolite of the coprophilous fungus *Sporormiella teretispora*.³⁵⁷ Compounds **520–522** displayed mild cytotoxic activity towards the A-549 cell line.



The fungus *Penicillium janthinellum*, isolated from the Chinese soft coral *Dendronephthya* sp., was the source of two unusual *N*-oxygenated diketopiperazines, janthinolides A (**523**) and B (**524**).³⁵⁸ A sediment-borne isolate of *Gliocladium* sp. from China was found to produce the new diketopiperazines, gliocladride (**525**).³⁵⁹ and the isomeric PJ147 (**526**) and PJ157 (**527**),³⁶⁰ of which **526** displayed cytotoxicity towards the human A375-S2 melanoma cell line. A later reinvestigation resulted in the discovery of the closely related gliocladrides A (**528**) and B (**529**), both of which exhibited moderate cytotoxicity against three cell lines.³⁶¹

The fungus *Penicillium bilaii* MST-MF667, obtained from a boat ramp on the Huon estuary in Tasmania, was identified as a source of new bis(methylthio)diketopiperazines, bilains A–C (**530–532**).³⁶² All three proved inactive when tested for antibacterial, antifungal, antiparasitic and cytotoxic properties. Compounds **530–532** are closely related to *cis*-bis(methylthio)silvatin, described from a terrestrial strain of *Aspergillus silvaticus*,³⁶³ while the corresponding *trans*-stereoisomer of the latter had previously been characterised from a saltwater culture of the terrestrial fungus *Coriolus consors*.³⁶⁴



Published on 12 January 2011. Downloaded by Pennsylvania State University on 13/05/2016 04:15:05.



The fungus *Pseudallescheria* sp., isolated from the Korean brown alga *Agarum cribrosum*, was found to produce a new gliotoxin congener, dehydroxybisdethiobis(methylthio)gliotoxin (533), besides gliotoxin and a further known derivative.³⁶⁵ All three compounds displayed antibacterial activity against methicillin-resistant and multidrug-resistant *Staphylococcus aureus*. The related congener 534 was reported from an isolate of *Aspergillus fumigatus* that was cultured from the marine sediments collected China, together with gliotoxin and three known derivatives.³⁶⁶ Only gliotoxin and one of the known analogues displayed cytotoxic activity, while 534 was inactive.



Alternaria raphani, a halotolerant fungal strain isolated from sediment collected in a Chinese sea salt field, produced a new diketopiperazine, alternarosin A (535), together with three new cerebrosides.³⁶⁷ Compound 535 displayed very weak antimicrobial activity against *Escherichia coli*, *Bacillus subtilis* and *Candida albicans*, but was devoid of cytotoxic or radical scavenging activity. Seawater-based fermentation of the fungus *Chromocleista* sp. isolated from a sediment sample collected in the Gulf of Mexico led to the discovery of 4'-hydroxyphenopyrrozin (536), two epimeric diketopiperazines 537 and 538 and their decomposition product 539, in addition to a series of known natural products including diketopiperazines.³⁶⁸ Only 536 displayed mild antifungal activity against *Candida albicans*, while none of the compounds was active against *Staphylococcus aureus* or possessed cytotoxic properties.



The new diketopiperazine (Z)-3-benzylidene-8,8a-dihydroxy-2-methylhexahydropyrrolo[1,2-*a*]pyrazine-1,4-dione (**540**) was reported as a metabolite of an unidentified endophytic fungus of the Chinese mangrove plant *Acanthus ilicifolius*, together with a new nicotinamide alkaloid (**653**).³⁶⁹ Three new dioxopiperazine metabolites **541–543** were isolated from a strain of *Aspergillus fumigatus* which was obtained from marine sediments collected in China. Compounds **541–543** were devoid of cytotoxic activity.³⁷⁰



A new diketopiperazine, (Z)-6-benzylidene-3-hydroxymethyl-1,4-dimethyl-3-methylsulfanylpiperazine-2,5-dione (544), was isolated from the culture broth of the fungal strain CRIF2 belonging to the order Pleosporales, obtained from an unidentified sponge in Thailand.¹³⁴ Chemically and biogenetically, 544 is closely related to (3R,6R)-bisdethiodi(methylthio)hyalodendrin, previously reported from a soil-borne Penicillium turbatum,³⁷¹ which was also obtained in the current study. Both compounds exhibited weak cytotoxic activity toward various cell lines. The fungus Aspergillus, isolated from an unidentified Japanese calcareous sponge, produced JBIR-74 (545) and JBIR-75 (546) which were devoid of cytotoxic or antimicrobial activity.372 Cyclo(N-MeVal-N-MeAla) (547) was obtained from the culture broth of an unidentified fungus from the mangrove Avicennia marina in Hong Kong, and its structure was established by X-ray diffraction analysis.¹⁶⁸



4.7 Alkaloids and other nitrogen-containing metabolites

Plectosphaeroic acids A-C (548-550) are complex fungal alkaloids produced by Plectosphaerella cucumerina, obtained from sediments collected in British Columbia.³⁷³ Biogenetically, they appear to be derived from derived either a serine or an alanine and four tryptophan units, processed in three distinctly different ways, *i.e.* a diketopiperazine, removal of the side chain, and catabolic degradation via the kynurenine pathway to give two 3hydroxyanthranilic acid units, which in turn are oxidatively coupled to give the phenoxazinone heterocycle present in 548-550. Compounds 548-550 inhibited indoleamine 2,3-dioxygenase (IDO), an emerging molecular target for treating cancer involved in immune escape.³⁷⁴ The endophytic fungus Penicillium sp., isolated from the Chinese mangrove plant Aegiceras corniculatum, produced eight new janthitrem-type indole triterpenes, shearinines D-K (551-558).375 Compound 551 is the 22-hydroxy derivative of the known shearinine A, which had previously been described from ascostromata of the terrestrial fungus Eupenicillium shearii.376 Shearinine A and the known ergot alkaloids paspaline³⁷⁷ and paspalitrem³⁷⁸ were likewise detected in the culture broth of the fungus under study. Biogenetically, shearinines are suggested to arise from paspaline, which upon prenylation would undergo oxidative ring formation to yield paspalitrem A. The occurrence of 551-558 would then require a further prenylation step, oxidative cyclisation to yield the pyrane ring, and various combinations of olefinic rearrangements or oxidative cleavage of the C-2-C-18 bond, the latter of which might occur spontaneously, since analogous reactions were observed during the NMR measurements. Compounds 551, 552 and (to a lower extent) 554 exhibited significant in vitro blocking activity on large-conductance calcium-activated potassium channels.375 In a parallel study, a marine isolate of fungus Penicillium janthinellum was isolated from a Russian







in fact identical to shearinine K (**558**), while **559** and **560** are obviously new compounds and thus should be renamed. Interestingly, the NMR data of the H-22,H-23-*cis* congener **560** differ significantly from shearinine D (**551**), suggesting that both H-22 and H-23 follow the β -orientation. Shearinine A, **559**, and **560** were found to induce apoptosis in human leukemia HL-60 cells, while **560** also inhibited EGF-induced malignant transformation of murine JB6 P⁺ Cl 41 cells, indicative of a potential cancer preventive effect.

A strain of *Aspergillus flavus*, isolated from the Chinese green alga *Enteromorpha tubulosa*, yielded a new alkaloid, iso- α -cyclopiazonic acid (561), together with α -cyclopiazonic acid, the 5-epimer of 561.³⁸¹ α -Cyclopiazonic acid is a known indole–tetramic acid mycotoxin from the terrestrial fungus, *Penicillium cyclopium*, and is of interest due to its Ca²⁺–ATPase inhibition.³⁸² 561 displayed moderate cytotoxic activity against four different cell lines, but was considerably less active than α -cyclopiazonic acid.



The sponge-derived fungus *Gymnascella dankaliensis*, isolated from *Halichondria japonica* collected off Osaka in Japan, had previously been identified as a source of gymnastatins and structurally unusual steroids. Further chemical investigation led to the discovery of gymnastatins F–H (**562–564**), and the fatty acid- or polyketide-derived gymnamide (**565**).³⁸³ Compounds **562** and **563** possess a unique bicyclo[3.3.1]nonane ring system which differs from that of previously reported gymnastatins, and were





found to exhibit pronounced cytotoxic activity against the P388 cancer cell line. In a subsequent communication, additional gymnastatin congeners gymnastatins Q (566) and R (567) as well as the related dankastatins A (568) and B (569) were observed, when soluble starch instead of glucose was added to the culture broth.³⁸⁴ Compounds 566–569 displayed growth inhibition against the P388 cancer cell line, and furthermore, 566 inhibited growth of BSY-1 (breast) and MKN7 (stomach) human cancer cell lines. When cultured in a bromine-containing medium, the same fungus yielded the brominated analogues gymnastatins I–K (570–572), all of them exhibiting potent cytotoxicity towards the murine P388 lymphocytic leukemia cell line. Moreover, 570 and 571 displayed pronounced activity against a panel of 39 human cancer cell lines.³⁸⁵

Awajanomycin (573) is a bicyclic metabolite featuring both a γ -lactone and a δ -lactam ring, produced by the fungus *Acremonium* sp. which was obtained from a sea mud sample in Japan.³⁸⁶ Compound 573 was also converted into its reduced congener **574** and its hydrolysis product **575**. Compounds **573** and **574** displayed moderate cytotoxic activity towards the A549 (human lung adenocarcinoma) cell line, but were devoid of antimicrobial activity.



A new tyrosine-derived metabolite, aspergillusol A (**576**) was isolated on a gram scale, together with a methyl ester of 4hydroxyphenylpyruvic acid oxime (**577**) and the known secalonic acid A, from the marine-derived fungus *Aspergillus aculeatus*, obtained from the Thai sponge *Xestospongia testudinaria*.³⁸⁷ Compound **576** exhibits a striking structural analogy to wellknown tyrosine alkaloids from Verongida sponges, most notably psammaplin A,³⁸⁸ which raises interesting questions regarding the biosynthetic origin of the sponge metabolites. Compound **577** had previously only been known synthetically, and not as a natural product, although it cannot be excluded that it actually represents an artefact derived from methanolysis of **576**. Compound **576** inhibited α -glucosidase from *Saccharomyces cerevisiae*, but was inactive towards α -glucosidase from *Bacillus stearothermophilus*.



Bioassay-guided fractionation of the culture broth of an Aspergillus fumigatus isolate which was obtained from a deepwater sediment collected off Vanuatu, using a yeast-halo assay with wild-type and cell-cycle mutant strains of the budding yeast Saccharomyces cerevisiae,389 led to the discovery of 11-Omethylpseurotin A (578),³⁹⁰ besides the known but inactive pseurotin A (579), initially characterised from Pseudoeurotium ovalis.³⁹¹ Compound 578 displayed the greatest activity differential versus the wild-type strain against the Hof1 (hof1 Δ) haploid deletion strain, carrying a mutation in a gene involved in cytokinesis. 14-Norpseurotin A (580) was discovered along with three new diketopiperazines and a new rearranged dammarane triterpene, from a culture of the fungus Aspergillus sydowi, obtained from a driftwood sample collected in China.²⁵⁰ Compound 580 displayed significant antibiotic activity towards Escherichia coli, Bacillus subtilis and Micrococcus lysoleikticus.

Likewise closely related to pseurotin A (**578**) is cephalimysin A (**581**), isolated from a strain of *Aspergillus fumigatus* which was originally separated from the marine fish *Mugil cephalus*.³⁹² Compound **581** exhibited significant cytotoxic activity against P388 and HL-60 cells.



The fungus *Aspergillus sydowi*, isolated from marine sediments collected in China, was identified as the producer of two additional pseurotin derivatives, azaspirofurans A (**582**) and B (**583**), characterized by the presence of an additional furan ring in the side chain.³⁹³ Compound **582** exhibited moderate cytotoxicity against the A549 cell line.



The fungus Spicaria elegans was isolated from marine sediments collected in China, and proved to represent a prolific source of new cytochalasin derivatives with various carbon skeletons. Cytochalasins Z_7 – Z_9 (584–586) are rare examples of cytochalasins with a 12-membered macrolactone ring, exhibiting mild cytotoxicity towards cell lines P388 and A-549 in the MTT assay.^{128,394} Reinvestigation of the same fungus resulted in the discovery of cytochalasins Z₁₀-Z₁₅ (587-592), representing the first cytochalasin congeners from nature with an open 8-carbon side chain instead of the usual 11-14-membered macrocyclic ring.³⁹⁵ 588 and 589 displayed moderate cytotoxicity toward A-549 cells, whereas the remaining compounds were inactive. By varying the culture conditions according to the OSMAC (one strain-many compounds) approach, a drastically altered metabolite profile was obtained, resulting in the discovery of the novel spicochalasin A (593) and aspochalasins M-Q (594-598), besides the known aspochalasins B and D.³⁹⁶ Compound 593 possesses an unusual pentacyclic ring system, and similar to 594 exhibited modest cytotoxic activity against HL-60 cells. On the other hand, treatment of the same fungus with the cytochrome P-450 inhibitor metapyrone resulted in the production of two new 7-deoxycytochalasins, 7-deoxy-cytochalasin Z_7 (599) and 7deoxy-cytochalasin Z_9 (600).³⁹⁷ It is interesting to note that although oxidation at C-7 was prevented, the "Baeyer-Villigerlike" oxidation at C-9 still occurred. Compound 599 displayed moderate cytotoxic activity towards the A-549 cell line, while 600 was inactive. Cytochalasins Z_{16} – Z_{20} (601–605) were subsequently reported from an endophytic strain of Aspergillus flavipes,





он

ΗŃ

|| 0

605

589 (R¹ = R³ = OH, R² = R⁴ = H) **590** (R¹, R² = O, R³ = H, R⁴ = OH) **591** (R¹ = H, R² = OH, R³, R⁴ = O) **592** (R¹ = OH, R² = H, R³, R⁴ = O)



587 (R¹ = R³ = OH, R² = R⁴ = H) **588** (R¹, R² = O, R³ = H, R⁴ = OH)









ОН

OCH₃

ö

604

OH

E HQ

0

H,

СНО

Ŕ¹∬ 0

606 ($R^1 = OCH_3$, $R^2 = OH$)

607 (R¹ = OH, R² = H)

608 (R¹ = OH, R² = OH)

R²







isolated from the Chinese mangrove plant, *Acanthus ilicifolius*.³⁹⁸ When tested for cytotoxicity towards various cell lines, **602** was moderately active, followed by **601** and **604**. A recent reinvestigation of *Spicaria elegans*, using a longer fermentation time, yielded additional aspochalasin congeners, aspochalasins R–T (**606–608**), all of which proved inactive in terms of cytotoxic effects.³⁹⁹

Xylarisin (609) is a new cytochalasin derivative with a highly substituted perhydroisoindole moiety linked to an 11-membered



ring that was produced by the fungus *Xylaria* sp., isolated from the Thai gorgonian sea fan, *Annella* sp.⁴⁰⁰ Compound **609** displayed mild antibacterial activity against *Staphylococcus aureus* and methicillin-resistant *S. aureus*. Chemical investigation of the endophytic fungus *Chaetomium globosum* derived from the Chinese green alga *Ulva pertusa* resulted in the discovery of seven new cytochalasan derivatives of the chaetoglobosin class, cytoglobosins A–G (**610–616**), together with the structurally related known compounds, isochaetoglobosin D and chaetoglobosin F_{ex} .⁴⁰¹ The latter two compounds are metabolites of a soil-borne *C. globosum*⁴⁰² and *C. subaffine*,⁴⁰³ respectively. Compounds **612** and **613** exhibited moderate cytotoxic activity against the A-549 cell line. The fungus *Exophiala* sp., isolated from the surface of the

Korean marine sponge Halichondria panicea, yielded circumdatin I (617), a member of the circumdatin class of alkaloids,⁴⁰⁴ together with the known circumdatins C405 and G,406 previously reported from a terrestrial and a marine-derived strain of Aspergillus ochraceus, respectively. All three compounds exhibited UV-A protecting activity which exceeded that of the positive control, the commercially applied sunscreen agent oxybenzone. From the culture of an Aspergillus ostianus strain isolated from an unidentified marine sponge at Pohnpei, Micronesia, the two known circumdatins A (618) and B (619) as well as the new circumdatin J (620) were obtained.⁴⁰⁷ Compound 618 and 619 were initially described as metabolites of from a terrestrial isolate of the fungus Aspergillus ochraceus, and their structures reported as zwitterions.⁴⁰⁵ In the present study, both structures were revised on the basis of X-ray crystallographic analysis, and 618 and 619 were now found to contain a very unusual oxepin framework in the "Southern" part. Compound 620 is the 15-deoxy congener of circumdatin D, likewise reported from A. ochraceus.408 Compound 618-620 exhibited neither antimicrobial nor cytotoxic properties. An endophytic strain of A. ochraceus, obtained from the Chinese brown alga Sargassum kjellmanianum, yielded the new 2-hydroxycircumdatin C (621), together with the anthranilic acid-containing diketopiperazine 622, which up to that point was only known synthetically.⁴⁰⁹ Compound 621 displayed significant DPPH radical-scavenging activity, and was more potent than butylated hydroxytoluene (BHT), which served as a positive control.

From the Chinese sponge *Mycale plumosa*, the fungus *Penicillium aurantiogriseum* was isolated, which was found to produce three new quinazoline alkaloids, aurantiomides A–C (**623–625**).⁴¹⁰ Compounds **624** and **625** displayed moderate cytotoxicity against several cell lines, while **623** was inactive. All three compounds are closely related to anacine, a metabolite previously described from terrestrial strains of *P. aurantiogriseum*⁴¹¹ and *P. verrucosum*.⁴¹²





An Australian marine-derived isolate of *Aspergillus versicolor*, recovered from beach sand, was grown on a cellophane raft over nutrient-rich media, and by this treatment was induced to produce a new quinazoline-type alkaloid, cottoquinazoline A (**626**), besides two new cyclopentapeptides.³¹⁸ Compound **626** is the 3-nor derivative of the known fumiquinazoline D, previously reported from a strain of *Aspergillus fumigatus* separated from the marine fish *Pseudolabrus japonicus*.⁴¹³



The fungal strain *Penicillium* sp., isolated from the red alga *Laurencia* sp. collected in the Bahamas Islands, yielded penilumamide (**627**), an unusual alkaloid combining a lumazine system with L-methionine sulfoxide and an anthranilic acid ester.⁴¹⁴ The 1,3-dimethyllumazine-6-carboxamide moiety within a natural product had previously only been reported from the freshwater leech *Limnatis nilotica*.⁴¹⁵ The structure elucidation of **627** was severely hampered by the lack of protons in the lumazine part, but was achieved by a combination of 1,1-ADEQUATE

and ¹H,¹⁵N HMBC experiments, use of the NMR-based structure generator COCON,⁴¹⁶ and detailed analysis of the ESI-TOF MS fragmentations. Compound **627** was inactive when tested for cytotoxic and antimicrobial properties, and also did not affect cellular Ca²⁺ signalling in neuroendocrine cells.

The dimeric alkaloid xylopyridine A (**628**) was isolated from the endophytic fungus *Xylaria* sp., obtained from an undisclosed mangrove in Hong Kong.⁴¹⁷ By fluorescence quenching and spectrophotometric titration experiments, **628** was demonstrated to possess strong DNA-binding affinity, presumably *via* an intercalation mechanism.



In an interesting study, it was shown that marinamide (**629**) and its methyl ester (**630**) are only produced when two unidentified fungal strains, obtained from an undisclosed plant growing in an estuarine mangrove in Hong Kong, were fermented together, but not when either of the two fungi were cultured on their own.⁴¹⁸ It is a bit unfortunate that so little detail about the identity and the origin of the two strains were reported, since while there are very few examples of competing co-culture of fungi and bacteria leading to the production of new secondary metabolites,^{419,420} this study seems to represent the first example involving two fungal strains.

The isomeric pyrrolyl 4-quinolinone alkaloid penicinoline (631) was isolated from the endophytic fungus *Penicillium* sp., isolated from the Chinese mangrove plant *Acanthus ilicifolius*.⁴²¹ Upon methylation, an *N*-methylated lactam was unexpectedly formed. Compound 631 exhibited potent cytotoxicity towards the 95-D and HepG2 cell lines, and also displayed strong insecticidal activity against *Aphis gossypii*.



The new isoquinoline alkaloid **632** was reported from the endophytic fungus *Phomopsis* sp., isolated from an undisclosed Chinese mangrove plant.⁴²² However, the trivial name assigned to **632**, phomopsin A, must be rejected, since it is already in use for a cyclic hexapeptide mycotoxin produced by *Phomopsis leptostromiformis*, the causative agent of the disease lupinosis.⁴²³ Compound **632** exhibited mild cytotoxicity towards two cell lines.

The structurally unique isocoumarin-substituted azepine alkaloid Sg17-1-4 (633) is a metabolite of the fungus *Alternaria*

tenuis, which was obtained from an undisclosed Chinese alga.⁴²⁴ Compound **633** displayed weak and moderate cytotoxic activity towards human malignant A375-S2 and human cervical cancer HeLa cells, respectively.



Sorbicillactone A (634), produced by marine-derived strains of *Penicillium chrysogenum*, represents the first example of a sorbicillin-derived alkaloid,⁴²⁵ and its production at a 100 g scale has been achieved, in order to provide sufficient amounts for preclinical evaluation and structure–activity relationship (SAR) studies.⁴²⁶ Compound 634 has raised considerable interest due to its cytostatic activity against murine leukemic lymphoblasts (L5178y) and the ability to protect human T cells against the cytopathic effects of HIV-1.⁴²⁷



The fungus Paecilomyces marquandii, isolated from an intertidal sediment sample collected in Argentina, produced an ureido Diels-Alder adduct of sorbicillinol (635).428 Even though a series of sorbicillinoids have been described from fungi previously, this seems to represent the first example of a Diels-Alder adduct of sorbicillinol with another non-sorbicillol-type moiety, in this case maleimide. However, the putative biogenetic precursors were not detected in this study. Interestingly, both the ESI as and the EI mass spectra of 635 were dominated by the corresponding fragments resulting from retro-Diels-Alder fragmentation. Similarly, from the culture broth of the fungus Aspergillus niger, endophytic to the Chinese brown alga Colpomenia sinuosa, an unusual steroid derivative, ergosterimide (636), was obtained.429 Formally, 636 is a Diels-Alder adduct of (22E,24R)-ergosta-5,7,14-trien-3β-ol and maleimide. The latter is widely used for technical applications, and due to its high reactivity is commonly used as a reactant for Diels-Alder reactions in synthetic laboratories. If maleimide indeed was a metabolite of the two fungal strains under study, 635 and 636 would represent the first natural Diels-Alder adducts of this type. The same isolate of Aspergillus niger also produced a new amino-substituted dihydrostyrylpyrone (637) which displayed moderate antifungal activity against Candida albicans.⁴³⁰ This type of fungal natural product seems to be restricted to Aspergillus section Nigri,431 and the only reported derivatives reported so far include pyrophen^{31,432} and aspernigrin B.^{433,434} Subsequently, isopyrophen (638) and aspergillusol (639) were reported from the same fungus⁴³⁵ Compound **638** was reported to be a stereoisomer of pyrophen, which would require the existence of stable *cis/trans* isomers at the amide bond.



The endophytic fungus *Penicillium* sp., isolated from the bark of the Chinese mangrove plant *Aegiceras corniculatum*, yielded a series of tetramic acids, penicillenol A₁ (640), A₂ (641), B₁ (642), B₂ (643), C₁ (644) and C₂ (645).⁴³⁶ 640–643 displayed moderate degrees of cytotoxicity towards HL-60 cells, while congeners 644 and 645, which possess an additional double bond near the chain terminus, were inactive.



Beauversetin (**646**) is a 3-decalinoyl tetramic acid derivative produced by the fungus *Beauveria bassiana*, isolated from the German sponge *Myxilla incrustans*.⁴³⁷ Compound **646** exhibited moderate cytotoxic activity towards a panel of 6 cell lines, and is structurally related to equisetin, a metabolite of *Fusarium equiseti*.⁴³⁸

The fungus *Pestalotiopsis* sp., isolated from the Chinese mangrove plant *Rhizophora mucronata*, was found to produce a new alkaloid, pestalotiopsoid A (647), in addition to a series of new cytosporones and coumarins (269–278).¹⁹⁸



The biosynthesis of the 2-azaanthraquinone, scorpinone (**648**), originally described from the marine sediment-derived fungus *Amorosia littoralis*,⁴³⁹ has been demonstrated to proceed *via* a regular polyketide pathway with a linear heptaketide as biosynthetic intermediate,⁴⁴⁰ which would also account for the presence of the known cyclic hemiketal herbarin, originally isolated from a terrestrial *Torula herbarum*.⁴⁴¹ Thus, it seems clear that at some point late in the biosynthesis of **648**, an exchange of an oxygen for a nitrogen atom has to occur, however, it is not possible to decide whether this nitrogen is derived from inorganic nitrogen or a nitrogen-containing organic precursor.

A sediment-borne *Aspergillus carbonarius* from China yielded carbonarone A (**649**), a new γ -pyrone, and the related α -pyridone derivative carbonarone B (**650**).⁴⁴² Both compounds displayed moderate cytotoxic activity against K562 cells.



A marine-derived *Penicillium* sp., isolated from the brown alga *Xiphophora gladiata* collected in New Zealand, produced two new *N*-deoxy analogues **651** and **652** of the known *N*-hydroxy-2-pyridone alkaloids, PF1140 and akanthomycin.⁴⁴³ PF1140, which was also detected in the culture broth, was previously reported from an *Eupenicillium* sp.,⁴⁴⁴ while akanthomycin is a metabolite of the entomopathogenic fungus *Akanthomyces gracilis*.⁴⁴⁵ PF1140 displayed modest antimicrobial and cytotoxic properties, whereas **651** and **652** were inactive.



The new alkaloid 4-hydroxy-6-(hydroxy(phenyl)methyl)-*N*-(3methylbutanoyl)nicotinamide (**653**) was reported as a metabolite of an unidentified endophytic fungus of the Chinese mangrove plant *Acanthus ilicifolius*, together with a new diketopiperazine.³⁶⁹

Pyranonigrin A (**654**) was the major UV-active compound in the extract of the fungus *Aspergillus niger*, which was obtained from mangrove wood in Hong Kong.⁴⁴⁶ Compound **654** had previously been reported from a sponge-derived *A. niger* strain,⁴³³ but based on extensive analysis of its spectroscopic properties including an ¹H–¹⁵N HMBC NMR experiment, its structure was revised as given in **654**. Since the initial discovery of **654** in a marine-derived *A. niger*, this metabolite has been shown to occur in at least six different terrestrial species of *Aspergillus* section *Nigri*,⁴⁴⁷ and also in rice mold starters used in the manufacturing process of fermented foods.^{448,449}



N-Formyl-2-(4-hydroxyphenyl)acetamide (**655**) was described as a metabolite of an unidentified fungus obtained from the Korean brown alga *Ishige okamurae*, together with further known hydroxylated phenylacetic acid derivatives.⁴⁵⁰ Compound **655** exhibited pronounced radical scavenging activity in the DPPH assay, and was more active than the positive control, ascorbic acid.

N-Methyl-1*H*-indole-2-carboxamide (**656**) from the fungus *Cladosporium cladosporioides*, isolated from the Chilean sponge *Cliona* sp., has apparently never been reported as a natural product before, but has now been described and its structure determined by single-crystal X-ray diffraction.⁴⁵¹

Two new indole derivatives, 2-(1H-indol-3-yl)ethyl 2-hydroxypropanoate (657) and 2-(1H-indol-3-yl)ethyl 5-hydroxypentanoate (658), were reported from the yeast *Pichia membranifaciens*, isolated from the Japanese sponge *Halichondria okadai*.⁴⁵² Chiral HPLC revealed 657 to occur as a 5 : 8 (*S/R*) mixture. Both compounds exhibited weak radical scavenging activity in the DPPH assay.



The fungal strain *Trichoderma atroviride*, isolated from sediment near the Chinese mangrove *Ceriops tagal*, produced the new compound 3-hydroxybutan-2-yl 4-(2-hydroxy-*N*-(3-oxobutan-2-yl)propanamido)butanoate (**659**).²¹¹

ZZF51(A) (660) is a copper complex of two fusaric acid (5butylpicolinic acid) moieties produced by the fungus *Fusarium* sp., an endophyte of the Chinese mangrove plant *Castaniopsis fissa.*⁴⁵³ Fusaric acid is a mycotoxin discovered in the 1930s in various *Fusarium* species, and has been shown to possess multiple effects on the mammalian nervous, cardiovascular and immune system.⁴⁵⁴ In culture, the producing strain was able to tolerate concentrations of Cu(II) ions up to 300 ppm, and showed active biosorption of this heavy metal. Compound **660** displayed mild antibiotic activity against 4 bacterial strains, and also displayed cytotoxicity towards 3 different tumour cell lines.

In the culture extract of the fungus *Botrytis* sp., obtained from the surface of the Korean green alga *Enteromorpha compressa*, the new nitrogen-containing cyclopentenone bromomyrothenone B (661) was detected, besides the new non-nitrogenated cyclopentenone botrytinone (299).²¹⁶ Compound 661 did not display significant activity during a screening for radical scavenging, tyrosinase inhibitory and antimicrobial properties. Previously, the debrominated congener of 661, myrothenone B, had been reported from two independent sources, an algicolous *Myrothecium* sp. from the same algal species,⁴⁵⁵ and interestingly, also from an endophytic *Streptomyces* sp. isolated from the Chinese mangrove plant *Aegiceras comiculatum*.⁴⁵⁶ Subsequently, 661 and 299 were also detected in the culture medium of *Rhizopus stolonifer*, isolated from the surface of the Korean brown alga, *Sargassum horneri*.²¹⁷



4.8 Shikimate-derived metabolites

An endophytic strain of *Aspergillus sydowii* was isolated from the inner tissue of the red alga *Acanthophora spicifera*, collected in India. Chemical analysis revealed the presence of two new chlorinated 2,5-diarylcyclopentenones, sydowin A (**662**) and B (**663**).⁴⁵⁷ Two aromatic butenolides, aspernolides A (**664**) and B (**665**), along with known metabolites including butyrolactone I, were obtained from the fermentation broth of the fungus *Aspergillus terreus*, which was isolated from the Indian soft coral *Sinularia kavarattiensis*.⁴⁵⁸ Butyrolactone I, assumed to be a biogenetic precursor of **664** and **665**, was originally described from a terrestrial isolate of *A. terreus*.⁴⁵⁹ Compound **664** was also shown to be formed from butyrolactone I by heating⁴⁵⁸ or treatment with dilute hydrochloric acid,⁴⁵⁹ and exhibited mild cytotoxicity against several cancer cell lines.





Phomoindene A (**666**), a rare example of an indene derivative from Nature, was described from the fungus *Phomopsis* sp., obtained from a sediment sample in a Chinese mangrove habitat, but was inactive when tested for cytotoxic properties.⁴⁶⁰

An isolate of *Aspergillus candidus*, obtained from a sediment sample collected in Japan, yielded prenylterphenyllin (**667**), 4"-deoxyprenylterphenyllin (**668**) and 4"-deoxyisoterprenin (**669**), together with the known 4"-deoxyterprenin, all of which exhibited moderate cytotoxic activity against KB3-1 cells.⁴⁶¹ Compounds **667–669** are analogues of terphenyllin⁴⁶² and terprenin,⁴⁶³ known metabolites from terrestrial strains of *A. candidus*. Through labelling studies with phenylalanine, it has been demonstrated that the biosynthesis of terphenyllin and the related volucrisporin involves self-condensation of two phenyl-propanoid units to give the terphenyl ring system.⁴⁶⁴



The biphenyl derivative 4-(3-hydroxypropyl)-5,6-dimethoxybiphenyl-3,4'-diol (670) was obtained from the culture broth of the fungus *Penicillium thomi*, an endophyte of the root of the Chinese mangrove *Bruguiera gymnorrhiza*.⁴⁶⁵ It displayed moderate cytotoxicity towards three human cancer cell lines, A549, HepG2 and HT29. From the saltwater culture of an *Arthrinium* sp., derived from a marine sediment collected at a depth of 550 m near the U.S. Virgin Islands, the known compound tyrosol (a common degradation product of tyrosine) and its new derivative, tyrosol carbamate (671), were obtained.⁴⁶⁶

The fungus *Penicillium griseofulvum*, isolated from the mangrove *Lumnitzera racemosa*, yielded 4-hydroxyphenethyl methyl succinate (**672**) and 4-hydroxyphenethyl 2-(4-hydroxyphenyl)acetate (**673**), both displaying moderate radical scavenging properties, while **673** also exhibited weak cytotoxic activity.⁴⁶⁷ The fungus *Nigrospora sphaerica* was obtained from mud in an intertidal zone in China, and was found to produce 1-(5-oxotetrahydrofuran-2-yl)ethyl 2-phenylacetate (**674**) and 3-hydroxybutan-2-yl 2-hydroxy-3-phenylpropanoate (**675**).⁴⁶⁸



Four unusual allenic ethers, (*E*)-methyl 3-(4-(buta-2,3-dienyloxy)-3-methoxyphenyl)acrylate (**676**), (*E*)-methyl 3-(4-(4-(buta-2,3-dienyloxy)benzyloxy)) acrylate (**677**), (*E*)-methyl 3-(4-(4-(buta-2,3-dienyloxy)benzyloxy)-3-methoxyphenyl)acrylate (**678**), and methyl 4-(4-(buta-2,3-dienyloxy)benzyloxy)benzoate (**679**) have been reported from the fungus *Xylaria* sp., obtained from the seeds of an undisclosed angiosperm mangrove tree from Hong Kong.⁴⁶⁹ The allenic ether moiety in **677–679** is also present in xyloallenolide **B** (**186**), a metabolite from the same fungus.¹⁴⁰

4.9 Lipids

The fungus Aspergillus niger, endophytic to the Chinese brown alga Colpomenia sinuosa, yielded asperamides A (680) and B



(681), a sphingolipid and its corresponding cerebroside.⁴⁷⁰ Compound 680 showed moderate antifungal activity against Candida albicans. While sphingolipids containing a 9-methyl-C₁₈-sphingosine moiety have been described from natural sources on various occasions,⁴⁷¹ 680 and 681 contain a hitherto unreported 9-methyl-C₂₀-sphingosine moiety. Structurally related cerebrosides, asperiamides B (682) and C (683), were obtained from another strain of Aspergillus niger, isolated from seawater in China, together with two known intermediates of aflatoxin biosynthesis, averufin and nidurufin. The latter two displayed moderate antiviral activity against tobacco mosaic virus, while 682 and 683 were inactive.⁴⁷² Alternaria raphani, a halotolerant fungal strain isolated from sediment collected in a Chinese sea salt field, produced three new cerebrosides, alternarosides A-C (684-686), together with a new diketopiperazine.367 Compounds 684-686 displayed very weak antimicrobial activity against Escherichia coli, Bacillus subtilis and Candida albicans, but were devoid of cytotoxic or radical scavenging activity.

A total of 16 fungal strains were isolated from various tissues of the fish, *Argyrosomus argentatus* (white croaker). Screening for antifungal activity against the human-pathogenic *Candida albicans*, *Aspergillus niger* and *Trichophyton rubrum* identified a *Myrothecium* sp. as the most active isolate, which produced mainly known trichothecenes, but also the new 4,5-ditridecyloctanedioic acid (687).⁴⁷³ Compound 687 was inactive in the antifungal bioassays. The fungus *Aureobasidium* sp., isolated from Neptune Grass, *Posidonia oceanica*, yielded an unusual diester, aureobasidin (688), along with two hydroxylated decanoic acids (689 and 690).⁴⁷⁴ Compounds 688 and 689 inhibited larval settlement of *Balanus amphitrite* larvae, while 690 displayed antifungal activity against *Candida albicans*. All three compounds exhibited antibiotic activity against *Staphylococcus aureus*, *Escherichia coli* and *Bacillus subtilis*.

5 Conclusion

Fungi from marine habitats are a prolific source of new chemical diversity, and so far have provided more than 1000 new natural products, some of them with clinically relevant pharmacological activity. Probably the most important representative is the diketopiperazine halimide, which initially was discovered by Bill Fenical's group in the 1990s,⁴⁷⁵ and which acts as a tubulindepolymerising agent. This molecule served as a lead structure for the closely related synthetic analogue plinabulin (NPI-2358), currently undergoing phase II clinical trials in patients with advanced non-small-cell lung cancer.476-478 Other important classes of metabolites include for examples the phomactins, the peribysins, or the tryprostatins, all of them mentioned in this review. The study of the secondary metabolite chemistry of marine-derived fungi has gained momentum in recent years, especially with the entry of Chinese researchers into the discipline, and there continues to be a steady increase in the total number of new structures reported in the chemical literature each year. In light of the pronounced discrepancy between the actual number of cultivated strains and the estimated biodiversity of fungi in marine habitats in general, it is to be expected that this trend will continue in the years to come.





6 References

- 1 T. S. Bugni and C. M. Ireland, Nat. Prod. Rep., 2004, 21, 143–163.
- 2 M. Saleem, M. S. Ali, S. Hussain, A. Jabbar, M. Ashraf and Y. S. Lee, *Nat. Prod. Rep.*, 2007, **24**, 1142–1152.
- 3 J. Kohlmeyer and E. Kohlmeyer, *Marine mycology: The higher fungi*, Academic Press, New York, 1979.
- 4 E. B. G. Jones, J. Sakayaroj, S. Suetrong, S. Somrithipol and K. L. Pang, *Fungal Diversity*, 2009, **35**, 1–187.
- 5 K. L. Pang, L. L. P. Vrijmoed, T. K. Goh, N. Plaingam and E. B. G. Jones, *Bot. Mar.*, 2008, **51**, 171–178.
- 6 S. Damare and C. Raghukumar, Microb. Ecol., 2008, 56, 168–177.
- 7 X. Lai, L. Cao, H. Tan, S. Fang, Y. Huang and S. Zhou, *ISME J.*, 2007, **1**, 756–762.
- 8 D. L. Hawksworth, Mycol. Res., 2001, 105, 1422-1432.
- 9 J. P. Schmit and G. M. Mueller, Biodiversity Conserv., 2007, 16, 99-111.
- 10 A. Zuccaro, C. L. Schoch, J. W. Spatafora, J. Kohlmeyer, S. Draeger and J. I. Mitchell, *Appl. Environ. Microbiol.*, 2008, 74, 931–941.
- 11 A. Zuccaro, B. Schulz and J. I. Mitchell, Mycol. Res., 2003, 107, 1451-1466.
- 12 A. Zuccaro, R. C. Summerbell, W. Gams, H. J. Schroers and J. I. Mitchell, *Stud. Mycol.*, 2004, 283–297.
- 13 L. Fieseler, M. Horn, M. Wagner and U. Hentschel, *Appl. Environ. Microbiol.*, 2004, **70**, 3724–3732.

- 14 U. Hentschel, J. Hopke, M. Horn, A. B. Friedrich, M. Wagner, J. Hacker and B. S. Moore, *Appl. Environ. Microbiol.*, 2002, 68, 4431–4440.
- 15 M. W. Taylor, R. Radax, D. Steger and M. Wagner, *Microbiol. Mol. Biol. Rev.*, 2007, **71**, 295–347.
- 16 P. Baker, J. Kennedy, A. Dobson and J. Marchesi, *Mar. Biotechnol.*, 2009, **11**, 540–547.
- 17 Q. Li and G. Wang, Microbiol. Res., 2009, 164, 233-241.
- 18 W. C. Liu, C. Q. Li, P. Zhu, J. L. Yang and K. D. Cheng, *Fungal Diversity*, 2010, 42, 1–15.
- 19 Z. Paz, M. Komon-Zelazowska, I. S. Druzhinina, M. M. Aveskamp, A. Shnaiderman, Y. Aluma, S. Carmeli, M. Ilan and O. Yarden, *Fungal Diversity*, 2010, **42**, 17–26.
- 20 M. Pivkin, S. Aleshko, V. Krasokhin and Y. Khudyakova, *Russ. J. Mar. Biol.*, 2006, **32**, 207–213.
- 21 G. Wang, Q. Li and P. Zhu, Antonie van Leeuwenhoek, 2008, 93, 163–174.
- 22 Z. Gao, B. Li, C. Zheng and G. Wang, *Appl. Environ. Microbiol.*, 2008, **74**, 6091–6101.
- 23 J. W. Blunt, B. R. Copp, M. H. G. Munro, P. T. Northcote and M. R. Prinsep, *Nat. Prod. Rep.*, 2010, **27**, 165–237.
- 24 S. Maiya, A. Grundmann, X. Li, S. M. Li and G. Turner, *ChemBioChem*, 2007, **8**, 1736–1743.
- 25 R. J. Cox, Org. Biomol. Chem., 2007, 5, 2010-2026.
- 26 H. Wei, T. Itoh, M. Kinoshita, N. Kotoku, S. Aoki and M. Kobayashi, *Tetrahedron*, 2005, **61**, 8054–8058.

- 28 G. Lang, J. Wiese, R. Schmaljohann and J. F. Imhoff, *Tetrahedron*, 2007, **63**, 11844–11849.
- 29 H. Jayasuriya, Z. Q. Guan, A. W. Dombrowski, G. F. Bills, J. D. Polishook, R. G. Jenkins, L. Koch, T. Crumley, T. Tamas, M. Dubois, A. Misura, S. J. Darkin-Rattray, L. Gregory and S. B. Singh, J. Nat. Prod., 2007, 70, 1364–1367.
- 30 F. Z. Wang, T. J. Zhu, M. Zhang, A. Q. Lin, W. M. Zhu and Q. Q. Gu, *Magn. Reson. Chem.*, 2010, 48, 155–158.
- 31 M. Varoglu and P. Crews, J. Nat. Prod., 2000, 63, 41-43.
- 32 D. H. Li, F. P. Wang, S. X. Cai, X. Zeng, X. Xiao, Q. Q. Gu and W. M. Zhu, *J. Antibiot.*, 2007, **60**, 317–320.
- 33 R. Andrade, W. A. Ayer and L. S. Trifonov, Can. J. Chem., 1996, 74, 371–379.
- 34 D. H. Li, F. P. Wang, X. Xiao, Y. C. Fang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, *Tetrahedron Lett.*, 2007, 48, 5235–5238.
- 35 D. H. Li, S. X. Cai, T. J. Zhu, F. P. Wang, X. A. Xiao and Q. Q. Gu, *Tetrahedron*, 2010, 66, 5101–5106.
- 36 L. Du, T. J. Zhu, L. Y. Li, S. X. Cai, B. Y. Zhao and Q. Q. Gu, *Chem. Pharm. Bull.*, 2009, 57, 220–223.
- 37 K. Neumann, A. Abdel-Lateff, A. D. Wright, S. Kehraus, A. Krick and G. M. König, *Eur. J. Org. Chem.*, 2007, 2268–2275.
- 38 J. Ueda, J. Hashimoto, S. Inaba, M. Takagi and K. Shin-ya, J. Antibiot., 2010, 63, 203–205.
- 39 A. Hirota, Y. Morimitsu and H. Hojo, *Biosci., Biotechnol., Biochem.*, 1997, **61**, 647–650.
- 40 X. Yang, T. T. Khong, L. Chen, H. D. Choi, J. S. Kang and B. W. Son, *Chem. Pharm. Bull.*, 2008, 56, 1355–1356.
- 41 W. H. Urry, H. L. Wehrmeister, E. B. Hodge and P. H. Hidy, *Tetrahedron Lett.*, 1966, 7, 3109–3114.
- 42 L. L. Zhao, Y. Gai, H. Kobayashi, C. Q. Hu and H. P. Zhang, *Chin. Chem. Lett.*, 2008, **19**, 1089–1092.
- 43 M. Isaka, A. Yangchum, S. Intamas, K. Kocharin, E. B. G. Jones, P. Kongsaeree and S. Prabpai, *Tetrahedron*, 2009, 65, 4396–4403.
- 44 M. Isaka, C. Suyarnsestakorn, M. Tanticharoen, P. Kongsaeree and Y. Thebtaranonth, *J. Org. Chem.*, 2002, **67**, 1561–1566.
- 45 P. Vongvilai, M. Isaka, P. Kittakoop, P. Srikitikulchai, P. Kongsaeree and Y. Thebtaranonth, J. Nat. Prod., 2004, 67, 457–460.
- 46 M. S. R. Nair and S. T. Carey, Tetrahedron Lett., 1980, 21, 2011-2012.
- 47 S. Barluenga, R. Jogireddy, G. K. Koripelly and N. Winssinger, ChemBioChem, 2010, 11, 1692–1699.
- 48 R.-y. Yang, C.-y. Li, Y.-c. Lin, G.-t. Peng, Z.-g. She and S.-n. Zhou, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 4205–4208.
- 49 H. Greve, P. J. Schupp, E. Eguereva, S. Kehraus, G. Kelter, A. Maier, H. H. Fiebig and G. M. König, *Eur. J. Org. Chem.*, 2008, 5085–5092.
- 50 K.-K. Li, Y.-J. Lu, X.-H. Song, Z.-G. She, X.-W. Wu, L.-K. An, C.-X. Ye and Y.-C. Lin, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 3326–3328.
- 51 J. Fuchser, S. Grabley, M. Noltemeyer, S. Philipps, R. Thiericke and A. Zeeck, *Liebigs Ann. Chem.*, 1994, 831–835.
- 52 J. Fuchser and A. Zeeck, Liebigs Ann./Recl., 1997, 87-95.
- 53 K. Kito, R. Ookura, S. Yoshida, M. Namikoshi, T. Ooi and T. Kusumi, J. Nat. Prod., 2007, 70, 2022–2025.
 54 M. Namikoshi, P. Namikoshi, H. Namikoshi, A. Davida, A. Dav
- 54 M. Namikoshi, R. Negishi, H. Nagai, A. Dmitrenok and H. Kobayashi, J. Antibiot., 2003, 56, 755–761.
- 55 K. Kito, R. Ookura, S. Yoshida, M. Namikoshi, T. Ooi and T. Kusumi, *Org. Lett.*, 2008, **10**, 225–228.
- 56 H. Greve, P. J. Schupp, E. Eguereva, S. Kehraus and G. M. König, J. Nat. Prod., 2008, 71, 1651–1653.
- 57 M. Tsuda, T. Mugishima, K. Komatsu, T. Sone, M. Tanaka, Y. Mikami and J. Kobayashi, *J. Nat. Prod.*, 2003, **66**, 412–415.
- 58 S. Grabley, E. Granzer, K. Hutter, D. Ludwig, M. Mayer, R. Thiericke, G. Till, J. Wink, S. Philips and A. Zeeck, J. Antibiot., 1992, 45, 56-65.
- 59 X. P. Du, C. H. Lu, Y. Y. Li, Z. G. Zheng, W. J. Su and Y. M. Shen, J. Antibiot., 2008, **61**, 250–253.
- 60 T. Yamada, M. Doi, H. Shigeta, Y. Muroga, S. Hosoe, A. Numata and R. Tanaka, *Tetrahedron Lett.*, 2008, **49**, 4192–4195.
- 61 M. Yasuhide, T. Yamada, A. Numata and R. Tanaka, *J. Antibiot.*, 2008, 61, 615–622.
 62 T. Varnada, M. Y. J. K. W. Stranger, M. Y. Stranger, M. S. Stranger, and S. S.
- 62 T. Yamada, M. Yasuhide, H. Shigeta, A. Numata and R. Tanaka, J. Antibiot., 2009, 62, 353–357.

- 63 T. Yamada, Y. Muroga and R. Tanaka, *Mar. Drugs*, 2009, **7**, 249–257.
- 64 Y. Muroga, T. Yamada, A. Numata and R. Tanaka, *Tetrahedron*, 2009, 65, 7580–7586.
 65 Y. Muran, T. Y. Kanaka, and K. Tanaka, *Tetrahedron*, 2009, 65, 7580–7586.

- 67 F. Almassi, E. L. Ghisalberti, M. J. Narbey and K. Sivasithamparam, J. Nat. Prod., 1991, **54**, 396–402.
- 68 E. L. Ghisalberti and C. Y. Rowland, J. Nat. Prod., 1993, 56, 1799– 1804.
- F. H. Song, H. Q. Dai, Y. J. Tong, B. A. Ren, C. X. Chen, N. Sun, X. Y. Liu, J. Bian, M. Liu, H. Gao, H. W. Liu, X. P. Chen and L. X. Zhang, *J. Nat. Prod.*, 2010, **73**, 806–810.
 R. W. Dunlop, A. Simon, K. Sivasithamparam and
- E. L. Ghisalberti, J. Nat. Prod., 1989, **52**, 67–74.
- 71 F. Hiramatsu, T. Miyajima, T. Murayama, K. Takahashi, T. Koseki and Y. Shiono, J. Antibiot., 2006, **59**, 704–709.
- 72 A. Evidente, L. Conti, C. Altomare, A. Bottalico, G. Sindona, A. L. Segre and A. Logrieco, *Nat. Toxins*, 1994, **2**, 4–13.
- 73 C. Osterhage, R. Kaminsky, G. M. König and A. D. Wright, J. Org. Chem., 2000, 65, 6412–6417.
- 74 S. F. Seibert, A. Krick, E. Eguereva, S. Kehraus and G. M. König, Org. Lett., 2007, 9, 239–242.
- 75 X. Li, Y. H. Yao, Y. A. Zheng, I. Sattler and W. H. Lin, *Arch. Pharmacal Res.*, 2007, **30**, 812–815.
- 76 X. Li, I. Sattler and W. H. Lin, J. Antibiot., 2007, 60, 191–195.
- 77 C. Kasettrathat, N. Ngamrojanavanich, S. Wiyakrutta, C. Mahidol,
 S. Ruchirawat and P. Kittakoop, *Phytochemistry*, 2008, **69**, 2621–2626.
- 78 T. Hosoe, K. Fukushima, K. Takizawa, T. Itabashi, K. Yoza and K. Kawai, *Heterocycles*, 2006, 68, 1949–1953.
- 79 R. F. Angawi, D. C. Swenson, J. B. Gloer and D. T. Wicklow, J. Nat. Prod., 2003, 66, 1259–1262.
- 80 H. P. Nguyen, D. H. Zhang, U. Lee, J. S. Kang, H. D. Choi and B. W. Son, J. Nat. Prod., 2007, 70, 1188–1190.
- 81 H. Kobayashi, R. Sunaga, K. Furihata, N. Morisaki and S. Iwasaki, J. Antibiot., 1995, **48**, 42–52.
- 82 Y. Gai, L. L. Zhao, C. Q. Hu and H. P. Zhang, *Chin. Chem. Lett.*, 2007, **18**, 954–956.
- 83 K. Trisuwan, V. Rukachaisirikul, Y. Sukpondma, S. Preedanon, S. Phongpaichit and J. Sakayaroj, *Phytochemistry*, 2009, **70**, 554– 557.
- 84 L. Du, T. J. Zhu, Y. C. Fang, H. B. Liu, Q. Q. Gu and W. M. Zhu, *Tetrahedron*, 2007, **63**, 1085–1088.
- 85 K. Tao, L. Du, X. Sun, M. Cai, T. Zhu, X. Zhou, Q. Gu and Y. Zhang, *Tetrahedron Lett.*, 2009, **50**, 1082–1085.
- 86 L. Du, T. J. Zhu, H. B. Liu, Y. C. Fang, W. M. Zhu and Q. Q. Gu, *J. Nat. Prod.*, 2008, **71**, 1837–1842.
- 87 J. F. Grove, J. Chem. Soc., Perkin Trans. 1, 1972, 2406-2411.
- 88 W. Wang, T. Zhu, H. Tao, Z. Lu, Y. Fang, Q. Gu and W. Zhu, J. Antibiot., 2007, 60, 603–607.
- 89 A. Krick, S. Kehraus, C. Gerhäuser, K. Klimo, M. Nieger, A. Maier, H. H. Fiebig, I. Atodiresei, G. Raabe, J. Fleischhauer and G. M. König, J. Nat. Prod., 2007, 70, 353–360.
- 90 A. Pontius, A. Krick, R. Mesry, S. Kehraus, S. E. Foegen, M. Müller, K. Klimo, C. Gerhäuser and G. M. König, J. Nat. Prod., 2008, 71, 1793–1799.
- 91 J. X. Yang, F. Xu, C. H. Huang, J. Li, Z. G. She, Z. Pei and Y. C. Lin, *Eur. J. Org. Chem.*, 2010, 3692–3695.
- 92 S. Lösgen, O. Schlörke, K. Meindl, R. Herbst-Irmer and A. Zeeck, *Eur. J. Org. Chem.*, 2007, 2191–2196.
- 93 F. M. Dean, R. A. Eade, R. A. Moubasher and A. Robertson, *Nature*, 1957, **179**, 366–366.
- 94 M. Aoki, Y. Itezono, H. Shirai, N. Nakayama, A. Sakai, Y. Tanaka, A. Yamaguchi, N. Shimma, K. Yokose and H. Seto, *Tetrahedron Lett.*, 1991, **32**, 4737–4740.
- 95 Y. Zhang, X.-M. Li and B.-G. Wang, J. Antibiot., 2007, 60, 204–210.
- 96 Y. P. Zhang, S. Ling, Y. C. Fang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, *Chem. Biodiversity*, 2008, 5, 93–100.
- 97 Y. Ohkawa, K. Miki, T. Suzuki, K. Nishio, T. Sugita, K. Kinoshita, K. Takahashi and K. Koyama, J. Nat. Prod., 2010, 73, 579–582.
- 98 Z. J. Huang, R. Y. Yang, Z. Y. Guo, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2010, **46**, 15–18.

- 99 K. Kanoh, A. Okada, K. Adachi, H. Imagawa, M. Nishizawa, S. Matsuda, Y. Shizuri and R. Utsumi, J. Antibiot., 2008, 61, 142– 148.
- 100 K. Krohn, A. Michel, U. Florke, H. J. Aust, S. Draeger and B. Schulz, *Liebigs Ann. Chem.*, 1994, 1093–1097.
- 101 K. Krohn, A. Michel, U. Florke, H. J. Aust, S. Draeger and B. Schulz, *Liebigs Ann. Chem.*, 1994, 1099–1108.
- 102 S. S. Gao, X. M. Li and B. G. Wang, Nat. Prod. Commun., 2009, 4, 1477–1480.
- 103 M. E. Stack, E. P. Mazzola, S. W. Page, A. E. Pohland, R. J. Highet, M. S. Tempesta and D. G. Corley, *J. Nat. Prod.*, 1986, **49**, 866–871.
- 104 A. Arnone, G. Nasini, L. Merlini and G. Assante, J. Chem. Soc., Perkin Trans. 1, 1986, 525–530.
- 105 J. Kjer, V. Wray, R. Edrada-Ebel, R. Ebel, A. Pretsch, W. Lin and P. Proksch, J. Nat. Prod., 2009, 72, 2053–2057.
- 106 T. K. Mitchell, F. Alejos-Gonzalez, H. S. Gracz, D. A. Danehower, M. E. Daub and W. S. Chilton, *Phytochemistry*, 2003, 62, 723–732.
- 107 T. K. Mitchell, W. S. Chilton and M. E. Daub, Appl. Environ. Microbiol., 2002, 68, 4173–4181.
- 108 L. Wen, X. L. Cai, F. Xu, Z. G. She, W. L. Chan, L. L. P. Vrijmoed, E. B. G. Jones and Y. C. Lin, *J. Org. Chem.*, 2009, **74**, 1093–1098.
- 109 A. Pontius, A. Krick, S. Kehraus, R. Brun and G. M. König, J. Nat. Prod., 2008, 71, 1579–1584.
- 110 Z. J. Wu, M. A. Ouyang and Q. W. Tan, Pest Manage. Sci., 2009, 65, 60–65.
- 111 C. L. Shao, C. Y. Wang, M. Y. Wei, Y. C. Gu, X. K. Xia, Z. G. She and Y. C. Lin, *Magn. Reson. Chem.*, 2008, 46, 1066–1069.
- 112 A. M. D. Marinho, E. Rodrigues, M. D. R. Moitinho and L. S. Santos, J. Braz. Chem. Soc., 2005, 16, 280–283.
- 113 Z. Y. Guo, F. Cheng, K. Zou, J. Z. Wang, Z. G. She and Y. C. Lin, *Nat. Prod. Commun.*, 2009, 4, 1481–1483.
- 114 X. Li, Y. N. Zheng, I. Sattler and W. H. Lin, Arch. Pharmacal Res., 2006, 29, 942–945.
 115 D. L. Li, X. M. Lingel D. C. W. L. K. K. Li, Phys. Rev. Lett. 2000.
- 115 D. L. Li, X. M. Li and B. G. Wang, J. Microbiol. Biotechnol., 2009, 19, 675–680.
- 116 Y. Li, X. Li, U. Lee, J. S. Kang, H. D. Choi and B. W. Son, *Chem. Pharm. Bull.*, 2006, 54, 882–883.
- 117 A. A. El-Beih, T. Kawabata, K. Koimaru, T. Ohta and S. Tsukamoto, *Chem. Pharm. Bull.*, 2007, **55**, 1097–1098.
- 118 C. L. Shao, C. Y. Wang, C. J. Zheng, Z. G. She, Y. C. Gu and Y. C. Lin, *Nat. Prod. Res.*, 2010, 24, 81–85.
- 119 C. L. Shao, C. Y. Wang, M. Y. Wei, S. D. Li, Z. G. She, Y. C. Gu and Y. C. Lin, *Magn. Reson. Chem.*, 2008, 46, 886–889.
- 120 J. Z. Xu, T. Nakazawa, K. Ukai, H. Kobayashi, R. E. P. Mangindaan, D. S. Wewengkang, H. Rotinsulu and M. Namikoshi, J. Antibiot., 2008, 61, 415–419.
- 121 X. K. Xia, H. R. Huang, Z. G. She, C. L. Shao, F. Liu, X. L. Cai, L. L. P. Vrijmoed and Y. C. Lin, *Magn. Reson. Chem.*, 2007, 45, 1006–1009.
- 122 Y. G. Chen, X. L. Cai, J. H. Pan, J. P. Gao, J. Li, J. Yuan, L. W. Fu, Z. G. She and Y. C. Lin, *Magn. Reson. Chem.*, 2009, **47**, 362–365.
- 123 H. B. Huang, Q. Li, X. J. Feng, B. Chen, J. Wang, L. Liu, Z. G. She and Y. C. Lin, *Magn. Reson. Chem.*, 2010, 48, 496–499.
- 124 J. H. Pan, J. J. Deng, Y. G. Chen, J. P. Gao, Y. C. Lin, Z. G. She and Y. C. Gu, *Helv. Chim. Acta*, 2010, **93**, 1369–1374.
- 125 F. H. Stodola, R. F. Vesonder, D. I. Fennell and D. Weisleder, *Phytochemistry*, 1972, 11, 2107–2108.
- 126 J. Sierankiewicz and S. Gatenbeck, Acta Chem. Scand., 1972, 26, 455–458.
- 127 C. A. Motti, D. G. Bourne, J. N. Burnell, J. R. Doyle, D. S. Haines, C. H. Liptrot, L. E. Llewellyn, S. Ludke, A. Muirhead and D. M. Tapiolas, *Appl. Environ. Microbiol.*, 2007, **73**, 1921–1927.
- 128 C. Y. Li, R. Y. Yang, Y. C. Lin and S. N. Zhou, *Chem. Nat. Compd.*, 2006, 42, 290–293.
- 129 C. Y. Li, R. Y. Yang, Y. C. Lin, Z. G. She and S. N. Zhou, J. Asian Nat. Prod. Res., 2007, 9, 285–291.
- 130 N. Tan, Y. W. Tao, J. H. Pan, S. Y. Wang, F. Xu, Z. G. She, Y. C. Lin and E. B. G. Jones, *Chem. Nat. Compd.*, 2008, 44, 296–300.
- 131 X. K. Xia, H. R. Huang, Z. G. She, J. W. Cai, L. Lan, J. Y. Zhang, L. W. Fu, L. L. P. Vrijmoed and Y. C. Lin, *Helv. Chim. Acta*, 2007, 90, 1925–1931.
- 132 J. Fuska, D. Uhrin, B. Proksa, Z. Voticky and J. Ruppeldt, J. Antibiot., 1986, 39, 1605–1608.
- 133 X. K. Xia, F. Liu, Z. G. She, L. G. Yang, M. F. Li, L. L. P. Vrijmoed and Y. C. Lin, *Magn. Reson. Chem.*, 2008, 46, 693–696.

- 134 V. Prachyawarakorn, C. Mahidol, S. Sureram, S. Sangpetsiripan, S. Wiyakrutta, S. Ruchirawat and P. Kittakoop, *Planta Med.*, 2008, 74, 69–72.
- 135 M. Makino, T. Endoh, Y. Ogawa, K. Watanabe and Y. Fujimoto, *Heterocycles*, 1998, 48, 1931–1934.
- 136 T. Ohzeki and K. Mori, Biosci., Biotechnol., Biochem., 2003, 67, 2240-2244.
- 137 X. Liu, F. Xu, Y. Zhang, L. Liu, H. Huang, X. Cai, Y. Lin and W. Chan, *Russ. Chem. Bull.*, 2006, 55, 1091–1092.
- 138 W. Q. Yin, Y. C. Lin, Z. G. She, L. L. P. Vrijmoed and E. B. G. Jones, *Chem. Nat. Compd.*, 2008, 44, 3–5.
- 139 G. S. Cockerill, P. C. Levett and D. A. Whiting, J. Chem. Soc., Perkin Trans. 1, 1995, 1103–1113.
- 140 F. Xu, Y. Zhang, J. J. Wang, J. Y. Pang, C. H. Huang, X. Y. Wu, Z. G. She, L. L. P. Vrijmoed, E. B. G. Jones and Y. H. Lin, *J. Nat. Prod.*, 2008, **71**, 1251–1253.
- 141 K. Trisuwan, V. Rukachaisirikul, Y. Sukpondma, S. Phongpaichit, S. Preedanon and J. Sakayaroj, *Chem. Pharm. Bull.*, 2009, 57, 1100–1102.
- 142 S. F. Seibert, E. Eguereva, A. Krick, S. Kehraus, E. Voloshina, G. Raabe, J. Fleischhauer, E. Leistner, M. Wiese, H. Prinz, K. Alexandrov, P. Janning, H. Waldmann and G. M. König, *Org. Biomol. Chem.*, 2006, 4, 2233–2240.
- 143 C. Kusnick, R. Jansen, K. Liberra and U. Lindequist, *Pharmazie*, 2002, 57, 510–512.
- 144 L. Colombo, C. Gennari, G. S. Ricca, C. Scolastico and F. Aragozzini, J. Chem. Soc., Perkin Trans. 1, 1980, 675–676.
- 145 S. F. Seibert, G. M. König, E. Voloshina, G. Rabbe and J. Fleischhauer, *Chirality*, 2006, 18, 413–418.
- 146 L. Wen, Z. Y. Guo, Q. Li, D. Z. Zhang, Z. G. She and L. L. P. Vrijmoed, *Chem. Nat. Compd.*, 2010, 46, 363–365.
- 147 M. Naganuma, M. Nishida, K. Kuramochi, F. Sugawara, H. Yoshida and Y. Mizushina, *Bioorg. Med. Chem.*, 2008, 16, 2939–2944.
- 148 Y. Kimura, T. Yoshinari, H. Koshino, S. Fujioka, K. Okada and A. Shimada, *Biosci., Biotechnol., Biochem.*, 2007, 71, 1896–1901.
- 149 W. A. Ayer and J. S. Racok, Can. J. Chem., 1990, 68, 2085-2094.
- 150 Z. H. Xin, T. Li, T. J. Zhu, W. L. Wang, L. Du, Y. C. Fang, Q. Q. Gu and W. M. Zhu, Arch. Pharmacal Res., 2007, 30, 816–819.
- 151 M. M. Chien, P. L. Schiff, D. J. Slatkin and J. E. Knapp, *Lloydia*, 1977, **40**, 301–302.
- 152 Z. Han, W. L. Mei, Y. X. Zhao, Y. Y. Deng and H. F. Dai, *Chem. Nat. Compd.*, 2009, **45**, 805–807.
- 153 Z. J. Huang, C. L. Shao, Y. G. Chen, Z. G. She, Y. C. Lin and S. N. Zhou, *Chem. Nat. Compd.*, 2007, 43, 655–658.
- 154 A. A. El-Beih, H. Kato, T. Ohta and S. Tsukamoto, *Chem. Pharm. Bull.*, 2007, 55, 953–954.
- 155 A. Pontius, I. Mohamed, A. Krick, S. Kehraus and G. M. König, J. Nat. Prod., 2008, 71, 272–274.
- 156 G. Y. Chen, Y. C. Lin, L. L. P. Vrijmoed and W. F. Fong, *Chem. Nat. Compd.*, 2006, **42**, 138–141.
- 157 F. Liu, X.-L. Cai, H. Yang, X.-K. Xia, Z.-Y. Guo, J. Yuan, M.-F. Li, Z.-G. She and Y.-C. Lin, *Planta Med.*, 2010, **76**, 185.
- 158 D. H. Zhang, X. G. Li, J. S. Kang, H. D. Choi, J. H. Jung and B. W. Son, *J. Microbiol. Biotechnol.*, 2007, **17**, 865–867.
- 159 R. F. Curtis, C. H. Hassall and M. Nazar, J. Chem. Soc., Perkin Trans. 1, 1968, 1, 85–93.
- 160 R. F. Curtis, C. H. Hassall and M. Nazar, Chem. Ind., 1966, 17, 702.
- 161 J. Barber, R. H. Carter, M. J. Garson and J. Staunton, J. Chem. Soc., Perkin Trans. 1, 1981, 2577–2583.
- 162 B. R. Clark, R. J. Capon, E. Lacey, S. Tennant and J. H. Gill, Org. Biomol. Chem., 2006, 4, 1520–1528.
- 163 K. Kanoh, K. Adachi, S. Matsuda, Y. Shizuri, K. Yasumoto, T. Kusumi, K. Okumura and T. Kirikae, J. Antibiot., 2008, 61, 192–194.
- 164 M. Elbandy, P. B. Shinde, H. T. Dang, J. Hong, K. S. Bae and J. H. Jung, J. Nat. Prod., 2008, 71, 869–872.
- 165 M. A. Abdel-Wahab, R. N. Asolkar, P. Inderbitzin and W. Fenical, *Phytochemistry*, 2007, 68, 1212–1218.
- 166 D. M. Roll, M. Tischler, R. T. Williamson and G. T. Carter, J. Antibiot., 2002, 55, 520–523.
- 167 W. A. Ayer and P. A. Craw, Can. J. Chem., 1992, 70, 1348-1355.
- 168 S. Y. Wang, Z. L. Xu, Z. G. She, H. Wang, C. R. Li and Y. C. Lin, J. Asian Nat. Prod. Res., 2008, 10, 622–626.

- 169 J. H. Sohn and H. Oh, *Bull. Korean Chem. Soc.*, 2010, **31**, 1695–1698. 170 X. P. Fang, J. E. Anderson, C. J. Chang, P. E. Fanwick and
- J. L. McLaughlin, J. Chem. Soc., Perkin Trans. 1, 1990, 1655–1661.
 171 Y. Takeda, Y. Okada, T. Masuda, E. Hirata, T. Shinzato, A. Takushi, Q. Yu and H. Otsuka, Chem. Pharm. Bull., 2000, 48, 752–754
- 172 G. Hao, Z. Qing-Hua, J. Miao-Miao, T. Jin-Shan, M. Cheng-Du, H. Kui, N. Michio, W. Nai-Li and Y. Xin-Sheng, *Magn. Reson. Chem.*, 2008, 46, 1148–1152.
- 173 M. A. M. Shushni, R. Mentel, U. Lindequist and R. Jansen, *Chem. Biodiversity*, 2009, **6**, 127–137.
- 174 Y. Kimura, T. Mizuno, H. Nakajima and T. Hamasaki, Biosci., Biotechnol., Biochem., 1992, 56, 1664–1665.
- 175 T. Yamada, M. Iritani, K. Minoura, K. Kawai and A. Numata, Org. Biomol. Chem., 2004, 2, 2131–2135.
- 176 T. Yamada, M. Doi, A. Miura, W. Harada, M. Hiramura, K. Minoura, R. Tanaka and A. Numata, J. Antibiot., 2005, 58, 185–191.
- 177 T. Yamada, M. Iritani, H. Ohishi, K. Tanaka, K. Minoura, M. Doi and A. Numata, Org. Biomol. Chem., 2007, 5, 3979–3986.
- 178 A. Numata, M. Iritani, T. Yamada, K. Minoura, E. Matsumura, T. Yamori and T. Tsuruo, *Tetrahedron Lett.*, 1997, 38, 8215– 8218.
- 179 Y. Usami, Y. Horibe, I. Takaoka, H. Ichikawa and M. Arimoto, Synlett, 2006, 1598–1600.
- 180 Y. Usami, K. Mizuki, H. Ichikawa and M. Arimoto, *Tetrahedron: Asymmetry*, 2008, 19, 1461–1464.
- 181 M. F. Elsebai, S. Kehraus, M. Gütschow and G. M. König, Nat. Prod. Commun., 2009, 4, 1463–1468.
- 182 V. Rukachaisirikul, N. Khamthong, Y. Sukpondma, S. Phongpaichit, N. Hutadilok-Towatana, P. Graidist, J. Sakayaroj and K. Kirtikara, *Arch. Pharmacal Res.*, 2010, 33, 375–380.
- 183 D. Zhang, X. Yang, J. S. Kang, H. D. Choi and B. W. Son, J. Nat. Prod., 2008, 71, 1458–1460.
- 184 K. Trisuwan, V. Rukachaisirikul, Y. Sukpondma, S. Preedanon, S. Phongpaichit, N. Rungjindamai and J. Sakayaroj, *J. Nat. Prod.*, 2008, **71**, 1323–1326.
- 185 A. Closse, R. Mauli and H. P. Sigg, Helv. Chim. Acta, 1966, 49, 204– 213.
- 186 Z. J. Lin, T. J. Zhu, Y. C. Fang, Q. Q. Gu and W. M. Zhu, *Phytochemistry*, 2008, **69**, 1273–1278.
- 187 A. Q. Lin, X. M. Lu, Y. C. Fang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, J. Antibiot., 2008, 61, 245–249.
- 188 L. Chen, Y. C. Fang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, J. Nat. Prod., 2008, 71, 66–70.
- 189 V. N. Nenkep, K. Yun, Y. Li, H. D. Choi, J. S. Kang and B. W. Son, J. Antibiot., 2010, 63, 199–201.
- 190 R. Liu, W. M. Zhu, Y. P. Zhang, T. J. Zhu, H. B. Liu, Y. C. Fang and Q. Q. Gu, J. Antibiot., 2006, 59, 362–365.
- 191 Y. W. Tao, C. B. Mou, M. A. Zeng, F. Xu, A. Cai, Z. G. She, S. N. Zhou and Y. C. Lin, *Magn. Reson. Chem.*, 2008, 46, 761–764.
 192 G. Lang, A. L. J. Cole, J. W. Blunt, W. T. Robinson and
- 192 G. Lang, A. L. J. Cole, J. W. Blunt, W. 1. Robinson and M. H. G. Munro, J. Nat. Prod., 2007, 70, 310–311.
- 193 C. Y. Li, W. J. Ding, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2008, 44, 163–165.
- 194 H. J. Yan, S. S. Gao, C. S. Li, X. M. Li and B. G. Wang, *Molecules*, 2010, 15, 3270–3275.
- 195 Z. J. Huang, Z. Y. Guo, R. Y. Yang, X. H. Yin, X. Y. Li, W. Q. Luo, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2009, **45**, 625–628.
- 196 Q. Y. Xu, J. F. Wang, Y. J. Huang, Z. H. Zheng, S. Y. Song, Y. M. Zhang and W. J. Su, *Acta Oceanol. Sin.*, 2004, 23, 541–547.
 197 J. Xu, J. Kjer, J. Sendker, V. Wray, H. S. Guan, R. Edrada,
- W. H. Lin, J. Wu and P. Proksch, J. Nat. Prod., 2009, 72, 662–665.
- 198 J. Xu, J. Kjer, J. Sendker, V. Wray, H. S. Guan, R. Edrada, W. E. G. Muller, M. Bayer, W. H. Lin, J. Wu and P. Proksch, *Bioorg. Med. Chem.*, 2009, **17**, 7362–7367.
- 199 S. F. Brady, M. M. Wagenaar, M. P. Singh, J. E. Janso and J. Clardy, Org. Lett., 2000, 2, 4043–4046.
- 200 A. M. Gamal-Eldeen, A. Abdel-Lateff and T. Okino, *Environ. Toxicol. Pharmacol.*, 2009, 28, 317–322.
- 201 D. H. Zhang, X. G. Li, J. S. Kang, H. D. Choi and B. W. Son, Bull. Korean Chem. Soc., 2007, 28, 887–888.
- 202 J. Thibonnet, M. Abarbri, J.-L. Parrain and A. Duchêne, J. Org. Chem., 2002, 67, 3941–3944.

- 203 P. Proksch, R. Ebel, R. Edrada, F. Riebe, H. Liu, A. Diesel, M. Bayer, X. Li, W. H. Lin, V. Grebenyuk, W. E. G. Müller, S. Draeger, A. Zuccaro and B. Schulz, *Bot. Mar.*, 2008, **51**, 209– 218.
- 204 T. O. Larsen, N. B. Perry and B. Andersen, *Tetrahedron Lett.*, 2003, 44, 4511–4513.
- 205 A. Abdel-Lateff, K. Fisch and A. D. Wright, Z. Naturforsch. C., 2009, 64, 186–192.
- 206 A. Schüffler, O. Sterner and H. Anke, Z. Naturforsch. C., 2007, 62, 169–172.
- 207 M. Elbandy, P. B. Shinde, J. Hong, K. S. Bae, M. A. Kim, S. M. Lee and J. H. Jung, *Bull. Korean Chem. Soc.*, 2009, **30**, 188–192.
- 208 H. H. Wu, L. Tian, G. Chen, N. Xu, Y. N. Wang, S. Sun and Y. H. Pei, J. Asian Nat. Prod. Res., 2009, 11, 748–751.
- 209 H. H. Wu, L. Tian, B. M. Feng, Z. F. Li, Q. H. Zhang and Y. H. Pei, J. Asian Nat. Prod. Res., 2010, 12, 15–19.
- 210 P. M. Scott, B. Kennedy and W. Vanwalbe, *Experientia*, 1972, 28, 1252–1252.
- 211 S. Sun, L. Tian, Z. H. Wu, G. Chen, H. H. Wu, Y. N. Wang and Y. H. Pei, J. Asian Nat. Prod. Res., 2009, 11, 898–903.
- 212 C. L. Shao, C. Y. Wang, M. Y. Wei, Z. B. Jia, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2009, **45**, 779–781.
- 213 C. Seo, J. H. Sohn, H. Oh, B. Y. Kim and J. S. Ahn, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 6095–6097.
- 214 K. Hamano, M. Kinoshitaokami, K. Minagawa, H. Haruyama, T. Kinoshita, T. Hosoya, K. Furuya, K. Kinoshita, K. Tabata, A. Hemmi and K. Tanzawa, J. Antibiot., 1993, 46, 1648–1657.
- 215 J. L. You, H. Q. Dai, Z. H. Chen, G. J. Liu, Z. X. He, F. H. Song, X. Yang, H. A. Fu, L. X. Zhang and X. P. Chen, *J. Ind. Microbiol. Biotechnol.*, 2010, **37**, 245–252.
- 216 X. F. Li, D. H. Zhang, U. Lee, X. G. Li, J. G. Cheng, W. L. Zhu, J. H. Jung, H. D. Choi and B. W. Son, *J. Nat. Prod.*, 2007, **70**, 307–309.
- 217 Z. Feng, A. S. Leutou, G. Yang, V. N. Nenkep, X. N. Siwe, H. D. Choi, J. S. Kang and B. W. Son, *Bull. Korean Chem. Soc.*, 2009, **30**, 2345–2350.
- 218 J. Z. Xu, A. Takasaki, H. Kobayashi, T. Oda, J. Yamada, R. E. P. Mangindaan, K. Ukai, H. Nagai and M. Namikoshi, J. Antibiot., 2006, 59, 451–455.
- 219 M. Cueto, J. B. MacMillan, P. R. Jensen and W. Fenical, *Phytochemistry*, 2006, 67, 1826–1831.
- 220 M. P. Lopez-Gresa, N. Cabedo, M. C. Gonzalez-Mas, M. L. Ciavatta, C. Avila and J. Primo, J. Nat. Prod., 2009, 72, 1348–1351.
- 221 J. P. Springer, J. W. Dorner, R. J. Cole and R. H. Cox, J. Org. Chem., 1979, 44, 4852–4854.
- 222 G. Y. Li, B. G. Li, T. Yang, J. H. Yin, H. Y. Qi, G. Y. Liu and G. L. Zhang, J. Nat. Prod., 2005, 68, 1243–1246.
- 223 Y. Li, D. Z. Ye, X. L. Chen, X. H. Lu, Z. Z. Shao, H. Zhang and Y. S. Che, *J. Nat. Prod.*, 2009, **72**, 912–916.
 224 F. A. Macias, R. M. Varela, A. M. Simonet, H. G. Cutler,
- 224 F. A. Macias, R. M. Varela, A. M. Simonet, H. G. Cutler, S. J. Cutler, F. M. Dugan and R. A. Hill, *J. Org. Chem.*, 2000, 65, 9039–9046.
- 225 F. A. Macias, R. M. Varela, A. M. Simonet, H. G. Cutler, S. J. Cutler, S. A. Ross, D. C. Dunbar, F. M. Dugan and R. A. Hill, *Tetrahedron Lett.*, 2000, **41**, 2683–2686.
- 226 I. E. Mohamed, H. Gross, A. Pontius, S. Kehraus, A. Krick, G. Kelter, A. Maier, H.-H. Fiebig and G. M. König, *Org. Lett.*, 2009, **11**, 5014–5017.
- 227 T. S. Bugni, D. Abbanat, V. S. Bernan, W. M. Maiese, M. Greenstein, R. M. Van Wagoner and C. M. Ireland, *J. Org. Chem.*, 2000, **65**, 7195–7200.
- 228 A. Kralj, S. Kehraus, A. Krick, E. Eguereva, G. Kelter, M. Maurer, A. Wortmann, H. H. Fiebig and G. M. König, *J. Nat. Prod.*, 2006, 69, 995–1000.
- 229 L. Wen, Y. C. Lin, Z. G. She, D. S. Du, W. L. Chan and Z. H. Zheng, J. Asian Nat. Prod. Res., 2008, 10, 133–137.
- 230 Z. J. Huang, R. Y. Yang, X. H. Yin, Z. G. She and Y. C. Lin, *Magn. Reson. Chem.*, 2010, 48, 80–82.
- 231 Z. J. Huang, R. Y. Yang, Z. Y. Guo, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2010, 46, 348–351.
- 232 S. Wang, X. M. Li, F. Teuscher, D. L. Li, A. Diesel, R. Ebel, P. Proksch and B. G. Wang, J. Nat. Prod., 2006, 69, 1622– 1625.

- 233 M. L. Ciavatta, M. Pilar Lopez-Gresa, M. Gavagnin, R. Nicoletti, E. Manzo, E. Mollo, Y.-W. Guo and G. Cimino, *Tetrahedron*, 2008, **64**, 5365–5369.
- 234 S. Stevens-Miles, M. A. Goetz, G. F. Bills, R. A. Giacobbe, J. S. Tkacz, R. S. L. Chang, M. Mojena, I. Martin, M. T. Diez, F. Pelaez, O. D. Hensens, T. Jones, R. W. Burg, Y. L. Kong and L. Y. Huang, J. Antibiot., 1996, 49, 119–123.
- 235 J.-H. Jang, K. Kanoh, K. Adachi and Y. Shizuri, J. Antibiot., 2006, 59, 428–431.
- 236 Z. Y. Lu, Y. Wang, C. D. Miao, P. P. Liu, K. Hong and W. M. Zhu, J. Nat. Prod., 2009, 72, 1761–1767.
- 237 H. G. Cutler, F. G. Crumley, J. P. Springer, R. H. Cox, R. J. Cole, J. W. Dorner and J. E. Thean, J. Agric. Food Chem., 1980, 28, 989– 991.
- 238 M. F. Elsebai, S. Kehraus, M. Gutschow and G. M. Konig, *Nat. Prod. Commun.*, 2010, 5, 1071–1076.
- 239 D. R. Sanson, H. Gracz, M. S. Tempesta, D. S. Fukuda, W. M. Nakatsukasa, T. H. Sands, P. J. Baker and J. S. Mynderse, *Tetrahedron*, 1991, 47, 3633–3644.
- 240 M. Izumikawa, S. T. Khan, H. Komaki, A. Nagai, S. Inaba, M. Takagi and K. Shin-Ya, *Biosci., Biotechnol., Biochem.*, 2009, 73, 2138–2140.
- 241 F. Liu, Q. Li, H. Yang, X. L. Cai, X. K. Xia, S. P. Chen, M. F. Li, Z. G. She and Y. C. Lin, *Magn. Reson. Chem.*, 2009, **47**, 453–455.
- 242 H. Oh, T. O. Kwon, J. B. Gloer, L. Marvanová and C. A. Shearer, J. Nat. Prod., 1999, 62, 580–583.
- 243 C. L. Shao, Z. Y. Guo, H. Peng, G. T. Peng, Z. J. Huang, Z. G. She, Y. C. Lin and S. N. Zhou, *Chem. Nat. Compd.*, 2007, 43, 377–379.
- 244 M. Y. Wei, X. L. Zhang, S. D. Li, C. L. Shao, C. Y. Wang, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2010, 46, 340–342.
- 245 P. Zhang, B. Bao, H. T. Dang, J. Hong, H. J. Lee, E. S. Yoo, K. S. Bae and J. H. Jung, J. Nat. Prod., 2009, 72, 270–275.
- 246 S. B. Singh, R. G. Ball, G. F. Bills, C. Cascales, J. B. Gibbs, M. A. Goetz, K. Hoogsteen, R. G. Jenkins, J. M. Liesch, R. B. Lingham, K. C. Silverman and D. L. Zink, J. Org. Chem., 1996, 61, 7727–7737.
- 247 L. Y. Li, I. Sattler, Z. W. Deng, I. Groth, G. Walther, K. D. Menzel, G. Peschel, S. Grabley and W. H. Lin, *Phytochemistry*, 2008, **69**, 511–517.
- 248 A. I. Laskin, P. Grabowich, C. De Lisle Meyers and J. Fried, J. Med. Chem., 1964, 7, 406–409.
- 249 N. Shirane, Y. Hashimoto, K. Ueda, H. Takenaka and K. Katoh, *Phytochemistry*, 1996, 43, 99–104.
- 250 M. Zhang, W. L. Wang, Y. C. Fang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, J. Nat. Prod., 2008, 71, 985–989.
- 251 H. Y. Chen, C. W. Lin, G. Y. Chen and G. C. Ou, Acta Crystallogr., Sect. E: Struct. Rep. Online, 2008, 64, O890–U2367.
- 252 T. Amagata, M. Tanaka, T. Yamada, M. Doi, K. Minoura, H. Ohishi, T. Yamori and A. Numata, J. Nat. Prod., 2007, 70, 1731–1740.
- 253 T. Amagata, M. Doi, M. Tohgo, K. Minoura and A. Numata, Chem. Commun., 1999, 1321–1322.
- 254 L. Canonica, B. Danieli, G. Lesma, G. Palmisano and A. Mugnoli, *Helv. Chim. Acta*, 1987, 70, 701–716.
- 255 F. Wang, Y. Fang, M. Zhang, A. Lin, A. Zhu, Q. Gu and W. Zhu, *Steroids*, 2008, **73**, 19–26.
- 256 H. Gao, K. Hong, X. Zhang, H. W. Liu, N. L. Wang, L. Zhuang and X. S. Yao, *Helv. Chim. Acta*, 2007, **90**, 1165–1178.
- 257 H. Gao, K. Hong, G. D. Chen, C. X. Wang, J. S. Tang, Y. Yu, M. M. Jiang, M. M. Li, N. L. Wang and X. S. Yao, *Magn. Reson. Chem.*, 2010, **48**, 38–43.
- 258 Y. Sun, L. Tian, J. Huang, W. Li and Y. H. Pei, Nat. Prod. Res., 2006, 20, 381–384.
- 259 M. Ishino, N. Kiyomichi, K. Takatori, T. Sugita, M. Shiro, K. Kinoshita, K. Takahashi and K. Koyama, *Tetrahedron*, 2010, 66, 2594–2597.
- 260 W. P. D. Goldring and G. Pattenden, Acc. Chem. Res., 2006, 39, 354–361.
- 261 M. Sugano, A. Sato, Y. Iijima, T. Oshima, K. Furuya, H. Kuwano, T. Hata and H. Hanzawa, J. Am. Chem. Soc., 1991, 113, 5463–5464.
- 262 M. Sugano, A. Sato, Y. Iijima, K. Furuya, H. Haruyama, K. Yoda and T. Hata, *J. Org. Chem.*, 1994, **59**, 564–569.
- 263 S. S. Afiyatullov, A. I. Kalinovsky, M. V. Pivkin, P. S. Dmitrenok and T. A. Kuznetsova, *Nat. Prod. Res.*, 2006, **20**, 902–908.

- 264 S. Yoshida, K. Kito, T. Ooi, K. Kanoh, Y. Shizuri and T. Kusumi, *Chem. Lett.*, 2007, 36, 1386–1387.
- 265 S. Dettrakul, P. Kittakoop, M. Isaka, S. Nopichai, C. Suyarnsestakorn, M. Tanticharoen and Y. Thebtaranonth, *Bioorg. Med. Chem. Lett.*, 2003, 13, 1253–1255.
 266 W. Pongcharoen, V. Rukachaisirikul, S. Phongpaichit,
- 266 W. Pongcharoen, V. Rukachaisirikul, S. Phongpaichit, N. Rungjindamai and J. Sakayaroj, J. Nat. Prod., 2006, 69, 856–858.
- 267 L. Du, D. H. Li, T. J. Zhu, S. X. Cai, F. P. Wang, X. Xiao and Q. Q. Gu, *Tetrahedron*, 2009, **65**, 1033–1039.
- 268 T. Roncal, S. Cordobes, U. Ugalde, Y. H. He and O. Sterner, *Tetrahedron Lett.*, 2002, **43**, 6799–6802.
- 269 T. Yamada, K. Minoura, R. Tanaka and A. Numata, J. Antibiot., 2006, **59**, 345–350.
- 270 T. Yamada, K. Minoura, R. Tanaka and A. Numata, J. Antibiot., 2007, 60, 370–375.
- 271 Y. F. Huang, L. Qiao, A. L. Lv, Y. H. Pei and L. Tian, *Chin. Chem. Lett.*, 2008, **19**, 562–564.
- 272 G. Schneider, H. Anke and O. Sterner, Nat. Prod. Lett., 1997, 10, 133–138.
- 273 K. Motohashi, J. Hashimoto, S. Inaba, S. T. Khan, H. Komaki, A. Nagai, M. Takagi and K. Shin-ya, J. Antibiot., 2009, 62, 247– 250.
- 274 C. Riche, C. Pascard-Billy, M. Devys, A. Gaudemer, M. Barbier and J.-F. Bousquet, *Tetrahedron Lett.*, 1974, 15, 2765–2766.
- 275 Y. Tirilly, J. Kloosterman, G. Sipma and J. J. Kettenes-van den Bosch, *Phytochemistry*, 1983, **22**, 2082–2083.
- 276 H. Oh, P. R. Jensen, B. T. Murphy, C. Fiorilla, J. F. Sullivan, T. Ramsey and W. Fenical, J. Nat. Prod., 2010, 73, 998–1001.
- 277 M. Isaka, U. Srisanoh, S. Veeranondha, W. Choowong and S. Lumyong, *Tetrahedron*, 2009, 65, 8808–8815.
 278 C. L. Shao, C. Y. Wang, C. Y. Li, Z. G. She, Y. C. Gu and Y. C. Lin,
- 278 C. L. Shao, C. Y. Wang, C. Y. Li, Z. G. She, Y. C. Gu and Y. C. Lin, *Nat. Prod. Res.*, 2009, **23**, 1579–1583.
- 279 U. Höller, G. M. König and A. D. Wright, J. Nat. Prod., 1999, 62, 114–118.
- 280 H. B. Liu, R. Edrada-Ebel, R. Ebel, Y. Wang, B. Schulz, S. Draeger, W. E. G. Müller, V. Wray, W. H. Lin and P. Proksch, *J. Nat. Prod.*, 2009, **72**, 1585–1588.
- 281 T. Ogawa, K. Ando, T. Tanaka, Y. Uosaki and Y. Matsuda, J. Antibiot., 1996, 49, 1–5.
- 282 Y. Uosaki, M. Yoshida, T. Ogawa and Y. Saitoh, J. Antibiot., 1996, 49, 6–12.
- 283 T. Ogawa, Y. Uosaki, T. Tanaka, E. Tsukuda, A. Mihara and Y. Matsuda, J. Antibiot., 1996, 49, 168–172.
- 284 M. Y. Wei, C. Y. Wang, Q. A. Liu, C. L. Shao, Z. G. She and Y. C. Lin, *Mar. Drugs*, 2010, 8, 941–949.
- 285 T. Hamasaki, K. Nagayama and Y. Hatsuda, Agric. Biol. Chem., 1978, 42, 37–40.
- 286 T. Hamasaki, Y. Sato and Y. Hatsuda, Agric. Biol. Chem., 1975, 39, 2337–2340.
- 287 T. Hamasaki, Y. Sato, Y. Hatsuda, M. Tanabe and L. W. Cary, Tetrahedron Lett., 1975, 16, 659–660.
- 288 Z. Y. Lu, H. J. Zhu, P. Fu, Y. Wang, Z. H. Zhang, H. P. Lin, P. P. Liu, Y. B. Zhuang, K. Hong and W. M. Zhu, J. Nat. Prod., 2010, 73, 911–914.
- 289 C. Almeida, S. Elsaedi, S. Kehraus and G. M. König, *Nat. Prod. Commun.*, 2010, 5, 507–510.
- 290 S. Nozoe, H. Kobayashi and N. Morisaki, *Tetrahedron Lett.*, 1976, 17, 4625–4626.
- 291 C. Almeida, E. Eguereva, S. Kehraus, C. Siering and G. M. König, J. Nat. Prod., 2010, 73, 476–478.
- 292 M. Katayama and S. Marumo, *Tetrahedron Lett.*, 1979, 20, 1773– 1776.
- 293 M. Katayama, S. Marumo and H. Hattori, *Tetrahedron Lett.*, 1983, 24, 1703–1706.
- 294 H. R. Huang, X. K. Xia, Z. G. She, Y. C. Lin, L. L. P. Vrijmoed and E. B. G. Jones, J. Asian Nat. Prod. Res., 2006, 8, 609–612.
- 295 C. M. Boot, K. Tenney, F. A. Valeriote and P. Crews, J. Nat. Prod., 2006, 69, 83–92.
- 296 C. M. Boot, T. Amagata, K. Tenney, J. E. Compton, H. Pietraszkiewicz, F. A. Valeriote and P. Crews, *Tetrahedron*, 2007, **63**, 9903–9914.
- 297 S. Gupta, S. B. Krasnoff, D. W. Roberts, J. A. A. Renwick, L. S. Brinen and J. Clardy, J. Org. Chem., 1992, 57, 2306–2313.
- 298 Y. Hayakawa, Y. Hattori, T. Kawasaki, K. Kanoh, K. Adachi, Y. Shizuri and K. Shin-ya, J. Antibiot., 2008, 61, 365–371.

View Article Online

- 299 P. Pruksakorn, M. Arai, N. Kotoku, C. Vilcheze, A. D. Baughn, P. Moodley, W. R. Jacobs and M. Kobayashi, *Bioorg. Med. Chem. Lett.*, 2010, 20, 3658–3663.
- 300 T. Arai, Y. Mikami, K. Fukushim, T. Utsumi and K. Yazawa, J. Antibiot., 1973, 26, 157–161.
- 301 K. Fuji, E. Fujita, Y. Takaishi, T. Fujita, I. Arita, M. Komatsu and N. Hiratsuka, *Experientia*, 1978, 34, 237–239.
- 302 J. W. Ren, C. M. Xue, L. Tian, M. J. Xu, J. Chen, Z. W. Deng, P. Proksch and W. H. Lin, *J. Nat. Prod.*, 2009, **72**, 1036–1044.
- 303 M. Mohamed-Benkada, M. Montagu, J. F. Biard, F. Mondeguer, P. Verite, M. Dalgalarrondo, J. Bissett and Y. F. Pouchus, *Rapid Commun. Mass Spectrom.*, 2006, **20**, 1176–1180.
- 304 N. Ruiz, G. Wielgosz-Collin, L. Poirier, O. Grovel, K. E. Petit, M. Mohamed-Benkada, T. R. du Pont, J. Bissett, P. Verite, G. Barnathan and Y. F. Pouchus, *Peptides*, 2007, 28, 1351– 1358.
- 305 L. J. Cruz, M. M. Insua, J. P. Baz, M. Trujillo, R. A. Rodriguez-Mias, E. Oliveira, E. Giralt, F. Albericio and L. M. Canedo, J. Org. Chem., 2006, 71, 3335–3338.
- 306 J. Zheng, H. Zhu, K. Hong, Y. Wang, P. Liu, X. Wang, X. Peng and W. Zhu, Org. Lett., 2009, 11, 5262–5265.
- 307 G. Lang, M. I. Mitova, G. Ellis, S. van der Sar, R. K. Phipps, J. W. Blunt, N. J. Cummings, A. L. J. Cole and M. H. G. Munro, *J. Nat. Prod.*, 2006, **69**, 621–624.
- 308 T. Amagata, B. I. Morinaka, A. Amagata, K. Tenney, F. A. Valeriote, E. Lobkovsky, J. Clardy and P. Crews, J. Nat. Prod., 2006, 69, 1560–1565.
- 309 D. C. Oh, C. A. Kauffman, P. R. Jensen and W. Fenical, J. Nat. Prod., 2007, 70, 515–520.
- 310 Y. M. Chiang, E. Szewczyk, T. Nayak, A. D. Davidson, J. F. Sanchez, H. C. Lo, W. Y. Ho, H. Simityan, E. Kuo, A. Praseuth, K. Watanabe, B. R. Oakley and C. C. C. Wang, *Chem. Biol.*, 2008, **15**, 527–532.
- 311 Z. G. Yu, G. Lang, I. Kajahn, R. Schmaljohann and J. F. Imhoff, J. Nat. Prod., 2008, 71, 1052–1054.
- 312 S. P. Lira, A. M. Vita-Marques, M. H. R. Seleghim, T. S. Bugni, D. V. LaBarbera, L. D. Sette, S. R. P. Sponchiado, C. M. Ireland and R. G. S. Berlinck, *J. Antibiot.*, 2006, **59**, 553–563.
- 313 A. M. de Vita-Marques, S. P. Lira, R. G. S. Berlinck, M. H. R. Seleghim, S. R. P. Sponchiado, S. M. Tauk-Tornisielo, M. Barata, C. Pessoa, M. O. de Moraes, B. C. Cavalcanti, G. G. F. Nascimento, A. O. de Souza, F. C. S. Galetti, C. L. Silva, M. Silva, E. F. Pimenta, O. Thiemann, M. R. Z. Passarini and L. D. Sette, *Quim. Nova*, 2008, **31**, 1099–1103.
- 314 A. Kralj, S. Kehraus, A. Krick, G. van Echten-Deckert and G. M. König, *Planta Med.*, 2007, 73, 366–371.
- 315 H. R. Huang, Z. G. She, Y. C. Lin, L. L. P. Vrijmoed and W. H. Lin, J. Nat. Prod., 2007, 70, 1696–1699.
- 316 D. C. Oh, P. R. Jensen and W. Fenical, *Tetrahedron Lett.*, 2006, **47**, 8625–8628.
- 317 M. Y. Kim, J. H. Sohn, J. S. Ahn and H. Oh, J. Nat. Prod., 2009, 72, 2065–2068.
- 318 L. J. Fremlin, A. M. Piggott, E. Lacey and R. J. Capon, J. Nat. Prod., 2009, 72, 666–670.
- 319 G. N. Belofsky, P. R. Jensen and W. Fenical, *Tetrahedron Lett.*, 1999, **40**, 2913–2916.
- 320 W. Gu, M. Cueto, P. R. Jensen, W. Fenical and R. B. Silverman, *Tetrahedron*, 2007, **63**, 6535–6541.
- 321 S. B. Singh, D. L. Zink, J. D. Polishook, A. W. Dombrowski, S. J. DarkinRattray, D. M. Schmatz and M. A. Goetz, *Tetrahedron Lett.*, 1996, 37, 8077–8080.
- 322 H. Tani, Y. Fujii and H. Nakajima, *Phytochemistry*, 2001, **58**, 305–310.
- 323 Y. Sun, L. Tian, Y. F. Huang, Y. Sha and Y. H. Pei, *Pharmazie*, 2006, **61**, 809–810.
- 324 K. Motohashi, S. Inaba, M. Takagi and K. Shin-Ya, *Biosci.*, *Biotechnol., Biochem.*, 2009, **73**, 1898–1900.
- 325 R. Myokei, A. Sakurai, C. F. Chang and Y. Kodaira, *Tetrahedron* Lett., 1969, **10**, 695–698.
- 326 R. Ratnayake, L. J. Fremlin, E. Lacey, J. H. Gill and R. J. Capon, J. Nat. Prod., 2008, 71, 403–408.
- 327 Y. M. Lee, H. T. Dang, J. Hong, C. O. Lee, K. S. Bae, D. K. Kim and J. H. Jung, *Bull. Korean Chem. Soc.*, 2010, **31**, 205–208.
- 328 H. Shigemori, S. Wakuri, K. Yazawa, T. Nakamura, T. Sasaki and J. Kobayashi, *Tetrahedron*, 1991, 47, 8529–8534.

- 329 Z. J. Lin, T. J. Zhu, Y. C. Fang and Q. Q. Gu, Magn. Reson. Chem., 2008, 46, 1212–1216.
- 330 T. Kagamizono, N. Sakai, K. Arai, K. Kobinata and H. Osada, *Tetrahedron Lett.*, 1997, 38, 1223–1226.
- 331 M. Izumikawa, J. Hashimoto, M. Takagi and K. Shin-ya, J. Antibiot., 2010, 63, 389–391.
- 332 H. Kato, T. Yoshida, T. Tokue, Y. Nojiri, H. Hirota, T. Ohta, R. M. Williams and S. Tsukamoto, *Angew. Chem., Int. Ed.*, 2007, 46, 2254–2256.
- 333 S. Tsukamoto, H. Kato, M. Samizo, Y. Nojiri, H. Onuki, H. Hirota and T. Ohta, J. Nat. Prod., 2008, 71, 2064–2067.
- 334 A. C. Whyte, J. B. Gloer, D. T. Wicklow and P. F. Dowd, J. Nat. Prod., 1996, 59, 1093–1095.
- 335 M. Yamazaki, E. Okuyama, M. Kobayashi and H. Inoue, *Tetrahedron Lett.*, 1981, 22, 135–136.
- 336 J. F. Qian-Cutrone, S. Huang, Y. Z. Shu, D. Vyas, C. Fairchild, A. Menendez, K. Krampitz, R. Dalterio, S. E. Klohr and Q. Gao, *J. Am. Chem. Soc.*, 2002, **124**, 14556–14557.
- 337 T. J. Greshock, A. W. Grubbs, S. Tsukamoto and R. M. Williams, Angew. Chem., Int. Ed., 2007, 46, 2262–2265.
- 338 S. Tsukamoto, H. Kato, T. J. Greshock, H. Hirota, T. Ohta and R. M. Williams, J. Am. Chem. Soc., 2009, 131, 3834.
- 339 S. Tsukamoto, T. Kawabata, H. Kato, T. J. Greshock, H. Hirota, T. Ohta and R. M. Williams, *Org. Lett.*, 2009, **11**, 1297–1300.
- 340 T. J. Greshock, A. W. Grubbs, P. Jiao, D. T. Wicklow, J. B. Gloer and R. M. Williams, *Angew. Chem.*, *Int. Ed.*, 2008, 47, 3573–3577.
- 341 K. Kito, R. Ookura, T. Kusumi, M. Namikoshi and T. Ooi, *Heterocycles*, 2009, 78, 2101–2106.
- 342 S. Tsukamoto, H. Umaoka, K. Yoshikawa, T. Ikeda and H. Hirota, J. Nat. Prod., 2010, 73, 1438–1440.
- 343 K. Nozawa and S. Nakajima, J. Nat. Prod., 1979, 42, 374-377.
- 344 J. C. Frisvad, J. Smedsgaard, T. O. Larsen and R. A. Samson, Stud. Mycol., 2004, 201–241.
- 345 F. Z. Wang, Y. C. Fang, T. J. Zhu, M. Zhang, A. Q. Lin, Q. Q. Gu and W. M. Zhu, *Tetrahedron*, 2008, 64, 7986–7991.
- 346 C. B. Cui, H. Kakeya and H. Osada, *Tetrahedron*, 1996, **52**, 12651– 12666.
- 347 C. B. Cui, H. Kakeya and H. Osada, J. Antibiot., 1996, 49, 832-835.
- 348 M. Yamazaki, K. Sasago and K. Miyaki, J. Chem. Soc., Chem. Commun., 1974, 408–409.
- 349 J. Fayos, D. Lokensga and J. Clardy, J. Am. Chem. Soc., 1974, 96, 6785–6787.
- 350 L. Du, X. Y. Yang, T. J. Zhu, F. P. Wang, X. Xiao, H. Park and Q. Q. Gu, Chem. Pharm. Bull., 2009, 57, 873–876.
- 351 M. Kusano, G. Sotoma, H. Koshino, J. Uzawa, M. Chijimatsu, S. Fujioka, T. Kawano and Y. Kimura, J. Chem. Soc., Perkin Trans. 1, 1998, 2823–2826.
- 352 Y. Kimura, A. Sawada, M. Kuramata, M. Kusano, S. Fujioka, T. Kawano and A. Shimada, J. Nat. Prod., 2005, 68, 237–239.
- 353 D. L. Li, X. M. Li, T. G. Li, H. Y. Dang and B. G. Wang, *Helv. Chim. Acta*, 2008, **91**, 1888–1893.
- 354 W.-L. Wang, Z.-Y. Lu, H.-W. Tao, T.-J. Zhu, Y.-C. Fang, Q.-Q. Gu and W.-M. Zhu, J. Nat. Prod., 2007, 70, 1558–1564.
- 355 C. Cardani, G. Casnati, F. Piozzi and A. Quilico, *Tetrahedron Lett.*, 1959, 1, 1–8.
- 356 C. B. Cui, H. Kakeya, G. Okada, R. Onose, M. Ubukata, I. Takahashi, K. Isono and H. Osada, J. Antibiot., 1995, 48, 1382– 1384.
- 357 Y. Wang, J. B. Gloer, J. A. Scott and D. Malloch, J. Nat. Prod., 1995, 58, 93–99.
- 358 C. M. Xue, T. Li, Z. W. Deng, H. Z. Fu and W. H. Lin, *Pharmazie*, 2006, **61**, 1041–1044.
- 359 Y. Yao, L. Tian, J. Q. Cao and Y. H. Pei, *Pharmazie*, 2007, **62**, 478–479.
- 360 Y. F. Huang, L. Tian, H. M. Hua and Y. H. Pei, J. Asian Nat. Prod. Res., 2007, 9, 197–201.
- 361 Y. Yao, L. Tian, J. Li, J. Q. Cao and Y. H. Pei, *Pharmazie*, 2009, 64, 616–618.
- 362 R. J. Capon, M. Stewart, R. Ratnayake, E. Lacey and J. H. Gill, J. Nat. Prod., 2007, 70, 1746–1752.
- 363 N. Kawahara, K. Nozawa, S. Nakajima and K. Kawai, J. Chem. Soc., Perkin Trans. 1, 1987, 2099–2101.
- 364 G.-Y.-S. Wang, L. M. Abrell, A. Avelar, B. M. Borgeson and P. Crews, *Tetrahedron*, 1998, 54, 7335–7342.

- 366 W. Y. Zhao, T. J. Zhu, X. X. Han, G. T. Fan, H. B. Liu, W. M. Zhu and Q. Q. Gu, *Nat. Prod. Res.*, 2009, **23**, 203–207.
- 367 W. L. Wang, Y. Wang, H. W. Tao, X. P. Peng, P. P. Liu and W. M. Zhu, J. Nat. Prod., 2009, 72, 1695–1698.
- 368 Y. C. Park, S. P. Gunasekera, J. V. Lopez, P. J. McCarthy and A. E. Wright, J. Nat. Prod., 2006, 69, 580–584.
- 369 Y. G. Chen, C. L. Shao, Z. J. Huang, Y. Zhang, X. L. Cai, Z. G. She, S. N. Zhou and Y. C. Lin, *Magn. Reson. Chem.*, 2009, 47, 92–95.
- 370 W. Y. Zhao, T. J. Zhu, G. T. Fan, H. B. Liu, Y. C. Fang, Q. Q. Gu and W. M. Zhu, *Nat. Prod. Res.*, 2010, **24**, 953–957.
- 371 K. H. Michel, M. O. Chaney, N. D. Jones, M. M. Hoehn and R. Nagaraja, J. Antibiot., 1974, 27, 57–64.
- 372 M. Takagi, K. Motohashi and K. Shin-ya, J. Antibiot., 2010, 63, 393-395.
- 373 G. Carr, W. Tay, H. Bottriell, S. K. Andersen, A. G. Mauk and R. J. Andersen, Org. Lett., 2009, 11, 2996–2999.
- 374 G. C. Prendergast, Oncogene, 2008, 27, 3889-3900.
- 375 M. J. Xu, G. Gessner, I. Groth, C. Lange, A. Christner, T. Bruhn, Z. W. Deng, X. Li, S. H. Heinemann, S. Grabley, G. Bringmann, I. Sattler and W. H. Lin, *Tetrahedron*, 2007, **63**, 435–444.
- 376 G. N. Belofsky, J. B. Gloer, D. T. Wicklow and P. F. Dowd, *Tetrahedron*, 1995, **51**, 3959–3968.
- 377 J. P. Springer and J. Clardy, Tetrahedron Lett., 1980, 21, 231-234.
- 378 J. W. Dorner, R. J. Cole, R. H. Cox and B. M. Cunfer, J. Agric. Food Chem., 1984, 32, 1069–1071.
- 379 O. F. Smetanina, A. I. Kalinovsky, Y. V. Khudyakova, M. V. Pivkin, P. S. Dmitrenok, S. N. Fedorov, H. Ji, J. Y. Kwak and T. A. Kuznetsova, *J. Nat. Prod.*, 2007, **70**, 906–909.
- 380 O. F. Smetanina, A. I. Kalinovsky, Y. V. Khudyakova, P. S. Dmitrenok, S. N. Federov, H. Ji, J. Y. Kwak and T. A. Kuznetsova, J. Nat. Prod., 2007, 70, 2054–2054.
- 381 A. Q. Lin, L. Du, Y. C. Fang, F. Z. Wang, T. J. Zhu, Q. Q. Gu and W. M. Zhu, *Chem. Nat. Compd.*, 2009, 45, 677–680.
- 382 G. A. Burdock and W. G. Flamm, *Int. J. Toxicol.*, 2000, **19**, 195–218.
 383 T. Amagata, K. Minoura and A. Numata, *J. Nat. Prod.*, 2006, **69**,
- 1384–1388.
 384 T. Amagata, M. Tanaka, T. Yamada, K. Minoura and A. Numata, *J. Nat. Prod.*, 2008, **71**, 340–345.
- 385 T. Amagata, K. Takigawa, K. Minoura and A. Numata, *Heterocycles*, 2010, **81**, 897–907.
- 386 J.-H. Jang, K. Kanoh, K. Adachi and Y. Shizuri, J. Nat. Prod., 2006, 69, 1358–1360.
- 387 N. Ingavat, J. Dobereiner, S. Wiyakrutta, C. Mahidol, S. Ruchirawat and P. Kittakoop, *J. Nat. Prod.*, 2009, **72**, 2049–2052.
- 388 E. Quiñoà and P. Crews, *Tetrahedron Lett.*, 1987, 28, 3229–3232.
 389 N. C. Gassner, C. M. Tamble, J. E. Bock, N. Cotton, K. N. White, K. Tenney, R. P. S. Onge, M. J. Proctor, G. Giaever, C. Nislow, R. W. Davis, P. Crews, T. R. Holman and S. R. Lokey, *J. Nat. Prod.*, 2007, 70, 383–390.
- 390 C. M. Boot, N. C. Gassner, J. E. Compton, K. Tenney, C. M. Tamble, R. S. Lokey, T. R. Holman and P. Crews, *J. Nat. Prod.*, 2007, **70**, 1672–1675.
- 391 P. Bloch, C. Tamm, P. Bollinger, T. J. Petcher and H. P. Weber, *Helv. Chim. Acta*, 1976, **59**, 133–137.
- 392 T. Yamada, E. Imai, K. Nakatuji, A. Numata and R. Tanaka, *Tetrahedron Lett.*, 2007, **48**, 6294–6296.
- 393 H. Ren, R. Liu, L. Chen, T. J. Zhu, W. M. Zhu and Q. Q. Gu, Arch. Pharmacal Res., 2010, 33, 499–502.
- 394 R. Liu, Q. Q. Gu, W. M. Zhu, C. B. Cui, G. T. Fan, Y. C. Fang, T. J. Zhu and H. B. Liu, J. Nat. Prod., 2006, 69, 871–875.
- 395 R. Liu, Z. J. Lin, T. J. Zhu, Y. C. Fang, Q. Q. Gu and W. M. Zhu, J. Nat. Prod., 2008, 71, 1127–1132.
- 396 Z. J. Lin, T. J. Zhu, H. J. Wei, G. J. Zhang, H. Wang and Q. Q. Gu, *Eur. J. Org. Chem.*, 2009, 3045–3051.
- 397 Z. J. Lin, T. J. Zhu, G. J. Zhang, H. J. Wei and Q. Q. Gu, *Can. J. Chem.*, 2009, **87**, 486–489.
- 398 Z. J. Lin, G. J. Zhang, T. J. Zhu, R. Liu, H. J. Wei and Q. Q. Gu, *Helv. Chim. Acta*, 2009, 92, 1538–1544.
- 399 Z. J. Lin, T. J. Zhu, L. Chen and Q. Q. Gu, *Chin. Chem. Lett.*, 2010, 21, 824–826.
- 400 V. Rukachaisirikul, N. Khamthong, Y. Sukpondma, C. Pakawatchal, S. Phongpaichit, J. Sakayaroj and K. Kirtikara, *Chem. Pharm. Bull.*, 2009, **57**, 1409–1411.

- 401 C. M. Cui, X. M. Li, C. S. Li, P. Proksch and B. G. Wang, J. Nat. Prod., 2010, 73, 729–733.
- 402 S. Kanokmedhakul, K. Kanokmedhakul, N. Phonkerd, K. Soytong, P. Kongsaeree and A. Suksamrarn, *Planta Med.*, 2002, 68, 834–836.
- 403 H. Oikawa, Y. Murakami and A. Ichihara, Biosci., Biotechnol., Biochem., 1993, 57, 628–631.
- 404 D. H. Zhang, X. D. Yang, J. S. Kang, H. D. Choi and B. W. Son, J. Antibiot., 2008, 61, 40–42.
- 405 L. Rahbæk, J. Breinholt, J. C. Frisvad and C. Christophersen, J. Org. Chem., 1999, 64, 1689–1692.
- 406 J. R. Dai, B. K. Carté, P. J. Sidebottom, A. L. S. Yew, S. B. Ng, Y. C. Huang and M. S. Butler, J. Nat. Prod., 2001, 64, 125–126.
- 407 R. Ookura, K. Kito, T. Ooi, M. Namikoshi and T. Kusumi, J. Org. Chem., 2008, 73, 4245–4247.
- 408 L. Rahbæk and J. Breinholt, J. Nat. Prod., 1999, 62, 904-905.
- 409 C. M. Cui, X. M. Li, C. S. Li, H. F. Sun, S. S. Gan and B. G. Wang, *Helv. Chim. Acta*, 2009, **92**, 1366–1370.
- 410 Z. H. Xin, Y. C. Fang, L. Du, T. J. Zhu, L. Duan, J. Chen, Q. Q. Gu and W. M. Zhu, J. Nat. Prod., 2007, 70, 853–855.
- 411 J. M. Boyeskorkis, K. A. Gurney, J. Penn, P. G. Mantle, J. N. Bilton and R. N. Sheppard, *J. Nat. Prod.*, 1993, **56**, 1707–1717.
- 412 T. O. Larsen, H. Franzyk and S. R. Jensen, J. Nat. Prod., 1999, 62, 1578–1580.
- 413 C. Takahashi, T. Matsushita, M. Doi, K. Minoura, T. Shingu, Y. Kumeda and A. Numata, J. Chem. Soc., Perkin Trans. 1, 1995, 2345–2353.
- 414 S. W. Meyer, T. F. Mordhorst, C. Lee, P. R. Jensen, W. Fenical and M. Kock, Org. Biomol. Chem., 2010, 8, 2158–2163.
- 415 G. Voerman, S. Cavalli, G. A. van der Marel, W. Pfleiderer, J. H. van Boom and D. V. Filippov, J. Nat. Prod., 2005, 68, 938–941.
- 416 J. Junker, W. Maier, T. Lindel and M. Köck, Org. Lett., 1999, 1, 737–740.
- 417 F. Xu, J. Y. Pang, B. T. Lu, J. J. Wang, Y. Zhang, Z. G. She, L. L. P. Vrijmoed, E. B. Gareth Jones and Y. C. Lin, *Chin. J. Chem.*, 2009, **27**, 365–368.
- 418 F. Zhu and Y. C. Lin, Chin. Sci. Bull., 2006, 51, 1426-1430.
- 419 M. Cueto, P. R. Jensen, C. Kauffman, W. Fenical, E. Lobkovsky and J. Clardy, J. Nat. Prod., 2001, 64, 1444–1446.
- 420 D. C. Oh, P. R. Jensen, C. A. Kauffman and W. Fenical, *Bioorg. Med. Chem.*, 2005, **13**, 5267–5273.
- 421 C.-L. Shao, C.-Y. Wang, Y.-C. Gu, M.-Y. Wei, J.-H. Pan, D.-S. Deng, Z.-G. She and Y.-C. Lin, *Bioorg. Med. Chem. Lett.*, 2010, 20, 3284–3286.
- 422 Y. W. Tao, X. J. Zeng, C. B. Mou, J. Li, X. L. Cai, Z. G. She, S. I. Zhou and Y. C. Lin, *Magn. Reson. Chem.*, 2008, 46, 501–505.
- 423 C. C. J. Culvenor, P. A. Cockrum, J. A. Edgar, J. L. Frahn, C. P. Gorst-Allman, A. J. Jones, W. F. O. Marasas, K. E. Murray, L. W. Smith, P. S. Steyn, R. Vleggaar and P. L. Wessels, J. Chem. Soc., Chem. Commun., 1983, 1259–1262.
- 424 Y. F. Huang, L. H. Li, L. Tian, L. Qiao, H. M. Hua and Y. H. Pei, J. Antibiot., 2006, **59**, 355–357.
- 425 G. Bringmann, G. Lang, J. Mühlbacher, K. Schaumann, S. Steffens, P. G. Rytik, U. Hentschel, J. Morschhauser and W. E. G. Müller, *Prog. Mol. Subcell. Biol.*, 2003, **37**, 231–253.
- 426 G. Bringmann, T. A. M. Gulder, G. Lang, S. Schmitt, R. Stöhr, J. Wiese, K. Nagel and J. F. Imhoff, *Mar. Drugs*, 2007, 5, 23–30.
- 427 G. Bringmann, G. Lang, T. A. M. Gulder, H. Tsuruta, J. Mühlbacher, K. Maksimenka, S. Steffens, K. Schaumann, R. Stöhr, J. Wiese, J. F. Imhoff, S. Perović-Ottstadt, O. Boreiko and W. E. G. Müller, *Tetrahedron*, 2005, **61**, 7252–7265.
- 428 G. M. Cabrera, M. Butler, M. A. Rodriguez, A. Godeas, R. Haddad and M. N. Eberlin, *J. Nat. Prod.*, 2006, **69**, 1806–1808.
- 429 Y. Zhang, X. M. Li, P. Proksch and B. G. Wang, *Steroids*, 2007, 72, 723–727.
- 430 Y. Zhang, X. M. Li, C. Y. Wang and B. G. Wang, *Chin. Chem. Lett.*, 2007, **18**, 951–953.
- 431 J. Varga, S. Kocsube, B. Toth, J. C. Frisvad, G. Perrone, A. Susca, M. Meijer and R. A. Samson, *Int. J. Syst. Evol. Microbiol.*, 2007, 57, 1925–1932.
- 432 C. L. Barnes, J. R. Steiner, E. Torres, R. Pacheco and H. Marquez, Int. J. Pept. Protein Res., 1990, 36, 292–296.
- 433 J. Hiort, K. Maksimenka, M. Reichert, S. Perović-Ottstadt, W. H. Lin, V. Wray, K. Steube, K. Schaumann, H. Weber, P. Proksch, R. Ebel, W. E. G. Müller and G. Bringmann, J. Nat. Prod., 2004, 67, 1532–1543.

- Prod., 2005, 68, 1821–1821.
 435 Y. Zhang, X. M. Li, Y. Feng and B. G. Wang, Nat. Prod. Res., 2010, 24, 1036–1043.
- 436 Z. H. Lin, Z. Y. Lu, T. H. Zhu, Y. C. Fang, Q. Q. Gu and W. M. Zhu, *Chem. Pharm. Bull.*, 2008, 56, 217–221.
- 437 K. Neumann, S. Kehraus, M. Gutschow and G. M. Koning, Nat. Prod. Commun., 2009, 4, 347–354.
- 438 R. F. Vesonder, L. W. Tjarks, W. K. Rohwedder, H. R. Burmeister and J. A. Laugal, J. Antibiot., 1979, 32, 759–761.
- 439 A. Miljkovic, P. G. Mantle, D. J. Williams and B. Rassing, J. Nat. Prod., 2001, 64, 1251–1253.
- 440 R. M. Van Wagoner, P. G. Mantle and J. L. C. Wright, J. Nat. Prod., 2008, **71**, 426–430.
- 441 R. Nagarajan, N. Narasimhachari, M. V. Kadkol and K. S. Gopalkrishnan, J. Antibiot., 1971, 24, 249–252.
- 442 Y. P. Zhang, T. J. Zhu, Y. C. Fang, H. B. Liu, Q. Q. Gu and W. M. Zhu, J. Antibiot., 2007, 60, 153–157.
- 443 E. D. de Silva, A. S. Geiermann, M. I. Mitova, P. Kuegler, J. W. Blunt, A. L. J. Cole and M. H. G. Munro, *J. Nat. Prod.*, 2009, **72**, 477–479.
- 444 Y. Fujita, H. Oguri and H. Oikawa, J. Antibiot., 2005, 58, 425-427.
- 445 M. M. Wagenaar, D. M. Gibson and J. Clardy, Org. Lett., 2002, 4, 671–673.
- 446 G. Schlingmann, T. Taniguchi, H. Y. He, R. Bigelis, H. Y. Yang, F. E. Koehn, G. T. Carter and N. Berova, *J. Nat. Prod.*, 2007, **70**, 1180–1187.
- 447 R. A. Samson, J. Houbraken, A. F. A. Kuijpers, J. M. Frank and J. C. Frisvad, *Stud. Mycol.*, 2004, 45–61.
- 448 Y. Miyake, C. Ito, M. Itoigawa and T. Osawa, *Biosci., Biotechnol., Biochem.*, 2007, 71, 2515–2521.
- 449 Y. Miyake, M. Mochizuki, C. Ito, M. Itoigawa and T. Osawa, Biosci., Biotechnol., Biochem., 2008, 72, 1580–1585.
- 450 X. F. Li, S. K. Kim, J. S. Kang, H. D. Choi and B. W. Son, J. Microbiol. Biotechnol., 2006, 16, 637–638.
- 451 V. Manriquez, A. Galdamez, B. Veliz, J. Rovirosa, A. R. Diaz-Marrero, M. Cueto, J. Darias, C. Martinez and A. San-Martin, J. *Chil. Chem. Soc.*, 2009, 54, 314–316.
- 452 Y. Sugiyama, Y. Ito, M. Suzuki and A. Hirota, J. Nat. Prod., 2009, 72, 2069–2071.
- 453 N. Tan, J. H. Pan, G. T. Peng, C. B. Mou, Y. W. Tao, Z. G. She, Z. L. Yang, S. N. Zhou and Y. C. Lin, *Chin. J. Chem.*, 2008, 26, 516–521.
- 454 H. X. Wang and T. B. Ng, Life Sci., 1999, 65, 849-856.
- 455 X. F. Li, M. K. Kim, U. Lee, S. K. Kim, J. S. Kang, H. D. Choi and B. W. Son, *Chem. Pharm. Bull.*, 2005, **53**, 453–455.

- 456 L. Y. Li, W. H. Lin, H. Z. Fu, I. Sattler, X. S. Huang and S. Grabley, J. Antibiot., 2005, 58, 594–598.
- 457 F. Teuscher, W. Lin, V. Wray, R. Edrada, K. Padmakumar, P. Proksch and R. Ebel, *Nat. Prod. Commun.*, 2006, 1, 927–933.
- 458 R. R. Parvatkar, C. D'Souza, A. Tripathi and C. G. Naik, *Phytochemistry*, 2009, **70**, 128–132.
- 459 N. Kiriyama, K. Nitta, Y. Sakaguchi, Y. Taguchi and Y. Yamamoto, *Chem. Pharm. Bull.*, 1977, 25, 2593–2601.
- 460 Y. G. Chen, J. H. Pan, F. Xu, F. Liu, J. X. Yang, C. H. Huang, C. L. Xu, Y. J. Lu, X. L. Cai, Z. G. She and Y. C. Lin, *Chem. Nat. Compd.*, 2010, **46**, 230–232.
- 461 H. Wei, H. Inada, A. Hayashi, K. Higashimoto, P. Pruksakorn, S. Kamada, M. Arai, S. Ishida and M. Kobayashi, J. Antibiot., 2007, 60, 586–590.
- 462 R. Marchelli and L. C. Vining, J. Antibiot., 1975, 28, 328-331.
- 463 T. Kamigauchi, R. Sakazaki, K. Nagashima, Y. Kawamura, Y. Yasuda, K. Matsushima, H. Tani, Y. Takahashi, K. Ishii, R. Suzuki, K. Koizumi, H. Nakai, Y. Ikenishi and Y. Terui, *J. Antibiot.*, 1998, **51**, 445–450.
- 464 P. Chandra, G. Read and L. C. Vining, Can. J. Biochem. Cell Biol., 1966, 44, 403–413.
- 465 G. Chen, Y. Zhu, H. Z. Wang, S. J. Wang and R. Q. Zhang, J. Asian Nat. Prod. Res., 2007, 9, 159–164.
- 466 J. T. Gautschi, K. Tenney, J. Compton and P. Crews, *Nat. Prod. Commun.*, 2007, 2, 541–546.
- 467 Y. N. Wang, L. Tian, H. M. Hua, X. Lu, S. Sun, H. H. Wu and Y. H. Pei, J. Asian Nat. Prod. Res., 2009, 11, 912–917.
- 468 Q. H. Zhang, L. Tian, L. D. Zhou, Y. Zhang, Z. F. Li, H. M. Hua and Y. H. Pei, J. Asian Nat. Prod. Res., 2009, 11, 962–966.
- 469 S. Y. Wang, Z. L. Xu, W. W. Mao, Z. G. She, N. Tan, C. R. Li and Y. C. Lin, *Nat. Prod. Res.*, 2008, **22**, 612–617.
- 470 Y. Zhang, S. Wang, X.-M. Li, C.-M. Cui, C. Feng and B.-G. Wang, *Lipids*, 2007, 42, 759–764.
- 471 R. X. Tan and J. H. Chen, Nat. Prod. Rep., 2003, 20, 509-534.
- 472 Z. J. Wu, M. A. Ouyang, R. K. Su and Y. H. Kuo, *Chin. J. Chem.*, 2008, **26**, 759–764.
- 473 J. Y. Liu, L. L. Huang, Y. H. Ye, W. X. Zou, Z. J. Guo and R. X. Tan, J. Appl. Microbiol., 2006, 100, 195–202.
- 474 A. Abdel-Lateff, E. S. Elkhayat, M. A. Fouad and T. Okino, Nat. Prod. Commun., 2009, 4, 389-394.
- 475 W. Fenical, P. R. Jensen and X. C. Cheng, US Pat. 6069146, 1998.
- 476 D. G. I. Kingston, J. Nat. Prod., 2009, 72, 507-515.
- 477 http://www.clinicaltrials.gov/ct2/show/NCT00322608?term=npi-2358&rank=2, accessed 30/10/2010.
- 478 O. Aren, L. Matamala, M. Reyes, A. Santini, K. McArthur, G. K. Lloyd and M. A. Spear, J. Thorac. Oncol., 2010, 5, S91– S92.