Terpenoids and Phenols from Taiwania flousiana

XIANG Ying, YANG Sheng-Ping, ZHAN Zha-Jun, YUE Jian-Min*

(State Key Laboratory of Drug Research, Institute of Materia Medica, Shanghai Institutes for Biological Sciences, The Chinese Academy of Sciences, Shanghai 201203, China)

Abstract: Two new terpenoids, namely taiwaniatriol (1) and senecrassidiol-9-O- β -*D*-glucopyranoside (8), along with fifteen known compounds including five triterpenoids (2-6), one diterpene (7) and nine phenols (9-17), were isolated from the root bark of *Taiwania flousiana* Gaussen. Taiwaniatriol was elucidated as (24*S*)-3 β -methoxy-5 α -lanost-9(11)-ene-7 β , 24, 25-triol. Their structures were established mainly by spectral methods.

Key words: Taiwania flousiana; Taxodiaceae; triterpenoids; sesquiterpene glycoside; phenols

The genus *Taiwania* belonging to the family Taxodiaceae has only two species. A number of sesquiterpenes (Cheng *et al.*, 1967; Kuo *et al.*, 1969; He *et al.*, 1997), diterpenes (Lin *et al.*, 1995; Lin *et al.*, 1996; Lin *et al.*, 1998a), cycloadducts of diterpenes (Lin *et al.*, 1996; Lin *et al.*, 1997), sterols (Lin *et al.*, 1998a), lignans (Lin *et al.*, 1967; Lin *et al.*, 1998b) and biflavones (Kamil *et al.*, 1977; Kamil *et al.*, 1981) were isolated from *Taiwania cryptomerioides*. Among these compounds, α -cadinol showed selectively cytotoxic activity against the human colon adenocarcinoma (HT-29) cell line with an ED₅₀ value of 0.778 µg/mL (He *et al.*, 1997). Ferruginol and taiwanin C exhibited significant antifungal activity (Chang *et al.*, 1999).

T. flousiana Gaussen, an evergreen tree with linear-triangular leaves mainly distributed in China and northern Burma, has not been previously investigated chemically. In an effort to understand the chemical constituents of this plant, seventeen compounds, including six triterpenoids, one sesquiterpene glycoside, one diterpenoid, and nine phenols were isolated from the root bark of *T. flousiana*. Two of them are new compounds, namely (24*S*)-3β-methoxy-5α-lanost-9(11)-ene-7β,24,25-triol (1) and senecrassidiol-9-O-β-*D*-glucopyranoside (5). A trivial name taiwaniatriol was given to compound 1. Their structures were established mainly by spectral methods. Herein, we present the isolation and structural elucidation of these compounds (Fig.1).

1 Results and Discussion

(24S)-3 β -Methoxy-5 α -lanost-9(11)-ene-7 β ,24,25-triol (1), obtained as white powder, $[\alpha]_D^{20}$ + 42.0° (*c* 0.14, CHCl₃). The molecular formula C₃₁H₅₄O₄ was determined by HR-

EI-MS at m/z 490.400 8 [M]⁺ (calcd. 490.402 2). Its IR spectrum showed absorption bands at 3 547 cm^{-1} and 3 452 cm⁻¹ ascribable to hydroxyl groups, and a medium absorption band at 1 628 cm⁻¹ assignable to a double bond. In the ¹H-NMR (see **Experimental**) and ¹³C-NMR spectra (Table 1), it exhibited signals for one secondary methyl, seven tertiary methyls, one methoxyl group, three oxygenated tertiary carbons, one oxygenated quaternary carbon, and one trisubstituted double bond. The spectral data of compound 1 were remarkably similar to those of compound 2 except for the presence of one more hydroxyl group attached to a tertiary carbon. Compared with compound 2, the carbon signals of C-6, C-7 and C-8 in compound 1 resonating at δ 31.9, 72.5 and 50.4 were down-field shifted **D** 8 10.7, 44.5 and 8.6, respectively, indicating the presence of a C-7-OH (Fig. 1). The C-5 signal (δ 49.2) of compound **1** was up-field shifted $D\delta$ 3.8 compared with that of compound 2 caused by a typical γ -gauche effect of the C-7-OH, suggesting that the C-7-OH took a β -orientation, which was confirmed by the large coupling constants of H-7 (dt, J = 10.6, 4.9 Hz) taking an axial position. Compared with the known compound vietchiolide (Tanaka and Matshunaga, 1990) with a 7β -OH, the carbon signals of the B-ring and the coupling constants of 7α -H in compound 1 were dramatically matched with those reported for vietchiolide. 2D-NMR experiment (HMQC and HMBC) further confirmed the structure of compound 1 (Fig.2). The structure of compound 1 was thus elucidated to be (24S)-3 β -methoxy-5 α -lanost-9(11)-en-7 β , 24, 25-triol.

Senecrassidiol-9-O- β -*D*-glucopyranoside (**8**) obtained as pale gum, $[\alpha]_D^{20}$ -34.0° (*c* 0.18, CH₃OH), has the molecular formula C₂₁H₃₆O₇ determined by HR-EI-MS at *m/z*

Received 23 Oct. 2003 Accepted 21 Jun. 2004

Supported by the National Natural Science Foundation of China (30025044) and the State Key Program of Basic Research of China ("973" project, 2002CB512807).

^{*} Author for correspondence. Tel.: +86 (0)21 50806718; Fax: +86 (0)21 50807088; E-mail: <jmyue@mail.shcnc.ac.cn>.



Fig.1. Structures of compounds 1 - 17.

Table 1	C-NMR data of compounds 1-4 (CDC13, 400 MHZ)									
No.	1	2	3	4	No.	1	2	3	4	
1	36.2	36.0	36.1	36.7	17	50.2	51.0	51.0	50.9	
2	28.6	28.1	29.7	33.6	18	14.3	14.4	14.4	14.4	
3	88.4	88.6	78.9	217.3	19	22.0	22.2	22.2	22.0	
4	38.9	39.0	39.1	47.7	20	36.4	36.4	36.4	36.4	
5	49.2	53.0	52.5	53.4	21	18.6	18.5	18.5	18.5	
6	31.9	21.2	21.4	22.5	22	33.7	33.9	33.9	34.9	
7	72.5	28.0	28.1	28.0	23	28.8	28.7	28.7	27.7	
8	50.4	41.8	41.8	41.9	24	79.6	79.6	79.6	79.6	
9	146.3	148.7	148.5	147.1	25	73.3	73.2	73.2	73.2	
10	39.1	39.4	39.4	39.1	26	26.6	26.5	26.5	25.6	
11	117.2	114.7	114.9	116.2	27	23.3	23.2	23.2	23.2	
12	36.9	37.2	37.1	37.2	28	18.3	18.5	18.5	18.5	
13	45.1	44.3	44.3	44.3	29	28.2	28.2	28.1	26.5	
14	46.5	47.0	47.0	47.0	30	16.3	16.4	15.6	21.8	
15	36.7	33.6	33.9	33.9	OCH ₃	57.6	57.5			
16	22.6	22.5	28.1	28.7						

Table 1	¹³ C-NMR	data of con	npounds 1	-4	(CDCl ₃ ,	400 MHz)
---------	---------------------	-------------	-----------	----	----------------------	----------

382.237 0 [M-H₂O]⁺ (calcd. 382.235 5) and the ¹³C-NMR spectra. A broad IR absorption band at 3 415 cm⁻¹ was ascribed to hydroxyl groups. The ¹H-NMR spectrum showed three angular methyl signals at δ 0.87, 0.94 and 1.12

(each 3H, s). A typical fragmentation ion at m/z 220 [M-62-H₂O]⁺ and ¹³C-NMR spectral data suggested that the compound **8** was likely a sesquiterpenoid glucoside. The ¹H-NMR and ¹³C-NMR (see **Experimental**) indicated the sugar



Fig.2. Selected HMBC correlations of compound 1.

moiety was glucose. The proton and carbon signals of the anomeric center of the glucose ($\delta_{\rm H}$ 4.31, 1H, d, J = 7.9 Hz; $\delta_{\rm C}$ 100.7) suggested a β -configuration. The ¹H-NMR and ¹³C-NMR data of the aglycone moiety of compound **8** were closely matched with those of senecrassidiol (Bohlman and Ziesche, 1981; Iwabuchi *et al.*, 1990), except for the C-9 at δ 79.0 and the C-10 at δ 22.8, the former was down-field shifted ca. $D\delta$ 5, and the latter was up-field shifted ca. $D\delta$ 4 compared with those of senecrassidiol resulting from glycosylation, indicating that the sugar moiety was definitely linked to C-9. The structure of compound **8** was thus elucidated as senecrassidiol-9-O- β -D-glucopyranoside.

Compounds 2 and 3 were respectively identified as (24S)-3 β -methoxy-5 α -lanost-9(11)-ene-24,25-diol and (24S)- 5α -lanost-9(11)-ene-3 β ,24,25-triol on the basis of spectral data (Kutney et al., 1981). As no ¹³C-NMR spectral data have been reported for compounds 2 and 3 before, the detail assignments of ¹³C-NMR data for compounds 2 and **3** were thus made for the first time (Table 1). Compound **4** was identified as 3-oxo-lanost-9(11)-ene-24S,25-diol (Wada et al., 2001). The known serratene triterpenes, serrat-14-en- 3β ,21 β -diol (5) (Fang *et al.*, 1991) and 29-acetoxy-3 β methoxyserrat-14-en-21 α -ol (6) (Wada *et al.*, 2001) were identified by comparison of the spectral data with those reported. The known diterpenoid was identified as 9B,13Bendoperoxide-abieta-8 (14)-en-18-oic acid (7) by comparison of its ¹H- and ¹³C-NMR spectral data with those reported values of its methyl ester (Monaco et al., 1987; Barrero et al., 1991). This paper deals with the spectral data in its original form (see experimental).

The known lignans, icariside E4 (9) (Miyase *et al.*, 1989), (2*R*, 3*R*)-2,3-dihydro-7-hydroxy-2-(4'-hydroxy-3'methoxyphenyl)-3-hydroxymethyl-5-benzofuranpropanol 4'-O-(3-O-methyl- α -*L*-rhamnopyranoside) (10) (Pan and Lundgren, 1995), (2*R*, 3*R*)-2, 3-dihydro-7-hydroxy-2-(4'-hydroxy-3'-methoxyphenyl)-3-hydroxymethyl-5benzofuranpropanol-4'-O- α -*L*-rhamnopyranoside (11) (Popoff and Theander, 1975), and dihydrodehydrodiconiferyl alcohol 4'-O- β -*D*-glucoside (12) (Abe and Yamauchi, 1986) were identified by spectral data.

The known compounds, monoaryl glycosides β -hydroxypropiovanillone 3-O- β -*D*-glucopyranoside (**13**) (Anderson and Lundgren, 1988), 3,4-dimethoxyphenyl-2-O-(3-O-methyl- α -*L*-rhamnopyranosyl)- β -*D*-glucopyranoside (**14**) (Pan and Lundgren, 1995), and three known flavonoids taxifolin 3'-O- β -*D*-glucopyranoside (**15**), taxifolin (**16**) (Shen and Theander, 1985), and quercetin-3'-O- β -*D*-glucopyranoside (**17**) were also identified by comparison of their spectral data with those reported.

2 Experimental

2.1 General

Optical rotations were measured on a Perkin-Elmer 341 polarimeter (Na filter, λ =589 nm). IR spectra were recorded on a Perkin-Elmer 577 spectrometer. NMR spectra were recorded on a Bruker AM-400 (400 MHz) spectrometer with TMS as internal standard. Mass spectra including high-resolution mass spectra were recorded on a Finnigan MAT 95 mass spectrometer. All solvents used were of analytical grade (Shanghai Chemical Plant). Silica gel (200–300 mesh) was used for column chromatography, and pre-coated silica gel GF254 plates (Qingdao Marine Chemical Plant) were used for TLC. C-18 reversed-phase silica gel (150-200 mesh, Merck) and MCI GEL CHP20P (75–150 μ) (Mitsubishi Chemical Industry Ltd.) were also used for column chromatography.

2.2 Plant material

The root bark of *Taiwania flousiana* Gaussen was collected in Lichuan, Hubei, China (May, 2001) and was authenticated by Prof. ZHANG Chang-Gong (Tongji Medical College, Huazhong University of Sciences and Technology, Wuhan, China). A voucher specimen (20010102) has been deposited in the Faculty of Pharmacognosy, Tongji Medical College, Huazhong University of Sciences and Technology.

2.3 Extraction and isolation

The dried root bark (1.15 kg) was powdered and extracted with 95% ethanol to give crude extract (270 g). The crude was dissolved in H₂O and partitioned with petroleum ether and EtOAc successively to afford petroleum ether and EtOAc soluble fractions, respectively. The petroleum ether soluble part (21 g) was subjected to silica gel chromatography using a gradient mixture of petroleum ether-Me₂CO (from 1:0 to 1:1) as eluting solvent to give four fractions. Fraction 1 was mainly composed of waxy materials. Fraction 2 was separated by CC on silica gel with a gradient mixture of petroleum ether-CHCl₃ (1:1 to CHCl₃) to give compound 5 (55 mg), and two sub-fractions 2a and 2b. Sub-fraction 2a was subjected to a C-18 reversed-phase silica gel CC using a gradient solvent H₂O-Me₂CO to give compound 6 (2.3 mg). Sub-fraction 2b was separated by CC on silica gel with mixture of petroleum ether-Me₂CO (4:1) to give compounds 2 (42 mg) and 4 (49 mg). Fraction 3 was subjected to silica gel CC using petroleum ether with increasing amount of iso-PrOH as solvent to give two subfractions 3a and 3b, which were then subjected to C-18 reversed-phase silica gel CC to yield compounds 3 (8 mg) and 7 (12 mg), respectively. Fraction 4 was subjected to silica gel CC using petroleum ether with increasing amount of iso-PrOH as solvent to give a major fraction, which was further purified by C-18 reversed-phase silica gel CC to obtain compound 1 (4.3 mg).

The ethyl acetate soluble part (42 g) was subjected to silica gel CC using a gradient mixture of petroleum ether-Me₂CO (3:1 to Me₂CO) as solvent to give eight fractions. Fraction 4 was chromatographed over MCI-gel and silica gel columns to give compound 16 (40 mg). Fraction 7 was separated on column of MCI-gel, and then to column of C-18 reversed-phase silica gel to give compounds 10 (80 mg), 15 (650 mg) and 17 (120 mg). Fraction 8 was separated by column of MCI-gel and then silica gel CC with a gradient mixture of CHCl3-MeOH to give eight sub-fractions (8a to 8h). Sub-fraction 8b was purified over column of C-18 reversed-phase silica gel to give compound 9 (10 mg). Subfraction 8c was separated by silica gel CC to give a mixture of two compounds, which were then purified by preparative TLC to obtain compounds 11 (90 mg) and 12 (20 mg). Sub-fractions 8e, 8f and 8h were purified by Sephadex LH-20 to give compounds 8 (2.2 mg), 13 (9 mg) and 14 (15 mg), respectively.

2.4 Identification

Compound 1 White powder, $C_{31}H_{54}O_4$, $[\alpha]_D^{20}+42.0^{\circ}(c 0.14, CHCl_3)$. ¹H-NMR δ (in CDCl₃): 0.68 (3H, s, Me-18), 0.81 (3H, s, Me-30), 0.84 (3H, s, Me-28), 0.88 (1H, t, J = 6.9 Hz, H-5), 0.92 (3H, d, J = 6.5 Hz, Me-21), 0.98 (3H, s, Me-29), 1.07 (3H, s, Me-19), 1.11 (1H, m, H-23), 1.17 (3H, s, Me-26), 1.22 (3H, s, Me-27), 1.30 (1H, m, H-16), 1.31 (1H, m, H-2), 1.32 (1H, m, H-1), 1.40 (2H, m, H-15), 1.53 (1H, br d, J = 12.6 Hz, H-6), 1.63 (1H, m, H-20), 1.66 (1H, m, H-17), 1.79 (1H, m, H-23), 1.94 (1H, m, H-1), 1.98 (2H, m, H-16 and H-6), 1.99 (1H, m, H-2), 2.11 (1H, br d, J = 9.5 Hz, H-12), 2.17 (1H, br d, J = 10.6 Hz, H-8), 2.20 (1H, m, H-12), 2.34 (2H, m, H-22), 2.65 (1H, dd, J = 11.7, 4.1 Hz, H-3), 3.29 (1H, dd, J = 10.0, 1.7 Hz, H-24), 3.37 (3H, s, OMe), 3.69 (1H, dt, J = 10.6, 4.9 Hz, H-7), 5.30 (1H, d,

 $J=6.2 \text{ Hz}, \text{H-11}). {}^{13}\text{C-NMR}: \text{see Table 1. IR (KBr)} \nu_{\text{max}}: 3547, 3452, 2918, 1716, 1628, 1464, 1371, 1159, 1097, 1070, 978. EI-MS$ *m*/*z*: 490 (M⁺, 2), 472 (23), 457 (54), 454 (62), 439 (53), 425 (18), 414 (26), 407 (40), 399 (26), 367 (25), 343 (100), 327 (31), 285 (21), 260 (22), 225 (23), 183 (38), 173 (49), 159 (34), 151 (33), 147 (32), 145 (38), 133 (45), 123 (45), 121 (68), 109 (63). HR-EI-MS: 490.400 8 [M⁺] (calcd. 490.402 2 for C₃₁H₅₄O₄).

Compound 2 White powder, $C_{31}H_{54}O_3$, ¹H-NMR δ (in CDCl₃): 0.64 (3H, s, Me-18), 0.73 (3H, s, Me-28), 0.79 (3H, s, Me-30), 0.89 (3H, d, J = 6.4 Hz, Me-21), 0.96 (3H, s, Me-29), 1.02 (3H, s, Me-19), 1.13 (3H, s, Me-26), 1.21 (3H, s, Me-27), 2.64 (1H, dd, J = 11.4, 4.1 Hz, H-3), 3.28 (1H, dd, J = 10.1, 1.8 Hz, H-24), 3.36 (3H, s, OMe), 5.21 (1H, d, J = 6.0 Hz, H-11). ¹³C-NMR: see Table 1.

Compound 3 White powder, $C_{30}H_{52}O_3$, ¹H-NMR δ (in CDCl₃): 0.65 (3H, s, Me-18), 0.74 (3H, s, Me-28), 0.82 (3H, s, Me-30), 0.90 (3H, d, J = 6.5 Hz, Me-21), 0.98 (3H, s, Me-29), 1.04 (3H, s, Me-19), 1.16 (3H, s, Me-26), 1.22 (3H, s, Me-27), 3.22 (1H, dd, J = 11.4, 4.2 Hz, H-3), 3.29 (1H, dd, J = 10.2, 2.1 Hz, H-24), 5.22 (1H, d, J = 6.2 Hz, H-11). ¹³C-NMR: see Table 1.

Compound 4 White powder, $C_{30}H_{50}O_3$, ¹H-NMR δ (in CDCl₃): 0.65 (3H, s, Me-18), 0.74 (3H, s, Me-28), 0.90 (3H, d, J = 6.4 Hz, Me-21), 1.06 (3H, s, Me-30), 1.07 (3H, s, Me-29), 1.22 (3H, s, Me-26), 1.22 (3H, s, Me-27), 3.29 (1H, br d, J = 9.3 Hz, H-24), 5.28 (1H, d, J = 5.9 Hz, H-11). ¹³C-NMR: see Table 1.

Compound 5 White powder, $C_{30}H_{50}O_2$, ¹H-NMR δ (in CDCl₃): 0.67 (3H, s, Me-28), 0.74 (3H, s, Me-24), 0.77 (3H, s, Me-25), 0.81 (3H, s, Me-26), 0.86 (3H, s, Me-29), 0.91 (3H, s, Me-30), 0.95 (3H, s, Me-23), 3.17 (1H, dd, J = 11.8, 4.9 Hz, H-3), 3.43 (1H, dd, J = 2.9, 2.4 Hz, H-21), 5.30 (1H, br s, H-15). ¹³C-NMR δ (in CDCl₃): 13.3 (C-28), 15.4 (C-26), 15.7 (C-23), 18.9 (C-6), 19.8 (C-24), 21.8 (C-30), 24.0 (C-16), 25.2 (C-2), 25.4 (C-11), 27.2 (C-12), 27.6 (C-20), 27.7 (C-29), 28.1 (C-25), 31.2 (C-19), 35.9 (C-10), 37.1 (C-18), 37.5 (C-22), 38.2 (C-4), 38.6 (C-1), 39.0 (C-8), 43.4 (C-17), 45.2 (C-7), 55.8 (C-5), 56.3 (C-27), 56.9 (C-13), 62.9 (C-9), 76.2 (C-21), 78.9 (C-3), 122.1 (C-15), 138.5 (C-14).

Compound 6 White powder, $C_{33}H_{54}O_4$, ¹H-NMR δ (in CDCl₃): 0.65 (3H, s, Me-18), 0.74 (3H, s, Me-28), 0.82 (3H, s, Me-30), 0.90 (3H, d, J = 6.5 Hz, Me-21), 0.98 (3H, s, Me-29), 1.04 (3H, s, Me-19), 1.16 (3H, s, Me-26), 1.22 (3H, s, Me-27), 3.22 (1H, dd, J = 11.4, 4.2 Hz, H-3), 3.29 (1H, dd, J = 10.2, 2.1 Hz, H-24), 5.22 (1H, d, J = 6.2 Hz, H-11). ¹³C-NMR δ (in CDCl₃): 14.0 (C-28), 15.7 (C-25), 16.2 (C-24), 18.8 (C-6), 19.8 (C-26), 21.1 (OAc), 21.6 (C-30), 22.4 (C-2), 24.3 (C-16), 25.5 (C-11), 27.2 (C-12), 27.8 (C-20), 28.1 (C-23), 35.9 (C-18), 37.2

(C-8), 37.3 (C-19), 38.2 (C-10), 38.5 (C-1), 38.9 (C-4), 42.0 (C-22), 45.2 (C-7), 50.6 (C-17), 56.0 (C-27), 56.3 (C-5), 57.1 (C-13), 57.5 (OMe), 62.8 (C-9), 64.7 (C-29), 79.4 (C-21), 88.5 (C-3), 121.6 (C-15), 138.3 (C-14), 171.0 (OAc).

Compound 7 White powder, $C_{20}H_{30}O_4$, ¹H-NMR δ (in Me₂CO-*d*₆): 0.93 (3H, d, *J* = 6.9 Hz, H-16), 0.95 (3H, d, *J* = 6.9 Hz, H-17), 1.08 (3H, s, H-20), 1.30 (3H, s, H-19), 6.17 (1H, t, *J* = 2.2 Hz, H-14). ¹³C-NMR δ (in Me₂CO-*d*₆): 17.1 (C-17), 17.3 (C-16), 17.6 (C-19), 18.0 (C-2), 18.1 (C-20), 21.0 (C-6), 24.1 (C-11), 25.6 (C-12), 28.5 (C-7), 32.6 (C-15), 34.1 (C-1), 37.8 (C-3), 38.9 (C-10), 40.7 (C-5), 47.1 (C-4), 79.6 (C-13), 82.0 (C-9), 127.1 (C-14), 144.4 (C-8), 179.3 (C-18).

Compound 8 Pale gum, $C_{21}H_{36}O_7$, $[\alpha]_D^{20}$ - 34.0° (*c* 0.18, CH₃OH). ¹H-NMR δ (in CD₃OD): 0.87 (3H, s, Me-13), 0.94 (3H, s, Me-15), 1.03 (1H, dd, J = 13.9, 12.6 Hz, H-8), 1.12 (3H, s, Me-14), 1.29 (1H, m, H-11), 1.37 (1H, br d, J = 13.6 Hz, H-12), 1.38 (2H, m, H-6), 1.45 (1H, m, H-8), 1.57 (1H, m, H-3), 1.63 (1H, m, H-10), 1.77 (1H, m, H-5), 1.78 (1H, m, H-12), 1.79 (1H, m, H-3), 1.80 (1H, m, H-10), 1.94 (1H, m, H-11), 2.42 (1H, dd, J=11.4, 7.5, 4.2 Hz, H-2), 3.19 (1H, m, H-2'), 3.26 (1H, m, H-3'), 3.30(1H, m, H-4'), 3.37(1H, m, H-9), 3.38(1H, m, H-5'), 3.62 (1H, dd, J = 11.6, 5.4 Hz, H-6'), 3.78 (1H, dd, J = 11.6, 2.6 Hz, H-6'), 4.31 (1H, d, J=7.9 Hz, H-1'). ¹³C-NMR δ (in CD₃OD): 22.6 (C-6), 22.8 (C-10), 24.6 (C-13), 29.0 (C-14), 31.0 (C-15), 34.2 (C-4), 35.5 (C-3), 36.6 (C-11), 37.0 (C-7), 37.4(C-8), 38.8(C-12), 42.6(C-2), 49.7(C-5), 62.8(C-6'), 71.7 (C-4'), 72.4 (C-1), 74.7 (C-2'), 77.1 (C-3'), 77.7 (C-5'), 79.0 (C-9), 100.7 (C-1'). IR (KBr) v_{max}: 3415, 2926, 1464, 1383, 1078, 1041. EI-MS m/z: 382 ([M-H₂O]⁺, 7), 327 (12), 220 (12), 203 (100), 182 (24), 165 (35), 147 (46), 123 (86), 119 (24), 107 (36). HR-EI-MS: 382.237 0 (calcd. 382.235 5 for C₂₁H₃₄O₆ ([M- $H_2O]^+)).$

Compound 9 White powder, $C_{26}H_{34}O_{10}$, ¹H-NMR δ (in CD₃OD): 1.05 (3H, d, J = 6.2 Hz, Me-6″), 1.64 (2H, m, H-8), 2.45 (2H, dd, J = 8.1, 7.3 Hz, H-7), 3.62 (3H, s, OMe), 3.68 (3H, s, OMe), 5.17 (1H, d, J = 1.5 Hz, H-1″), 5.38 (1H, d, J = 5.9 Hz, H-7′), 6.54 (1H, s, H-6), 6.56 (1H, s, H-2), 6.74 (1H, dd, J = 8.4, 1.8 Hz, H-6′), 6.86 (1H, d, J = 1.8 Hz, H-2′), 6.91 (1H, d, J = 8.4 Hz, H-5′). ¹³C-NMR δ (in CD₃OD): 18.4 (C-6″), 33.4 (C-7), 36.3 (C-8), 56.1 (C-8′), 56.9 (OMe), 57.2 (OMe), 62.7 (C-9), 65.5 (C-9′), 71.3 (C-5″), 72.5 (C-2″), 72.6 (C-3″), 74.3 (C-4″), 89.0 (C-7′), 101.8 (C-1″), 111.7 (C-2′), 114.5 (C-2), 118.4 (C-6), 119.6 (C-6′), 120.0 (C-5′), 130.0 (C-1), 137.5 (C-5), 139.2 (C-1′), 145.7 (C-3), 147.0 (C-4), 147.9 (C-4′), 152.5 (C-3′).

Compound 10 White powder, $C_{26}H_{34}O_{10}$, ¹H-NMR δ (in Me₂CO-*d*₆): 1.16 (2H, d, J = 6.2 Hz, Me-6"), 1.74 (2H, m, H-8), 2.52 (2H, dd, J = 8.1, 7.3 Hz, H-7), 3.44 (3H, s, 3"-OMe), 3.81 (3H, s, 3'-OMe), 4.27 (1H, t, J = 2.2 Hz, H-2"), 5.39 (1H,

d, J = 1.5 Hz, H-1″), 5.57 (1H, d, J = 5.9 Hz, H-7′), 6.59 (1H, s, H-6), 6.61 (1H, d, J = 1.5 Hz, H-2), 6.96 (1H, dd, J = 8.4, 1.8 Hz, H-6′), 7.10 (1H, d, J = 8.4 Hz, H-5′), 7.12 (1H, d, J = 1.8 Hz, H-2′). ¹³C-NMR δ (in Me₂CO- d_6): 17.7 (C-6″), 31.9 (C-7), 35.2 (C-8), 55.0 (C-8′), 55.9 (3′-OMe), 56.8 (3″-OMe), 61.3 (C-9), 64.4 (C-9′), 67.1 (C-2″), 69.9 (C-5″), 71.5 (C-4″), 81.3 (C-3″), 87.1 (C-7′), 100.6 (C-1″), 110.8 (C-2′), 115.8 (C-2), 116.4 (C-6), 118.3 (C-6′), 119.1 (C-5′), 129.0 (C-1), 136.0 (C-5), 138.4 (C-1′), 141.3 (C-3), 145.5 (C-4), 145.5 (C-4′), 151.2 (C-3′).

Compound 11 White powder, $C_{25}H_{32}O_{10}$, ¹H-NMR δ (in Me₂CO-*d*₆): 1.17 (3H, d, J = 6.2 Hz, Me-6″), 1.74 (2H, m, H-8), 2.53 (2H, dd, J = 8.0, 7.3 Hz, H-7), 3.80 (3H, s, 3'-OMe), 5.37 (1H, s, H-1″), 5.56 (1H, d, J = 5.9 Hz, H-7), 6.60 (1H, s, H-6), 6.61 (1H, s, H-2), 6.95 (1H, d, J = 8.0 Hz, H-6′), 7.10 (1H, d, J = 8.0 Hz, H-5′), 7.11 (1H, s, H-2′). ¹³C-NMR δ (in Me₂CO-*d*₆): 17.7 (C-6″), 32.1 (C-7), 35.5 (C-8), 55.0 (C-8′), 56.0 (3′-OMe), 61.5 (C-9), 64.6 (C-9′), 69.9 (C-5″), 71.4 (C-2″), 71.9 (C-3″), 73.2 (C-4″), 87.4 (C-7′), 100.5 (C-1″), 111.1 (C-2′), 116.0 (C-2), 116.6 (C-6), 118.6 (C-6′), 119.0 (C-5′), 129.2 (C-1), 136.1 (C-5), 138.1 (C-1′), 141.3 (C-3), 145.6 (C-4), 145.9 (C-4′), 151.3 (C-3′).

Compound 12 White powder, $C_{25}H_{32}O_{11}$, ¹H-NMR δ (in CD₃OD): 1.80 (2H, m, H-8), 2.62 (2H, dd, J=8.1, 7.3 Hz, H-7), 3.82 (3H, s, OMe), 3.85 (3H, s, OMe), 4.84 (1H, d, J=7.2 Hz, H-1"), 5.54 (1H, d, J=5.9 Hz, H-7'), 6.71 (1H, s, H-6), 6.73 (1H, s, H-2), 6.92 (1H, dd, J=8.4, 1.8 Hz, H-6'), 7.02 (1H, d, J=1.8 Hz, H-2'), 7.13 (1H, d, J=8.4 Hz, H-5'). ¹³C-NMR δ (in CD₃OD): 33.4 (C-7), 36.3 (C-8), 56.1 (C-8'), 57.2 (OMe), 57.2 (OMe), 62.7 (C-9), 63.0 (C-6"), 65.5 (C-9'), 71.8 (C-4"), 75.4 (C-2"), 78.3 (C-5"), 78.6 (C-3"), 89.0 (C-7'), 103.2 (C-1"), 111.7 (C-2'), 114.6 (C-2), 118.4 (C-6), 118.4 (C-6'), 119.9 (C-5'), 130.1 (C-1), 137.6 (C-5), 138.8 (C-1'), 145.7 (C-3), 148.1 (C-4), 148.1 (C-4'), 151.4 (C-3').

Compound 13 White powder, $C_{16}H_{22}O_9$, ¹H-NMR δ (in CD₃OD): 3.17 (1H, dd, J=8.8, 8.1 Hz, H-2"), 3.30 (overlapped with the signals of solvents, H-3" and H-4"), 3.34 (2H, s, H-2), 3.38 (1H, m, H-5"), 3.68 (1H, dd, J=13.2, 4.4 Hz, H-6"), 3.85 (1H, dd, J=13.2, 5.9 Hz, H-6"), 3.89 (3H, s, OMe), 3.97 (1H, m, H-3), 4.26 (1H, m, H-3), 4.34 (1H, d, J=8.1 Hz, H-1"), 6.83 (1H, d, J=8.4 Hz, H-5'), 7.53 (1H, s, H-2'), 7.59 (1H, d, J= 8.4 Hz, H-6'). ¹³C-NMR δ (in CD₃OD): 39.6 (C-2), 56.8 (OMe), 63.9 (C-6"), 66.8 (C-3), 71.9 (C-4"), 75.4 (C-2"), 78.4 (C-5"), 104.9 (C-1"), 112.3 (C-5'), 116.7 (C-2'), 125.7 (C-6'), 137.1 (C-1'), 150.0 (C-4'), 155.7 (C-3'), 199.9 (C-1).

Compound 14 White powder, $C_{19}H_{28}O_{12}$, ¹H-NMR δ (in CD₃OD): 1.30 (3H, d, J = 6.3 Hz, Me-6"), 3.31 (1H, m, H-3'), 3.35 (1H, m, H-4'), 3.40 (3H, s, 3"-OMe), 3.41 (1H, m, H-

5'), 3.47 (1H, t, J=9.5 Hz, H-4"), 3.58 (1H, dd, J=9.1, 8.4 Hz, H-3'), 3.63 (1H, d, J=7.7 Hz, H-2'), 3.68 (1H, br d, J=12.7 Hz, H-6'), 3.77 (3H, s, 4-OMe), 3.81 (3H, s, 3-OMe), 3.90 (1H, dd, 12.1, 2.2 Hz, H-6'), 4.12 (1H, m, H-5"), 4.15 (1H, m, H-2"), 4.89 (1H, d, J=7.8 Hz, H-1'), 5.32 (1H, d, J=1.8 Hz, H-1"), 6.64 (1H, dd, J=8.8, 2.9 Hz, H-6), 6.75 (1H, d, J=2.9 Hz, H-2), 6.85 (1H, d, J=8.8 Hz, H-5). ¹³C-NMR δ (in CD₃OD): 18.7 (C-6"), 56.9 (3-OMe), 57.7 (4-OMe), 57.7 (3'-OMe), 63.1 (C-6'), 68.5 (C-2"), 70.4 (C-5"), 72.1 (C-4'), 73.2 (C-4"), 78.6 (C-5'), 79.4 (C-2'), 79.7 (C-3'), 82.4 (C-3"), 102.0 (C-1'), 102.8(C-1"), 104.0(C-2), 108.9(C-5), 114.5 (C-6), 146.4(C-1), 151.7 (C-4), 154.2(C-3).

Compound 15 White powder, $C_{21}H_{22}O_{12}$, ¹H-NMR δ (in CD₃OD): 3.21 – 3.38 (4H, m, H-2" – H-5"), 3.50 (1H, dd, J = 12.1, 5.9 Hz, H-6"), 3.73 (1H, dd, J = 12.1, 1.8 Hz, H-6"), 4.41 (1H, d, J = 11.7 Hz, H-3), 4.67 (1H, d, J = 7.3 Hz, H-1"), 4.81 (1H, d, J = 11.7 Hz, H-2), 5.71 (1H, d, J = 2.2 Hz, H-6), 5.75 (1H, d, J = 2.2 Hz, H-8), 6.73 (1H, d, J = 8.4 Hz, H-5'), 6.93 (1H, dd, J = 8.4, 1.8 Hz, H-6'), 7.21 (1H, d, J = 1.8 Hz, H-2'). ¹³C-NMR δ (in CD₃OD): 63.0 (C-6"), 71.9 (C-4"), 73.9 (C-3), 75.3 (C-2"), 78.0 (C-3"), 78.7 (C-5"), 85.3 (C-2), 96.8 (C-8), 97.8 (C-6), 102.3 (C-10), 104.4 (C-1"), 117.4 (C-5'), 118.6 (C-2'), 125.1 (C-6'), 130.4 (C-1'), 147.0 (C-3'), 149.4 (C-4'), 164.8 (C-9), 165.7 (C-7), 169.1 (C-5), 198.8 (C-4).

Compound 16 White powder, $C_{15}H_{12}O_7$, ¹H-NMR δ (in Me₂CO-*d*₆): 4.59 (1H, d, *J* = 11.4 Hz, H-3) 4.99 (1H, d, *J* = 11.4 Hz, H-2), 5.92 (1H, d, *J* = 1.8 Hz, H-6), 5.97 (1H, d, *J* = 1.8 Hz, H-8), 6.87 (2H, m, H-5', H-6'), 7.05 (1H, d, *J* = 1.1 Hz, H-2'). ¹³C-NMR δ (in DMCO-d₆): 72.6 (C-3), 84.1 (C-2), 95.6 (C-8), 96.6 (C-6), 100.9 (C-10), 115.4 (C-2'), 115.4 (C-5'), 120.3 (C-6'), 129.1 (C-1'), 145.4 (C-3'), 146.2 (C-4'), 163.6 (C-9), 164.2 (C-7), 167.7 (C-5), 197.7 (C-4).

Compound 17 Yellow powder, $C_{21}H_{20}O_{12}$, ¹H-NMR δ (in CD₃OD): 3.33 – 3.46 (4H, m, H-2" – H-5"), 3.64 (1H, dd, J = 11.7, 3.3 Hz, H-6"), 3.81 (1H, br d, J = 11.7 Hz, H-6"), 4.71 (1H, d, J = 7.0 Hz, H-1"), 5.99 (1H, d, J = 1.7 Hz, H-6), 6.25 (1H, d, J = 1.7 Hz, H-8), 6.78 (1H, d, J = 8.4 Hz, H-5'), 7.69 (1H, d, J = 8.4 Hz, H-6'), 7.96 (1H, s, H-2'). ¹³C-NMR δ (in CD₃OD): 62.9 (C-6"), 71.7 (C-4"), 75.3 (C-2"), 78.1 (C-3"), 78.9 (C-5"), 95.1 (C-8), 99.9 (C-6), 104.9 (C-1"), 104.9 (C-10), 117.6 (C-2'), 118.4 (C-5'), 124.8 (C-1'), 125.5 (C-6'), 138.0 (C-3), 147.2 (C-2), 147.8 (C-3'), 150.9 (C-4'), 158.7 (C-9), 163.0 (C-5), 166.5 (C-7), 177.9 (C-4).

Acknowledgements: The authors are grateful to Prof. ZHANG Chang-Gong (Tongji Medical College, Huazhong University of Sciences and Technology, Wuhan, China) for the plant material collection and identification.

References:

- Abe F, Yamauchi T. 1986. Lignans from *Trachelospermum* asiaticum (Tracheolospermum). Chem Pharm Bull, 34: 4340–4345.
- Andersson R, Lundgren L. 1988. Monoryl and cyclohexenone glycosides from needles of *Pinus sylvestris*. *Phytochemistry*, 27: 559–562.
- Barrero A R, Sanchez J F, Alvarez-Manzaneda R E J, Muñoz Dorado M. 1991. Endoperoxide diterpenoids and other constituents from *Abies marocana*. *Phytochemistry*, **30**: 593–562.
- Bohlmann F, Ziesche J. 1981. Sesquiterpenes from three Senecio species. Phytochemistry, 20: 469–472.
- Chang S T, Wang S Y, Wu C L, Su Y C, Kuo Y H. 1999. Antifungal compounds in the ethyl acetate soluble fraction of the extractives of *Taiwania (Taiwania cryptomerioides* Hayata) heartwood. *Holzforschung*, **53**: 487–490.
- Cheng Y S, Kuo Y H, Lin Y T. 1967. Extractive components from the wood of *Taiwania cryptomerioides* Hayata: the structure of "T-cadinol" and "T-muurolol". *J Chem Soc Chem Commun*, (12): 565–566.
- Fang J M, Tsai W Y, Cheng Y S. 1991. Serratene triterpenes from *Pinus armandii* bark. *Phytochemistry*, **30**: 1333–1336.
- He K, Zeng L, Shi G, Zhao G X, Kozlowski J F, McLaughlin J L.
 1997. Bioactive compounds from *Taiwania cryptomerioides*. *J Nat Prod*, **60**: 38–40.
- Iwabuchi H, Kato N, Yoshikura M. 1990. Studies on the sesquiterpenoids of *Panax gingseng* C. A. Meyer. . *Chem Pharm Bull*, 38: 1405–1407.
- Kamil M, Ilyas M, Rahman W, Hasaka N, Okigawa M, Kawano N. 1977. Taiwaniaflavone: a new series of naturally occurring biflavones from *Taiwania cryptomerioides*. *Chem Ind*, (14): 160.
- Kamil M, Ilyas M, Rahman W, Hasaka N, Okigawa M, Kawano N. 1981. Taiwaniaflavone and its derivatives: a new series of biflavones from *Taiwania cryptomerioides* Hayata. *J Chem Soc Perkin Trans*, (2): 553–559.
- Kuo Y H, Cheng Y S, Lin Y T. 1969. Extractive components from the wood of *Taiwania cryptomerioides* Hayata: three new sesquiterpene alcohols, muurolane-3-ene-9β-ol-2-one, muurolane-2α,9β-diol, and muurolane-2β,9β-diol-3-one. *Tetrahedron Lett*, **10**: 2375–2377.
- Kutney J P, Eigendorf G, Worth B A, Rowe J W, Conner A H, Nagasampagi B A. 1981. New triterpenes from the bark of western white pine (*Pinus monticola* Dougl.). *Helv Chim Acta*, 64: 1183–1207.
- Lin W H, Fang J M, Cheng Y S. 1995. Uncommon diterpenoids with the skeleton of six-five-six fused-rings from *Taiwania cryptomerioids*. *Phytochemistry*, **40**: 871–873.

- Lin W H, Fang J M, Cheng Y S. 1996. Diterpenoids and related cycloadducts from *Taiwania cryptomerioids*. *Phytochemistry*, 42: 1657–1663.
- Lin W H, Fang J M, Cheng Y S. 1997. Cycloadducts of terpene quinines from *Taiwania cryptomerioids*. *Phytochemistry*, 46: 169–173.
- Lin W H, Fang J M, Cheng Y S. 1998a. Diterpenoids and steroids from *Taiwania cryptomerioids*. *Phytochemistry*, **48**: 1391– 1397.
- Lin W H, Fang J M, Cheng Y S. 1998b. Lignans from *Taiwania* cryptomerioids. Phytochemistry, **50**: 653–658.
- Lin Y T, Lo T B, Wang K T, Weinstein W. 1967. Phytochemical studies. . The structures of taiwanins C and E. *Tetrahedron Lett*, 8: 849–852.
- Miyase T, Ueno A, Takizawa N, Kobayashi H, Oguchi H. 1989. Ionone and lignan glycosides from *Epimedium diphyllum*. *Phytochemistry*, **28**: 3483–3485.

- Monaco P, Parrilli M, Previtera L. 1987. Two endoperoxide diterpenes from *Elodes canadensis*. *Tetrahedron Lett*, 28: 4609–4610.
- Pan H, Lundgren L. 1995. Phenolic extractives from root bark of Picea abies. Phytochemistry, 39: 1423–1428.
- Popoff T, Theander O. 1975. Two glycosides of a new dilignol from *Pinus silvestris*. *Phytochemistry*, **14**: 2065–2066.
- Tanaka R, Matshunaga S. 1990. Veitchiolide, a tetracyclic triterpene lactone from *Abies veitchii*. *Phytochemistry*, 29: 3267–3269
- Shen Z, Theander O. 1985. Flavonoid glycosides from needles of Pinus massoniana. Phytochemistry, 24: 155–158.
- Wada S, Iida A, Tanaka R. 2001. Triterpene constituents from the stem bark of *Pinus luchuensis* and their DNA topoisomerase inhibitory effect. *Planta Med*, 67: 659– 664.

(Managing editor: WANG Wei)