www.nature.com/onc

Bcl-2 family proteins and cancer

KW Yip and JC Reed

Burnham Institute for Medical Research, La Jolla, CA, USA

BCL-2 was the first anti-death gene discovered, a milestone with far reaching implications for tumor biology. Multiple members of the human Bcl-2 family of apoptosis-regulating proteins have been identified, including six antiapoptotic, three structurally similar proapoptotic proteins and several structurally diverse proapoptotic interacting proteins that operate as upstream agonists or antagonists. These proteins, in turn, are regulated through myriad post-translational modifications and interactions with other proteins. Bcl-2-family proteins regulate all major types of cell death, including apoptosis, necrosis and autophagy, thus operating as nodal points at the convergence of multiple pathways with broad relevance to oncology. Experimental therapies targeting Bcl-2family mRNAs or proteins are currently in clinical testing, raising hopes that a new class of anticancer drugs may soon be available.

Oncogene (2008) 27, 6398–6406; doi:10.1038/onc.2008.307

Keywords: Bcl-2; cancer; apoptosis; autophagy; necrosis; therapeutics

Introduction

The average adult human produces and in parallel eradicates ~ 60 billion cells daily, with new cells formed by cell division and old cells eliminated principally by apoptosis, thus striking a balance under normal circumstances. The ability to control cell numbers at both the points of entry and exit allows flexibility to more rapidly respond to stress, injury and physiological cues. However, it also creates a liability in terms of neoplasia, as genes that normally suppress or induce physiological cell death often become dysregulated in cancers, with defective cell death mechanisms now recognized as one of the six hallmarks of cancer (Hanahan and Weinberg, 2000). Bcl-2-family proteins play central roles in cell death regulation and are capable of regulating diverse cell death mechanisms that encompass apoptosis, necrosis and autophagy (Cory et al., 2003; Levine and Kroemer, 2008; Reed, 2008). Alterations in their expression and function contribute to the pathogenesis and progression of human cancers,

Correspondence: JC Reed, Burnham Institute for Medical Research, La Jolla, CA 92037, USA. E-mail: reedoffice@burnham.org thus providing targets for drug discovery that are currently being explored in human clinical trials.

Altered expression of BCL-2-family genes in cancer

Abundant examples exist where the regulation of genes encoding either antiapoptotic or proapoptotic Bcl-2family proteins is altered in cancers. In fact, the founding member of the gene family, BCL-2, was discovered because of its involvement in t(14;18)chromosomal translocations observed in non-Hodgkin's lymphomas (Tsujimoto et al., 1985). In t(14;18) translocations, the BCL-2 gene from chromosome 18 becomes fused with the immunoglobulin heavy-chain locus on chromosome 14, bringing the juxtaposed BCL-2 gene under the control of the immunoglobulin heavy-chain enhancer, and thereby dysregulating BCL-2 gene expression at a transcriptional level. In addition to chromosomal translocations as a mechanism for activation of the BCL-2 gene in human malignancies, BCL-2 gene amplification is also found in non-Hodgkin's lymphomas and small cell lung cancers (Ikegaki et al., 1994; Monni et al., 1997). Besides changes to BCL-2 gene structure or copy number, many additional mechanisms contribute to elevated gene expression, which is estimated to occur in perhaps as many as half of all human cancers. Among the contributing mechanisms are (a) loss of endogenous microRNAs (miRs) that normally repress BCL-2 gene expression (Cimmino et al., 2005), which has been documented in chronic lymphocytic leukemia, where the genes encoding miR15 and miR16 become deleted or inactivated by mutations in >70% of these leukemia, and (b) gene hypomethylation, implying altered epigenetic regulation of BCL-2 in some malignancies (Hanada et al., 1993). Altered expression of other antiapoptotic members of the BCL-2 gene family has also been documented in human cancers and leukemias (see for example, Tron et al., 1995; Krajewski et al., 1995a, b; Brousset et al., 1996; Khanna et al., 1996; Krajewska et al., 1996a, 2008; Yip et al., 2006), although no somatic mutations have been discovered in these genes to date. Loss of miR-29, which represses antiapoptotic family member MCL-1, can occur in chronic lymphocytic leukemia and colon cancers (Calin et al., 2005; Cummins et al., 2006; Mott et al., 2007), suggesting at least one responsible mechanism for elevated expression of relatives of BCL-2.

Defects in the expression of proapoptotic members of the BCL-2 family also occur in cancer, resulting in loss of the tumor suppressor function of these killer genes. The best documented is BAX, where homozygous deletions or inactivating mutations have been identified, particularly in cancers that arise with microsatellite instability because of defective DNA mismatch repair (Rampino et al., 1997; Meijerink et al., 1998). In this regard, the human BAX gene contains a homopolymeric stretch of eight guanosine residues in the sense strand that is a target for frame-shift mutations. Defective expression of proapoptotic BCL-2-family genes also occurs in the setting of loss of p53 function. Among the direct targets of the p53 transcription factor are BAX, BID, PUMA and NOXA (Miyashita et al., 1994; Miyashita and Reed, 1995; Oda et al., 2000; Sax et al., 2002; Yu et al., 2003), thus demonstrating strong connections between genome surveillance by p53 and cell death genes of the BCL-2 family. More recently, cytosolic interactions of p53 protein with pro- and antiapoptotic Bcl-2-family proteins have been observed, directly modulating the bioactivities of the p21-Bax and p26-Bcl-2 proteins, and suggesting that p53 regulates the Bcl-2 family at both transcriptional and post-transcriptional levels (Chipuk et al., 2004; Deng et al., 2006). The activities of additional proapoptotic members of the Bcl-2 family are also suppressed through post-translational modifications. For example, proapoptotic protein BAD is phosphorylated by Akt (PKB) and other protein kinases known to be hyperactive in cancers, resulting in its sequestration by 14-3-3 (Zha et al., 1996).

Bcl-2 and resistance to chemotherapy

Overexpression of the Bcl-2 and related antiapoptotic proteins has been demonstrated to inhibit cell death induced by many stimuli, including growth factor deprivation, hypoxia and oxidative stress. However, it is the ability of antiapoptotic Bcl-2-family proteins to suppress cell death induced by cytotoxic anticancer drugs that makes these proteins particularly interesting as potential targets for cancer drug discovery. Regardless of the primary mode of action, whether single or double-strand DNA breaks, whether microtubule depolymerization or aggregation, whether nuclear hormone receptor activation (glucocorticoid receptor) or inhibition (estrogen and androgen receptors), essentially all traditional anticancer drugs appear to depend in large measure on Bcl-2/Bax-dependent mechanisms for killing cancer cells (reviewed by Debatin et al., 2002; Reed, 2008). Thus, Bcl-2 operates at a distal point in a conserved cell death pathway utilized by most anticancer drugs, constituting a form of intrinsic chemoresistance, distinct from the previously identified mechanisms involving drug efflux, drug metabolism, drug inactivation and related mechanisms. This observation presumably explains why expression of a variety of Bcl-2-family proteins has been shown to be of prognostic significance for many types of cancer and

6300

leukemia treated by chemotherapy (see for example, Yunis *et al.*, 1989; Tang *et al.*, 1994; Hermine *et al.*, 1996; Hill *et al.*, 1996; Krajewska *et al.*, 1996a, b, 2008; Gascoyne *et al.*, 1997; Pedersen *et al.*, 2002).

Cellular functions of Bcl-2-family proteins: guardians of organellar cell life and death

Core members of the Bcl-2 family share structural similarity with the pore-forming domains of bacterial toxins, emphasizing the relevance of these proteins to membrane biology (reviewed by Schendel *et al.*, 1998). Several Bcl-2 family proteins insert into intracellular membranes, particularly membranes of mitochondria and endoplasmic reticulum (ER), operating as guardians of these organelles.

Mitochondria

The central pathway involved in daily ('normal') programmed cell death in most tissues involves mitochondria, energy-producing organelles that play critical roles in both cell life and death (Green and Kroemer, 2004). Several Bcl-2-family proteins, both antiapoptotic and proapoptotic, have C-terminal transmembrane domains that insert in the outer membrane of mitochondria. Proapoptotic Bcl-2-family proteins such as Bax and Bak induce mitochondrial outer membrane permeabilization (MOMP), causing the release of caspaseactivating proteins and other cell death mediators, whereas antiapoptotic proteins such as Bcl-2 serve as guardians of the outer membrane and preserve its integrity by opposing Bax and Bak (Figure 1a). Induction of MOMP correlates with oligomerization of Bax and/or Bak in the outer mitochondrial membrane, which is opposed by antiapoptotic proteins such as Bcl-2 and Bcl-X_L (Eskes et al., 2000; Wei et al., 2000; Chipuk and Green, 2008). In fact, these observations have been reconstituted in synthetic liposomes, showing that Bax can be induced to oligomerize, permeabilizing liposomes in vitro, through a Bcl-X_L-inhibitable mechanism (Kuwana et al., 2002). Thus, Bax and structurally related proapoptotic proteins (Bak; Bok) are hypothesized to form proteinacous or lipidic pores upon oligomerization in mitochondrial membranes, alone or in conjunction with resident mitochondrial proteins, thereby allowing escape of various apoptogenic proteins from these organelles (reviewed by Reed, 2006; Chipuk and Green, 2008). Gene ablation studies in mice have demonstrated that either Bax or Bak is necessary for MOMP, establishing these pore-forming proapoptotic proteins as the distal elements controlling permeability of mitochondrial membranes (Wei et al., 2001), and thus the governors of life/decisions in the mammalian mitochondrial pathway for cell death.

Mitochondria induce apoptosis by releasing proteins that participate in caspase activation (for example, cytochrome c) and that neutralize endogenous inhibitors of caspases (for example, SMAC; OMI/Htra2, which inhibit IAPs), as well as unleashing several mediators of



Bcl-X (Atg6) **Figure 1** Cellular phenotypes of Bcl-2-family proteins. Bcl-2 family proteins (*in boxes*) insert into membranes of mitochondria (a) and endoplasmic reticulum (ER) (b), promoting life (green) or death (*red*) cellular events. Bcl-2-family proteins do not insert directly into membranes of lysosomes (c), but do affect lysosomalrelated processes such as autophagy (*blue*). See text for details. Dotted lines represent transcriptional regulation, thick lines (for example, between Irel and XBP-1) represent mRNA splicing alterations. Note that some Bcl-2-family proteins are not included

here, such as Bok and Bcl-rambo.

caspase-independent necrotic cell death (Reed, 2002). For example, MOMP releases several proteins that contribute to non-apoptotic cell death, including DNase, endonuclease G and AIF, a flavoprotein reported to enter the nucleus and promote genome destruction (Penninger and Kroemer, 2003). MOMP also results secondarily in elaboration of reactive oxygen species, causing lipid peroxidation and membrane damage, which impair normal ion-homeostasis, causing cellular swelling and plasma membrane rupture, as well as rupture of lysosomes and release of hydrolytic enzymes that destroy proteins, nucleic acids and lipids.

Cross-talk of the mitochondrial pathway with other cell death pathways probably occurs through a variety of mechanisms. A prominent example utilized by TNFfamily death receptors involves caspase-mediated cleavage and activation of Bid, which then targets mitochondrial membranes where it binds Bcl-2/Baxfamily proteins and modulates their activities (Korsmeyer *et al.*, 2000). Integration of mitochondria into cell death pathways initiated from other organelles such as ER and lysosomes has also been documented (Ferri and Kroemer, 2001; Jaattela *et al.*, 2004; Xu *et al.*, 2005; Kroemer *et al.*, 2007), thus placing mitochondriaassociated Bcl-2-family proteins at a downstream point of convergence of many cell death pathways.

Endoplasmic reticulum (ER)

During times of cellular stress, the ER can initiate molecular events that lead to either caspase-dependent or caspase-independent cell death (reviewed by Xu et al., 2005). In this regard, the accumulation of unfolded proteins in the lumen of the ER ('ER stress') triggers an evolutionarily conserved series of signaling events, termed the unfolded protein response (reviewed by Kim et al., 2006; Ron and Walter, 2007). ER stress is induced by several stimuli of relevance to tumor biology, including hypoxia, oxidative stress and nutrient insufficiency, and markers of ER stress have been identified in the centers of tumors with insufficient vasculature supply (Blais et al., 2004; Daneshmand et al., 2007; Dong et al., 2008; Zheng et al., 2008). Factors that perturb normal Ca²⁺ homeostasis in the ER also cause accumulation of unfolded proteins, presumably due to the Ca²⁺ dependence of many ER chaperones, including Grp78 (Hsp70 homolog), Grp94 (Hsp90 homolog) and Calreticulin. Overexpression of antiapoptotic Bcl-2family proteins (for example, Bcl-2; Bcl-X_L) has been shown to protect cells against cell death induced by ER stress, whereas proapoptotic Bcl-2-family proteins (for example, Bax; Bak) are required (Scorrano et al., 2003; Thomenius and Distelhorst, 2003) (Figure 1b). Precisely how Bcl-2-family proteins regulate ER-initiated cell death mechanisms is unclear. Effects of Bcl-2-family proteins on ER Ca²⁺ regulation are among the likely contributors. In this regard, Bcl-2 and Bcl-X_L reduce basal Ca^{2+} concentrations in the ER, apparently through effects on inositol-3-phosphate receptors (IP3Rs)-second messenger-gated Ca2+ channels (He et al., 1997; Kuo et al., 1998; Pinton et al., 2000; Li et al., 2002). The ability of Bcl-X_L to reduce resting ER Ca^{2+} concentrations is dependent on BI-1, a cytoprotective ER protein that associates with Bcl-2 and Bcl-X_L (Xu and Reed, 1998; Xu et al., 2008). Bax and Bak have opposing effects on ER Ca²⁺ concentrations.

The full implications of these changes in ER Ca²⁺ regulation by Bcl-2/Bax-family proteins are not entirely understood, but myriad Ca2+-dependent cell death mechanisms exist that may be of relevance to circumstances where cellular stress causes the ER to dump Ca^{2+} into the cytosol, including (a) Ca^{2+} -induced mitochondrial membrane permeability transition, releasing apoptogenic proteins from mitochondria and stimulating reactive oxygen species production (Bernardi *et al.*, 2006); (b) Ca^{2+} -sensitive mitochondrial fission protein Drp-1 (Szabadkai et al., 2004), which has been implicated in Bax-induced release of cytochrome c from mitochondria; (c) Ca^{2+} -sensitive phosphatases such as calcineurin, (PPase-B), which regulates activity of proapoptotic Bcl-2-family protein BAD (Wang et al., 1999) and which dephosphorylates NFAT-family transcription factors, allowing entry into the nucleus and transactivation of proapoptotic genes encoding Fasligand and Nur77/TR3 (Youn et al., 1999); (d) Ca²⁺ -dependent phospholipases and phospholipid scramblases, the latter of which have been suggested to transfer cardiolipin from the inner to the outer membrane of mitochondria (a signal for targeted

6400

insertion of proapoptotic Bcl-2-family proteins Bid and Bax into mitochondrial membranes) (Lutter et al., 2000; Kuwana et al., 2002; McMillin and Dowhan, 2002); (e) Ca²⁺-sensitive isoforms of nitric oxide synthase, thereby generating reactive nitrogen species and contributing to oxidative stress (reviewed by Orrenius et al., 2003; Benhar et al., 2006); (g) Ca²⁺-binding protein TCTP (fortilin), a putative modulator of antiapoptotic Bcl-2/ Bax-family proteins such as Mcl-1 (Liu *et al.*, 2005); (h) $Ca^{2+}/calmodulin-induced$ activation of the peptidyl proylyl isomerase, FKBP38, which binds to Bcl-2 and induces apoptosis (Edlich et al., 2005); (i) calpain-family cysteine proteases, Ca2+-dependent proteases implicated in many pathological cell death scenarios and whose substrates include Bax and Bid (which are activated) (Wood et al., 1998; Wood and Newcomb, 1999; Chen et al., 2001), Bcl-2 and Bcl-X_L (which are inhibited), several caspases, and autophagy protein Atg5, which binds and inhibits Bcl-X_L upon cleavage (reviewed by Breckenridge et al., 2003; Yousefi et al., 2006); and (j) death-associated kinase (DAP kinase) and its close relative DRP-1, which contain calmodulin-binding domains (reviewed by Shohat et al., 2002), where DAP kinase can induce either apoptosis or autophagy (reviewed by Bialik and Kimchi, 2004). Moreover, Ca²⁺-mediated activation of protein kinase C-theta (PKC- Θ) and of calmodulin-dependent kinase kinase- β induce autophagy in the context of ER stress (Hoyer-Hansen et al., 2007; Sakaki et al., 2008), having further implications for cell life and death mechanisms of relevance to circumstances where ER stress plays a role.

In addition to regulating ER Ca²⁺ homeostasis, Bcl-2family proteins directly regulate Irel, one of the three principal ER signal-transducing proteins involved in unfolded protein response (reviewed by Xu et al., 2005; Kim et al., 2006; Ron and Walter, 2007). Ire1 is a transmembrane ER resident protein that oligomerizes upon release by ER chaperones when unfolded proteins accumulate. Ire1 possesses an intrinsic serine/threonine kinase domain and triggers activation of a kinase pathway that includes Ask1, JNK and p38 MAPK. In addition, Irel has an endoribonuclease domain that processes mRNAs encoding XBP-1, a transcription factor involved in inducing expression of downstream transcription factor genes (for example, CHOP) that control the expression of BCL-2-family genes, such as BIM and BCL-2, to promote cell death. It has been reported that Bax and Bak directly bind and activate Ire1, whereas antiapoptotic Bcl-2 family proteins oppose this effect (McCullough et al., 2001; Hetz et al., 2006; Puthalakath et al., 2007) (Figure 1b).

Lysosomes

Lysosomes participate in cell life and death in at least two contexts—necrosis and autophagy. Rupture or deterioration of lysosomal membranes is a hallmark of necrotic cell death, and although Bcl-2 family proteins do not appear to insert into lysosomal membranes to an appreciable degree, they have been reported to suppress downstream sequeli of lysosomal enzyme release into

the cytosol, particularly as pertains to effects on mitochondria (reviewed by Ferri and Kroemer, 2001; Kroemer et al., 2007). Lysosomes are also critically involved in autophagy, where autophagic vesicles fuse with lysosomes to accomplish the degradation of the contents for purposes of generating fuel for sustaining energy. In this regard, antiapoptotic Bcl-2 family proteins are well known for their ability to prolong survival of growth factor-dependent cells when deprived of the obligate growth factors. In fact, the antiapoptotic function of Bcl-2 was first elucidated in gene transfection studies where interleukin-3 (IL-3)-dependent murine hematopoietic cells were shown to cease division but to survive for prolonged periods in the absence of IL-3 when Bcl-2 was constitutively overexpressed (Vaux et al., 1988). Growth factor deprivation can also lead to nutrient deprivation, because expression of cell surface glucose transporters and amino-acid transporters depends on them. Prolonged nutrient deprivation invokes autophagy, an evolutionarily conserved response for catabolizing macromolecules and organelles, thereby generating substrates for ATP production (Edinger and Thompson, 2004; Lum et al., 2005). Autophagy is initially induced to prolong cell survival, but when taken to extremes, it may cause cell death. Indeed, circumstances have been identified where autophagy is induced and where Bcl-2 or $Bcl-X_L$ either suppress or promote cell death, through mechanisms requiring components of the autophagic machinery (Baehrecke, 2005; Meier and Vousden, 2007). In this regard, Bcl-2 and Bcl-X_L suppress autophagy by binding the protein beclin 1 (Atg6), an essential component of the mammalian autophagy system that marks autophagic vesicles for fusion with lysosomes (Shimizu et al., 2004; Pattingre et al., 2005) (Figure 1c). Beclin 1 has been touted as a tumor suppressor gene, becoming haploinsufficient in some types of human cancers (Aita et al., 1999; Liang et al., 1999). Interestingly, proteintargeting studies suggest that the anti-autophagic function of Bcl-2 is conveyed from the ER (Pattingre et al., 2005).

Bcl-2-family protein interaction networks: dueling dimers

Bcl-2 family proteins contain up to four conserved Bcl-2 homology (BH) domains (Figure 2). All antiapoptotic members of the human Bcl-2 family contain BH1, BH2, BH3 and BH4, including Bcl-2, Bcl-X_L, Mcl-1, Bcl-W, Bfl-1 and Bcl-B (Figure 2). Mammalian proapoptotic proteins Bax, Bak and Bok contain BH1, BH2 and BH3 (but not BH4) and have been termed 'multi-domain' proapoptotic proteins (reviewed by Reed, 2006) (Figure 2). Where elucidated to date, the antiapoptotic and multi-domain proapoptotic Bcl-2 family proteins have been shown to share the same basic threedimensional protein fold, comprised of a bundle of amphipathic α -helices surrounding a core pair of hydrophobic α -helices, bearing striking similarity to the poreforming domains of bacterial toxins such as diphtheria toxin and the colicins (reviewed by Schendel et al., 1998).



Figure 2 Comparisons of domain structures of Bcl-2-family proteins. Bcl-2-family proteins have at least one of four Bcl-2 homology (BH) domains (BH1, BH2, BH3 or BH4), and typically also possess a transmembrane domain (TM). Antiapoptotic Bcl-2-family members contain all four BH domains. Proapoptotic Bcl-2-family members can be separated into 'multi-domain' or 'BH3-only' proteins. Some BH3-only proteins do not have a TM (dotted line). Note that some BH3-only proteins have a limited set of BH domains (for example, Bcl-G has BH2 and BH3).

Bcl-2-family proteins dimerize with each other through BH3 domain-dependent interactions (Chittenden *et al.*, 1995). Many proapoptotic proteins share sequence homology only in the BH3 domain ('BH3only' proteins), including Bid, Bim, Hrk, Bik, Puma, Noxa, Bcl-G_S, Bmf and possibly Spike (reviewed by Strasser, 2005) (Figure 2). These BH3-only proteins uniformly operate as antagonists of antiapoptotic Bcl-2family proteins in a BH3-dependent manner that correlates with BH3-dependent binding. However, some BH3-only proteins, namely Bid, Bim and Puma, have dual functions as both antagonists of antiapoptotic Bcl-2 family members and as agonists of proapoptotic multidomain proteins Bax and Bak (reviewed by Korsmeyer *et al.*, 2000) (Figure 1a).

Structural studies have shown that the BH3 peptide is an amphipathic α -helix of ~16–20 amino-acids length that binds a hydrophobic pocket on the surface of antiapoptotic Bcl-2-family proteins, thus revealing the structural basis for antagonism of pro- and antiapoptotic members of the family and setting the stage for subsequent drug discovery strategies based on mimicking BH3 peptides with chemical compounds that bind the same pocket (Muchmore et al., 1996; Sattler et al., 1997; Oltersdorf et al., 2005). Several chemical inhibitors of antiapoptotic Bcl-2-family proteins based on mimicking BH3 peptides have been described (Wang et al., 2000a, b; Degterev et al., 2001; Enyedy et al., 2001; Tzung et al., 2001; Kutzki et al., 2002; Leone et al., 2003; Becattini et al., 2004; Pellecchia and Reed, 2004; Walensky et al., 2004; Oltersdorf et al., 2005; Mohan et al., 2007; Nguyen et al., 2007; Kitada et al., 2008), and at least three of these agents are currently being tested in human clinical trials for cancer (reviewed by Reed and Pellecchia, 2005). The structural basis for activation of Bax and Bak by BH3 peptides has not yet been reported. Agonistic BH3 peptides induce Bax and Bak oligomerization in mitochondrial membranes and in synthetic liposomes, and thus chemicals that mimic their actions could also provide another path forward towards therapeutics (Letai et al., 2002). A variety of posttranslational modifications and protein interactions have been reported to control the availability of BH3only proteins for dimerization with other Bcl-2-family members. Thus, compounds that target the relevant signal-transduction pathways might also find utility as activators of BH3-only proteins. Indeed, this notion embodies the observation that longer isoforms of the proapoptotic Bim protein associate through the dynein light chain with microtubules, requiring release to translocate to intracellular membranes and dimerize with other members of the Bcl-2 family (Puthalakath *et al.*, 1999), hence, suggesting a probable role for Bim in the cytotoxic mechanisms of antimicrotubule drugs.

Phenotypic conversion of Bcl-2

Bcl-2 is best known for its ability to suppress apoptosis, but a variety of studies have suggested that it may undergo conversion from protector to killer under some circumstances. For example, proteolytic removal of N-terminal sequences by caspase-mediated cleavage reverses the phenotype of Bcl-2 (Cheng et al., 1997). Interestingly, mutations in translocated BCL-2 alleles have been identified in human lymphomas that ablate the asparatic acid residue required for caspase cleavage (Tanaka et al., 1992), suggesting that some tumors may evolve strategies for avoiding Bcl-2 phenotype reversal. In addition, the orphan nuclear receptor Nur77 can be induced to translocate from nucleus to cytosol, binding Bcl-2, and inducing a conformational change in Bcl-2 that probably mimics what happens during caspase cleavage, exposing the normally buried BH3 domain of Bcl-2 and causing it to function as a proapoptotic protein (Lin et al., 2004). A potentially similar mechanism was identified for Bcl-XL, showing that lipid modifications of K-Ras can promote its association with Bcl-X_L on mitochondria and induce apoptosis (Bivona et al., 2006). Thus, depending on with what proteins Bcl-2 and Bcl-X_L interact, their phenotypes can be converted from anti- to proapoptotic, revealing an additional level of complexity to these proteins that has clear therapeutic implications for malignancies that overexpress these Bcl-2-family proteins. Perhaps phenotypic conversion underlies the recent discovery of compounds that show superior cytotoxic activity in Bcl-2/Bcl-X_L overexpressing cells compared to control cells (Schwartz et al., 2007), as well as providing a potential explanation for why high levels of Bcl-2 expression are sometimes associated with better patient prognosis (see for example, Silvestrini et al., 1994).

Bcl-2-family proteins-at the center of the action

While much research has focused on homo- and heterodimerization among Bcl-2-family proteins, many if not most of these proteins have other interaction partners that regulate their activity and that link them to a wide variety of cellular pathways, giving the impression that Bcl-2-family proteins operate as critical nodes in complex networks to integrate information and make ultimate life/death decisions. For the more extensively studied members of the family, such as Bcl-2, Bcl-X_L and Bax, an impressive list of potential interacting proteins has accumulated (reviewed by Reed, 2008). The structural basis for most of these non-familial protein interactions has not been elucidated, and their roles in physiology are not always well understood, but the extensive repertoire of partners suggests that diverse signaling, developmental and metabolic pathways converge on Bcl-2-family members.

A few examples exist where Bcl-2 family proteins serve as regulators of other types of proteins, rather than the converse, where the interacting proteins regulate the activity of the Bcl-2-family members. For example, proapoptotic protein BAD has been reported to associate with a protein complex containing hexose kinase and regulate glucose-stimulated insulin secretion by pancreatic β cells (Danial *et al.*, 2008). In addition, Bcl-2 and Bcl-X_L bind to and suppresses NLR-family protein NLRP1 (NALP1), an activator of inflammatory caspases involved in proteolytic processing and maturation of pro-IL-1 β , pro-IL-18 and pro-IL-33 (Bruey et al., 2007). The aforementioned interaction of Bcl-2 and Bcl-X_L with autophagy protein beclin 1 (Liang et al., 1998) is another example of an effector function achieved through interactions with non-homologous proteins, in this case linking Bcl-2 to suppression of autophagy (Pattingre et al., 2005). Thus, Bcl-2 family proteins appear to regulate aspects of metabolism, innate immunity and autophagy, in addition to their central roles in cell life and death.

Current and future therapeutic opportunities

The fruits of over three decades of research on Bcl-2family proteins have yielded new strategies for therapeutic intervention, some of which have advanced to clinical testing in humans (Table 1). The most advanced of these candidate therapeutics is a nucleaseresistant phosphorothioate antisense oligonucleotide targeting Bcl-2 mRNA (oblimersen sodium), which has shown promising activity for chronic lymphocytic leukemia and malignant melanoma in randomized phase III trials (Reed *et al.*, 1990; Klasa *et al.*, 2002; Bedikian *et al.*, 2006; O'Brien *et al.*, 2007). Results for myeloma, AML and hormone refractory prostate cancer however were not encouraging, perhaps due to overexpression of additional antiapoptotic members of the family besides

References

- Aita VM, Liang XH, Murty VV, Pincus DL, Yu W, Cayanis E *et al.* (1999). Cloning and genomic organization of beclin 1, a candidate tumor suppressor gene on chromosome 17q21. *Genomics* **59**: 59–65.
- Baehrecke EH. (2005). Autophagy: dual roles in life and death? *Nat Rev Mol Cell Biol* 6: 505–510.

Bcl-2 family proteins and cancer KW Yip and JC Reed

 Table 1
 Summary of candidate therapeutics targeting Bcl-2 family proteins

| Active agent | Sponsoring company | Stage |
|--|----------------------------|-------------|
| Genasense (oblimersen sodium: <i>antisense</i>) | Genta | Phase III |
| ABT-737/ABT-263 | Abbott Laboratories | Phase I/II |
| Epigallocatechin gallate | Mayo Clinic | Phase I/II |
| Gossypol | Ascenta/NCI | Phase I/II |
| GX-15-070 (obatoclax) | Gemin X | Phase I/II |
| Polyphenon E | Mayo Clinic (Mitsui Norin) | Phase I |
| Antimycin A3 | University of Washington | Preclinical |
| Apogossypol | Burnham Institute for | Preclinical |
| | Medical Research/NCI | |
| Apogossypolone | University of Michigan | Preclinical |
| BH3I-1/BH3I-2 | Harvard University | Preclinical |
| Compound 6 | University of Michigan | Preclinical |
| CPM-1285 analogs | Raylight Chemokine | Preclinical |
| | Pharmaceuticals | |
| HA14-1 analogs | Raylight Chemokine | Preclinical |
| | Pharmaceuticals | |
| SAHBs | Harvard University | Preclinical |
| Terphenyl derivative | Yale University | Preclinical |
| Theaflavin | Burnham Institute for | Preclinical |
| | Medical Research | |
| | | |

Abbreviation: NCI, National Cancer Institute.

Bcl-2 in these malignancies. Several small-molecule chemical antagonists of antiapoptotic Bcl-2-family proteins have been described that bind the same pocket occupied by proapoptotic BH3 domains (reviewed by Reed and Pellecchia, 2005). These compounds have different potencies and variable spectra of activity against the six antiapoptotic members. Compounds that appear to convert Bcl-2, Bcl-X_L or their relatives from protectors to killers have been identified, including molecules that directly bind these proteins (Manion et al., 2004; Schwartz et al., 2007) and others that trigger Nur77 translocation from nucleus to cytosol, where it can bind Bcl-2, Bfl-1 and Bcl-B (Han et al., 2006; Luciano et al., 2007). Compounds targeting the non-BH3 site on Bcl-2 and Bcl-B possibly where Nur77 binds have also been recently reported, providing a firm starting point for drug optimization (Yip et al., 2008). Altogether, these activities raise hope that a new class of anticancer drugs may soon be available based on targeting Bcl-2-family proteins.

Acknowledgements

We thank M Hanaii and T Siegfried for manuscript preparation, and the NIH for generous support.

- Becattini B, Kitada S, Leone M, Monosov E, Chandler S, Dayong Z et al. (2004). Rational design and real time in-cell detection of the pro-apoptotic activity of a novel compound targeting Bcl-Xl. Chem Biol 11: 389–395.
- Bedikian AY, Millward M, Pehamberger H, Conry R, Gore M, Trefzer U et al. (2006). Bcl-2 antisense (oblimersen sodium) plus

dacarbazine in patients with advanced melanoma: the Oblimersen Melanoma Study Group. *J Clin Oncol* **24**: 4738–4745.

- Benhar M, Forrester MT, Stamler JS. (2006). Nitrosative stress in the ER: a new role for S-nitrosylation in neurodegenerative diseases. ACS Chem Biol 1: 355–358.
- Bernardi P, Krauskopf A, Basson E, Petronilli V, Blachly-Dyson E, Di Lisa F *et al.* (2006). The mitochondrial permeability transition from *in vitro* artifact to disease target. *FEBS J* 273: 2077–2099.
- Bialik S, Kimchi A. (2004). DAP-kinase as a target for drug design in cancer and diseases associated with accelerated cell death. *Semin Cancer Biol* 14: 283–294.
- Bivona TG, Quatela SE, Bodemann BO, Ahearn IM, Soskis MJ, Mor A *et al.* (2006). PKC regulates a farnesyl-electrostatic switch on K-Ras that promotes its association with Bcl-XL on mitochondria and induces apoptosis. *Mol Cell* **21**: 481–493.
- Blais JD, Filipenko V, Bi M, Harding HP, Ron D, Koumenis C et al. (2004). Activating transcription factor 4 is translationally regulated by hypoxic stress. *Mol Cell Biol* 24: 7469–7482.
- Breckenridge DG, Germain M, Mathai JP, Nguyen M, Shore GC. (2003). Regulation of apoptosis by endoplasmic reticulum pathways. *Oncogene* **22**: 8608–8618.
- Brousset P, Krajewski S, Meggetto F, Laurent G, Pris J, Delsol G *et al.* (1996). Frequent expression of the cell death-inducing gene Bax in Reed-Sternberg cells of Hodgkin's disease. *Blood* **87**: 2470–2475.
- Bruey JM, Bruey-Sedano N, Luciano F, Zhai D, Balpai R, Xu C *et al.* (2007). Bcl-2 and Bcl-XL regulate proinflammatory caspase-1 activation by interaction with NALP1. *Cell* **129**: 45–56.
- Calin GA, Ferracin M, Cimmino A, Di Leva G, Shimizu M, Wojcik SE *et al.* (2005). A microRNA signature associated with prognosis and progression in chronic lymphocytic leukemia. *N Engl J Med* 353: 1793–1801.
- Chen M, He H, Zhan S, Krajewski S, Reed JC, Gottlieb RA. (2001). Bid is cleaved by calpain to an active fragment *in vitro* and during myocardial ischemia/reperfusion. *J Biol Chem* **276**: 30724–30728.
- Cheng E, Clem R, Ravi R, Kirsch D, Kastan M, Bedi A *et al.* (1997). Conversion of Bcl-2 to a Bax-like death effector by caspases. *Science* **278**: 1966–1968.
- Chipuk JE, Green DR. (2008). How do BCL-2 proteins induce mitochondrial outer membrane permeabilization? *Trends Cell Biol* 18: 157–164.
- Chipuk JE, Kuwana T, Bouchier-Hayes L, Droin NM, Newmeyer DD, Schuler M *et al.* (2004). Direct activation of Bax by p53 mediates mitochondrial membrane permeabilization and apoptosis. *Science* **303**: 1010–1014.
- Chittenden T, Flemington C, Houghton AB, Ebb RG, Gallo GJ, Elangovan B *et al.* (1995). A conserved domain in Bak, distinct from BH1 and BH2, mediates cell death and protein binding functions. *EMBO J* 14: 5589–5596.
- Cimmino A, Calin GA, Fabbri M, Iorio MV, Ferracin M, Shimizu M et al. (2005). miR-15 and miR-16 induce apoptosis by targeting BCL2. Proc Nat Acad Sci USA **102**: 13944–13949.
- Cory S, Huang DC, Adams JM. (2003). The Bcl-2 family: roles in cell survival and oncogenesis. *Oncogene* 22: 8590–8607.
- Cummins JM, He Y, Leary RJ, Pagliarini R, Diaz Jr LA, Sjoblom T et al. (2006). The colorectal microRNAome. Proc Natl Acad Sci USA 103: 3687–3692.
- Daneshmand S, Quek ML, Lin E, Lee C, Cote RJ, Hawes D *et al.* (2007). Glucose-regulated protein GRP78 is up-regulated in prostate cancer and correlates with recurrence and survival. *Hum Pathol* **38**: 1547–1552.
- Danial NN, Walensky LD, Zhang CY, Choi CS, Fisher JK, Molina AJ et al. (2008). Dual role of proapoptotic BAD in insulin secretion and beta cell survival. Nat Med 14: 144–153.
- Debatin KM, Poncet D, Kroemer G. (2002). Chemotherapy: targeting the mitochondrial cell death pathway. *Oncogene* **21**: 8786–8803.
- Degterev A, Lugovskoy A, Cardone M, Mulley B, Wagner G, Mitchison T *et al.* (2001). Identification of small-molecule inhibitors of interaction between the BH3 domain and Bcl-xL. *Nature Cell Biol* **3**: 173–182.

- Deng X, Gao F, Flagg T, Anderson J, May WS. (2006). Bcl2's flexible loop domain regulates p53 binding and survival. *Mol Cell Biol* 26: 4421–4434.
- Dong D, Ni M, Li J, Xiong S, Ye W, Virrey JJ et al. (2008). Critical role of the stress chaperone GRP78/BiP in tumor proliferation, survival, and tumor angiogenesis in transgene-induced mammary tumor development. Cancer Res 68: 498–505.
- Edinger AL, Thompson CB. (2004). Death by design: apoptosis, necrosis and autophagy. *Curr Opin Cell Biol* 16: 663–669.
- Edlich F, Weiwad M, Erdmann F, Fanghanel J, Jarczowski F, Rahfeld JU *et al.* (2005). Bcl-2 regulator FKBP38 is activated by Ca(2+)/calmodulin. *EMBO J* **24**: 2688–2699.
- Enyedy IJ, Ling Y, Nacro K, Tomita Y, Wu X, Cao Y *et al.* (2001). Discovery of small-molecule inhibitors of Bcl-2 through structure-based computer screening. *J Med Chem* **44**: 4313–4324.
- Eskes R, Desagher S, Antonsson B, Martinou JC. (2000). Bid induces the oligomerization and insertion of Bax into the outer mitochondrial membrane. *Mol Cell Biol* **20**: 929–935.
- Ferri KF, Kroemer G. (2001). Organelle-specific initiation of cell death pathways. *Nat Cell Biol* **3**: E255–E263.
- Gascoyne RD, Adomat SA, Krajewski S, Krajewska M, Horsman DE, Tolcher AM *et al.* (1997). Prognostic significance of Bcl-2 protein expression and Bcl-2 gene rearrangement in diffuse aggressive non-Hodgkin's lymphoma. *Blood* **90**: 244–251.
- Green DR, Kroemer G. (2004). The pathophysiology of mitochondrial cell death. *Science* **305**: 626–629.
- Han YH, Cao X, Lin B, Lin F, Kolluri SK, Stebbins J *et al.* (2006). Regulation of Nur77 nuclear export by c-Jun N-terminal kinase and Akt. *Oncogene* **25**: 2974–2986.
- Hanada M, Delia D, Aiello A, Stadtmauer E, Reed J. (1993). Bcl-2 gene hypomethylation and high-level expression in B-cell chronic lymphocytic leukemia. *Blood* 82: 1820–1828.
- Hanahan D, Weinberg RA. (2000). The hallmarks of cancer. *Cell* **100**: 57–70.
- He H, Lam M, McCormick TS, Distelhorst CW. (1997). Maintenance of calcium homeostasis in the reticulum by Bcl-2. J Cell Biol 138: 1219–1228.
- Hermine O, Haioun C, Lepage E, d'Agay M-F, Briere J, Lavignac C et al. (1996). Prognostic significance of Bcl-2 protein expression in aggressive non-Hodgkin's lymphoma. *Blood* 87: 265–272.
- Hetz C, Bernasconi P, Fisher J, Lee AH, Bassik MC, Antonsson B et al. (2006). Proapoptotic BAX and BAK modulate the unfolded protein response by a direct interaction with IRE1alpha. Science 312: 572–576.
- Hill ME, MacLennan KA, Cunningham DC, Hudson BV, Burke M, Clarke P et al. (1996). Prognostic significance of BCL-2 expression and bcl-2 major breakpoint region rearrangement in diffuse large cell non-Hodgkin's lymphoma: a British National Lymphoma Investigation Study. *Blood* 88: 1046–1051.
- Hoyer-Hansen M, Bastholm L, Szyniarowski P, Campanella M, Szabadkai G, Farkas T *et al.* (2007). Control of macroautophagy by calcium, calmodulin-dependent kinase kinase-beta, and Bcl-2. *Mol Cell* 25: 193–205.
- Ikegaki N, Katsumata M, Minna J, Tsujimoto Y. (1994). Expression of Bcl-2 in small cell lung carcinoma cells. *Cancer Res* 54: 6–8.
- Jaattela M, Cande C, Kroemer G. (2004). Lysosomes and mitochondria in the commitment to apoptosis: a potential role for cathepsin D and AIF. *Cell Death Differ* **11**: 135–136.
- Khanna KK, Wie T, Burrows SR, Moss DJ, Krajewski S, Reed JC *et al.* (1996). Expression of p53, Bcl-2, Bax, Bcl-X₂ and c-myc in radiation-induced apoptosis in Burkitt's lymphoma cells. *Cell Death Differ* **3**: 315–322.
- Kim R, Emi M, Tanabe K, Murakami S. (2006). Role of the unfolded protein response in cell death. *Apoptosis* **11**: 5–13.
- Kitada S, Kress CL, Krajewska M, Jia L, Pellecchia M, Reed JC. (2008). Bcl-2 antagonist apogossypol (NSC736630) displays single-agent activity in Bcl-2-transgenic mice and has superior efficacy with less toxicity compared with gossypol (NSC19048). *Blood* 111: 3211–3219.
- Klasa RJ, Gillum AM, Klem RE, Frankel SR. (2002). Oblimersen Bcl-2 antisense: facilitating apoptosis in anticancer treatment. *Antisense Nucleic Acid Drug Dev* **12**: 193–213.

6404

- Korsmeyer SJ, Wei MC, Saito M, Weiler S, Oh KJ, Schlesinger PH. (2000). Pro-apoptotic cascade activates BID, which oligomerizes BAK or BAX into pores that result in the release of cytochrome *c. Cell Death Differ* **7**: 1166–1173.
- Krajewska M, Kitada S, Winter JN, Variakojis D, Lichtenstein A, Zhai D *et al.* (2008). Bcl-B expression in human epithelial and nonepithelial malignancies. *Clin Cancer Res* **14**: 3011–3021.
- Krajewska M, Krajewski S, Epstein JI, Shabaik A, Sauvageot J, Song K *et al.* (1996a). Immunohistochemical analysis of Bcl-2, Bax, Bcl-X and Mcl-1 expression in prostate cancers. *Am J Pathol* 148: 1567–1576.
- Krajewska M, Moss S, Krajewski S, Song K, Holt P, Reed JC. (1996b). Elevated expression of Bcl-X and reduced Bak in primary colorectal adenocarcinomas. *Cancer Res* 56: 2422–2427.
- Krajewski S, Blomvqvist C, Franssila K, Krajewska M, Wasenius V-M, Niskanen E *et al.* (1995a). Reduced expression of pro-apoptotic gene Bax is associated with poor response rates to combination chemotherapy and shorter survival in women with metastatic breast adenocarcinoma. *Cancer Res* 55: 4471–4478.
- Krajewski S, Chatten J, Hanada M, Womer R, Reed JC. (1995b). Immunohistochemical analysis of the Bcl-2 oncoprotein in human neuroblastomas. *Lab Invest* 71: 42–54.
- Kroemer G, Galluzzi L, Brenner C. (2007). Mitochondrial membrane permeabilization in cell death. *Physiol Rev* 87: 99–163.
- Kuo TH, Kim HR, Zhu L, Yu Y, Lin HM, Tsang W. (1998). Modulation of endoplasmic reticulum calcium pump by Bcl-2. *Oncogene* 17: 1903–1910.
- Kutzki O, Park HS, Ernst JT, Orner BP, Yin H, Hamilton AD. (2002). Development of a potent Bcl-x(L) antagonist based on alpha-helix mimicry. J Am Chem Soc 124: 11838–11839.
- Kuwana T, Mackey MR, Perkins G, Ellisman MH, Latterich M, Schneiter R *et al.* (2002). Bid, bax, and lipids cooperate to form supramolecular openings in the outer mitochondrial membrane. *Cell* **111**: 331–342.
- Leone M, Zhai D, Sareth S, Kitada S, Reed JC, Pellecchia M. (2003). Cancer prevention by tea polyphenols is linked to their direct inhibition of anti-apoptotic Bcl-2-family proteins. *Cancer Res* 63: 8118–8121.
- Letai A, Bassik MC, Walensky LD, Sorcinelli MD, Weiler S, Korsmeyer SJ. (2002). Distinct BH3 domains either sensitize or activate mitochondrial apoptosis, serving as prototype cancer therapeutics. *Cancer Cell* **2**: 183–192.
- Levine B, Kroemer G. (2008). Autophagy in the pathogenesis of disease. *Cell* **132**: 27–42.
- Li C, Fox CJ, Master SR, Bindokas VP, Chodosh LA, Thompson CB. (2002). Bcl-X(L) affects Ca(2+) homeostasis by altering expression of inositol 1,4,5-trisphosphate receptors. *Proc Natl Acad Sci USA* **99**: 9830–9835.
- Liang XH, Jackson S, Seaman M, Brown K, Kempkes B, Hibshoosh H et al. (1999). Induction of autophagy and inhibition of tumorigenesis by beclin 1. Nature 402: 672–676.
- Liang XH, Kleeman LK, Jiang HH, Gordon G, Goldman JE, Berry G *et al.* (1998). Protection against fatal Sindbis virus encephalitis by beclin, a novel Bcl-2-interacting protein. *J Virol* **72**: 8586–8596.
- Lin B, Kolluri SK, Lin F, Liu W, Han Y-H, Cao X et al. (2004). Conversion of Bcl-2 from protector to killer by interaction with nuclear orphan receptor TR3/NGFI-B/Nur77. Cell 116: 527–540.
- Liu H, Peng HW, Cheng YS, Yuan HS, Yang-Yen HF. (2005). Stabilization and enhancement of the antiapoptotic activity of mcl-1 by TCTP. *Mol Cell Biol* 25: 3117–3126.
- Luciano F, Krajewska M, Ortiz-Rubio P, Krajewski S, Zhai D, Faustin B *et al.* (2007). Nur77 converts phenotype of Bcl-B, an antiapoptotic protein expressed in plasma cells and myeloma. *Blood* **109**: 3849–3855.
- Lum JJ, DeBerardinis RJ, Thompson CB. (2005). Autophagy in metazoans: cell survival in the land of plenty. *Nat Rev Mol Cell Biol* 6: 439–448.
- Lutter M, Fang M, Luo X, Nishijima M, Xie X-s, Wang X. (2000). Cardiolipin provides specificity for targeting of tBid to mitochondria. *Nat Cell Biol* 2: 754–756.

- Manion MK, O'Neill JW, Giedt CD, Kim KM, Zhang KY, Hockenbery DM. (2004). Bcl-XL mutations suppress cellular sensitivity to antimycin A. J Biol Chem 279: 2159–2165.
- McCullough KD, Martindale JL, Klotz LO, Aw TY, Holbrook NJ. (2001). Gadd153 sensitizes cells to endoplasmic reticulum stress by down-regulating Bcl2 and perturbing the cellular redox state. *Mol Cell Biol* 21: 1249–1259.
- McMillin JB, Dowhan W. (2002). Cardiolipin and apoptosis. Biochem Biophys Acta 1585: 97–107.
- Meier P, Vousden KH. (2007). Lucifer's labyrinth—ten years of path finding in cell death. *Mol Cell* 28: 746–754.
- Meijerink JP, Mensink EJ, Wang K, Sedlak TW, Sloetjes AW, de Witte T *et al.* (1998). Hematopoietic malignancies demonstrate loss-of-function mutations of BAX. *Blood* **91**: 2991–2997.
- Miyashita T, Krajewski S, Krajewska M, Wang HG, Lin HK, Hoffman B *et al.* (1994). Tumor suppressor p53 is a regulator of BCL-2 and BAX in gene expression *in vitro* and *in vivo. Oncogene* **9**: 1799–1805.
- Miyashita T, Reed JC. (1995). Tumor suppressor p53 is a direct transcriptional activator of human Bax gene. Cell 80: 293–299.
- Mohan KV, Gunasekaran P, Varalakshmi E, Hara Y, Nagini S. (2007). *In vitro* evaluation of the anticancer effect of lactoferrin and tea polyphenol combination on oral carcinoma cells. *Cell Biol Int* 31: 599–608.
- Monni O, Joensuu H, Franssila K, Klefstrom J, Alitalo K, Knuutila S. (1997). BCL2 overexpression associated with chromosomal amplification in diffuse large B-cell lymphoma. *Blood* **90**: 1168–1174.
- Mott JL, Kobayashi S, Bronk SF, Gores GJ. (2007). mir-29 regulates Mcl-1 protein expression and apoptosis. Oncogene 26: 6133–6140.
- Muchmore SW, Sattler M, Liang H, Meadows RP, Harlan JE, Yoon HS *et al.* (1996). X-ray and NMR structure of human Bcl-XL, an inhibitor of programmed cell death. *Nature* **381**: 335–341.
- Nguyen M, Marcellus RC, Roulston A, Watson M, Serfass L, Murthy Madiraju SR *et al.* (2007). Small molecule obatoclax (GX15-070) antagonizes MCL-1 and overcomes MCL-1-mediated resistance to apoptosis. *Proc Natl Acad Sci USA* **104**: 19512–19517.
- O'Brien S, Moore JO, Boyd TE, Larratt LM, Skotnicki A, Koziner B *et al.* (2007). Randomized phase III trial of fludarabine plus cyclophosphamide with or without oblimersen sodium (Bcl-2 antisense) in patients with relapsed or refractory chronic lymphocytic leukemia. *J Clin Oncol* **25**: 1114–1120.
- Oda E, Ohki R, Murasawa H, Nemoto J, Shibue T, Yamashita T et al. (2000). Noxa, a BH3-only member of the Bcl-2 family and candidate mediator of p53-induced apoptosis. *Science* 288: 1053–1058.
- Oltersdorf T, Elmore SW, Shoemaker AR, Armstrong RC, Augeri DJ, Belli BA et al. (2005). An inhibitor of Bcl-2-family proteins induces regression of solid tumors. *Nature* 435: 677–681.
- Orrenius S, Zhivotovsky B, Nicotera P. (2003). Regulation of cell death: the calcium-apoptosis link. Nat Rev Mol Cell Biol 4: 552–565.
- Pattingre S, Tassa A, Qu X, Garuti R, Liang XH, Mizushima N et al. (2005). Bcl-2 antiapoptotic proteins inhibit beclin 1-dependent autophagy. Cell 122: 927–939.
- Pedersen IM, Kitada S, Leoni LM, Zapata JM, Karras J, Tsukada N et al. (2002). Protection of CLL B-cells by a follicular dendritic cell line is dependent on induction of Mcl-1. *Blood* 100: 1795–1801.
- Pellecchia M, Reed JC. (2004). Inhibition of anti-apoptotic Bcl-2 family proteins by natural polyphenols: new avenues for cancer chemoprevention and chemotherapy. *Curr Pharm Des* 10: 1387–1398.
- Penninger JM, Kroemer G. (2003). Mitochondria, AIF and caspases rivaling for cell death execution. *Nat Cell Biol* 5: 97–99.
- Pinton P, Ferrari D, Magalhaes P, Schulze-Osthoff K, Di Virgilio F, Pozzan T et al. (2000). Reduced loading of intracellular Ca²⁺ stores and downregulation of capacitative Ca²⁺ influx in Bcl-2overexpressing cells. J Cell Biol 148: 857–862.
- Puthalakath H, Huang D, O'Reilly L, King S, Strasser A. (1999). The proapoptotic activity of the Bcl-2 family member bim is regulated by interaction with the dynein motor complex. *Mol Cell* 3: 287–296.
- Puthalakath H, O'Reilly L, Gunn P, Lee L, Kelly P, Huntington N et al. (2007). ER stress triggers apoptosis by activating BH3-only protein bim. Cell 129: 1337–1349.

- Rampino N, Yamamoto H, Ionov Y, Li Y, Sawai H, Reed JC et al. (1997). Somatic frameshift mutations in the BAX gene in colon cancers of the microsatellite mutator phenotype. Science 275: 967–969.
- Reed JC. (2002). Apoptosis-based therapies. Nat Rev Drug Disc 1: 111-121.
- Reed JC. (2006). Proapoptotic multidomain Bcl-2/Bax-family proteins: mechanisms, physiological roles, and therapeutic opportunities. *Cell Death Differ* 13: 1378–1386.
- Reed JC. (2008). Bcl-2-family proteins and hematologic malignancies: history and future prospects. *Blood* **111**: 3322–3330.
- Reed JC, Pellecchia M. (2005). Apoptosis-based therapies for hematological malignancies. *Blood* 106: 408–418.
- Reed JC, Stein C, Subasinghe C, Haldar S, Croce CM, Yum S *et al.* (1990). Antisense-mediated inhibition of BCL2 proto-oncogene expression and leukemic cell growth and survival: comparisons of phosphodiester and phosphorothioate oligodeoxynucleotides. *Cancer Res* **50**: 6565–6570.
- Ron D, Walter P. (2007). Signal integration in the endoplasmic reticulum unfolded protein response. *Nat Rev Mol Cell Biol* **8**: 519–529.
- Sakaki K, Wu J, Kaufman RJ. (2008). Protein Kinase C{theta} is required for autophagy in response to stress in the endoplasmic reticulum. *J Biol Chem* **283**: 15370–15380.
- Sattler M, Liang H, Nettesheim D, Meadows RP, Harlan JE, Eberstadt M et al. (1997). Structure of Bcl-xL-Bak peptide complex: recognition between regulators of apoptosis. Science 275: 983–986.
- Sax JK, Fei P, Murphy ME, Bernhard E, Korsmeyer SJ, El-Deiry WS. (2002). BID regulation by p53 contributes to chemosensitivity. *Nat Cell Biol* **4**: 842–849.
- Schendel S, Montal M, Reed JC. (1998). Bcl-2 family proteins as ionchannels. Cell Death Differ 5: 372–380.
- Schwartz PS, Manion MK, Emerson CB, Fry JS, Schulz CM, Sweet IR *et al.* (2007). 2-Methoxy antimycin reveals a unique mechanism for Bcl-x(L) inhibition. *Mol Cancer Ther* **6**: 2073–2080.
- Scorrano L, Oakes SA, Opferman JT, Cheng EH, Sorcinelli MD, Pozzan T et al. (2003). BAX and BAK regulation of endoplasmic reticulum Ca²⁺: a control point for apoptosis. *Science* **300**: 135–139.
- Shimizu S, Kanaseki T, Mizushima N, Mizuta T, Arakawa-Kobayashi S, Thompson CB *et al.* (2004). Role of Bcl-2 family proteins in a non-apoptotic programmed cell death dependent on autophagy genes. *Nat Cell Biol* 6: 1221–1228.
- Shohat G, Shani G, Eisenstein M, Kimchi A. (2002). The DAP-kinase family of proteins: study of a novel group of calcium-regulated death-promoting kinases. *Biochim Biophys Acta* **1600**: 45–50.
- Silvestrini R, Veneroni S, Daidone MG, Benini E, Boracchi P, Mezzetti M et al. (1994). The Bcl-2 protein: a prognostic indicator strongly related to p53 protein in lymph node-negative breast cancer patients. J Natl Cancer Inst 86: 499–504.
- Strasser A. (2005). The role of BH3-only proteins in the immune system. *Nat Rev Immunol* **5**: 189–200.
- Szabadkai G, Simoni AM, Chami M, Wieckowski MR, Youle RJ, Rizzuto R. (2004). Drp-1-dependent division of the mitochondrial network blocks intraorganellar Ca²⁺ waves and protects against Ca²⁺-mediated apoptosis. *Mol Cell* **16**: 59–68.
- Tanaka S, Louie D, Kant J, Reed J. (1992). Frequent somatic mutations in translocated BcL2 genes of non-Hodgkin's lymphoma patients. *Blood* **79**: 229–237.
- Tang SC, Visser L, Hepperle B, Hanson J, Poppema S. (1994). Clinical significance of bcl-2-MBR gene rearrangement and protein expression in diffuse large-cell non-Hodgkin's lymphoma: an analysis of 83 cases. J Clin Oncol 12: 149–154.
- Thomenius MJ, Distelhorst CW. (2003). Bcl-2 on the endoplasmic reticulum: protecting the mitochondria from a distance. *J Cell Sci* **116**: 4493–4499.
- Tron VA, Krajewski S, Klein-Parker H, Li G, Ho VC, Reed JC. (1995). Immunohistochemical analysis of Bcl-2 protein regulation in cutaneous melanoma. *Am J Pathol* **146**: 643–650.
- Tsujimoto Y, Cossman J, Jaffe E, Croce C. (1985). Involvement of the Bcl-2 gene in human follicular lymphoma. *Science* **228**: 1440–1443.

- Tzung S, Kim KM, Basanez G, Giedt CD, Simon J, Zimmerberg J et al. (2001). Antimycin: A mimics a cell-death-inducing Bcl-2 homology domain 3. Nat Cell Biol 3: 183–192.
- Vaux DL, Cory S, Adams JM. (1988). Bcl-2 gene promotes haemopoietic cell survival and cooperates with c-myc to immortalize pre-B cells. *Nature* 335: 440–442.
- Walensky LD, Kung AL, Escher I, Malia TJ, Barbuto S, Wright RD et al. (2004). Activation of apoptosis in vivo by a hydrocarbonstapled BH3 helix. Science 305: 1466–1470.
- Wang HG, Pathan N, Ethell IM, Krajewski S, Yamaguchi Y, Shibasaki F *et al.* (1999). Ca2+-induced apoptosis through calcineurin dephosphorylation of BAD. *Science* **284**: 339–343.
- Wang J-L, Liu D, Zhang Z-J, Shan S, Han X, Srinivascula AM et al. (2000a). Structure-based discovery of an organic compound that binds Bcl-2 protein and induces apoptosis of tumor cells. Proc Natl Acad Sci USA 97: 7124–7129.
- Wang J-L, Zhang Z-J, Choksi S, Shan S, lu Z, Croce CM *et al.* (2000b). Cell permeable Bcl-2 binding peptides: a chemical approach to apoptosis induction in tumor cells. *Cancer Res* 60: 1498–1502.
- Wei MC, Lindsten T, Mootha VK, Weiler S, Gross A, Ashiya M et al. (2000). tBID, a membrane-targeted death ligand, oligomerizes BAK to release cytochrome c. *Genes Dev* 14: 2060–2071.
- Wei MC, Zong WX, Cheng EH, Lindsten T, Panoutsakopoulou V, Ross AJ et al. (2001). Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. Science 292: 727–730.
- Wood DE, Newcomb EW. (1999). Caspase-dependent activation of calpain during drug-induced apoptosis. *J Biol Chem* 274: 8309–8315.
- Wood DE, Thomas A, Devi LA, Berman Y, Beavis RC, Reed JC et al. (1998). Bax cleavage is mediated by calpain during drug-induced apoptosis. Oncogene 17: 1069–1078.
- Xu C, Bailly-Maitre B, Reed JC. (2005). Endoplamic reticulum stress: cell life and death decisions. *J Clinical Invest* **115**: 2656–2664.
- Xu C, Xu W, Palmer AE, Reed JC. (2008). BI-1 regulates endoplasmic reticulum Ca2+ homeostasis downstream of Bcl-2-family proteins. *J Biol Chem* 283: 11477–11484.
- Xu Q, Reed JC. (1998). BAX inhibitor-1, a mammalian apoptosis suppressor identified by functional screening in yeast. *Mol Cell* 1: 337–346.
- Yip KW, Godoi PH, Zhai D, Garcia X, Cellitti JF, Cuddy M et al. (2008). A TR3/Nur77 peptide-based high-throughput fluorescence polarization screen for small molecule bcl-b inhibitors. J Biomol Screen (E-pub ahead of print).
- Yip KW, Shi W, Pintilie M, Martin JD, Mocanu JD, Wong D *et al* (2006). Prognostic significance of the Epstein–Barr virus, p53, Bcl-2, and survivin in nasopharyngeal cancer. *Clin Cancer Res* **12**: 5726–5732.
- Youn H, Sun L, Prywes R, Liu J. (1999). Apoptosis of T cells mediated by ca²⁺-induced release of the transcription factor mef2. *Science* 286: 790–793.
- Yousefi S, Perozzo R, Schmid I, Ziemiecki A, Schaffner T, Scapozza L et al. (2006). Calpain-mediated cleavage of Atg5 switches autophagy to apoptosis. Nat Cell Biol 8: 1124–1132.
- Yu J, Wang Z, Kinzler KW, Vogelstein B, Zhang L. (2003). PUMA mediates the apoptotic response to p53 in colorectal cancer cells. *Proc Natl Acad Sci USA* 100: 1931–1936.
- Yunis JJ, Mayer MG, Arensen MA, Aeppli DP, Oken MM, Frizzera G. (1989). Bcl-2 and other genomic alterations in the prognosis of large-cell lymphomas. N Engl J Med 320: 1047–1054.
- Zha J, Harada H, Yang E, Jockel J, Korsmeyer SJ. (1996). Serine phosphorylation of death agonist BAD in response to survival factor results in binding to 14-3-3 not BCL-X_L. *Cell* 87: 619–628.
- Zheng HC, Takahashi H, Li XH, Hara T, Masuda S, Guan YF *et al.* (2008). Overexpression of GRP78 and GRP94 are markers for aggressive behavior and poor prognosis in gastric carcinomas. *Hum Pathol* **39**: 1042–1049.

5406