

Modeling Respiratory Depression Induced by Remifentanyl and Propofol during Sedation and Analgesia Using a Continuous Noninvasive Measurement of pCO₂^S

Jacqueline A. Hannam, Xavier Borrat, Iñaki F. Trocóniz, José F. Valencia, Erik W. Jensen, Angela Pedroso, Jenifer Muñoz, Sergi Castellví-Bel, Antoni Castells, and Pedro L. Gambús

Pharmacometrics and Systems Pharmacology, Department of Pharmacy and Pharmaceutical Technology, School of Pharmacy, University of Navarra, Pamplona, Spain (J.A.H., I.F.T.); Department of Anaesthesiology, School of Medicine, University of Auckland, Auckland, New Zealand (J.A.H.); Systems Pharmacology Effect Control and Modeling Research Group, Department of Anesthesia, Hospital CLINIC de Barcelona, Barcelona, Spain (X.B., J.F.V., A.P., P.L.G.); Electronic Engineering Program, School of Engineering, Universidad de San Buenaventura, Cali, Colombia (J.F.V.); Centre for Biomedical Engineering Research, Universitat Politècnica de Catalunya, Barcelona, Spain (E.W.J.); Gastroenterology Department, Hospital CLINIC, CIBERehd, Institut d'investigacions Biomèdiques August Pi i Sunyer, University of Barcelona, Barcelona, Spain (J.M., S.C.-B., A.C.); Department of Anesthesia and Perioperative Care, University of California, San Francisco, California (P.L.G.); and Neuroimmunology Research Group, Institut d'investigacions Biomèdiques August Pi i Sunyer, University of Barcelona, Barcelona, Spain (P.L.G.)

Received June 18, 2015; accepted December 10, 2015

ABSTRACT

Respiratory depression is a common adverse effect of propofol and remifentanyl. We aimed to develop a model for respiratory depressant effects of propofol with remifentanyl in patients undergoing endoscopy with sedation. Data were available for 136 patients undergoing endoscopy with sedation. Participants randomly received infusions of propofol and remifentanyl. Predicted plasma concentrations, outputted by infusion pumps, were available. Transcutaneous arterial pressure of carbon dioxide (pCO₂) was measured. Data were analyzed using nonlinear mixed-effects modeling methods. Covariate relationships were investigated for age, noxious stimuli (endoscopy tube insertion), and A118G genotype for the μ -opioid receptor (OPRM1). Participants had a median (range) age of 64.0 (25.0–88.0) years, weight of 70.0 (35.0–98.0) kg, and height of 164.0 (147.0–190.0) cm. Seven

percent were recessive homozygous for OPRM1 polymorphism. An indirect-effect model with a “modulator” compartment best described pCO₂ data ($P < 0.001$) over a direct-effect model. Remifentanyl inhibited pCO₂ removal with an IC₅₀ of 1.13 ng/ml and first-order rate constant (k_{e0}) of 0.28 minute⁻¹. Propofol affected the modulator compartment with an IC₅₀ of 4.97 μ g/ml (no effect-site compartment). Propofol IC₅₀ and remifentanyl k_{e0} were reduced with increasing age. Noxious stimuli and genotype were not significant covariates. An indirect-effect model with a rebound mechanism can describe remifentanyl- and propofol-induced changes in pCO₂ in patients undergoing noxious procedures. The model may be useful for identifying optimal dosing schedules for these drugs in a combination that provides adequate sedation but avoids respiratory depression.

Introduction

Sedation with analgesia is used as an anesthetic technique to allow diagnostic or therapeutic procedures without pain or distress for patients. Combining sedation and analgesia provides optimal conditions for endoscopic diagnosis and intervention, and better success rates (Ootaki et al., 2012). Anesthesiologists must administer hypnotic and/or analgesic drugs, observe the effect induced, evaluate possible unwanted side effects, take action if required, and adjust dosing to the individual's response. While being sedated, patients breathe spontaneously with little airway support, and recover quickly to their preprocedure conditions. Most drugs used for sedation and analgesia also have respiratory depressant effects that occur in a concentration-dependent fashion.

Several methods of measuring ventilatory depression are currently available, but all have advantages and disadvantages:

Primary laboratory of origin:

The Pharmacometrics and Systems Pharmacology, Department of Pharmacy and Pharmaceutical Technology, School of Pharmacy, University of Navarra, Pamplona, Spain (responsible for modeling and analysis)

Systems Pharmacology Effect Control and Modeling (SPEC-M) Research Group, Department of Anesthesia, Hospital CLINIC de Barcelona, Barcelona, Spain (responsible for study design, conduct and data).

This work was supported by the Hospital CLINIC de Barcelona, Spain [Grant 3664]; the Fondo de Investigaciones Sanitarias, Health Department, Government of Spain [Grants PI/050072, PS09/01209, and FIS PI14/00173]; the Ministerio de Economía y Competitividad [Grant 2010-19273]; the Agència de Gestió d'Ajuts Universitaris i de Recerca [Grant 2009 849]; and the Research Agency of the Government of Colombia, COLCIENCIAS [Grant 297-2015].

This work was previously presented as follows: [Poster] Hannam JA, Trocóniz IF, Borrat X, and Gambús PL (2015) A model for the respiratory effects of remifentanyl and propofol during sedation and analgesia. *24th Population Approach Group in Europe Meeting*; June 2-5, 2015. Hersonissos, Crete, Greece. dx.doi.org/10.1124/jpet.115.226977.

^S This article has supplemental material available at (jpet.aspetjournals.org).

ABBREVIATIONS: BIS, bispectral index; IIV, interindividual variability; -2LL, $-2 \times \log$ likelihood; NOX, noxious stimulation; OBJ, objective function; SNP, single nucleotide polymorphism; TCI, targeted controlled infusion; VPC, visual predictive check.

oxygen saturation might show adequate levels during severe apnea; respiratory rate is difficult to measure objectively and clinically and, without an accurate evaluation of tidal volume, is hardly effective in assessing adequate ventilation; $p\text{CO}_2$ changes reflect respiratory function but must be measured by arterial blood sampling (invasive and noncontinuous) or through capnography, which may be susceptible to false negatives. Transcutaneous CO_2 monitors are based on arterialization of the capillary bed through the local application of heat. The use of Stow-Severinghaus electrodes provides information on the transcutaneous CO_2 tension continuously and non-invasively and with good correlation with arterial $p\text{CO}_2$. Transcutaneous measurement of arterial $p\text{CO}_2$ allows us to study respiratory depression by analyzing the time course of $p\text{CO}_2$ in individual patients undergoing sedation analgesia with propofol and remifentanyl. Measuring and predicting $p\text{CO}_2$ levels is clinically relevant since $p\text{CO}_2$ reflects the level of respiratory depression. Very high levels of $p\text{CO}_2$ may be associated with severe consequences, such as narcosis or cerebral edema (Joyce and McGee, 2011; Spindelboeck and Moser, 2012).

Several models of respiratory effects have been reported for individual drugs commonly used during sedation (propofol and the opioids remifentanyl and alfentanil) (Bouillon et al., 1999, 2003, 2004a; Caruso et al., 2007). Few reports exist for models of combined effects of propofol with remifentanyl on respiratory depression, despite the frequency with which agents are combined in anesthesia, and those that do are based on data derived from healthy volunteers (Nieuwenhuijs et al., 2003; Olofsen et al., 2010). Respiratory control is determined by multiple processes, in which intrinsic feedback is provided by arterial pH levels and concentrations of O_2 and CO_2 (Lloyd et al., 1958; Dahan et al., 1990; Ward and Karan, 2002). Feedback mechanisms regulate respiratory drive, which changes the alveolar minute ventilation. This makes it difficult to isolate and quantify key components of the system, and consequently, many of the current models have been developed in highly controlled conditions (Bouillon et al., 1999) and in healthy volunteers (Bouillon et al., 2003, 2004a; Caruso et al., 2007; Olofsen et al., 2010). This may limit their ability to predict respiratory depression in patient populations and the clinical environment.

A model has previously been reported for the effects of propofol and remifentanyl on bispectral index (BIS) in patients undergoing endoscopy under sedation and analgesia (Borrat et al., 2013). In that study, the effect of noxious stimulation on BIS was quantified, and the influence of the A118G single nucleotide polymorphism (SNP) of the OPRM1 gene (which encodes the μ -opioid receptor) on remifentanyl potency was investigated. In the present study, we aimed to develop a model to describe respiratory changes during propofol-remifentanyl sedation in the same patients using continuously and non-invasively measured levels of $p\text{CO}_2$. A secondary aim was to test the influence of noxious stimulation on CO_2 elimination and of the A118G SNP genotype on respiratory changes in response to remifentanyl.

Materials and Methods

This study was approved by the institutional review board of the Hospital CLINIC de Barcelona, Spain (reference 2007/3664). All participants gave written, informed consent before being enrolled in the project. The data were a subset of a larger study in which the influence of the A118G SNP genotype on opioid requirements during

sedation for endoscopy was investigated (Borrat et al., 2013). Study methods are described in brief, and have been reported in detail previously (Borrat et al., 2013).

Patients and Drug Administration. Two hundred and seven patients undergoing sedation and analgesia for ultrasonographic upper gastrointestinal endoscopy were enrolled; the aim was to include between 20 and 40 patients who could have the A118G SNP, since the expected prevalence of A118G in the OPRM1 gene has been estimated to be around 10–19% in the general population (Lotsch and Geisslinger, 2005). All patients received a combination of propofol and remifentanyl.

Participants were randomized to one of four groups. Each group received a fixed targeted controlled infusion (TCI) of 2.0 $\mu\text{g/ml}$ propofol, 3.0 $\mu\text{g/ml}$ propofol, 1.0 ng/ml remifentanyl, or 2.0 ng/ml remifentanyl. Infusions were given via a TCI system (Base Primea; Fresenius Kabi AG, Bad Homburg, Germany) set to target the desired concentration in the effect compartment. Parameter estimates as reported by Schnider et al. (1998, 1999) and Minto et al. (1997) were used for propofol and remifentanyl infusions, respectively. For each participant, the infusion of the second drug began after some data collection with the allocated drug only. The target effect-site concentration of the second drug was then determined by the nausea (or “gag”) response of the previous participant according to the Dixon up-down method (Dixon, 1991), and the second infusion was started. Gag response to insertion of the endoscopy tube was considered positive when nausea, cough, and/or fight against the introduction of the endoscopy probe was observed (evaluated by the endoscopist responsible for the procedure). In the two propofol groups, a positive response resulted in an increase of the target remifentanyl concentration by 0.5 ng/ml . In the remifentanyl groups, the corresponding increase in targeted propofol concentration was 0.5 $\mu\text{g/ml}$. A negative response to endoscopy tube insertion resulted in a reduction of the targeted concentration in the subsequent participant by the same magnitude. Once the response to endoscopy was observed, TCI targets for both drugs were altered according to clinical requirements as per standard clinical practice.

Response Measurements. Arterial blood pressure, pulse oximetry data, and respiratory rate were monitored noninvasively for all participants. In addition, electroencephalograph data from BIS (Bispectral Index A2000; Covidien, Boulder, CO) were recorded.

$p\text{CO}_2$ was measured using a SenTec Digital Monitor (SenTec, Therwil, Aarberg, BL, Switzerland). $p\text{CO}_2$ is measured with a sensor containing a Severinghaus-type pH-sensitive electrode bathed in an electrolytic solution protected by a permeable membrane. The sensor is warmed to a constant surface temperature of 42°C, increasing CO_2 permeability. CO_2 crosses the sensor membrane and modifies the pH in the electrolyte solution, which is sensed by the Severinghaus electrode. pH changes and, therefore, proportional electrode signal are directly related to $p\text{CO}_2$ concentration. The sensor was calibrated and prepared according to the manufacturer recommendations, then placed in the earlobe of the patient and secured with special adhesive and an ear clip. An equilibration period of about 5 minutes was observed before the monitor was ready to give accurate measures. Measurements were recorded online every second using specific software.

Data from $p\text{CO}_2$, drug infusion, predicted plasma concentrations, BIS, hemodynamics, noxious stimulation, and other relevant events were synchronized offline for further analysis, with a resolution of one datum every 30 seconds. Before beginning the study, a single venous blood sample was drawn for genotyping of the A118G SNP, as described elsewhere (Borrat et al., 2013). Prior to any drug administration, a 5-minute period was observed in which the patient rested in a quiet environment while baseline data were collected.

Data Analysis. Data were analyzed using a population approach in NONMEM version 7.2 (Icon Development Solutions, Ellicott City, MA). The stochastic approximation expectation maximization algorithm, followed by importance sampling, was used. Model selection was based on inspection of visual plots [including prediction-corrected visual predictive checks (VPCs)] (Bergstrand et al., 2011) and the change in the minimum value of the objective function (OBJ) provided by

NONMEM. The minimum OBJ approximately equals the $-2 \times \log$ likelihood ($-2LL$). A reduction in the OBJ between nested models suggests an improvement in model fit. A statistically significant improvement was required for inclusion of one additional parameter (one degree of freedom), equating to a reduction >3.84 based on a χ^2 distribution ($\alpha < 0.05$). Interindividual variability (IIV) was modeled exponentially, and residual error was determined using an additive error model. Subject-specific magnitude of residual error and the nondiagonal elements of the Ω variance-covariance were also tested for significance.

Model Building. Plasma drug concentration data were not available, so TCI system-predicted plasma concentrations were used as the pharmacokinetic basis of the model. For each drug, we tested the inclusion of a hypothetical effect-site compartment to describe the delay in effect onset (Sheiner et al., 1979). Thus, the time course of the predicted concentrations in the effect site was described as:

$$\frac{dC_e}{dt} = k_{e0} \times (C_p - C_e) \quad (1)$$

where C_p is the concentration predicted by the TCI system, C_e is the predicted concentration in the effect site, and k_{e0} is the first-order rate constant governing the disequilibrium in drug distribution between the central (plasma) and effect-site compartments. For both drugs, the presence of the effect compartment has been widely documented (Minto et al., 1997; Schnider et al., 1999; Babenco et al., 2000; Bouillon et al., 2003, 2004a).

In the current evaluation, the framework of the indirect and turnover response models including rebound mechanisms (Dayneka et al., 1993; Wakelkamp et al., 1996) was used to describe the time course of pCO_2 as the pharmacodynamic endpoint. pCO_2 levels are the result of the contribution of 1) CO_2 production and removal rates (i.e., removal from the lung alveolar via the process of respiration), as represented by the zero and first-order rate constants K_{in} and K_{deg} , respectively, and 2) feedback mechanisms represented by the modulator M (eqs. 2 and 3):

$$\frac{dpCO_2}{dt} = K_{in} - K_{deg} \times M \times pCO_2 \quad (2)$$

$$\frac{dM}{dt} = K_{mod} \times \left(\frac{pCO_{2(t)}}{pCO_{2(0)}} \right)^\alpha - K_{mod} \times M \quad (3)$$

where K_{mod} is the turnover rate constant governing M dynamics, and α scales the effect of the change in pCO_2 over time ($pCO_{2(t)}$) with respect to baseline ($pCO_{2(0)}$) on the production rate of M . In baseline conditions, the rate of CO_2 production is in equilibrium with its removal, then $dpCO_{2(0)}/dt = 0$, $K_{in} = pCO_{2(0)} \times K_{deg}$, and $pCO_{2(t)}$ equals $pCO_{2(0)}$.

The amount in the modulator compartment feeds back to the pCO_2 compartment to modulate the rate of pCO_2 removal (for example, via increasing or decreasing respiratory rate). Note that, in this model, rebound is parameterized as a fraction from baseline, so that in homeostatic conditions ($t = 0$), the amount in the modulator compartment is equal to 1, and no modulation of pCO_2 removal occurs.

Drug effects were modeled as follows. Remifentanyl is known to suppress ventilation (Dershwitz et al., 1996; Babenco et al., 2000), and this mechanism of action was incorporated in the model as a reduction of the K_{deg} parameter, as represented in eq. 4:

$$\frac{dpCO_2}{dt} = K_{in} - (K_{deg} \times M \times pCO_2 \times E_{REM}) \quad (4)$$

E_{REM} represents a function accounting for the remifentanyl drug effects, which takes the general form represented by eq. 5:

$$E_{REM} = 1 - I_{MAX} \frac{C_{eR}^\gamma}{C_{eR}^\gamma + IC_{50R}^\gamma} \quad (5)$$

where IC_{50R} is the concentration of remifentanyl in the effect site (C_{eR}) that causes 50% of the maximal inhibition in K_{deg} (I_{MAX}), and γ is a slope parameter governing the slope of the K_{deg} versus C_{eR} relationship. I_{MAX} was constrained between 0 and 1, and during model development, other models for drug effects, such as the linear model, were also tested.

Propofol effects (E_{PROPO}) were incorporated in the model by modifying the feedback mechanism affecting removal of pCO_2 (represented in eq. 6) following the observation that propofol alters the slope of the ventilation response to rising arterial CO_2 (Blouin et al., 1991). Subsequently, we incorporated propofol effects through the modulator compartment as inhibition of K_{mod} :

$$\frac{dM}{dt} = K_{mod} \times E_{PROPO} \times \left(\frac{pCO_{2(t)}}{pCO_{2(0)}} \right)^\alpha - K_{mod} \times M \quad (6)$$

E_{PROPO} has a structure similar to E_{REM} in eq. 5, and as in the case of remifentanyl, additional models for E_{PROPO} were tested during the model-building process. In addition, propofol has been shown to suppress CO_2 production in tissues by up to 30% in steady-state, controlled respiratory studies (Pavlin et al., 1996). To avoid bias in our parameter estimates, we included a correction factor on CO_2 production as suggested by Bouillon et al. (2004a) and Caruso et al. (2007, 2008) assuming an I_{MAX} of 0.3 for propofol effects on K_{in} (eq. 7):

$$\frac{dpCO_2}{dt} = (K_{in} \times E_{PROPO}) - (K_{deg} \times M \times pCO_2) \quad (7)$$

The model described in eqs. 1–7 reflects the observations that both drugs independently cause depression of the respiratory system. A schematic representation of the model developed for respiratory depression effects of remifentanyl and propofol in combination is provided in Fig. 1.

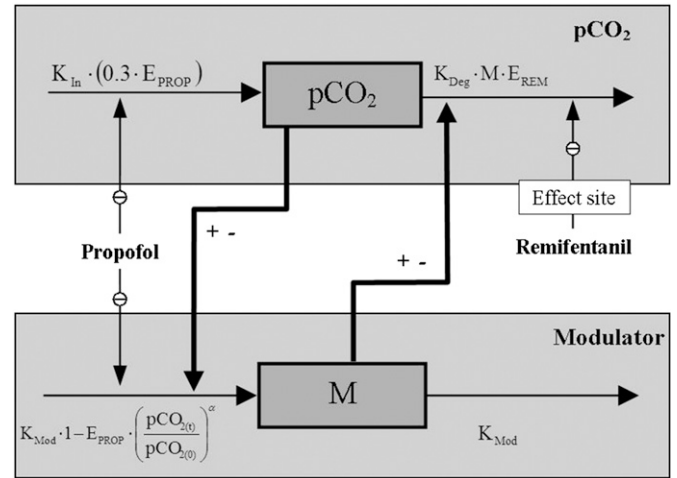


Fig. 1. Model of propofol and remifentanyl effects on pCO_2 . The model is based on two compartments: the main compartment describing changes in CO_2 , and a modulator compartment (M) representing feedback processes (such as control of ventilation rate) that work to maintain system homeostasis. Changes in CO_2 concentration in the main compartment modify the rate into M (by K_{mod}), and changes in M modify the rate of CO_2 removal from the main compartment (by K_{deg}). These primary relationships of the system are indicated by the heavy bold arrows. The influence of CO_2 on K_{mod} is determined by the ratio of pCO_2 at time t ($pCO_{2(t)}$) to that at baseline ($pCO_{2(0)}$), so during homeostasis, this term is equal to 1 and no system modulation occurs. Propofol reduces K_{mod} (thereby reducing the rate into M and inhibiting the feedback response to rising pCO_2), and has a small effect on metabolic CO_2 production (represented by K_{in} , $\leq 30\%$ reduction). Remifentanyl acts via an effect-site compartment to reduce K_{deg} . Drug effects for both remifentanyl (E_{REM}) and propofol (E_{PROPO}) are indicated in the figure by light arrows. α Is an amplification factor for the system feedback.

Covariate Model Selection. Effects of several covariates were explored for significance. We tested the effect of age on the IC_{50} parameters of both drugs, and on the k_{e0} of remifentanyl, based on the results obtained from previous analyses performed by Minto et al. (1997) and Schnider et al. (1999). A118G SNP was tested as a binary covariate for an influence on the IC_{50} of remifentanyl, as individuals carrying the A118G genotype are known to display reduced sensitivity to opioids for some endpoints (Skarke et al., 2003; Klepstad et al., 2004; Borrat et al., 2013). The third covariate explored was that of noxious stimulation (NOX). We hypothesized that noxious stimulation, or pain, is likely to increase respiration rate; therefore, we explored NOX effects as an increase in the K_{deg} parameter. NOX was introduced as a binary covariate (endoscopy tube inserted or not inserted) that varied within the period of endoscopy, as done in previous work focusing on sedation levels in which a significant influence of this covariate was detected on propofol and remifentanyl requirements (Borrat et al., 2013). We tested each covariate individually, requiring a statistically significant improvement ($\alpha < 0.05$) in model fit as judged by the $-2LL$ value for inclusion. For the final model, all significant covariates were included, and the model was reduced by removing those that failed to contribute to model fit. In addition to investigating covariates as described earlier, we also checked to see whether scaling to body weight was required for any parameters (this did not require the addition of a parameter to be estimated, so model improvement was evaluated using VPCs).

Results

Data were available for 136 of the 207 participants studied, providing a total of 38,761 pCO_2 observations. Seventy-one participants were excluded due to inadequate recordings of pCO_2 levels for the following reasons: unfinished signal stabilization despite more than 10 minutes waiting, sensor dislodged from the earlobe, excessive movement of the patient, poor quality of the signal, and problems with the data collection software. The final numbers of participants by group were $N = 36$ in the 2.0 $\mu g/ml$ propofol group, $N = 29$ in the 3.0 $\mu g/ml$ propofol group, $N = 29$ in the 1.0 ng/ml remifentanyl group, and $N = 31$ in the 2.0 ng/ml remifentanyl group. Demographic characteristics for the group are summarized in Table 1, whereas characteristics of the data are summarized in Table 2.

Model Building. Given the complexity of the mechanisms involved in the regulation of respiratory depression, as represented in previous published pharmacokinetic-pharmacodynamic models, and the observational characteristics of our data, we used the following techniques/approaches to during the model building process develop our selected model: 1) deterministic simulations with the aid software

Berkeley-Madonna (Macey and Oster, 2010) to find proper initial estimates of the model parameters, and 2) sequential model building where data from each drug was analyzed separately first, and combination data were then incorporated into the analysis. In addition, we experienced convergence issues with several models. All model features represented in eqs. 1–6 were supported by a significant reduction in $-2LL$. The main results obtained during model building ranked on the absolute decrease in $-2LL$, and the results of sensitivity analysis using simulation for each parameter in the final model are provided in the (Supplemental Material).

Considering the presence of an effect-site compartment for remifentanyl reduced the value of $-2LL$ by over 500 points ($P < 0.001$). In contrast, our data did not support the prediction of effect-site concentrations of propofol ($P > 0.05$); therefore, the effect of E_{PROP} on K_{mod} and K_{in} (eqs. 6 and 7) is driven by predicted plasma concentrations of propofol. With respect to the pharmacodynamic relationships (i.e., eq. 5), I_{MAX} was not found to be significantly different from 1 for the effects of remifentanyl and propofol on K_{deg} and K_{mod} , respectively ($P > 0.05$). As explained in the *Materials and Methods* section, the I_{MAX} corresponding to E_{PROP} on K_{in} was fixed (i.e., not estimated) to 0.3 according to literature estimates (Bouillon et al., 2004a; Caruso et al., 2007, 2008). Sigmoidicity was absent in the pharmacodynamic relationship of propofol (γ parameter not significantly different from 1; $P > 0.05$); in the case of remifentanyl, the estimate of γ was 2.75.

The inclusion of a modulator compartment (represented by eqs. 2 and 3) was highly significant, indicating a strong regulatory mechanism. The final model uses the ratio between current and baseline value of pCO_2 as the driving force triggering the regulatory mechanism. Other parameterizations were tested, such as that used by Olofsen et al., (2010), but their parametrization worsened the fit in our case. In addition, we obtained an estimate of the α parameter significantly different from 1 ($P < 0.001$). E_{PROP} effects on K_{mod} also resulted in significance, supporting the observation that propofol by itself has an effect of respiratory function. During model building, other model alternatives were also explored, such as including propofol effects on K_{deg} (with and without an interaction term between propofol and remifentanyl) and as an allosteric modulator of E_{REM} , but as these did not result in model improvements, they were not investigated further.

The following parameters in the model were associated with interpatient variability: $pCO_{2(0)}$, K_{deg} , and IC_{50R} . IIV was not supported by the data for the remaining parameters, despite individual testing. As stated in the *Materials and Methods* section, IIV was described with an exponential model. However, the distribution of the random effect for $pCO_{2(0)}$ was better described using the Box-Cox transformation (Box and Cox, 1964), which improved model performance as judged by visual inspection of the predictive checks. Results also indicated a significant patient-specific magnitude of residual error. The population model selected included covariance for the random effects associated with $pCO_{2(0)}$, K_{deg} , and IC_{50R} . We scaled $pCO_{2(0)}$ by weight, as this corrected a persistent misspecification in our VPCs.

A118G SNP in the OPRM1 genotype caused a small increase in the remifentanyl IC_{50} , from 1.12 ng/ml in normal patients to 1.32 ng/ml (18%) in those who were recessive homozygous for the GG SNP on the OPRM1 gene. However, this effect was neither statistically nor clinically significant.

TABLE 1
Participant demographics

Values are the median (range) unless otherwise indicated. Concentrations given for propofol and remifentanyl are those predicted by the TCI pump in the plasma compartment and for the full data set.

Participants	Value
Count of participants	136
Age (years)	64.0 (25.0–88.0)
Height (cm)	164.0 (147.0–190.0)
Weight (kg)	70.0 (35.0–98.0)
Gender (count, male/female)	84 / 52
OPRM1 ^a (count, %)	7 (5.4)
Propofol concentration ($\mu g/ml$)	2.72 (0, 13.0)
Remifentanyl concentration (ng/ml)	1.50 (0, 9.8)

^aRecessive homozygous (GG) for the SNP on the OPRM1 gene.

TABLE 2

Summary of baseline, infusion, and noxious stimulation conditions

Values are the median (range) durations, given in minutes. Median (range) predicted plasma concentrations for both drugs are also provided for each condition.

	Data Points	Duration (min)	Predicted Plasma Concentrations ^a	
			Propofol	Remifentanil
			$\mu\text{g/ml}$	ng/ml
Baseline (no drug)	970	2.5 (0–19.4)	—	—
Propofol infusion	2010	1.5 (0–19.1)	4.2 (0.004–10.6)	—
Remifentanil infusion	2647	2.9 (0–13.9)	—	3.1 (0.01–8.2)
Combination infusion	33,134	66.9 (15.1–142.2)	2.5 (0.002–13.0)	1.5 (0.004–9.8)
NOX = 0	17,223	22.5 (4.0–68.1)	2.7 (0–13.0)	1.2 (0–9.8)
NOX = 1	21,538	45.3 (1.85–126.9)	2.5 (0–8.9)	1.3 (0–5.9)

^aPlasma concentrations are predicted by the TCI system used in effect-site targeting mode. NOX is noxious stimulation as caused by insertion of the endoscopy tube.

When introduced individually, significant covariate effects were identified for age on remifentanil IC_{50} and k_{e0} , age on propofol IC_{50} , and NOX on K_{deg} (see Supplemental Table S1). To identify the final model, all significant covariates were included, and those that failed to estimate (indicating no effect) were removed. The final selected model included covariate effects for age on remifentanil k_{e0} ($\text{Age}_{k_{e0R}}$) and propofol IC_{50} ($\text{Age}_{\text{IC}_{50P}}$).

Table 3 lists the model parameter estimates corresponding to the selected model for the interaction of propofol and remifentanil in respiratory depression. Some parameters (α , $\text{Age}_{k_{e0R}}$, and $\text{Age}_{\text{IC}_{50P}}$) showed a high standard error, indicating that they were not fully identifiable. The percentage of η - and ε -shrinkage was lower than 5%.

Figure 2 shows the results of model performance. The panels corresponding to the prediction-corrected VPCs indicate that the mean tendency and the dispersion of data are well captured by the model, regardless of the independent variable used to check model performance (time or predicted concentrations). Similarly, conditional weighted residuals versus the three

different independent variables reveals that there were no systematic deviations from the perfect fit (i.e., conditional weighted residuals = 0), indicating an absence of major model misspecifications. Conditional weighted residuals versus time data points are visible for propofol alone, remifentanil alone, and the combination (Fig. 2B).

Figure 3 gives the profiles for predicted drug plasma concentrations for both drugs, the predicted effect site concentrations for remifentanil, and the observed and model-predicted pCO_2 levels for six patients selected at random.

Through typical simulations, Fig. 4 demonstrates the contribution of the different elements of the selected model to the time course of respiratory depression. Drug pharmacokinetic profiles (Fig. 4A) are simulated using standard population models given in the literature (see *Materials and Methods*). The kinetic profiles in Fig. 4B show that the model elements with greater impact on pCO_2 are E_{REM} and the modulator. Age appears to have a marginal effect on respiratory response, as shown in Fig. 4C. The effect of remifentanil on K_{deg} is more pronounced than the effect propofol exerts on K_{mod} and K_{in}

TABLE 3

Final parameter estimates for the final model

IIV is expressed as CV(%) with 95% confidence intervals given in square brackets. $\text{pCO}_{2(0)}$ is baseline pCO_2 , estimated per kilogram. K_{deg} is a rate constant describing the rate of pCO_2 removal from the main system compartment, K_{mod} describes the rate of synthesis and degradation from the modulator compartment, and α describes amplification of the feedback system in responding to changes in pCO_2 . IC_{50P} and IC_{50R} are the concentrations of propofol and remifentanil, respectively, that cause 50% the maximal drug effect. γ_R is a shape parameter describing the shape of the remifentanil concentration-response curve, and k_{e0R} describes the transfer of remifentanil between the plasma and effect-site compartments. Minimal IIV terms were added and fixed to a low value for all parameters not already associated with IIV (indicated by “—”) to improve NONMEM efficiency during stochastic approximation expectation maximization estimation methods with MU referencing.

Parameter	Estimate	[5th–95th]	Shrinkage	IIV
	CV%		%	CV%
System parameters				
$\text{pCO}_{2(0)}$ (mm Hg/kg)	36.4 (0.52)	[0.49–0.56]	0	29.2 ^a (27.6)
K_{deg} (min^{-1})	0.057 (39.1)	[0.01–0.10]	0.4	204.7 (32.7)
K_{mod} (min^{-1})	0.45 (43.0)	[0.07–0.83]	—	—
α	3.82 (94.8)	[–3.28–10.92]	—	—
Residual error (mm Hg)	1.98 (11.4)	[1.54–2.42]	1.9	52.82 (11.7)
Drug parameters				
IC_{50R} (ng/ml)	1.13 (44.0)	[0.16–2.10]	4.0	80.0 (25.2)
γ_R	2.75 (18.3)	[1.77–3.73]	—	—
k_{e0R} (min^{-1})	0.28 (37.3)	[0.07–0.48]	—	—
^b $\text{Age}_{k_{e0R}}$	0.12 (73.4)	[–0.05–0.29]	—	—
IC_{50P} ($\mu\text{g/ml}$)	4.97 (17.3)	[3.28–6.66]	—	—
^b $\text{Age}_{\text{IC}_{50P}}$	2.73 (51.3)	[–0.01–5.47]	—	—

CV, coefficient of variation.

^aIIV for $\text{pCO}_{2(0)}$ was best modeled using a Box-Cox transformation, and the Box-Cox parameter λ (CV%, 5th–95th) of –1.18 (11.4%, –1.43 to 0.92).

^bAge covariate effects, introduced as $\theta_{\text{Ind}} = \theta_{\text{pop}} - (\text{AGE}/64) * \theta_{\text{Age}}$.

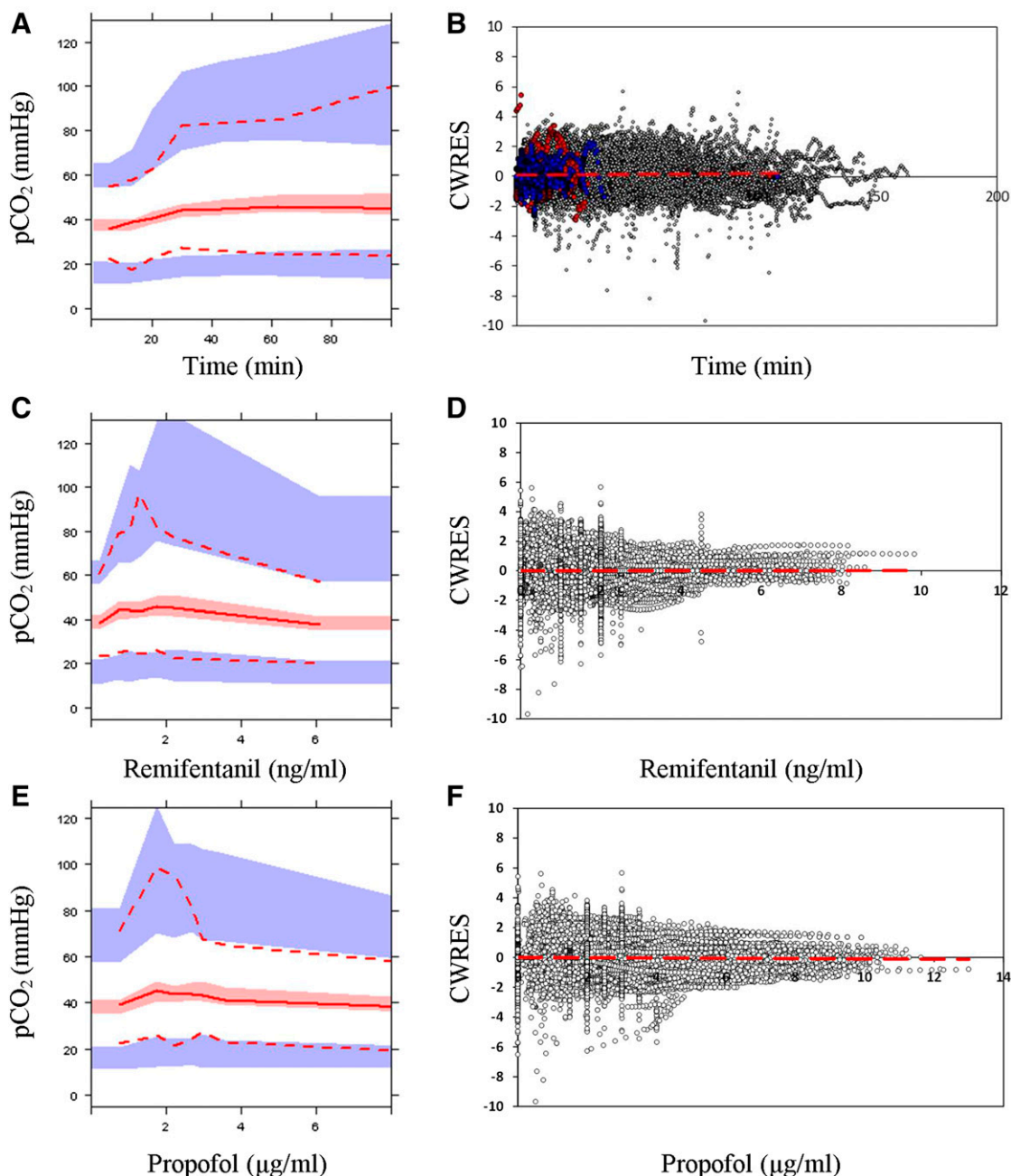


Fig. 2. Goodness-of-fit plots. The left panels give prediction-corrected VPCs, whereas the right panels give conditional weighted residuals (CWRES). Goodness of fit is given for $p\text{CO}_2$ versus time (A and B), pump-predicted remifentanyl concentrations in the plasma (C and D), and pump-predicted propofol concentrations in the plasma (E and F). The prediction-corrected VPC plots show median and 90% observation intervals (solid and dashed lines, respectively), overlaid with prediction percentiles (10, 50, and 90%, solid shaded areas). CWRES plots show the ideal fit (horizontal gray line, $\text{CWRES} = 0$) and the actual fit (red broken line). For the CWRES-versus-time plot, CWRES data points that pertain to propofol alone are given by red circles, remifentanyl by blue circles, and the combination by open circles. VPCs were constructed using 1000 simulations.

(Fig. 4D). Note that, although propofol does not affect K_{deg} directly, it indirectly reduces it through its action on M .

Figures 5 and 6 are simulations, restricted to the concentration range adequately covered by our data (remifentanyl ≤ 3.0 ng/ml and propofol ≤ 4.0 $\mu\text{g/ml}$). Figure 5 shows isobolograms corresponding to a 10 and 20% increase in $p\text{CO}_2$ from baseline once steady-state conditions are achieved, suggesting a synergistic relationship between propofol and remifentanyl. Figure 6 gives the time course of recovery following termination of an infusion ($t = 0$ is steady state). Note that, at time 0, the system

is assumed to be at steady state. Predicted $p\text{CO}_2$ returns to near baseline levels within 30 minutes for most concentration combinations, although some fluctuations exist due to the effect of the modulator/feedback components of the model.

Discussion

Propofol with remifentanyl is a popular hypnotic-opioid combination commonly used for anesthesia and sedation. Although several models for respiratory depression exist for

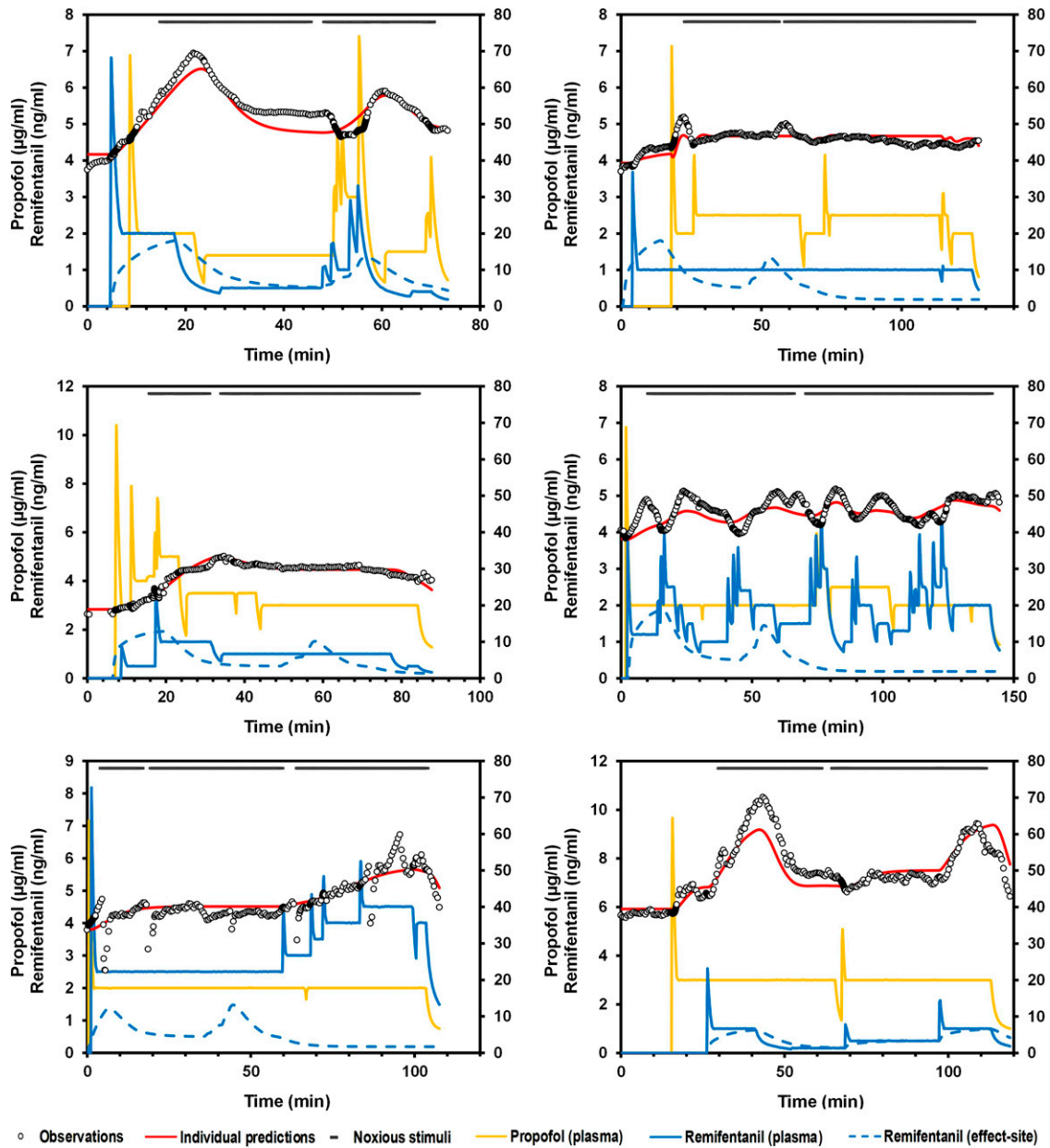


Fig. 3. Plots of individual fits for six participants, selected at random. Predicted plasma concentrations are given for propofol (yellow line), and for predicted plasma and effect-site concentrations for remifentanyl (blue solid and broken lines, respectively). Observed $p\text{CO}_2$ are open black circles, with individual model predictions in solid red lines. Durations of noxious stimuli are indicated by the horizontal black lines visible at the top of each plot.

healthy volunteers, or patients receiving just one of these drugs, a model for their combined effects on respiratory depression in patients undergoing noxious procedures has yet to be reported. We developed an indirect-effect model with system feedback to describe changes in $p\text{CO}_2$ induced by propofol and remifentanyl. OPRM1 genotype and noxious stimuli were not significant covariates in our data set. A combination of propofol 1.8 $\mu\text{g}/\text{ml}$ propofol and remifentanyl 1.5 ng/ml , which induces a sedation level where the patient is not responsive to verbal command but is rousable, has an expected $p\text{CO}_2$ response of 55.7 mm Hg (assuming steady-state conditions, basal $p\text{CO}_2$ of 39 mm Hg in a 65-year-old, 70-kg male).

We found remifentanyl potently inhibits $p\text{CO}_2$ removal, with an effect-site IC_{50} of 1.13 ng/ml . This is similar to that reported

in healthy volunteers (0.92–1.6 ng/ml) (Babenco et al., 2000; Bouillon et al., 2003; Olofsen et al., 2010). Onset of remifentanyl effects was slow, with a k_{e0} of 0.28 minute^{-1} ($t_{1/2k_{e0}}$ of 2.48 minutes) that increased with age. Others suggest somewhat faster onset (k_{e0} 0.34–1.3 minutes^{-1} , $t_{1/2k_{e0}}$ 0.53–2.03 minutes) for respiratory depressant effects (Babenco et al., 2000; Bouillon et al., 2003; Olofsen et al., 2010). This difference may be partly due to our older patient population (median age of 64.0 years in comparison with healthy volunteers aged <45 years). Slower onset with increasing age has also been reported for remifentanyl electroencephalograph pharmacodynamics (Minto et al., 1997). Propofol had an IC_{50} of 4.97 $\mu\text{g}/\text{ml}$ in plasma. Older individuals were more sensitive to propofol, with age-adjusted IC_{50} estimates of 2.65 and 1.9 $\mu\text{g}/\text{ml}$ in 50 and 65 year olds, respectively. An IC_{50} for propofol in the effect site of 1.33 $\mu\text{g}/\text{ml}$

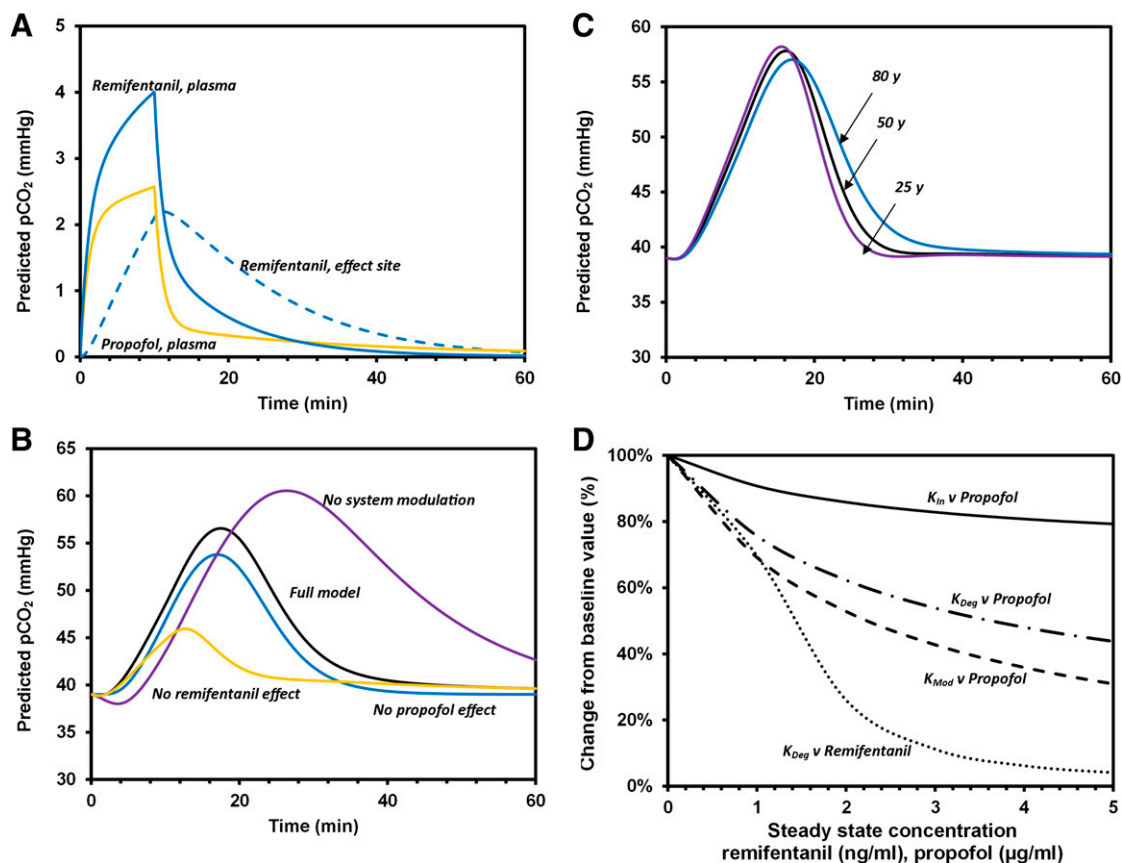


Fig. 4. Contribution of different elements of the final model over time. Simulation shows the time course of drug concentrations for a 10-minute fixed infusion of 2.0 $\mu\text{g/ml}$ propofol and 2.5 ng/ml remifentanyl (based on literature population pharmacokinetic models; see *Materials and Methods*) (A), and the corresponding change in predicted pCO_2 for 1) the full model (solid black line), 2) ignoring the contribution of remifentanyl, 3) ignoring the contribution of propofol, and 4) ignoring the contribution of the modulator compartment (B). (C) The contribution of age on pCO_2 for the same infusion inputs. (D) The percentage change from baseline value for K_{deg} and K_{mod} parameters with increasing steady-state concentrations of either drug alone.

was reported in healthy young adults (Bouillon et al., 2004a). Our estimate is higher, partly because we did not include an effect-site compartment for propofol. The corresponding IC_{50} in the effect site will be lower, as the drug is transferred more slowly and in smaller amounts to this compartment (dictated by the k_{e0} parameter). Propofol effects on tidal volume have a reported IC_{50} of 3.0 $\mu\text{g/ml}$ in children undergoing sedation for endoscopy (Hahn et al., 2011). Remifentanyl-propofol effects on ventilatory response to stepped increases in pCO_2 have been studied in healthy volunteers (Nieuwenhuijs et al., 2003). In these controlled, steady-state conditions, propofol predominantly suppressed the slope of the ventilatory response (IC_{50} of 1.0 $\mu\text{g/ml}$) and had a much smaller effect on reducing the set-point of that response. Our estimate of baseline pCO_2 was less than that typically reported (36.4 mm Hg/70 kg vs. 40.9–42.4 mm Hg in other studies) (Bouillon et al., 2003, 2004a; Nieuwenhuijs et al., 2003; Caruso et al., 2007). Elevated ventilation rate in study participants as a result of preinduction anxiety sometimes occurs (Goodman et al., 1987) and may also be true of our patients, accounting for our lower baseline pCO_2 . We also scaled baseline pCO_2 to weight; this was mandated by our data and a persistent misspecification in our checks of model performance. There are neither literature data nor a physiologic basis that we are aware of that supports the covariate effect of body weight on the baseline pCO_2 parameter. However, with this covariate in the selected model, model

performance represented by visual predictive checks was greatly improved over the model without its inclusion. We recognize that such part of our model indicated some degree of model misspecification, probably at a different level from baseline, that could not be handled in another way.

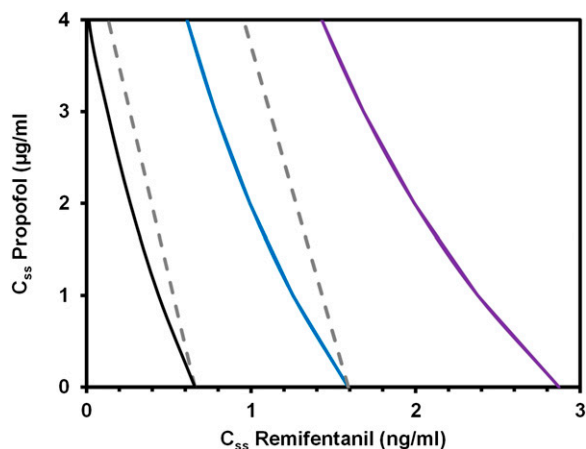


Fig. 5. Isoboles for steady-state concentrations of remifentanyl and propofol that cause 10 and 20% increases in pCO_2 from baseline. Broken lines indicate additive effects, whereas solid lines show model predictions and bow toward the plot origin, suggesting a synergistic relationship.

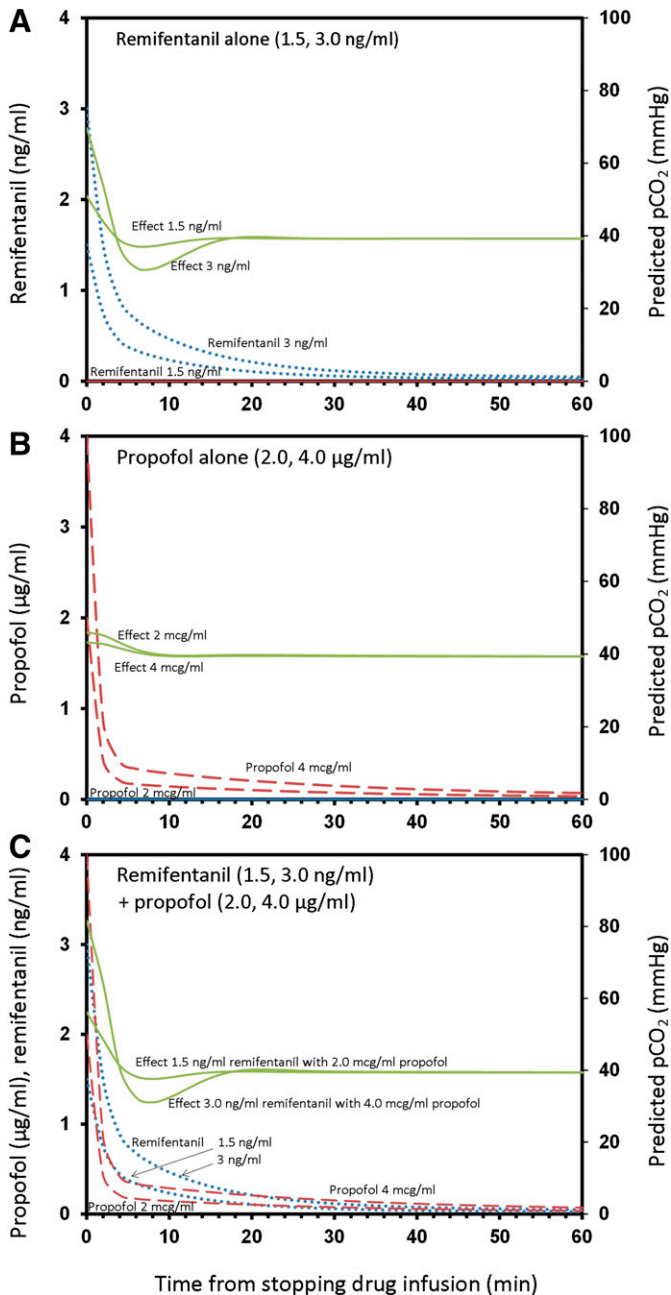


Fig. 6. Simulated time to recovery following termination of drug administration, from steady-state conditions. Plasma profiles for propofol (red broken lines) and remifentanyl (blue broken lines) are simulated using Schnider and Minto pharmacokinetic models, respectively. Predicted $p\text{CO}_2$ profiles are given by solid lines. The panels show profiles for: A) remifentanyl given alone, B) propofol given alone, and C) combined administration of remifentanyl and propofol. The system is assumed to be at steady state at time = 0.

The remifentanyl IC_{50} estimate for bispectral index suppression in the same patients was much larger than that estimated for $p\text{CO}_2$ (19.6 ng/ml) (Borrat et al., 2013). The inability of remifentanyl to substantively impact bispectral index leading to high IC_{50} estimates is well documented (Nieuwenhuijs et al., 2003; Manyam et al., 2007) and is indicative of its low impact on sedation levels (Bouillon et al., 2004b). Conversely, we saw a smaller IC_{50} estimate for propofol for bispectral index (3.86 $\mu\text{g}/\text{ml}$ in the effect site)

than that estimated for $p\text{CO}_2$, in line with propofol's potent sedative and anesthetic effects and smaller impact on the respiratory system.

Our model most closely resembles that of Bouillon et al. They described single-drug effects using CO_2 arterial and effect-site compartments (Bouillon et al., 2003, 2004a). Drug concentration indirectly affects CO_2 elimination from the arterial compartment (estimated at 0.08–0.11 minute^{-1} in volunteers, similar to our K_{deg} parameter at 0.06 minute^{-1}) (Bouillon et al., 2003, 2004a). They also applied system feedback to CO_2 elimination (using an equivalent function to eq. 3), the delay of which was dependent on the parameter describing the CO_2 transfer rate between compartments (k_{el,CO_2} , 0.9 minute^{-1}) (Bouillon et al., 1999, 2003, 2004a). In our model, feedback delay is described by K_{mod} (0.45 minute^{-1}). Our estimate of gain in the system response to increasing $p\text{CO}_2$ (α), at 3.82, was close to reported values of 4.3–4.37 established in single-drug studies in volunteers (Bouillon et al., 2003, 2004a). The large confidence intervals surrounding this parameter estimate reflect the uncontrolled, non-steady-state conditions of our study.

Olofsen et al. (2010) also used two compartments (tissue and alveolar) to describe CO_2 pharmacokinetics, with remifentanyl reducing inspired ventilation. Their model reflects the observation that opioids alter the baseline (or set-point) of the ventilatory response to rising $p\text{CO}_2$, whereas propofol alters the slope of that response (Nieuwenhuijs et al., 2003). They included both remifentanyl and propofol effects, but delay in system feedback was not estimated and propofol was incorporated as a (binary) covariate effect on system and remifentanyl parameters. Unlike these previous models, we grouped $p\text{CO}_2$ kinetics into a single compartment and described system modulation using compartmental kinetics. Propofol effects were applied to the rate of synthesis in the modulator compartment, thereby affecting the magnitude of the response to rising $p\text{CO}_2$. Remifentanyl effects were applied directly to the parameter describing $p\text{CO}_2$ removal, as done by others for opioids (usually minute ventilation, in our model K_{deg}) (Bouillon et al., 1999, 2003; Caruso et al., 2008; Olofsen et al., 2010). Thus, we include independent, concentration-based drug effects for both propofol and remifentanyl on $p\text{CO}_2$.

We modeled $p\text{CO}_2$ as an objective biomarker of respiratory depression. Previous work has established the correlation between $p\text{CO}_2$ and alveolar $p\text{CO}_2$ (Chhajed et al., 2010; Rollins et al., 2014). An absolute value above 75 mm Hg, in the severe hypercapnia range, can affect several organs and systems and may cause decreased cerebral blood flow, increased plasma catecholamine levels, and increased cardiac output and arterial blood pressure predisposing to severe arrhythmias. Hypercapnic pulmonary vasoconstriction augments hypoxic pulmonary vasoconstriction and may worsen right heart function. Values above 150 mm Hg have been associated with stupor and coma. Hypercapnia cannot easily be diagnosed clinically but is obvious with the aid of a quantitative CO_2 measurement system (Lumb, 2000). The trend of continuous measures of $p\text{CO}_2$ gives an idea of the global performance of the respiratory drive. Using this monitor in the clinical setting might be advantageous, particularly in patients breathing spontaneously, where capnography, transthoracic impedance measurement of respiratory rate, or estimation of tidal volume methods is not reliable. We found that we often had issues maintaining sensor contact in lightly sedated patients who

frequently moved. Consequently, data were unavailable for 71 of 207 participants, usually due to an unstable connection or signal. We note that newer sensors are now available that can be securely fixed to the chest using tape, and these may provide a more stable method of measuring transcutaneous pCO₂. Arterial blood sampling, the gold standard for pCO₂, is not a continuous measure, nor is it practical in this setting for obvious reasons.

We could not detect altered pCO₂ response for A118G polymorphic patients. Similarly, Romberg et al. (2005) did not detect differences in respiratory effects despite an increase in analgesic requirements. Noxious stimulation is usually associated with increased respiratory rate, which should decrease pCO₂. Although there was a trend, NOX was not included in the model based on our a priori criteria for covariate inclusion. The effect of age suggests that CO₂ washout is slower in older patients.

This model could be used to explore concentration ranges previously proposed as optimal for sedation, and to simulate expected pCO₂ levels while incorporating covariate and interindividual variability factors. This would help define rational and safe sedation ranges that avoid or minimize the consequences of respiratory depression and increased pCO₂. Automatic control closed-loop systems have already been used for adjusting propofol and remifentanyl to hypnotic endpoints using the BIS (Liu et al., 2011). Sedation and analgesia techniques might benefit from an automatic system able to use two different endpoints—hypnotic level on one side and pCO₂ as Bouillon et al. (2003) proposed for remifentanyl and pCO₂ (Caruso et al., 2006).

Several limitations of our work should be acknowledged. Modulation of the respiratory system occurs via several physiologic processes (Lloyd et al., 1958; Dahan et al., 1990; Ward and Karan, 2002). This makes estimation of model parameters difficult, even in controlled conditions and ventilation studies. We studied patients undergoing an uncomfortable procedure with anesthetic polypharmacy in non-steady-state conditions and all components of the respiratory system in play. Although an advantage is that our data reflect the clinical environment, this impedes our ability to identify and quantify system factors. Hypercarbic and hypoxic respiratory drives vary among individuals (Sahn et al., 1977). We did not establish individual sensitivity to rising CO₂, and our population may include outlier individuals. We modeled all processes of system modulation together as one process (in one compartment), which is physiologically inaccurate but does provide an adequate description of our data. An inhibitory effect of hypnotics on CO₂ production has been documented (Pavlin et al., 1996) and should be included to avoid biased parameter estimates (Bouillon et al., 2004a). We assumed only propofol inhibits CO₂ production, up to 30% of baseline (Bouillon et al., 2004a; Caruso et al., 2007, 2008). Of course, this assumption may be incorrect, particularly where multiple drugs are administered. We did notice parameter estimates were better aligned with literature values once this correction was included. We also had a high rate of dropouts as discussed earlier, although these were fairly random across the four groups (with perhaps some increased dropout in those individuals receiving remifentanyl first; see *Results*).

Using clinical data from patients undergoing sedation with analgesia, with noninvasively and continuously measured

pCO₂, we developed a pharmacokinetic-pharmacodynamic model characterizing a synergistic relationship for propofol and remifentanyl for respiratory depression. Neither A118G SNP in the OPRM1 gene nor noxious stimulation influenced the respiratory effects of remifentanyl in our data set. Age significantly affected the propofol and remifentanyl relationship with pCO₂, with older patients more prone to respiratory depression. Context-sensitive decrement times show that recovery from hypercapnia is fast, and within 15 minutes, pCO₂ nears baseline irrespective of the residual drug concentrations.

Authorship Contributions

Participated in research design: Borrat, Trocóniz, Castells, Gambús.

Conducted experiments: Borrat, Valencia, Jensen, Pedroso, Muñoz, Castellví-Bel, Castells, Gambús.

Performed data analysis: Hannam, Trocóniz, Valencia, Gambús.

Wrote or contributed to the writing of the manuscript: Hannam, Trocóniz, Gambús, Borrat.

References

- Babenco HD, Conard PF, and Gross JB (2000) The pharmacodynamic effect of a remifentanyl bolus on ventilatory control. *Anesthesiology* **92**:393–398.
- Bergstrand M, Hooker AC, Wallin JE, and Karlsson MO (2011) Prediction-corrected visual predictive checks for diagnosing nonlinear mixed-effects models. *AAPS J* **13**: 143–151.
- Blouin RT, Conard PF, and Gross JB (1991) Time course of ventilatory depression following induction doses of propofol and thiopental. *Anesthesiology* **75**:940–944.
- Borrat X, Trocóniz IF, Valencia JF, Rivadulla S, Sendino O, Llach J, Muñoz J, Castellví-Bel S, Jospin M, and Jensen EW, et al. (2013) Modeling the influence of the A118G polymorphism in the OPRM1 gene and of noxious stimulation on the synergistic relation between propofol and remifentanyl: sedation and analgesia in endoscopic procedures. *Anesthesiology* **118**:1395–1407.
- Bouillon T, Bruhn J, Radu-Radulescu L, Andresen C, Cohane C, and Shafer SL (2003) A model of the ventilatory depressant potency of remifentanyl in the non-steady state. *Anesthesiology* **99**:779–787.
- Bouillon T, Bruhn J, Radu-Radulescu L, Andresen C, Cohane C, and Shafer SL (2004a) Mixed-effects modeling of the intrinsic ventilatory depressant potency of propofol in the non-steady state. *Anesthesiology* **100**:240–250.
- Bouillon T, Schmidt C, Garstka G, Heimbach D, Stafforst D, Schwilden H, and Hoefl A (1999) Pharmacokinetic-pharmacodynamic modeling of the respiratory depressant effect of alfentanil. *Anesthesiology* **91**:144–155.
- Bouillon TW, Bruhn J, Radulescu L, Andresen C, Shafer TJ, Cohane C, and Shafer SL (2004b) Pharmacodynamic interaction between propofol and remifentanyl regarding hypnosis, tolerance of laryngoscopy, bispectral index, and electroencephalographic approximate entropy. *Anesthesiology* **100**:1353–1372.
- Box GEP and Cox DR (1964) An analysis of transformations. *J R Stat Soc, B* **26**:211–252.
- Caruso A, Bouillon T, Zanderigo A, Schumacher PM, and Luginbühl M (2006) Closed loop administration of remifentanyl for monitored anesthesia care using PCO₂ as an endpoint. Proceedings of the ASA annual meeting. *Anesthesiology* **105**:A1198.
- Caruso AL, Bouillon TW, Schumacher PM, Luginbühl M, and Morari M (2007) Drug-induced respiratory depression: an integrated model of drug effects on the hypercapnic and hypoxic drive. *Conf Proc IEEE Eng Med Biol Soc* **2007**:4259–4263.
- Caruso AL, Bouillon TW, Schumacher PM, and Morari M (2008) On the modeling of drug induced respiratory depression in the non-steady-state. *Conf Proc IEEE Eng Med Biol Soc* **2008**:5564–5568.
- Chhajed PN, Miedinger D, Baty F, Bernasconi M, Heuss LT, Leuppi JD, and Tamm M (2010) Comparison of combined oximetry and cutaneous capnography using a digital sensor with arterial blood gas analysis. *Scand J Clin Lab Invest* **70**:60–64.
- Dahan A, DeGoede J, Berkenbosch A, and Olivevier IC (1990) The influence of oxygen on the ventilatory response to carbon dioxide in man. *J Physiol* **428**:485–499.
- Dayneka NL, Garg V, and Jusko WJ (1993) Comparison of four basic models of indirect pharmacodynamic responses. *J Pharmacokinetic Biopharm* **21**:457–478.
- Dershwitz M, Hoke JF, Rosow CE, Michalowski P, Connors PM, Muir KT, and Dienstag JL (1996) Pharmacokinetics and pharmacodynamics of remifentanyl in volunteer subjects with severe liver disease. *Anesthesiology* **84**:812–820.
- Dixon WJ (1991) Staircase bioassay: the up-and-down method. *Neurosci Biobehav Rev* **15**:47–50.
- Goodman NW, Black AM, and Carter JA (1987) Some ventilatory effects of propofol as sole anaesthetic agent. *Br J Anaesth* **59**:1497–1503.
- Hahn JO, Khosravi S, Dosani M, Dumont GA, and Mark Ansermino J (2011) Pharmacodynamic modeling of propofol-induced tidal volume depression in children. *J Clin Monit Comput* **25**:275–284.
- Joyce RR and McGee WT (2011) Hypercapnic cerebral edema presenting in a woman with asthma: A case report. *J Med Case Rep* **5**:192.
- Klepstad P, Rakvåg TT, Kaasa S, Holthe M, Dale O, Borchgrevink PC, Baar C, Vikan T, Krokan HE, and Skorpen F (2004) The 118 A > G polymorphism in the human mu-opioid receptor gene may increase morphine requirements in patients with pain caused by malignant disease. *Acta Anaesthesiol Scand* **48**:1232–1239.

- Liu N, Chazot T, Hamada S, Landais A, Boichut N, Dussaussoy C, Trillat B, Beydon L, Samain E, and Sessler DI, et al. (2011) Closed-loop coadministration of propofol and remifentanyl guided by bispectral index: a randomized multicenter study. *Anesth Analg* **112**:546–557.
- Lloyd BB, Jukes MG, and Cunningham DJ (1958) The relation between alveolar oxygen pressure and the respiratory response to carbon dioxide in man. *Q J Exp Physiol Cogn Med Sci* **43**:214–227.
- Lötsch J and Geisslinger G (2005) Are mu-opioid receptor polymorphisms important for clinical opioid therapy? *Trends Mol Med* **11**:82–89.
- Lumb AB, ed, ed (2000) *Changes in the carbon dioxide tension, Nunn's applied respiratory physiology*, Ed. 5th. Butterworth-Heinemann, Oxford.
- Macey R and Oster G. University of California, Berkeley, USA. (2010) Berkeley madonna™ version 8.3.18.
- Manyam SC, Gupta DK, Johnson KB, White JL, Pace NL, Westenskow DR, and Egan TD (2007) When is a bispectral index of 60 too low?: Rational processed electroencephalographic targets are dependent on the sedative-opioid ratio. *Anesthesiology* **106**:472–483.
- Minto CF, Schnider TW, and Shafer SL (1997) Pharmacokinetics and pharmacodynamics of remifentanyl. II. Model application. *Anesthesiology* **86**:24–33.
- Nieuwenhuijs DJ, Olofsen E, Romberg RR, Sarton E, Ward D, Engbers F, Vuyk J, Mooren R, Teppema LJ, and Dahan A (2003) Response surface modeling of remifentanyl-propofol interaction on cardiorespiratory control and bispectral index. *Anesthesiology* **98**:312–322.
- Olofsen E, Boom M, Nieuwenhuijs D, Sarton E, Teppema L, Aarts L, and Dahan A (2010) Modeling the non-steady state respiratory effects of remifentanyl in awake and propofol-sedated healthy volunteers. *Anesthesiology* **112**:1382–1395.
- Ootaki C, Stevens T, Vargo J, You J, Shiba A, Foss J, Borkowski R, and Maurer W (2012) Does general anesthesia increase the diagnostic yield of endoscopic ultrasound-guided fine needle aspiration of pancreatic masses? *Anesthesiology* **117**:1044–1050.
- Pavlin DJ, Coda B, Shen DD, Tschanz J, Nguyen Q, Schaffer R, Donaldson G, Jacobson RC, and Chapman CR (1996) Effects of combining propofol and alfentanil on ventilation, analgesia, sedation, and emesis in human volunteers. *Anesthesiology* **84**:23–37.
- Rollins MD, Feiner JR, Lee JM, Shah S, and Larson M (2014) Pupillary effects of high-dose opioid quantified with infrared pupillometry. *Anesthesiology* **121**:1037–1044.
- Romberg RR, Olofsen E, Bijl H, Taschner PE, Teppema LJ, Sarton EY, van Kleef JW, and Dahan A (2005) Polymorphism of mu-opioid receptor gene (OPRM1:c.118A>G) does not protect against opioid-induced respiratory depression despite reduced analgesic response. *Anesthesiology* **102**:522–530.
- Sahn SA, Zwillich CW, Dick N, McCullough RE, Lakshminarayan S, and Weil JV (1977) Variability of ventilatory responses to hypoxia and hypercapnia. *J Appl Physiol* **43**:1019–1025.
- Schnider TW, Minto CF, Gambus PL, Andresen C, Goodale DB, Shafer SL, and Youngs EJ (1998) The influence of method of administration and covariates on the pharmacokinetics of propofol in adult volunteers. *Anesthesiology* **88**:1170–1182.
- Schnider TW, Minto CF, Shafer SL, Gambus PL, Andresen C, Goodale DB, and Youngs EJ (1999) The influence of age on propofol pharmacodynamics. *Anesthesiology* **90**:1502–1516.
- Sheiner LB, Stanski DR, Vozeh S, Miller RD, and Ham J (1979) Simultaneous modeling of pharmacokinetics and pharmacodynamics: application to d-tubocurarine. *Clin Pharmacol Ther* **25**:358–371.
- Skarke C, Darimont J, Schmidt H, Geisslinger G, and Lötsch J (2003) Analgesic effects of morphine and morphine-6-glucuronide in a transcutaneous electrical pain model in healthy volunteers. *Clin Pharmacol Ther* **73**:107–121.
- Spindelboeck W and Moser A (2012) Spontaneous tension pneumothorax and CO₂ narcosis in a near fatal episode of chronic obstructive pulmonary disease exacerbation. *Am J Emerg Med* **30**:1664.e3–1664.e4.
- Wakelkamp M, Alván G, Gabrielsson J, and Paintaud G (1996) Pharmacodynamic modeling of furosemide tolerance after multiple intravenous administration. *Clin Pharmacol Ther* **60**:75–88.
- Ward DS and Karan S (2002) Effects of pain and arousal on the control of breathing. *J Anesth* **16**:216–221.

Address correspondence to: Dr. Pedro L. Gambús, Anesthesiology Department, Hospital CLINIC de Barcelona, C/ Villarroel 170, 08036 Barcelona, Spain. E-mail: plgambus@hospitalclinic.org
