Antioxidants and Myocardial Ischemia: Reperfusion Injuries

Jan-Kan Chen, PhD; Shu-Er Chow¹, PhD

Animal studies have demonstrated that restoration of blood flow to severely ischemic myocardium is a prerequisite for myocardial salvage. However, it has been shown that the restoration of blood flow to ischemic myocardium may be associated with deleterious changes of the myocardium, including arrhythmias, enzyme release, and contractile dysfunction. These changes were considered to be additional injuries to the myocardium manifested at the time of reperfusion. The reperfusion was accompanied by a burst of oxygen free radical generation and their role as main mediators of the reperfusion injury have been well accepted. Reactive oxygen species (ROS) and cellular redox status regulate many important cellular activities. The role of antioxidant as a therapy for reperfusion injury has thus been tried with mixed and mostly negative results. Further studies are needed if the antioxidant therapies for ischemia reperfusion injury were to be effective.



Prof. Jan-Kan Chen

(Chang Gung Med J 2005;28:369-77)

Key words: polyphenolics, ROS, infarction.

It is well known that there is a positive correlation between the levels of dietary saturated fat and the mortality from coronary heart disease (CHD).⁽¹⁾ An interesting statistics however, showed that compared to other developed countries such as the USA, France has a much lower incidence of CHD despite the fact that they consume comparable amounts of dietary fat. This phenomenon, termed the French Paradox,⁽²⁾ is thought to result from a higher consumption of red wines by the French.^(3,4) Although some researchers suggest that the beneficial effects of alcohol are probably due to its hemostatic activity,^(5,6) it is now clear that wine, particularly red wine, is rich in phenolic compounds, and some of which have significant cardioprotective activities.^(7,8) The natural phenolic compounds include two major classes, the flavonoids and non-flavonoids. The non-flavonoid compounds in wine include stilbene, hydroxyl cinnamates and hydroxybenzoates. The compound that is responsible for possible cardiovascular benefits is the stilbene resveratrol.^(7.9) Resveratrol (RSV, trans-3, 5, 4'-trihydroxystilbene) was first isolated from the roots of the oriental herb and medicinal plant *Polygonum Cuspidatum*.⁽¹⁰⁾ It was also synthesized by leaf tissue from grapevine in response to fungal infection.⁽¹¹⁾ In a grape, RSV is found mainly in the skin. Among red wines, RSV contents varied from 2 to 10 µg/ml, while white wines had concentrations around 1 to 2 µg/ml.

Many studies have shown that RSV is a potent

From the Department of Physiology, College of Medicine; 'Center for General Studies, Chang Gung University, Taoyuan. Received: Mar. 30, 2005; Accepted: Apr. 12, 2005

Address for reprints: Prof. Jan-Kan Chen, Department of Physiology, College of Medicine, Chang Gung University. No. 259, Wen-Hwa 1st Rd., Gueishan Shiang, Taoyuan 333, Taiwan, R.O.C. Tel.: 886-3-2118800 ext. 5077; Fax: 886-3-2118700; E-mail: jkc508@mail.cgu.edu.tw

antioxidant.⁽¹²⁻¹⁴⁾ It prevents copper ion induced lipid peroxidation of the low density lipoprotein.^(12,15) In addition to its antioxidant activity, RSV has also been shown to suppress cell proliferation,⁽¹⁶⁻¹⁸⁾ promote cell differentiation,⁽¹⁹⁾ induce apoptosis,⁽²⁰⁻²²⁾ inhibit inflammation,^(13,23) scavange reactive oxygen species,⁽²⁴⁾ inhibit platelet aggregation,⁽²⁵⁾ and have cancer chemopreventive activity.⁽²⁶⁾ More recently, RSV was found to protect endothelial cells from oxidative damage and ameliorate ischemia-reperfusion injuries in a number of experimental models. In the present article, we review the roles of ROS in ischemia-reperfusion injury (IR injury) of the heart and the effect of some antioxidants in ameriolating the myocardial IR injury.

Redox imbalance and myocardial IR Injury

Inadequate blood supply to a region of the body for a certain period followed by the resumption of blood flow is termed ischemia-reperfusion. Ischemia-reperfusion results in varying degrees of tissue damage depending on the duration and extent of the hypoperfusion.

Myocardial damage induced by ischemia-reperfusion is due, at least in part, to the generation of ROS.⁽²⁷⁻³¹⁾ Evidence supporting ROS as a culprit of myocardial IR injury came from several direct and indirect observations. There have been reports showing a close correlation between the production of ROS and simultaneous consumption of endogenous antioxidants.(32-35) Indirect evidence consistent with this view is the cardioprotective effects of free radical scavengers and antioxidant supplements.⁽³⁶⁻³⁸⁾ In addition, direct genetic manipulations to overexpress or underexpress genes participating in the antioxidant defense also exhibit profound influence on the outcome of IR injury.⁽³⁹⁻⁴⁵⁾ It would seem that the link between ROS and IR injury has appeared to have been clear-cut, however, contradicting results have been reported regarding the effects of antioxidants on IR injury.

Inadequate perfusion of a tissue/organ leads to oxygen (O_2) and adenosine triphosphate (ATP) depletion, and the accumulation of toxic metabolites. Another effect of hypoperfusion is the conversion of xanthin dehydrogenase to xanthin oxidase, which upon reperfusion, catalyzes the conversion of hypoxanthine to xanthine with the concomitant production of ROS.

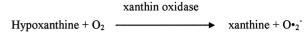


Fig. 1 Production of an oxygen radical by xanthine oxidase

Oxygen radicals (O_{2}^{-}) are also produced by the electron transport system of the mitohondria and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. The highly toxic ROS are converted to hydrogen peroxide (H₂O₂) by superoxide dismutase (SOD), and then to H₂O by catalase and/or glutathione oxidase (Fig. 2). However, under ischemic conditions, the endogenous antioxidant system is eroded and the tendency for metal ion assisted conversion of H₂O₂ into the destructive hydroxyl radical (OH•⁻) is increased.⁽⁴⁶⁾

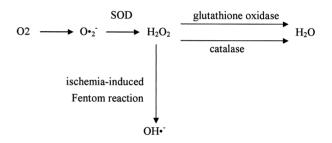


Fig. 2 ROS metabolism under normal and ischemic conditions

In addition to IR injuries, ROS have also been implicated in many clinical conditions including atherosclerosis, autoimmune diseases, alcoholic liver disease, and various inflammation related disorders. Accumulated evidence has shown that ROS production is a key event in reperfusion injury when oxygen is reintroduced to ischemic tissues.⁽⁴⁶⁻⁴⁹⁾ ROS, especially hydroxyl radical, cause the oxidation of proteins, lipids and nucleic acids, resulting in the structural and functional changes of proteins, disruption of membrane intergrity, and genetic mutations, respectively. ROS also cause severe functional and metabolic disorders, and such effects can be systemic, leading to multi-organ failure. ROS mediated reperfusion injury has been observed in heart, liver, lung, kidney and intestine. The IR injury of the heart is discussed below.

IR injury to vascular endothelium

ROS increase in concentration upon reperfusion

of the ischemic myocardium.⁽⁵⁰⁾ The formation of ROS exerts oxidative stress to the myocardium that may cause heart failure. The major ROS that are responsible for the oxidative stress are superoxide anion (O_{2^-}) , hydroxyl radical (OH^{-}) and H_2O_2 In the vascular walls, the enzyme systems involved in the production of these radicals including xanthine oxidase, NADPH oxidase and the endothelial nitric oxide (NO) synthase (eNOS). Because of its location, the endothelium is probably the prime target for ROS damage.

The endothelium-dependent vasorelaxation activity is highly sensitive to IR injury. Elevated levels of ROS reduce the bioavailability of NO through reacting with NO to form peroxynitrite.⁽⁵¹⁾ The reduced NO availability aggravates local oxidative stress by the formation of peroxynitrite and a reduced blood flow due to decreased NO availability. ROS have also shown to disrupt the integrity of the endothelial cell junctions leading to increased endothelial permeability, tissue edema and protein leakage.⁽⁵²⁾ Additional endothelial dysfunction related to ROS include production of proinflammatory cytoicines, activation of complement system, decreased production of prostacyclin (PGI₂), increased production of platelet activation factor (PAF), and thromboxane A₂ (TXA₂) and increased expression of adhesion molecules.⁽⁵³⁾

The decreased PGI₂ and increased PAF and TXA₂ productions by endothelium would certainly compromise the nonthrombogenic nature of the vascular surface. In an in vitro study, we showed that oxidized low density lipoprotein (oxLDL) dose-dependently reduced the ability of endothelial cells (EC) to stabilize platelets from adenosine diphosphate-induced (ADP-induced) aggregation and platelet [Ca⁺²]i rise (Fig. 3). Treatment of EC with RSV (from 5 μ M to 20 μ M) prior to oxLDL exposure effectively preserved the anti-platelet aggregation activity of the EC. The exposure time required for the maximal effect of RSV appeared to be short; 30 min incubation and 2 hr incubation were equally effective.

IR injury to myocardium

In the myocardium, O_2 is reduced via (1) the mitochondrial electron transport system (cytochrome oxidase system), which reduces 95% of O_2 to H_2O by tetravalent reduction without the pro-

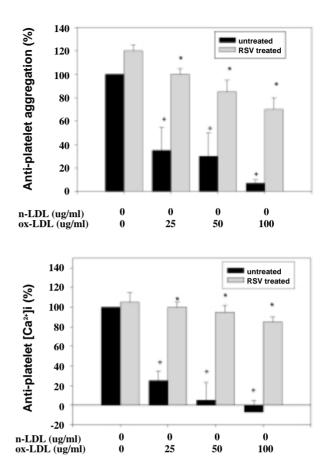


Fig. 3 RSV attenuates oxLDL-induced reduction of the antiplatelet aggregation activity of the endothelial cells. EC were treated with oxLDL as indicated for 1 hr with or without a 30 min prexposure to RSV before they were coincubated with platelets and ADP (4 μ M). Platelet activation was measured by aggregation (Panel A) and intraplatelet Ca⁺² rise (Panel B).

duction of ROS,⁽⁵⁴⁾ (2) the univalent pathway in which ROS are produced (Fig. 2). The endogenous antioxidant systems endow tissue with substantial ability to balance the ROS effect under normoxic conditions. However, under ischemia followed by reperfusion, the antioxidant defense is undermined and the oxidative damage to the tissue is ensured.

Metabolic disorder

A shortage of oxygen supply at the mitochondria level results in intracellular acidosis and an increased concentration of inorganic phosphate due to the breakdown of high energy phosphates. These early metabolic changes weaken the contractility of the ischemic zone.⁽⁵⁵⁾ The onset of anaerobic respiration and lactate release is another metabolic alteration which allows production of a small amount of ATP without the consumption of oxygen. However, prolonged ischemia results in a decrease in lactate production because anaerobic glycolysis is inhibited by further intracellular acidosis, and thus, a further drop in the intracellular ATP and creatine phosphate concentrations. At this stage of ischemia, the ionic conditions of the myocytes are altered, with an increase of the intracellular Na⁺¹ and a decrease of K⁺¹ and Mg⁺². Sodium ion extrusion through sarcolemmal $Na^{_{+1}}/K^{_{+1}}$ - ATPase is inhibited by a lack of ATP. Na⁺¹ influx leads to Ca⁺² influx since the Na⁺¹/Ca⁺² exchanger operates in reverse mode in Na⁺¹ -overloaded, depolarized cells. As a result, ischemic cells develop cytosolic Ca⁺² -overload. A Ca⁺² -overloaded ischemic cardiomyocyte may develop uncontrolled activation of the contractile machinery (contracture) upon reoxygenation.(56,57)

It has been shown that ischemic, Ca⁺² –overloaded cardiomyocytes develop hypercontracture immediately upon reperfusion.^(56,58) Reperfusion brings about the rapid recovery of the oxidative ATP production if the cytochrome oxidase system of the mitochondria was not damaged during the ischemic period. ATP provides energy for cardiomyocytes to recover from cytosolic ion imbalance and reactivate the contractile function. However, the contractile activation is usually faster than Ca⁺² recovery and it leads to a Ca⁺² - dependent hypercontracture.

ROS may damage sarcoplasmic reticulum causing Ca⁺² release and an increase of the cytosolic Ca⁺² levels.⁽⁵⁹⁾ This suggests that ROS formation and Ca⁺² surge might be involved in the contractile dysfunction of the ischemic myocardium (Fig. 4).

• Antioxidants and myocardial IR injury

It has become clear that redox balance is implicated in cell metabolism, signal transduction and gene expression.^(60,61) Cellular redox imbalance may compromise cell function and even cause cell death. Oxidative damage to protein, lipid and nucleic acid by ROS is well recognized. These oxidation reactions are believed to be implicated in numerous diseases in many organ systems including the cardiovascular system. However, antioxidants have also

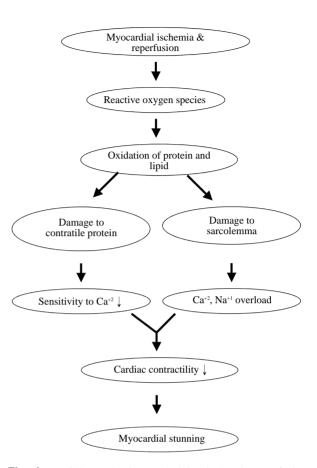


Fig. 4 Possible mechanism underlying ischemia reperfusion induced myocardial contractile stunning.

been shown to cause apoptosis of both normal and transformed cells. These observations strongly suggest that normal cellular function requires an optimal redox environment. The endogenous antioxidants in an ischemic tissue are believed to be eroded along the duration of the ischemia. To justify the use of antioxidants to prevent or ameriolate IR injury of a tissue or organ, it is necessary to establish a timedependent change of the antioxidant profile of the tissue following ischemia-reperfusion. Animal studies have consistently shown a depletion of myocardial nonenzymatic antioxidant levels. However, changes in the enzymatic antioxidants have been controversial,^(34,62-64) a decrease, increase, or maintaining no change in activities have been reported. In humans, the formation and release of oxidized glutathione (GSSG) in the coronary sinus following myocardial IR has been reported.⁽⁶⁵⁾ The release of GSSG was positively correlated with the duration of the ischemic period suggesting the consumption of glutathione (GSH) during cardiac ischemia. Work by De Vecchi et al., (1998)⁽⁶⁶⁾ showed massive reduction of glutathione in myocardium during bypass surgery and such glutathionine loss might be related to left ventricle dysfunction in ischemic human heart.

• Antioxidants as therapeutics

Numerous studies have evaluated the effects of antioxidants on IR injury in animals or in patients undergoing bypass surgery. Treatment of intestinal IR injury by antioxidants (Vitamins C and E, manitol and methyl prednisolone) in an animal model has been reported.⁽⁶⁷⁾ They showed that treatment with vit. C and manitol attenuated the IR injury, while treatment with vit. E and methyl prednisolone had no significant effect. Vitamins C, E, and thiol compounds, either alone or in combinations, have been used to evaluate the therapeutic effect on IR injury in patients undergoing cardiopulmonary bypass.⁽⁶⁸⁻⁷¹⁾ The results were mixed, as follows: protection by a high dose of vit. C (250 mg/kg bw) was observed in one study,(71) other studies showed that vitamin supplement was correlated with reduction in IR injuryrelated biochemical parameters, however, they were not consistently correlated with a more functional recovery, or clinical improvement.(68-70)

• Resveratrol ameriolates cardiac IR injury-Animal study

The effects of RSV administration on IRinduced cardiac injury have been studied in Sprague-Dawley rats subjected to myocardial ischemia by a temporary occlusion of the left main coronary artery.⁽⁷²⁾ In studies by Hung and colleagues, animals were infused with a boulus of RSV, at the desired doses, from the jugular vein 15 min before coronary occlusion.⁽¹⁴⁾ Adminstration of RSV was found to have no effect on the hemodynamic parameters of the sham operated animals.^(15,73,74) Occlusion of the coronary artery induced severe ventricular arrhythmias in animals of the vehicle group, which began after 6-7 min of occlusion, peaked after 8-12 min, and usually subsided at approximately 15 min. In the vehicle group, 100% of the animals developed ventricular tarchycardia (VT) and from 63% to 73% of the animals developed ventricular fibrillation (VF). Administration of RSV at 2.3x10⁻⁵ g/kg had no effect on ischemia-induced arrhythmias, or on mortality.⁽¹⁴⁾

A 5-min period of left main coronary artery occlusion, followed by 30-min reperfusion induced rhythm disturbances, and the severity of the disturbances is positively correlated with the ischemic duration. IR protocol-induced arrhythmias have been reported as relative to superoxide anion production.^(75,76) Administration of RSV at 15 min before IR, effectively reduced IR induced VT, VF, and the mortality rate of the experimental animals.⁽¹⁴⁾

In the same series of studies, Hung and colleagues evaluated myocardial damage by measuring plasma lactate dehydrogenase (LDH) level and the infarct size of the occluded zone. The left main coronary artery was occluded for 5-min followed by a 30min reperfusion period. Blood samples were taken at the end of the reperfusion. They found that RSV pretreatment reduced plasma LDH activity by more than 50% compared to the control animals.

To evaluate the effect of RSV on the infarct size, the left main coronary artery was occluded for 1 hr and reperfused for 3 hrs, or occluded for 4 hrs without reperfusion, and the infarct area was identified by staining with triphenyl tetrazolium chloride and –Evans blue.⁽⁷⁷⁾ Hung and colleagues found that RSV pretreatment reduced the infarct size, again, by more than 50%⁽⁷⁴⁾ compared to the control animals. These results clearly indicated that RSV possesses cardioprotective effect against IR induced injury of the myocardium.

Conclusions

Despite the fact that the *in vivo* measurement of ROS is rather difficult, the elevated production of ROS during IR is generally believed to be associated with myocardial tissue damage. Multiple enzymes and cell types are responsible for the accelerated ROS formation. However, because cells contain numerous antioxidant activities, it is therefore, uncertain whether an increase in ROS production is an accurate indication of oxidative damage. In addition, although oxidative damage of protein, lipids and nucleic acid by ROS have been well established, it has not been clear whether ROS produced at the time of reperfusion directly damage the myocardium.

The incomplete understanding of the roles of ROS in the IR injury plus the seemingly inconsistent results regarding the antioxidant effects in IR have hampered their therapeutic applications. Although vit. C, vit. E, and several thio compounds have been evaluated for alleviating IR injury in patients undergoing cardiopulmonary bypass, either alone or in combinations,⁽⁶⁸⁻⁷¹⁾ their beneficial effects still await further establishement.

Reports by Hung and colleagues^(14,15,73,74) clearly demonstrated that RSV administered 15 min before occlusion effectively alleviates ischemia-reperfusion induced rhythm disturbances and mortality. They found that the protective effects of RSV against ventricular arrhymia and mortality rate are NO-independent, while the protective effect against cardiomyocyte damage (LDTT and creatine kinase release) and infarction size are NO-dependent.⁽⁷⁴⁾ However, the mechanism underlying the protective effects of RSV against ischemia-reperfusion injury to date has not been fully elucidated, or their possible cytotoxic effect been evaluated. In addition, the possible existence of interspecies heterogeneity of the response to IR between human and rat also needs to be clarified. Before we can address all these issues. RSV and its derivatives will not be able to find their way into clinical applications in order to prevent or treat myocardial IR injury.

Acknowledgements

We are indebt to Dr. L-M Hung for part of the literature search during the preparation of this manuscript, and to Ms. H-W Huang for the typing work.

Supported by NSC grant 93-2320-B-182-002 from National Science Council, Taiwan.

REFERENCES

- 1. Renaud S, Gueguen R. The French paradox and wine drinking. Novartis Found Symp 1998;216:208-22.
- 2. Kopp P. Resveratrol, a phytoestrogen found in red wine. A possible explanation for the conundrum of the 'French paradox'? Eur J Endocrinol 1998;138(6):619-20.
- 3. Criqui MH, Ringel BL. Does diet or alcohol explain the French paradox? Lancet 1994;344:1719-23.
- 4. Renaud, S, de Lorgeril, M. Wine, alcohol, platelets, and the French paradox for coronary heart disease. Lancet 1992;339:1523-6.
- 5. Renaud S, Ruf CJ. Effects of alcohol on platelet function. Clin Chimica Acta 1996;246:77-89.
- Renaud SC, Gueguen, Schenker J, d'Hountaud A. Alcohol and Mortality in Middle-Aged Men from Eastern France. Epidemiology 1998;9:184-188.
- 7. Soleas GJ, Diamandis EP, Goldberg DM. Resveratrol: A molecule whose time has come? And gone? Clin Biochem

1997;30:91-113.

- 8. Pervaiz S. Resveratrol: from grapevines to mammalian biology. FASEB J 2003;14:1975-85.
- 9. Teissedre PL, Waterhouse AL, Walzem RL, German JB, Frankel EN, Ebeler SE, Clifford AJ. Phenolic Compounds of Grape and Wine and Health. Vin et Maladies Cardiovasculaires. Cahiers Scientifiques et Techniques. Office International de la Vigne et de Vin. 1997.
- Nonomura S, Kanagawa H, Makimoto A. Chemical constituents of polygonaceous plants. I. Studies on the components of ko-jo-kon. (polygonum cuspidatum sieb. Et zucc.) Yakugaku Zasshi 1963;83:988-90.
- 11. Langcake P. and Pryce RJ. The production of resveratrol by Vitis rinfera and other members of the Vitacea as a response to infection or injury. Physiological Plant Pathology 1976;9:77-86.
- 12. Belguendouz I, Fremont, Gozzalino MT. Interaction of transresveratrol with plasma lipoproteins. Biochem Pharmacol 1998;55:811-6.
- Rotondo S, Rajtar G, Manarini S, Celardo A, Rotilio D, DE Gaetano G, Evangelista V, Cerletti C: Effect of transresveratrol, a natural polyphenolic compound, on human polymorphonuclear leucocyte function. Br J Pharmacol 1998;123:1691-9.
- Hung LM, Chen JK, Huang SS, Lee RS, Su MJ. Cardioprotective effect of resveratrol, a natural antioxidant derived from grapes. Cardio Res 2000;47:549-55.
- Hung LM, Su MJ, Chu WK, Chiao CW, Chan WF, Chen JK. The protective effect of resveratrols on ischaemiareperfusion injuries of rat hearts is correlated with antioxidant efficacy. Br J Pharmacol 2002;135:1627-33.
- 16. Haworth RS, Avkiran M. Inhibition of protein kinase D by resveratrol. Biochem Pharmacol 2001;62:1647-51.
- 17. Tou JS, Urbizo C. Resveratrol inhibits the formation of phosphatidic acid and diglyceride in chemotactic peptideor phorbol ester-stimulated human neutrophils. Cell Signal 2001;13:91-7.
- Pozo-Guisado E, Alvarez-Barrientos A, Mulero-Navarro S, Santiago-Josefat B, Fernandez-Salguero PM. The antiproliferative activity of resveratrol results in apoptosis in MCF-7 but not in MDA-MB-231 human breast cancer cells: cell-specific alteration of the cell cycle. Biochem Pharmacol 2002;64(9):1375-86.
- 19. Mizutani K, Ikeda K, Kawai Y, Yamori Y. Resveratrol stimulates the proliferation and differentiation of osteoblastic MC3T3-E1 cells. Biochem Biophys Res Commun 1998;253(3):859-63.
- 20. Clement MV, Hirpara JL, Chawdhury SH, Pervaiz S. Chemopreventive agent resveratrol, a natural product derived from grapes, triggers CD95 signaling-dependent apoptosis in human tumor cells. Blood 1998;92(3):996-1002.
- Huang C, Ma WY, Goranson A, Dong Z. Resveratrol suppresses cell transformation and induces apoptosis through a p53-dependent pathway. Carcinogenesis 1999;20(2): 237-42.

- 22. Hsieh TC, Wu JM. Differential effects on growth, cell cycle arrest, and induction of apoptosis by resveratrol in human prostate cancer cell lines. Exp Cell Res 1999;249(1):109-15.
- Wadsworth TL, Koop DR. Effects of the wine polyphenolics quercetin and resveratrol on pro-inflammatory cytokine expression in RAW 264.7 macrophages. Biochem Pharmacol 1999;57(8):941-9.
- 24. Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. J Immunol 2000;64(12):6509-19.
- 25. Radomski MW, Palmer RM, Moncada S. The anti-aggregating properties of vascular endothelium: interactions between prostacyclin and nitric oxide. Br J Pharmacol 1987;3:639-46.
- 26. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, Fong HH, Farnsworth NR, Kinghorn AD, Mehta RG, Moon RC, Pezzuto JM. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. Science 1997;275(5297):218-20.
- Poltronieri R, Cevese A, Sbarbati A. Protective effect of selenium in cardiac ischemia and reperfusion. Cardioscience 1992;3(3):155-60.
- Gross GJ, Farber NE, Hardman HF, Warltier DC. Beneficial actions of superoxide dismutase and catalase in stunned myocardium of dogs. Am J Physiol 1986;250:372-7.
- 29. Opie LH. Reperfusion injury and its pharmacologic modification. Circulation 1989;80(4):1049-62.
- Kloner RA, Przyklenk K, Whittaker P. Deleterious effects of oxygen radicals in ischemia/reperfusion. Resolved and unresolved issues. Circulation 1989;80(5):1115-27.
- Kilgore KS, Lucchesi BR. Reperfusion injury after myocardial infarction: the role of free radicals and the inflammatory response. Clin Biochem 1993;26(5):359-70.
- 32. Godin DV and Garnett ME. Altered antioxidant status in the ischemic/reperfused rabbit myocardium: effects of allopurinol. Can J Cardiol 1989;5(7):365-71.
- 33. Pyles LA, Fortney JE, Kudlak JJ, Gustafson RA, Einzig S. Plasma antioxidant depletion after cardiopulmonary bypass in operations for congenital heart disease. J Thorac Cardiovasc Surg 1995;110(1):165-71.
- Ko KM, Garnett ME, Godin DV. Altered antioxidant status in ischemic/reperfused rabbit myocardium: reperfusion time-course study. Can J Cardiol 1990;6(7):299-304.
- Leichtweis S, Ji LL. Glutathione deficiency intensifies ischaemia-reperfusion induced cardiac dysfunction and oxidative stress. Acta Physiol Scand 2001;172(1):1-10.
- 36. Steare SE, Yellon DM. The potential for endogenous myocardial antioxidants to protect the myocardium against ischaemia-reperfusion injury: refreshing the parts exogenous antioxidants cannot reach? J Mol Cell Cardiol 1995;27(1):65-74.
- 37. Alberola A, Such L, Gil F, Zaragoza R, Morcillo EJ.

Protective effect of N-acetylcysteine on ischaemiainduced myocardial damage in canine heart. Naunyn Schmiedebergs Arch Pharmacol 1991;343(5):505-10.

- 38. Nishinaka Y, Sugiyama S, Yokota M, Saito H, Ozawa T. The effects of a high dose of ascorbate on ischemia-reperfusion-induced mitochondrial dysfunction in canine hearts. Heart Vessels 1992;7(1):18-23.
- Li G, Chen Y, Saari JT, Kang YJ. Catalase-overexpressing transgenic mouse heart is resistant to ischemia-reperfusion injury. Am J Physiol 1997;273(3 Pt 2):H1090-5.
- 40. Yoshida T, Maulik N, Engelman RM, Ho YS, Das DK. Targeted disruption of the mouse Sod I gene makes the hearts vulnerable to ischemic reperfusion injury. Circ Res 2000;86(3):264-9.
- Wang P, Chen H, Qin H, Sankarapandi S, Becher MW, Wong PC, Zweier JL. Overexpression of human copper, zinc-superoxide dismutase (SOD1) prevents postischemic injury. Proc Natl Acad Sci 1998;95(8):4556-60.
- Asimakis GK, Lick S, Patterson C. Post-ischemic recovery of contractile function is impaired in SOD2 (+/-) but not SOD1 (+/-) mouse hearts. Circulation 2002;105:981-6.
- 43. Chen EP, Bittner HB, Davis RD, Van Trigt P, Folz RJ. Physiologic effects of extracellular superoxide dismutase transgene overexpression on myocardial function after ischemia and reperfusion injury. J Thorac Cardiovasc Surg 1998;115(2):450-8.
- 44. Jones SP, Hoffmeyer MR, Sharp BR, Ho YS, Lefer DJ. Role of intracellular antioxidant enzymes after in vivo myocardial ischemia and reperfusion. Am J Physiol Heart Circ Physiol 2003;284(1):H277-82.
- 45. Sharp BR, Jones SP, Rimmer DM, Lefer DJ. Differential response to myocardial reperfusion injury in eNOS-deficient mice. Am J Physiol Heart Circ Physiol 2002;282(6):H2422-6.
- 46. Hess ML, Manson NH. Molecular oxygen: firend and foe. The role of the oxygen free radical system in the calcium paradox, the oxygen paradox and ischemia / reperfusion injury. J Mol Cell Cardiol 1984;16(11):969-85.
- 47. Park JL, Lucchesi BR. Mechanisms of myocardial reperfusion injury. Ann Thorac Surg 1999;68(5):1905-12.
- Ambrosio G, Tritto I. Reperfusion injury: experimental evidence and clinical implications. Am Heart J 1999;138 (2 Pt. 2)I:69-75.
- Weisfeldt ML, Zweier J, Ambrosio G, Becker LC, Flaherty JT. Evidence that free radicals result in reperfusion injury in heart muscle. Basic Life Sci 1988;49:911-99.
- 50. Garlick PB, Davies MJ, Davies Hearse DJ, Slater TF. Direct detection of free radicals in the reperfused rat heart using electron spin resonance spectroscopy. Circ Res 1987;61(5):757-60.
- 51. Moncada S, Higgs A. L-Arginine-nitric oxide pathway. N Engl J Med 1993;329:2002-2012.
- 52. Gilmont RR, Dardano A, Young M, Engle JS, Adamson BS, Smith DJ Jr, Rees RS. Effects of glutathione deple-

tion on oxidant-induced endothelial cell injury. J Surg Res 1998;80(1):62-8.

- Aktan AÖ, Yalcin AS. Ischemia-reperfusion Injury, Reactive oxygen metabolites, and the Surgeon Turk. J Med Sci 1998;28:1-5.
- 54. Ferrari R, Ceconi C, Curello S, Cargnoni A, Pasini E, De Giuli F, Albertini A. Role of oxygen free radicals in ischemic and reperfused myocardium. Am J Clin Nutr 1991;53:215S-22S.
- Ferrari R, Curello S, Cargnoni A, Condorelli E, Belloli S, Albertini A, Visioli O, Metabolic changes during postischemic reperfusion. J Mol Cell Cardiol 1988;20:113-9.
- 56. Schafer C, Ladilov Y, Inserte J, Schafer M, Haffner S, Garcia-Dorado D, Piper HM. Role of the reverse mode of the Na1 / Ca2 exchanger in reoxygenation-induced cardiomyocyte injury. Cardiovasc Res 2001;51:241-50.
- Piper HM, Abdallah Y, Schafer C. The first minutes of reperfusion: a window of opportunity for cardioprotection. Cardiovascular Research 2004;61:365-71.
- Siegmund B, Schlack W, Ladilov YV, Balser C, Piper HM. Halothane protects cardiomyocytes against reoxygenation-induced hypercontracture. Circulation 1997;96:4372-9.
- 59. Holmberg SR, Cumming DV, Kusama Y, Hearse DJ, Poole-Wilson PA, Shattock MJ, Williams AJ. Reactive oxygen species modify the structure and function of the cardiac sarcoplasmic reticulum calcium-release channel. Cardioscience 1991;2(1):19-25.
- Martin KR, Barrett JC. Reactive oxygen species as double-edged swords in cellular processes: low-dose cell signaling versus high-dose toxicity. Hum Exp Toxicol 2002;21(2):71-5.
- 61. Haddad JJ. Antioxidant and prooxidant mechanisms in the regulation of redox(y)-sensitive transcription factors. Cell Signal 2002;14:879-97.
- 62. Pietri S, Culcasi M, Stella L, Cozzone PJ. Ascorbyl free radical as a reliable indicator of free-radical-mediated myocardial ischemic and post-ischemic injury. A realtime continuous-flow ESR study. Eur J Biochem 1990;193(3):845-54.
- 63. Leichtweis S, Leeuwenburgh C, Bejma J, Ji LL. Aged rat hearts are not more susceptible to ischemia-reperfusion injury in vivo: role of glutathione. Mech Ageing Dev 2001;122(6):503-18.
- 64. Chatham JC, Seymour AL, Harmsen E, Radda GK. Depletion of myocardial glutathione: its effects on heart function and metabolism during ischaemia and reperfusion. Cardiovasc Res 1988;22(11):833-89.
- 65. Ferrari R, Alfieri O, Curello S, Ceconi C, Cargnoni A, Marzollo P, Pardini A, Caradonna E, Visioli O. Occurrence of oxidative stress during reperfusion of the human heart. Circulation 1990;81(1):201-11.
- 66. De Vecchi E, Pala MG, Di Credico G, Agape V, Paolini

G, Bonini PA, Grossi A, Paroni R. Relation between left ventricular function and oxidative stress in patients undergoing bypass surgery. Heart 1998;79(3):242-7.

- 67. Gunel E, Caglayan F, Caglayan O, Dilsiz A, Duman S, Aktan M. Treatment of intestinal reperfusion injury using antioxidant agents. J Pediatr Surg 1998.
- Barta E, Pechan I, Cornak V, Luknarova O, Rendekova V, Verchovodko P. Protective effect of alpha-tocopherol and L-ascorbic acid against the ischemic-reperfusion injury in patients during open-heart surgery. Bratisl Lek Listy 1991;92(3-4):174-83.
- 69. Westhuyzen J, Cochrane AD, Tesar PJ, Mau T, Cross DB, Frenneaux MP, Khafagi FA, Fleming SJ. Effect of preoperative supplementation with alpha-tocopherol and ascorbic acid on myocardial injury in patients undergoing cardiac operations. J Thorac Cardiovasc Surg 1997;113(5):942-8.
- Demirag K, Askar FZ, Uyar M, Cevik A, Ozmen D, Mutaf I, Bayindir O. The protective effects of high dose ascorbic acid and diltiazem on myocardial ischaemiareperfusion injury. Middle East J Anesthesiol 2001;16(1):67-79.
- Dingchao H, Zhiduan Q, Liye H, Xiaodong F. The protective effects of high-dose ascorbic acid on myocardium against reperfusion injury during and after cardiopulmonary bypass. Thorac Cardiovasc Surg 1994;42:276-8. 33(10):1536-1539.
- 72. Smith EF 3rd, Griswold DE, Egan JW, Hillegass LM, DiMartino MJ. Reduction of myocardial damage and polymorphonuclear leukocyte accumulation following coronary artery occlusion and reperfusion by the thromboxane receptor antagonist BM 13.505. J Cardiovasc Pharmacol 1989;5:715-22.
- 73. Hung LM, Chen JK, Lee RS, Liang HC, Su MJ. Beneficial effects of astringinin, a resveratrol analogue, on the ischemia and reperfusion damage in rat heart. Free Radic Biol Med 2001;30(8):877-83.
- Hung LM, Su MJ, Chen JK. Resveratrol protects myocardial ischemia-reperfusion injury through both NO-dependent and NO-independent mechanisms. Free Radic Biol Med 2004;36(6):774-81.
- 75. Manning AS, Coltart DJ, Hearse DJ. Ischemia and reperfusion-induced arrhythmias in the rat. Effects of xanthine oxidase inhibition with allopurinol. Circ Res 1984;55(4):545-8.
- Kusama Y, Bernier M, Hearse DJ. Exacerbation of reperfusion arrhythmias by sudden oxidant stress. Circ Res 1990;2:481-9.
- 77. Klein HH, Puschmann S, Schapper J, Schapper W. The mechanism of the tetrazolium reaction in identifying experimental myocardial infarction. Arch Pathol Anat 1981;393:287-97.

抗氧化劑與心肌缺血再灌流傷害

陳君侃 周淑娥1

動物實驗明顯指出,恢復血流是降低缺氧心臟心肌損傷的先決要件。但是各種缺血再灌 流的研究指出,缺氧的心臟恢復血流時,往往伴隨著明顯的心肌功能上和結構上的損傷;包 括心律不整,細胞傷害致胞内蛋白釋出,及收縮功能異常等。這些變化被認爲是心臟再灌流 時引起的二度傷害。由於再灌流時會伴隨著激烈的自由基產生,故一般認爲再灌流時的心肌 二度傷害,是因爲氧化壓力造成。細胞內許多重要的生化活動受到自由基及細胞氧化還原狀 態的調節,氧化還原狀態的失衡,是引起細胞功能異常的重要原因之一。唯利用抗氧化劑治 療缺血再灌流傷害的研究,各方的結果並不一致,且以負面的結果居多。故抗氧化劑治療的 臨床利用,仍有待進一步研究。(長庚醫誌 2005;28:369-77)

關鍵字:自由基,缺血再灌流,心肌損傷,抗氧化劑。

長庚大學 醫學院生理暨藥理學系, ¹通識中心 受文日期:民國94年3月30日;接受刊載:民國94年4月12日。 索取抽印本處:陳君侃教授,長庚大學 醫學院生理暨藥理系。桃園縣龜山鄉文化一路259號。Tel.: (03)2118800轉5077; Fax: (03)2118700; E-mail: jkc508@mail.cgu.edu.tw