Hidden Markov Models in Bioinformatics with Application to Gene Finding in Human DNA 308-761 Machine Learning Project

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The goal of this paper is to review the theory of Hidden Markov Models (HMM), to introduce problems in computational biology that can be solved using model-based approaches and to focus on the problem of gene finding. The paper also contains discussion of a small HMM experiment.

Introduction

A Hidden Markov Model (HMM) is a stochastic model that captures the statistical properties of observed real world data. A good HMM accurately models the real world source of the observed data and has the ability to simulate the source. Machine Learning techniques based on HMMs have been successfully applied to problems including speech recognition, optical character recognition, and as we will examine, problems in computational biology. The main computational biology problems with HMM-based solutions are protein family profiling, protein binding site recognition and the problem that is the topic of this paper, gene finding in DNA.

Genefinding or gene prediction in DNA has become one of the foremost computational biology problems for two reasons. First, because completely sequenced genomes have become readily available. And most importantly, because of the need to extract actual biological knowledge from this data to explain the molecular interactions that occur in cells and to define important cellular pathways. Discovering the location of genes on the genome is the first step towards building a such a body of knowledge. This paper will introduce several different statistical and algorithmic methods for gene finding, with a focus on the statistical model-based approach using HMMs.

Hidden Markov Models

A basic Markov model of a process is a model where each state corresponds to an observable event and the state transition probabilities depend only on the current and predecessor state. This model is extended to a Hidden Markov model for application to more complex processes, including speech recognition and computational genefinding. A generalized Hidden Markov Model (HMM) consists of a finite set of states, an alphabet of output symbols, a set of state transition probabilities and a set of emission probabilities. The emission probabilities specify the distribution of output symbols that may be emitted from each state. Therefore in a hidden model, there are two stochastic processes; the process of moving between states and the process of emitting an output sequence. The sequence of state transitions is a hidden process and is observed through the sequence of emitted symbols.

Let us formalize the definition of an HMM in the following way, taken from an HMM tutorial by Lawrence Rabiner [14]. An HMM is defined by the following elements:

- 1. Set S of N states, $S = S_1 S_2 \dots S_N$
- 2. Set V of M observation symbols, the output alphabet. $V = v_1v_2 \ldots v_M$.
- 3. Set A of state transition probabilities, $A = a_{ij}$ where a_{ij} is the probability of moving from state i to state i .

$$
a_{ij} = P(q_{t+1} = S_j | q_t = S_i), 1 \le i, j \le N
$$

4. Set B of observation symbol probabilities at state j, $B = b_i(k)$, where $b_i(k)$ is the probability of emitting symbol k at state i .

$$
b_j(k) = P(v_k|q_t = S_j), 1 \le j \le N, 1 \le k \le M.
$$

5. Set π , the initial state distribution $\pi = \pi_i$ where π_i is the probability that the start state is state i.

$$
\pi_i = P(q_1 = S_i), 1 \le i \le N.
$$

Given the definitions above, the notation of a model is $\lambda = (A, B, \pi)$.

Figure 1: A simple HMM $\lambda = (A, B, \pi)$, where $N = 3$, $M = 3$, a_{12}, a_{23}, a_{32} are non-zero, $b_1(a), b_2(t), b_3(g) = 1$ and $\pi = 1, 0, 0$. Note that states can be 'null' states that do not emit any symbol.

Model Architecture

The set of states S , the output symbol alphabet X and the connections between the states constitute the architecture of a model. The architecture of a HMM is problem dependent. The model is constructed to correspond to the properties and constraints of the observed sequences and of the process itself. HMM architecture can also be learned from the data, but in most computational biology problems, it is advantageous to use known constraints that characterize the processes. Further in this paper, we will examine how knowledge of the molecular mechanism of gene transcription guides the design of gene finding HMMs.

3

HMM Training and Decoding

Once the architecture of an HMM has been decided, an HMM must be trained to closely fit the process it models. Training involves adjusting the transition and output probabilities until the model sufficiently fits the process. These adjustments are performed using standard machine learning techniques to optimize $P(O|\lambda)$, the probability of observed sequence $O = O_1O_2 \ldots O_T$, given model λ over a set of training sequences. The most common and straightforward algorithm for HMM training is expectation maximization $(EM)^1$ which adapts the transition and output parameters by continually reestimating these parameters until $P(O|\lambda)$ has been locally maximized.

HMM decoding involves the prediction of hidden states given an observed sequence. The problem is to discover the best sequence of states $Q = q_1 q_2 \ldots q_T$ visited that accounts for an emitted sequence $O = O_1 O_2 \ldots O_T$ and a model λ . There may be several different ways to define a best sequence of states. A common decoding algorithm is the Viterbi algorithm². The Viterbi algorithm uses a dynamic programming approach to find the most likely sequence of states Q given an observed sequence O and model λ .

HMMs in Computational Biology

The field of computational biology involves the application of computer science theories and approaches to biological and medical problems. Computational biology is motivated by newly available and abundant raw molecular datasets gathered from a variety of organisms. Though the availability of this data marks a new era in biological research, it alone does not provide any biologically significant knowledge. The goal of computational biology is then to elucidate additional information regarding protein coding, protein function and many other cellular mechanisms from the raw datasets. This new information is required for drug design, medical diagnosis, medical treatment and countless fields of research.

The majority of raw molecular data used in computational biology corresponds to sequences of nucleotides corresponding to the primary structure of DNA and RNA or sequences of amino acids corresponding to the primary structure of proteins. Therefore the problem of inferring knowledge from this data belongs to the broader class of sequence analysis problems.

Two of the most studied sequence analysis problems are speech recognition and language processing. Biological sequences have the same left-to-right linear aspect as sequences of sounds corresponding to speech and sequences of words representing language. Consequently, the major computational biology sequence analysis problems can be mapped to linguistic problems. A common linguistic metaphor in computational biology is that of protein family classification as speech recognition. The metaphor suggests interpreting different proteins belonging to the same family as different vocalizations of the same word. Another metaphor is gene finding in DNA sequences as the parsing of language into words and semantically meaningful sentences. It follows that biological sequences can be treated linguistically with the same techniques used for speech recognition and language processing.

¹Please see references [1] and [14] for a complete specification of these and other HMM algorithms.

 2As in footnote 1.

Since the theory of HMMs was formalized in the late 1960s, several scientists have applied the theory to speech recognition and language processing. Just as HMMs were first introduced as mathematical models of language, HMMs can be used as mathematical models of molecular processes and biological sequences. In addition, HMMs have been applied to linguistics because they are suited for problems where the exact theory may be unknown but where there exists large amounts of data and knowledge derived from observation. As this is also the situation in biology, HMM-based approaches have been successfully applied to problems in computational biology. The main benefit is that an HMM provides a good method for learning the theory from the data and can provide a structured model of sequence data and molecular processes.

Application of HMMs to Specific Problems

It is clear that an HMM-based approach is a logical idea for tackling problems in computational biology. Much work has been performed and applications have been built using such an approach. Baldi and Brunak [1] define three main groups of problems in computational biology for which HMMs have been proven especially useful.

First, HMMs can be used for multiple alignments of DNA sequences, which is a difficult task to perform using a naive dynamic programming approach. Second, the structure of trained HMMs can uncover patterns in biological data. Such patterns have been used to discover periodicities within specific regions of the data and to help predict regions in sequences prone to forming specific structures. Third is the large set of classification problems. HMM based approaches have been applied to structure prediction - the problem of classifying each nucleotide according to which structure it belongs. HMMs have also been used in protein profiling to discriminate between different protein families and predict a new protein's family or subfamily. HMM-based approaches have also been successful when applied to the problem of gene finding in DNA. This is the problem of classifying DNA bases according to which type of job they perform during transcription. The remainder of this paper is concerned with this last problem of computational gene finding.

The Problem with Finding Genes

The problem of finding genes in DNA has been studied for many years. It was one of the first problems tackled once sufficient genomic data became available. The problem is given a sequence of DNA, determine the locations of genes, which are the regions containing information that code for proteins. At a very general level, nucleotides can be classified as belonging to coding regions in a gene, non-coding regions in a gene or intergenic regions. The problem of gene finding can then be stated as follows:

Input: A sequence of DNA $X = (x_1, ... x_n) \in \Sigma^*$, where $\Sigma = A, C, G, T$. **Output:** Correct labeling of each element in X as a belonding to a coding region, non-coding region or intergenic region.

Gene finding becomes complicated when the problem is approached in more biological detail. A eukaryotic gene contains coding regions called exons which may be interupted by non-coding regions called introns. The exons and introns are separated by splice sites. Regions outside genes are called intergenic. The goal of gene finding is then to annotate the sets of genomic data with the

location of genes and within these genes, specific areas such as promoter regions, introns and exons.

Many effective tools based on HMMs have been created for the purpose of gene finding. Among the most successful tools are Genie, GeneID and HMMGene [8]. Though each tool has a slightly different model, they each use the technique of combining several specialized submodels into a larger framework. The submodels correspond directly to different regions of DNA defined according to their function in the process of gene transcription. Most of the gene finding tools are hybrid models that include neural network components. In these tools, instead of an HMM, a neural network models certain regions, such as splice sites [12]. The overall framework of an HMM-based gene finder combines the submodels into a larger model corresponding to the organization of a gene in DNA and its functional roles. The following figure shows the different regions of DNA separated according to their function in transcription and translation.

Figure 2: The basic structure of a gene and also the basic schematic of a gene finding model. The units correspond to functionally different regions of DNA and in terms of HMM-based gene finders, to the specialized submodels that compose the complete model.

The HMM-based approach to gene finding has been well studied and well implemented. There are many tools available online, comprehensive analyses available and a comparison of the tools on a benchmark dataset by Burset and Guigo [2]. An extensive list of HMM-based gene finding resources is contained in [8]. Currently, gene finding tools are very good at classifying regions as coding or non-coding, but are not yet able to accurately predict the exact boundaries of exons. There are also complicated situations which cause poor gene finding performance such as multiple exon genes, nested genes and unusual genes. New work in gene finding applications involves the utilization of information contained within the data sets of known genes. With a large number of known genes in many organisms, there is a high chance that a newly discovered gene will be related to an already known gene. Therefore, gene finding can be attempted by matching new sequences of DNA against sets of known genes.

Experimenting with HMMs

My goal in this project is to learn about HMMs and their applications to problems in computational biology. In order to gain a strong understanding of the algorithms involved, I had plans to implement an HMM to perform gene finding. As my research progressed³ I decided to learn about HMM

 3 As I read the material and investigated the available datasets, I realized that gene finding had been done many, many times over and in much greater detail than I would be able to manage before the due date of this project. Also, even the smallest datasets for gene finding are huge! My poor connection at home couldn't cope.

algorithms by studying a simplified subproblem of gene finding. I chose to build and compare two HMMs for identify coding and non-coding regions in genes.

HMM Algorithms

The following section briefly describes the HMM algorithms I used in this experiment. For a complete explanation of these algorithms and their correctness, see [14]. The algorithms and notations used below follow directly from [6].

The basic task of building an HMM for a specific problem is to infer a set of parameters θ for a model λ from a set of data D. For simplicity of notation, assume that there is one sequence in the training set and D is a sequence of values d_1, \ldots, d_L . In this experiment, the problem is a classification problem and therefore each data element d_i can be represented as a value x_i and its corresponding class label y_i . D is then (X, Y) where $X = x_1, \ldots, x_L$ be the sequence of values and $Y = y_1, \ldots, y_L$ be the corresponding sequence of class labels. With respect to the problem of distinguishing coding and non-coding regions in DNA, X is a sequence of DNA and Y is a sequence identifying each DNA base as belonging to a coding region or non-coding region.

HMM Training Strategies

I implement two different strategies for performing the inference of HMM parameters for this problem. The first is the Maximum Likelihood (ML) approach, which infers the parameters by maximizing $P(X, Y | \lambda)$, the probability of the data set over all parameter sets for model λ .

$$
\theta^{ML} = maxP(X, Y | \theta, \lambda), \forall \theta
$$

The ML approach to modeling a classification problem involves combining several separately trained submodels into a larger model framework. Each submodel corresponds to a distinct class and is trained with data belonging to its respective class. It follows that all the elements of a training sequence for a specific submodel share the same class label.

The second is the Conditional Maximum Likelihood (CML) approach, which infers the parameters by maximizing $P(Y|X,\lambda)$, the probability of the labeling over all parameter sets for model λ.

$$
\theta^{CML} = maxP(Y|X, \theta_i, \lambda), \forall \theta
$$

The CML approach does not train submodels separately. Instead, it trains the entire model with a single class-labeled dataset.

Parameter Estimation using Baum-Welch Algorithm

Expectation maximization (EM) is the standard algorithm for maximizing the HMM parameters when the paths through the model for each training sequence are unknown. The standard EM algorithm is the Baum-Welch algorithm. This iterative algorithm has two steps, the expectation step and the maximization step. First, it calculates the expected number of times each transition and emission is used for the training set. Then, the transition and emission parameters are updated using reestimation formulas.

Recall the definition of HMM elements from the beginning of the paper. For ML and CML, the reestimation formulas are:

- $\bar{\pi}_i$ = expected number of times in state S_i at time $t = 1$.
- \bar{a}_{ij} = expected number of transitions from state S_i to state S_j / expected number of transitions leaving state S_i
- $\bar{b}_i(k)$ = expected number of times in state S_j and observing symbol v_k / expected number of times in state S_i

The expectation values are calculated using quantities determined in the forward and backward algorithms. The forward algorithm determines $f_k(i) = P(x_1 \dots x_i, q_i = S_k | \lambda)$, the probability of the observed sequence up to and including the *i*th term and being at state S_k at time *i*. The backward algorithm is very similar. It determines $b_k(i) = P(x_{i+1} \dots x_L | q_i = S_k, \lambda)$, the probability of observing sequence from $i + 1$ to the end given that the state at time i is S_k . For the complete forward and backward recusions and algorithms see [14].

ML and CML differ in the way that the expected values are tabulated. Though they both use the forward and backward tems, only correctly labeled paths are considered in CML. So, the forward and backward terms for states l that emit a different label than the observed label y_i are $f_l(i) = 0$ and $b_l(i) = 0$.

HMM Architecture

I constructed a very simple model structure of 8 states. The model consists of two submodels corresponding to coding and non-coding regions respectively. This model is oversimplified and to be scientifically useful, as it should include states corresponding to promoter regions and splice sites.

Figure 3: The letter labeling each state corresponds to the emission symbol of that state. The structure on the left is a possbible Markov chain for DNA. The structure on the right is the architecture of the coding/non-coding classification HMMs built in this experiment. The top set of states recongnizes the coding regions, the bottom set recognizes the non-coding regions. Note that this structure is a combination of two left hand structures and that the transitions within each set are not shown.

Using a so labeled architecture means that the emission probabilities did not have to be learned. This is often the approach taken when learning simple probabilistic properties of DNA sequences, but would not be effective in larger models, such as those used for gene finding.

Method

I downloaded HMM algorithms from [10] and rewrote them to deal with training sets with multiple sequences. I also wrote code to implement the CML approach described above by ensuring that only the desired class paths are considered when determining the reestimation values.

The datasets used were created from publicly available coding and non-coding vertebrate sequence data [2]. I created 5 sets of coding data, with each set containing 50 vertebrate coding sequences of length 300. I also created 5 sets of non-coding data, with each set containing 50 vertebrate non-coding sequences of length 350. The test data is a random combination of coding and non-coding sequences not seen during model training. The measure of goodness of each model is determined by the percentage of test set bases the model correctly labels.

Results and Discussion

The main result of my experimenting is that I learned the Baum-Welch and CML algorithms very well. I also clearly understand the computational difficulties faced by many bioinformaticians when dealing with large data sets like genomic sequence sets. Unfortunately, it has become clear that this experiment is simplified to the point of any measured results have little meaning. It is very difficult for me to distinguish between random and good behaviour of the HMMs, as each labeling had a 0.5 chance of being correct. I also found that though literature suggests the labeled architecture described in the previous section for such problems [6], it became very difficult to discern if the parameters were in fact being adjusted to maximize the probability of the training set given the model.

My incomplete results are quite random numbers, and I did not succeed in fully training a model using the CML approach. Perhaps the fact that the models did not convincingly represent the data is because of too little training data and an oversimplified model. I am convinced that the expectation and maximization algorithms functioned correctly, as I was successful in separately training the coding and non-coding submodels. However, I found it difficult to determine appropriate probabilities to govern the transitions between the two submodels.

From my newly acquired experience with HMM algorithms and training, I do believe that the CML approach is well suited to these types of classification problems and would be effective in a larger framework, as Krogh showed [11].

In conclusion, I believe that though I had difficulty training simple models, HMMs have been, and continue to be, an invaluable tool for infering biologically meaningful information from otherwise nearly incomprehensible DNA sequences.

References

- [1] Baldi, P. & Brunak S. "Bioinformatics The Machine Learning Approach". Massachusetts Institute of Technology, 1998.
- [2] Berkeley Drosophila Genome Project Representative Benchmark Data Sets of Human DNA Sequences - http://www.fruitfly.org/sequence/human-datasets.html.
- [3] Besemer, J. & Borodovsky, M. "Heuristic approach to deriving models for gene finding". Nucleic Acids Research, 1999, Vol. 27, No. 19, pp. 3911-3920.
- [4] Birney, E. "Hidden Markov Models in Biological Sequence Analysis". IBM Journal of Research and Development Volume 45, Numbers 3/4, 2001.
- [5] Burset & Guigo. "Evaluation of gene structure prediction programs". Genomics, 1996, Vol. 34, No. 3, pp. 353-367.
- [6] Durbin, R., Eddy, S., Krogh, A. and Mitchison, G. Biological Sequence Analysis Probabilistic Models of Proteins and Nucleic Acids. Cambridge University Press, 1998.
- [7] Ensembl Export View, Human Genome Sever http://www.ensembl.org/perl/exportview?tab=list.
- [8] Haussler, D. "Computational Genefinding". Trends in Biochemical Sciences, Supplementary Guide to Bioinformatics, 1998, pp. 12-15.
- [9] Henderson, J., Salzberg, S. and Fasman, K. "Finding Genes in DNA with a Hidden Markov Model". Journal of Computational Biology, Vol. 4, No. 2 (1997), pp. 127-141.
- [10] Kanungo, Tapas. HMM software learning toolkit. University of Maryland Institute for Advanced Computer Studies, Center for Automation Research, Language and Media Processing Lab - http://www.cfar.umd.edu/ kanungo/software/software.html.
- [11] Krogh, A. "Two Methods for Improving Performance of a HMM and Their Application for Gene Finding". Proceedings of the Fifth International Conference on Intelligent Systems for Molecular Biology, AAAI Press, Menlo Park, CA, 1997, pp. 179-186.
- [12] Kulp, D., Haussler, D., Reese, M. G. and Eeckman, F. H. "A Generalized Hidden Markov Model for the Recognition of Human Genes in DNA". Proceedings of the Fourth International Conference on Intelligent Systems for Molecular Biology, AAAI Press, Menlo Park, CA, 1996, pp. 134-142.
- [13] Pevzner, P. "Computational Molecular Biology: An Algorithmic Approach". The MIT Press, 2000.
- [14] Rabiner, Lawrence R. "A Tutorial on Hidden Markov Models and Selected Applications in Speech Recognition". *Proceedings of the IEEE*, Vol. 77, No. 2, February 1989, pp. 257-286.
- [15] Stormo, G. "Gene-Finding Approaches for Eukaryotes". Genome Research Vol. 10, Issue 4, April 2000, pp. 394-397.
- [16] University of California at Santa Cruz Human Genome Working Draft http://genome.ucsc.edu.