Free Radicals and Antioxidants: A Personal View

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Free radicals and other oxygen-derived species are constantly generated in vivo, both by "accidents of chemistry" and for specific metabolic purposes. The reactivity of different free radicals varies, but some can cause severe damage to biological molecules, especially to DNA, lipids, and proteins. Antioxidant defense systems scavenge and minimize the formation of oxygen-derived species, but they are not 100% effective. Hence, diet-derived antioxidants may be particularly important in diminishing cumulative oxidative damage and helping us to stay healthier for longer. Repair systems exist to deal with molecules that have been oxidatively damaged. Damage to DNA by hydroxyl radicals appears to occur in all aerobic cells, and might be a significant contributor to the age-dependent development of cancer. Lipid peroxidation probably contributes significantly to the development of atherosclerosis.

Why Do We Need Antioxidants?

When living organisms first appeared on the Earth, they did so under an atmosphere containing very little O_2 , i.e., they were essentially anaerobes. Anaerobic microorganisms still survive to this day, but their growth is inhibited and they can often be killed by exposure to 21% O₂, the current atmospheric level. As the O_2 content of the atmosphere rose (due to the evolution of organisms with photosynthetic water-splitting capacity) many primitive organisms may have died out. Present day anaerobes are presumably the descendants of those primitive organisms that followed the evolutionary path of "adapting" to rising atmospheric O₂ levels by restricting themselves to environments that the O_2 did not penetrate. However, other organisms began the evolutionary process of evolving antioxidant defense systems to protect against O_2 toxicity. In retrospect, this was a fruitful path to follow. Organisms that tolerated the presence of O_2 could also evolve to use it for metabolic transformations (oxidases, oxygenases, etc.) and for efficient energy production by using electron transport chains with O_2 as the terminal electron acceptor, such as those present in mitochondria. Our mitochondria make over 80% of the ATP we need, and the lethal effects of inhibiting this, e.g., by cyanide, show how important the mitochondria are.

It is interesting to note that we have developed antioxidant defenses to protect against 21% O₂, but no greater than that. This is evidenced by the fact that all aerobes suffer demonstrable injurious effects if exposed to O_2 at concentrations greater than 21%.1 For example, exposure of adult humans to pure O_2 at 1 atm pressure for as little as 6 hours causes chest soreness, cough, and sore throat in some subjects, and longer periods of exposure lead to alveolar damage. The incidence of ocular damage in babies known as retrolental fibroplasia (formation of fibrous tissue behind the lens) increased abruptly in the early 1940s among infants born prematurely and often led to blindness. Not until 1954 was it realized that retrolental fibroplasia is associated with the use of high O₂ concentrations in incubators for premature babies. More careful control of O₂ concentrations (continuous transcutaneous O₂ monitoring, with supplementary O₂ given only where necessary) and administration of the lipid-soluble antioxidant a-tocopherol have decreased its incidence. However, the problem has not disappeared, since many premature infants need increased levels of O_2 in order to survive at all.²

The damaging effects of elevated O_2 on aerobes vary considerably with the organism studied, age, physiological state, and diet. Different tissues are affected in different ways.¹ Thus, cold-blooded animals, such as turtles and crocodiles, are relatively resistant to O_2 at low environmental temperatures but become more sensitive at higher temperatures. Young rats resist lung damage in an atmosphere of 100% O_2 far more effectively than do adult rats.¹

The earliest suggestion made to explain O2 tox-

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Name	Formula	Comments
Hydrogen atom	H.	The simplest free radical.
Trichloromethyl	CCl3.	A carbon-centered radical (i.e., the unpaired electron resides on carbon). CCl_3 is formed during metabolism of CCl_4 in the liver and contributes to the toxic effects of this solvent. ⁵ Carbon-centered radicals usually react fast with O_2 to make peroxyl radicals, e.g., CCl_3 + $O_2 \rightarrow CCl_3O_2$.
Superoxide	O ₂	An oxygen-centered radical. Selectively reactive.
Hydroxyl	OH.	A highly reactive oxygen-centered radical. Attacks all molecules in the human body.
Thiyl	RS ⁻	A group of radicals with an unpaired electron residing on sulfur.
Peroxyl, alkoxyl	RO_2^{-} , RO^{-}	Oxygen-centered radicals formed (among other routes) during the breakdown of organic peroxides.
Oxides of nitrogen	NO', NO ₂ '	Nitric oxide is formed in vivo from the amino acid L-arginine. ⁶ Nitro- gen dioxide is made when NO reacts with O_2 and is found in pol- luted air and smoke from burning organic materials, e.g., cigarette smoke. ⁷

Table 1. Examples of Free Radicals

icity was that O_2 is a direct inhibitor of enzymes, thereby interfering with metabolism.¹ However, very few targets of direct damage by O_2 have been identified in aerobes. In 1954, Gerschman et al.³ proposed that the damaging effects of O_2 could be attributed to the formation of oxygen radicals. This hypothesis was popularized and converted into the "superoxide theory of O_2 toxicity" following the discovery of superoxide dismutase (SOD) enzymes by McCord and Fridovich.⁴ In its simplest form, this theory states that O_2 toxicity is due to excess formation of superoxide radical (O_2 .⁻⁻) and that the SOD enzymes are important antioxidant defenses.

What Is a Radical?

We need antioxidants to scavenge and prevent the formation of free radicals such as superoxide. But what exactly is a radical?

In the structure of atoms and molecules, electrons usually associate in pairs, with each pair moving within a defined region of space around the nucleus. This space is referred to as an atomic or molecular orbital. One electron in each pair has a spin quantum number of $+\frac{1}{2}$, the other $-\frac{1}{2}$. A free radical is any species capable of independent existence (hence the term "free") that contains one or more unpaired electrons. An unpaired electron is any electron that is alone in an orbital. The simplest free radical is an atom of the element hydrogen, with one proton and a single electron. Table 1 gives examples of other free radicals⁵⁻⁷ (a superscript dot is used to denote free radical species). The spectroscopic technique of electron spin resonance is specific for the detection and measurement of free radicals, recording the energy changes that occur as unpaired electrons respond to a magnetic field.8

What Radicals Are Made In Vivo?

Hydroxyl Radical

The chemical reactivity of free radicals varies. One of the most reactive is hydroxyl radical (OH[•]).⁹ Exposure of living organisms to ionizing radiation causes fission of O–H bonds in water:

$$H_2O \rightarrow H^+ + OH^-$$

to give H[•] and OH[•]. Hydroxyl radical reacts at a diffusion-controlled rate with almost all molecules in living cells.⁹ Hence, when OH[•] is formed in vivo, it damages whatever it is generated next to, as OH[•] cannot migrate any significant distance within the cell. Many, if not most, of the harmful effects of excess exposure to ionizing radiation upon living organisms are thought to be initiated by attack of OH[•] upon proteins, carbohydrates, DNA, and lipids.

Nitric Oxide

Whereas OH[•] is probably always harmful, other less reactive free radicals may often be useful in vivo. Free radicals are known to be produced metabolically in living organisms. For example, the free radical nitric oxide (NO[•]) is synthesized from the amino acid L-arginine by vascular endothelial cells, phagocytes, certain cells in the brain, and many other cell types.⁶ Nitric oxide is a vasodilator agent and possibly an important neurotransmitter. It may also be involved in the killing of parasites by macrophages in some mammalian species.⁶

Superoxide

Superoxide radical (O_2^{-}) is the one-electron reduction product of oxygen. It is produced by phagocytic cells (neutrophils, monocytes, macrophages,

eosinophils) and helps them to inactivate viruses and bacteria.¹⁰ Evidence is accumulating that O_2^{--} is also produced in vivo by several cell types other than phagocytes, including lymphocytes¹¹ and fibroblasts.^{12,13} Superoxide produced by such cells is often thought to be involved in intercellular signaling and growth regulation, and many experiments with cell cultures are consistent with this concept.¹⁴ However, it has not yet been proven as occurring in vivo. There are many reports^{15–17} that vascular endothelial cells generate O_2^{--} , but it is uncertain if they do this all the time in vivo or only after an insult, such as ischemia–reperfusion (or even the cell isolation process itself).

In addition to deliberate metabolic production of O_2 , this radical can also emanate from what may be called "accidents of chemistry." Superoxide and H₂O₂ may be generated by "autoxidation" reactions, in which such compounds as catecholamines, tetrahydrofolates, and reduced flavins react directly with O_2 to form $O_2^{\cdot-}$. The $O_2^{\cdot-}$ then oxidizes more of the compound and sets up a free radical chain reaction. In fact, most so-called autoxidation reactions in vitro depend on the presence of traces of catalytic transition metal ions. Superoxide might also be made in the mitochondria. The mitochondrial electron transport chain is a gradient of redox potential, from the highly reducing NADH/NAD+ couple to the oxidizing O₂. Thermodynamically, there is nothing to prevent constituents of the early part of the chain (nonheme-iron proteins, quinones, flavoproteins, cytochromes b) from reducing O₂ directly to make O_2 . Fortunately, such reactions are kinetically restricted, so that most of the electrons entering the mitochondrial electron transport arrive at cytochrome oxidase, and only a small percentage (perhaps only 1-3%) may leak. These studies of the leakiness of electron transport chains are based on experiments with bacteria and mitochondria respiring in air-saturated solutions⁴ and might be an overestimate. However, if we add in the deliberate O_2^{-} production from phagocytes, and other sources (such as electron leakage from other electron-transport chains and autoxidation reactions), a figure of 1-3% does not seem unreasonable. Although this figure may seem trivial, remember that humans are big animals and breathe in a lot of O₂. Thus, at rest, we may produce close to 2 kg of O_2^{-} per year (Figure 1).

Hydrogen Peroxide, a Nonradical

Most of the O_2 ⁻⁻ generated in vivo probably undergoes a nonenzymatic or SOD-catalyzed dismutation reaction, represented by the overall equation:

$$2O_2^{\cdot-} + 2H^+ - H_2O_2 + O^2.$$

This generates hydrogen peroxide (H₂O₂), a nonrad-

An adult at rest utilizes 3.5 mL O₂/kg/minute or 352.8 liters/day (assuming 70 kg body mass) or 14.7 moles/day. If 1% makes O₂⁻⁻ This is 0.147 moles/day or 53.66 moles/year or about 1.72 kg/year (of O₂⁻⁻). During bodily exertion, this would increase up to 10-fold (assuming that the 1% figure still applied).

Figure 1. How much superoxide is made in the human body?

ical. H_2O_2 resembles water in its molecular structure and is very diffusible within and between cells. As well as arising from O_2^{1-} , H_2O_2 is produced by the action of several oxidase enzymes in vivo, including amino acid oxidases and the enzyme xanthine oxidase.^{18,19} Xanthine oxidase catalyzes the oxidation of hypoxanthine to xanthine, and of xanthine to uric acid. Oxygen is simultaneously reduced both to O_2^{1-} and to H_2O_2 . Low levels of xanthine oxidase are present in many mammalian tissues, especially in the gastrointestinal tract. Levels of xanthine oxidase often increase when tissues are subjected to insult, such as trauma or deprivation of oxygen.¹⁹

Some metabolic roles for H₂O₂ are known, and others have been proposed.^{20–22} For example, H_2O_2 generated in the thyroid gland is used by a peroxidase enzyme to iodinate the thyroid hormones.²⁰ H_2O_2 may up-regulate the expression of certain genes by (directly or indirectly) leading to displacement of an inhibitory subunit from the cytoplasmic gene transcription factor NF-KB.23 Displacement of the inhibitory subunit causes the active factor to migrate to the nucleus and activate many different genes by binding to specific DNA sequences in enhancer and promoter elements.23 Thus, H2O2 and other peroxides can induce expression of genes controlled by NF-kB. This is of particular interest currently because in cell culture systems, H₂O₂ can activate NF-KB and induce the genetic expression of the provirus human immunodeficiency virus-1 (HIV-1). This virus is the most common cause of acquired immunodeficiency syndrome (AIDS), and its expression can be prevented by certain antioxidants.23 In addition, peroxides in human atherosclerotic lesions may activate NF-KB.24,25

Some Confusing Terminology: Oxygen-Derived Species, Reactive Oxygen Species, and Oxidants

Reactive oxygen species (ROS) is a collective term often used by scientists to include not only the oxy-

INITIATION of peroxidation usually occurs by the attack of any species (R[·]) capable of abstracting hydrogen from a polyunsaturated fatty acid side-chain in a membrane (such fatty acid side-chains are more susceptible to free radical attack than are saturated or monounsaturated side-chains).

$$-CH + R^{\cdot} \rightarrow -C^{\cdot} + RH$$

Species able to abstract hydrogen include OH and peroxyl radicals (Table 1). The carbon-centered radical reacts fast with O_2

$$-C' + O_2 \rightarrow -CO_2$$

A fatty acid side-chain **PEROXYL RADICAL** is formed. This can attack adjacent fatty acid side-chains and **PROPAGATE** lipid peroxidation.

$$-CO_2^{\cdot} + -CH \rightarrow -CO_2H + -C^{\cdot}$$

The chain reaction thus continues and **LIPID PEROXIDES** ($-CO_2H$) accumulate in the membrane. Lipid peroxides destabilize membranes and make them "leaky" to ions. Peroxyl radicals can attack not only lipids but also membrane proteins (e.g., damaging enzymes, receptors, and signal transduction systems) and oxidize cholesterol.

Figure 2. An outline of lipid peroxidation.

gen-centered radicals ($O_2^{\cdot-}$, and OH[•]) but also some nonradical derivatives of O_2 . These include H_2O_2 , hypochlorous acid (HOCl, an oxidizing and chlorinating agent produced by activated phagocytes),^{10,26} and ozone (O_3). "Reactive" is, of course, a relative term; neither $O_2^{\cdot-}$ nor H_2O_2 is particularly reactive in aqueous solution. Hence, some authors use the term "oxygen-derived species" instead. Another popular collective term is "oxidants." However, since $O_2^{\cdot-}$ and H_2O_2 can act as both oxidants and reductants in aqueous solution, I prefer to avoid that term.

How Do Radicals React?

Reactivity depends on the radical and what the radical is presented with. If two free radicals meet, they can join their unpaired electrons to form a covalent bond. Thus, atomic hydrogen forms diatomic hydrogen:

$$H^{\cdot} + H^{\cdot} \rightarrow H_2$$

A more biologically relevant example is the very fast²⁷ reaction of NO[•] and $O_2^{•-}$ to form a nonradical product, peroxynitrite²⁸:

NO' +
$$O_2^{\cdot-} \rightarrow ONOO^-$$
 (peroxynitrite)

However, when a free radical reacts with a nonradical, a new radical results, and a chain reaction is set up. Since most biological molecules are nonradicals, the generation of reactive radicals such as OH[•] in vivo usually sets off chain reactions.

For example, attack of reactive radicals upon fatty acid side chains in membranes and lipoproteins can abstract hydrogen, leaving a carbon-centered radical and initiating the process of lipid peroxidation (Figure 2).

When OH is generated adjacent to DNA, it attacks both the deoxyribose sugar and the purine and pyrimidine bases. Figure 3 shows the structures of some of the products generated by attack of OH' on the DNA bases. This wide range of products is characteristic of attack by OH⁻ and may be used to show that such attack has occurred in vivo. For example, if DNA is extracted from a tissue and most or all of the compounds shown in Figure 3 are present, this is good evidence²⁹ that the DNA has suffered attack by OH'. Such "OH'-fingerprint" experiments have been used to study the role of free radicals in DNA damage by radiation and toxic agents.²⁹ It has also been found that the amount of these OH -derived products in DNA from human cancerous tumors is greater than in adjacent noncancerous tissue.^{30,31} Whether this is due to increased OH⁻ formation or to decreased repair of the damage remains to be evaluated.

Toxicity of Superoxide, Hydrogen Peroxide, and Nitric Oxide

Experimental results clearly show that removal of $O_2^{\cdot-}$ and H_2O_2 by antioxidant defense systems is essential for healthy aerobic life.^{4,18,32,33} Why is this? In organic media, $O_2^{\cdot-}$ can be very reactive, but in aqueous media it is not, mainly acting as a moderate reducing agent.³⁴ However, superoxide can react very rapidly with a few targets. These include some bacterial iron–sulfur proteins, including enzymes essential to metabolism such as aconitase.^{35,36} Whether similar $O_2^{\cdot-}$ -sensitive targets exist in hu-



Figure 3. Products of attack of hydroxyl radicals upon purine and pyrimidine bases of DNA.

man cells remains to be established. However in isolated submitochondrial particles, $O_2^{\cdot-}$ has been claimed to inactivate the NADH dehydrogenase complex of the mitochondrial electron transport chain.³⁷

One important molecule in humans that reacts with O_2^{-} is NO^{\cdot}. NO^{\cdot} is useful in human metabolism, but an excess can be cytotoxic, and this mechanism of tissue injury has been implicated in several human diseases.6 Excess NO' can be directly toxic, e.g., by damaging iron-sulfur proteins,6 but evidence is increasing to show that NO'/O2'interactions are also involved.38,39 Since NO' acts upon smooth muscle cells in vessel walls to produce relaxation,⁶ then O_2^{-} , by removing NO⁻, can act as a vasoconstrictor, which might have deleterious effects in some clinical situations.^{40,41} For example, excess vascular O_2^{-} production could lead to hypertension⁴¹ and is implicated in the development of atherosclerosis.³⁹ Peroxynitrite, the product of NO[•] and O2.-, can cause direct biological damage by oxidizing -SH groups.^{38,42} In addition, at physiological pH, peroxynitrite can protonate and decompose to a range of noxious products, including nitrogen dioxide (NO_2), a powerful initiator of lipid peroxidation in biological fluids,⁴³ OH, and nitronium ion (NO_2), an agent active in nitrating aromatic rings, such as those of phenylalanine and tyrosine.^{38,44,45}

The protonated form of O_2^{--} , perhydroxyl radical (HO₂⁻) is much more reactive than O_2^{--} in vitro,³⁴ but very little O_2^{--} is protonated at pH 7.4. For example, HO₂⁻ can initiate peroxidation of polyunsaturated fatty acids, which O_2^{--} cannot^{46,47}:

$$L-H + HO_2^{-} \rightarrow L^{-} + H_2O_2.$$

However, HO_2 is not a very efficient initiator and there is no direct evidence that HO_2 exerts damaging effects in vivo.

Hydrogen peroxide at micromolar levels also appears poorly reactive, but higher (>50 μ M) levels of H₂O₂ can attack certain cellular targets.^{32,48} For example, it can oxidize an essential –SH group on the glycolytic enzyme glyceraldehyde-3-phosphate dehydrogenase, blocking glycolysis and interfering with other aspects of cell energy metabolism.⁴⁸

In 1970, Beauchamp and Fridovich⁴⁹ proposed

that the toxicity of $O_2^{\cdot-}$ and H_2O_2 could pertain to their conversion into OH[•]. Three mechanisms have been proposed to explain this. One is the interaction of $O_2^{\cdot-}$ and NO[•], as described above. An earlier proposal⁵⁰ was the superoxide-driven Fenton reaction:

and

$$Fe(II) + H_2O_2 \rightarrow OH^- + OH^- + Fe(III)$$

 $O_2^{\cdot-} + Fe(III) \rightarrow Fe(II) + O_2$

Despite repeated controversy in the literature, this author feels that the evidence for the formation of OH[•] when Fe²⁺ reacts with H₂O₂ is overwhelming.^{32,50,51} This does not, of course, preclude the formation of reactive species additional to OH[•], such as oxo-iron complexes (ferryl, perferryl). Copper ions also catalyze formation of OH[•] from H₂O₂.⁵⁰ A third mechanism for making OH[•] is the reaction of O₂^{•-} with hypochlorous acid:⁵²

$$O_2^{--} + HOCl \rightarrow OH^{-} + OH^{-} + O_2$$

This has a rate constant of $7.5 \times 10^6 \text{ M}^{-1} \text{ second}^{-1}$.

Do Metal Ion "Catalysts" Exist In Vivo?

Iron and copper ions in chemical forms that can decompose H_2O_2 to OH[•] are in very short supply in vivo. The human body is very careful to ensure that as much iron and copper as possible is kept safely bound to transport or storage proteins. Indeed, this "sequestration" of metal ions is an important anti-oxidant defense mechanism.^{32,53} Sequestration of metal ions deters the growth of bacteria in human body fluids⁵⁴ and also ensures that such fluids will, in general, not convert O_2^{--} and H_2O_2 into OH[•]. If iron or copper does become available to catalyze free radical reactions in body fluids, as happens in certain metal overload diseases, severe damage to many body tissues occurs.^{55,56}

Hence, a major determinant of the nature of the damage done by excess generation of reactive oxygen species in vivo may be the availability and location of metal ion catalysts of OH' radical formation.^{32,50} If, for example, "catalytic" iron salts are bound to DNA in one cell type and to membrane lipids in another, then excessive formation of H_2O_2 and O_2^{-} will, in the first case, damage the DNA and in the second case could initiate lipid peroxidation. Evidence for OH' formation in the nucleus of cells treated with H_2O_2 has been obtained by showing that all four DNA bases are modified in the way characteristic of OH⁻ attack (Figure 3).⁵⁷ If this OH' is formed by metal ion-dependent reactions, then the "catalytic" metal ions must be bound to the DNA itself.

Escherichia coli mutants lacking SOD activity are hypersensitive to damage by H_2O_2 ,³³ and if able

to enter the cell, extra SOD can protect cells against damage by H₂O₂.⁵⁸ These and much other data are consistent with a role of O_2^{-} in facilitating damage by H_2O_2 , and the Fenton reactions provide a possible explanation. However, many scientists are reluctant to believe that O_2^{-} serves only as a reducing agent for Fe(III), since in general, mammalian tissues are fairly good reducing environments containing thiols, NADH, NADPH, reduced folates, and ascorbate.⁵⁰ An additional, and possibly more important role for O_2^{-} may be in the provision of the iron required for OH' generation. Thus, O_2^{-} can reductively mobilize iron ions from the iron storage protein ferritin.59 Because the amount of O2.--releasable iron is very small, ferritin-bound iron is much safer than an equivalent amount of "free" iron.^{32,60} Superoxide might also release iron if it attacks iron-sulfur proteins.35,36

Antioxidant Defenses

All organisms suffer some exposure to OH', because it is generated by homolytic fission of O–H bonds in water driven by background ionizing radiation.⁹ This radical is so reactive with all biological molecules that it is impossible to evolve a specific scavenger of it. Almost everything in living organisms reacts with OH⁻ with second-order rate constants of 10^9-10^{10} M⁻¹ second⁻¹, so that collision of OH⁻ with the molecules almost always results in reaction.⁹ Once OH⁻ has been formed, damage caused by this radical is probably unavoidable and is dealt with by repair processes (Table 2).^{61–67} A large part of the body's antioxidant defenses serve to minimize any additional production of OH⁻.

Enzymes

Living organisms have evolved antioxidant defenses to remove excess O_2^{-} and H_2O_2 . SOD enzymes remove O_2^{-} by accelerating its conversion to H_2O_2 by about four orders of magnitude at pH 7.4. Mammalian cells have a SOD enzyme containing manganese (MnSOD) at its active site in the mitochondria. A SOD with copper and zinc (CuZnSOD) at the active site is also present, but largely in the cytosol.⁴ It has recently been shown that the familial dominant form of amyotrophic lateral sclerosis (ALS, Lou Gehrig's disease), a fatal degenerative disorder of motor neurons in the brain and spinal cord, is somehow related to mutations in the CuZn protein. These mutations decrease activity somewhat but may also cause the protein to become toxic^{68,69}, perhaps by releasing copper (a powerful prooxidant) and/or by acting as a peroxidase.⁶⁹

Because SOD enzymes generate H_2O_2 , they work in collaboration with H_2O_2 -removing enzymes. Catalases convert H_2O_2 to water and O_2

Substrate of Damage	Repair System	tive Recent References
DNA: All components of DNA can be attacked by OH', whereas singlet O_2 attacks guanine preferentially. H_2O_2 and O_2 . ⁻ do not attack DNA.	A wide range of enzymes exists that recognize abnormalities in DNA and remove them by excision, resynthesis, and rejoining of the DNA strand.	29,61
Proteins: Many reactive oxygen species can oxidize -SH groups. Hydroxyl radicals attack many amino acid residues. Proteins often bind transition metal ions, making them a target of attack by "site-spe- cific" OH generation.	Oxidized methionine residues may be repaired by methionine sulfoxide reductase. Dam- aged proteins may be recognized and pref- erentially destroyed by cellular proteases.	62-64
Lipids: Some reactive oxygen species (not including O_2^{-} or H_2O_2) can initiate lipid peroxidation.	Chain-breaking antioxidants (especially α -to- copherol) remove chain-propagating peroxyl radicals. Phospholipid hydroperoxide gluta- thione peroxidase can remove peroxides from membranes, as can some phospholi- pases. Normal membrane turnover can re- lease damaged lipids.	65-67

$$2H_2O_2 \rightarrow 2H_2O_2 + O_2$$

Catalases are present in the peroxisomes of mammalian cells, and probably serve to destroy H_2O_2 generated by oxidase enzymes located within these subcellular organelles.¹⁸ However, the most important H_2O_2 -removing enzymes in mammalian cells are the selenoprotein glutathione peroxidase (GSHPX) enzymes. A selenocysteine residue, essential for enzyme activity, is present at the active site. GSHPX enzymes remove H_2O_2 by using it to oxidize reduced glutathione (GSH) to oxidized glutathione (GSSG). Glutathione reductase, a flavoprotein (FAD-containing) enzyme, regenerates GSH from GSSG, with NADPH as a source of reducing power¹⁸ (Figure 4).

Sequestration of Transition Metals

As already emphasized, an additional important antioxidant defense system is that which allows organisms to generate metal ion storage and transport proteins. These are produced to keep iron and copper safely protein-bound whenever possible.

Low-Molecular Mass Antioxidants

GSH

In addition to antioxidant defense enzymes, some low-molecular mass free radical scavengers exist. GSH can scavenge various free radicals directly, as well as being a substrate for GSHPX enzymes.¹⁸

GLUTATHIONE PEROXIDASE

 $2\text{GSH} + \text{H}_2\text{O}_2 \rightarrow \text{GSSG} + 2\text{H}_2\text{O}$

2GSH + FATTY ACID-OOH \rightarrow GSSG + FATTY ACID-OH + H₂O

GLUTATHIONE REDUCTASE

 $\mathsf{GSSG} + \mathsf{NADPH} + \mathsf{H^{\scriptscriptstyle +}} \rightarrow \mathsf{2GSH} + \mathsf{NADP^{\scriptscriptstyle +}}$

Reduced glutathione is a tripeptide, glutamic acid–cysteine–glycine. It is present at millimolar concentrations in most mammalian cells and has multiple metabolic functions.¹⁸ In oxidized glutathione (GSSG), two tripeptides are linked by a disulfide bridge. Glutathione peroxidase can also destroy fatty acid (lipid–OOH) peroxides by converting them to alcohols (lipid–OH).¹⁸ Mammalian cells additionally contain a phospholipid hydroperoxide glutathione peroxidase that can apparently perform the same reaction upon lipid peroxides within membranes; how exactly it works is uncertain as yet.⁶⁶

Figure 4. The glutathione system.

Vitamin E

Alpha-tocopherol, the major constituent of the fatsoluble vitamin known as vitamin E, is the most important⁶⁵ (but by no means the only^{70,71}) free radical scavenger within membranes and lipoproteins. Alpha-tocopherol inhibits lipid peroxidation by scavenging peroxyl radicals (Table 1), which are intermediates in the chain reaction:

$$\alpha TH + LOO^{\cdot} \rightarrow \alpha T^{\cdot} + LOOH$$

The α -tocopherol radical (α T[']), although not completely unreactive,⁷² is less efficient at abstracting hydrogen than are peroxyl (LOO[']) radicals, so the chain reaction of peroxidation is slowed.⁶⁵ Several biological mechanisms may exist for recycling T['] back to α -tocopherol, although none of them has yet been proven rigorously to operate in vivo in humans.⁶⁵ Likely mechanisms include the reaction of T['] with ascorbic acid at the surface of membranes and lipoproteins:⁷¹

 $\alpha T'$ + ascorbate $\rightarrow \alpha TH$ + ascorbate^{*}

and/or with ubiquinol (reduced coenzyme Q) within membranes or lipoproteins:^{70,73}

 $CoQH_2 + \alpha T^{\cdot} \rightarrow \alpha TH + CoQH^{\cdot} + H^+.$

Vitamin E is known to be essential in the human diet. Severe deprivation produces neurological damage.⁷⁴

Ascorbic Acid

Ascorbic acid, the water-soluble vitamin C, plays several essential metabolic roles in vivo,75 such as in the synthesis of collagen. There is also frequent discussion of its importance as an antioxidant. Ascorbic acid is indeed a good scavenger of several reactive oxygen species^{76a} as summarized in Table 3, and it probably helps to recycle α -tocopherol in vivo. However, in the presence of transition metal ions (iron and copper), ascorbate can become prooxidant, acting as a reducing agent and generating O_2^{-} , H_2O_2 and OH^{-} . Normally, since such metal ions are available in very limited amounts in vivo, the antioxidant properties of ascorbate predominate.⁷⁶ However, ascorbate can be toxic if given to iron-overloaded patients without iron ion chelators.⁷⁷ In disease and tissue injury, transition metal ions do sometimes become more available, and the possibility that pro-oxidant actions of ascorbate might occur should not be ignored.50,53,78

Repair Systems

Normally, the production of ROS is approximately balanced by the antioxidant defense systems, i.e., the antioxidants are not present in great excess

Table 3. Ascorbic Acid as an Antioxidant

Scavenges O_2^- and HO_2^- (overall rate constant about $2.7 \times 10^5 \text{ M}^{-1} \text{ second}^{-1}$ at pH 7.4).

Scavenges water-soluble peroxyl (RO2⁺) radicals.

Scavenges thiyl and sulfenyl radicals.

- "Repairs," and so prevents damage by, radicals arising by attack of OH[•] upon uric acid.
- Can reduce carcinogenic nitrosamines to inactive products.
- Powerful scavenger of hypochlorous acid and an alternative substrate for the enzyme myeloperoxidase (possibly slowing HOCl formation).

Inhibits lipid peroxidation by hemoglobin– or myoglobin– H_2O_2 mixtures and prevents heme breakdown to release iron ions, by being preferentially oxidized.

- Powerful scavenger and quencher of singlet O_2 in aqueous solution.
- Regenerates α -tocopheryl radicals in membranes and lipoproteins.

Scavenges nitroxide radicals.

- Scavenges OH' radicals (rate constant $>10^9$ M⁻¹ second⁻¹).
- Protects plasma lipids against peroxidation induced by activated neutrophils.
- May protect against oxygen-derived species present in cigarette smoke.
- Powerful scavenger of O_3 and NO_2 in human body fluids, especially lung-lining fluids.

(hence the toxicity of extra O_2). One reason for this may be that production of some $O_2^{\cdot-}$ and H_2O_2 is useful in vivo, so that the body does not scavenge them with 100% efficiency. Indeed, there is good evidence for ongoing oxidative damage in the human body (Table 4),⁷⁹⁻⁸⁴ making clear the importance of repair systems.

Repair of oxidative DNA damage is particularly important,^{29,61} as the constant assault on our DNA by free radicals may contribute to the development of "spontaneous" human cancers.^{80,85} Indeed, it has been suggested that the aging process involves cumulative free radical damage over a lifetime.⁸⁶

Oxidative Stress

Because production of ROS and antioxidant defense are approximately balanced, it is easy to tip this balance in favor of the ROS and create the situation of oxidative stress.⁸⁷ Oxidative stress can result from:

- 1) depletions of antioxidants due to malnutrition^{87a} (e.g., through inadequate dietary intakes of α -tocopherol, ascorbic acid, sulfur-containing amino acids needed for GSH manufacture, or riboflavin) (needed to make the FAD cofactor in glutathione reductase);
- 2) excess production of reactive oxygen species,

Target of	
Damage	Evidence
DNA	Urinary excretion of DNA base damage products. ^{79,80} Low baseline levels of DNA damage products in DNA iso- lated from human cells (reviewed in Halliwell and Dizdaroglon ⁸¹).
Protein	Attack of free radicals upon proteins produces protein carbonyls. Low lev- els of these can be detected in human tissues and body fluids. ⁶²
Lipid	Accumulation of "age pigments" in tis- sues (reviewed in Halliwell and Gut- teridge ³²). Lipid peroxidation in ath- erosclerotic lesions. ⁸² Presence of end products of peroxidation in animal body fluids (for a recent review that examines the methodological prob- lems in measuring such products see Halliwell and Chirico ⁸³).
Uric acid	Attacked by several reactive oxygen species to generate allantoin, cyanuric acid, parabanic acid, oxonic acid, etc. These products are found in human body fluids and levels increase during oxidative stress. ⁸⁴

Table 4. Evidence that Oxidative Damage OccursIn Vivo

e.g., by exposure to elevated O_2 concentrations, the presence of toxins that are metabolized to produce free radicals, or excessive activation of "natural" radical-producing systems (e.g., inappropriate activation of phagocytic cells in chronic inflammatory diseases, such as rheumatoid arthritis and ulcerative colitis).^{32,88,89}

Cells can tolerate mild oxidative stress, which often results in up-regulation of the synthesis of antioxidant defense systems in an attempt to restore the balance. For example, if rats are gradually acclimatized to elevated O_2 , they can tolerate pure O_2 for much longer than control rats, apparently due to increased synthesis of antioxidant defense enzymes and of GSH in the lung.90,91 However, severe oxidative stress can produce major interdependent derangements of cell metabolism, including DNA strand breakage (often an early event), rises in intracellular "free" Ca2+, damage to membrane ion transporters and/or other specific proteins, and peroxidation of lipids. Cell injury and death may result.48,62,92,93 Oxidative stress can also lead to the release of transition metal ions able to promote such deleterious events as OH⁻ formation.^{32,93} It may, for example, cause Fe²⁺ release from mitochondria.⁹⁴ Superoxide can reductively mobilize small amounts of ferrous iron from ferritin.59,60 H₂O₂ can degrade hemoglobin, myoglobin, and probably other heme proteins to release iron ions from the heme ring.^{95,96}

Oxidative Stress and Human Disease

Does oxidative damage play a role in human disease? We have already seen that many of the biological consequences of excess radiation exposure may be due to OH⁻-dependent damage, and oxidative damage may account for damage to the retina of the eye (retinopathy of prematurity) in premature babies. Indeed, there are many papers in the biomedical literature suggesting a role for oxidative stress in other human diseases.⁹⁷

Tissue damage by disease, trauma, toxic agents, and other causes usually leads to formation of increased amounts of putative "injury mediators," such as prostaglandins, excess NO⁻, leukotrienes, interleukins, interferons, and tumor necrosis factors (TNFs). All of these at various times have been suggested as playing important roles in different human diseases. Currently, for example, there is much interest in the roles played by $TNF\alpha$, NO^{\cdot}, and interleukins in adult respiratory distress syndrome and septic shock. ROS can be placed in the same category, in that tissue damage will usually lead to increased ROS formation and oxidative stress. Figure 5 summarizes some of the reasons for this. Indeed, in most human diseases, oxidative stress is a secondary phenomenon, a consequence of the disease activity. That does not mean it is not important! Its importance varies in different disease states.⁸⁸ For example, oxidative damage to lipids in blood vessel walls seems to be a significant contributor to the development of atherosclerosis.^{24,25,82} Oxidative DNA damage may contribute to cancer and aging.^{80,85,86} Excess production of radicals probably contributes significantly to tissue damage in rheumatoid arthritis⁸⁸ and in inflammatory bowel diseases, such as Crohn's disease and ulcerative colitis.⁸⁹ There is growing evidence⁹⁸ that oxidative damage occurs in the neurodegenerative diseases (including the familial dominant form of ALS)^{68,69} and after traumatic brain injury.99

Since oxidative stress occurs to some degree after every tissue injury, the main question to be asked in evaluating its role in human disease is not "can we demonstrate oxidative stress?" but rather "does the oxidative stress that occurs make a significant contribution to disease activity?" The answer to the latter question appears to be "yes" in at least some cases. These include rheumatoid arthritis, the familial dominant form of ALS, and perhaps some other neurodegenerative diseases,⁹⁸ neurotrauma,⁹⁹ atherosclerosis,^{82,100} and ulcerative colitis.⁸⁹ However, it may well be "no" in many others. Elucidating the precise role played by free



Figure 5. How tissue damage can cause oxidative stress.

radicals has not been easy because they are difficult to measure, but the development of modern assay techniques is helping to solve this problem.⁸⁸

Conclusion: Nutritional Implications

We obtain certain essential antioxidants from the diet, primarily the tocopherols. Ascorbic acid may also have antioxidant functions. Many plant constituents have antioxidant activities in vitro, e.g., the flavonoids and other plant phenolics, but their bioavailability to humans and the significance of any role they might play are not clearly established, although research in this area is growing.¹⁰¹ Like ascorbate, many plant phenolics exert pro-oxidant effects in the presence of transition metal ions in vitro,¹⁰² but, again, the significance of this in vivo is uncertain. Epidemiologic evidence is accumulating to support the view that tocopherols decrease atherosclerosis and delay death from myocardial infarction, presumably by inhibiting lipid peroxidation.¹⁰³ Carotenoids, such as β -carotene and other plant pigments, may also have preventive effects against cancer and cardiovascular disease.¹⁰⁴ Several carotenoids can be made to exert antioxidant effects in vitro, but there is no evidence as yet that this mechanism produces any beneficial action in humans.105,106

Our endogenous antioxidant defenses are inad-

equate to prevent oxidative damage completely. Hence, sources of dietary antioxidants may be especially important to us. Determining their optimal intake is one of the greatest challenges in the nutrition and free radicals field today.

- 1. Balentine J. Pathology of oxygen toxicity. New York and London: Academic Press, 1982
- Ehrenkranz RA. Vitamin E and retinopathy of prematurity: still controversial. J Pediatr 1989;114: 801–3
- Gerschman K, Gilbert DL, Nye SW, Dwyer P, Fenn WO. Oxygen poisoning and X-irradiation: a mechanism in common. Science 1954;119:623–6
- Fridovich I. Superoxide dismutases. Methods Enzymol 1986;58:61–97
- Recknagel RO, Glende EA Jr, Dolak JA, Waller RL. Mechanisms of carbon tetrachloride toxicity. Pharmacol Ther 1989;43:139–54
- 6. Moncada S, Higgs A. The L-arginine-nitric oxide pathway. N Engl J Med 1993;329:2002-11
- Pryor WA, Stone K. Oxidants in cigarette smoke. Radicals, hydrogen peroxide, peroxynitrate and peroxynitrite. Ann NY Acad Sci 1993;686:12–28
- Janzen E, ed. Third international symposium on spin-trapping and aminoxyl radical chemistry. Free Radical Res Commun 1993;19(suppl 1):S1-230
- 9. von Sonntag C. *The chemical basis of radiation biology*. London: Taylor and Francis, 1987
- 10. Babior BM, Woodman RC. Chronic granulomatous disease. Semin Hematol 1990;27:247–59

- Maly FE. The B-lymphocyte: a newly-recognized source of reactive oxygen species with immunoregulatory potential. Free Radical Res Commun 1990;8:143–8.
- Meier B, Radeke H, Selle S, et al. Human fibroblasts release reactive oxygen species in response to treatment with synovial fluids from patients suffering from arthritis. Free Radical Res Commun 1990;8:149–60
- Murrell GAC, Francis MJO, Bromley L. Modulation of fibroblast proliferation by oxygen free radicals. Biochem J 1990;265:659–65
- 14. Burdon RH, Rice-Evans C. Free radicals and the regulation of mammalian cell proliferation. Free Radical Res Commun 1989;6:345–58
- Arroyo CM, Carmichael AJ, Bouscarel B, Liang JH, Weglicki WB. Endothelial cells as a source of oxygen-free radicals. An ESR study. Free Radical Res Commun 1990;9:287–96
- Babbs CF, Cregor MD, Turek JJ, Badylak SF. Endothelial superoxide production in buffer perfused rat lungs, demonstrated by a new histochemical technique. Lab Invest 1991;65:484–96
- Britigan BE, Roeder TL, Shasby DM. Insight into the nature and site of oxygen-centered free radical generation by endothelial cell monolayers using a novel spin trapping technique. Blood 1992;79:699–707
- Chance B, Sies H, Boveris A. Hydroperoxide metabolism in mammalian organs. Physiol Rev 1979;59:527–605
- Granger DN. Role of xanthine oxidase and granulocytes in ischemia-reperfusion injury. Am J Physiol 1988;255:H1269-75
- Dupuy C, Virion A, Ohayon R, Kaniewski J, Deme D, Pommier J. Mechanism of hydrogen peroxide formation catalyzed by NADPH oxidase in thyroid plasma membrane. J Biol Chem 1991;266:3739–43
- 21. Shapiro BM. The control of oxidative stress at fertilization. Science 1991;252:533-6
- Riley JCM, Behrman HR. Oxygen radicals and reactive oxygen species in reproduction. Proc Soc Exp Biol Med 1991;5:781-91
- Schreck R, Albermann KAJ, Baeuerle PA. Nuclear factor κB: an oxidative stress-responsive transcription factor of eukaryotic cells (a review). Free Radical Res Commun 1992;17:221–37
- Collins T. Endothelial nuclear factor-κB and the initiation of the atherosclerotic lesion. Lab Invest 1993;68:499–508
- Lusis AJ, Navab M. Lipoprotein oxidation and gene expression in the artery wall. Biochem Pharmacol 1993;46:2119–26
- Weiss SJ. Tissue destruction by neutrophils. N Engl J Med 1989;320:365–76
- Huie RE, Padmaja S. The reaction of NO with superoxide. Free Radical Res Commun 1993;18: 195–9
- Beckman JS, Beckman TW, Chen J, Marshall PA, Freeman BA. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. Proc Natl Acad Sci USA 1990;87:1620–4

- 29. Halliwell B, Aruoma OI, eds. DNA and free radicals. Chichester: Ellis-Harwood, 1993
- Malins DC. Identification of hydroxyl radical-induced lesions in DNA base structure: biomarkers with a putative link to cancer development. J Toxicol Environ Health 1993;40:247–61
- Olinksi R, Zastawny T, Budzbon J, Skokowski J, Zegarski W, Dizdaroglu M. DNA modification in chromatin of human cancerous tissues. FEBS Lett 1992;309:193–8
- 32. Halliwell B, Gutteridge JMC. *Free radicals in biology and medicine*, 2nd ed. Oxford, England: Clarendon Press, 1989
- Touati D. The molecular genetics of superoxide dismutase in *E. coli*. An approach to understanding the biological role and regulation of SODs in relation to other elements of the defense system against oxygen toxicity. Free Radical Res Commun 1989;8: 1–9
- Bielski BHJ. Reactivity of HO₂/O₂⁻ radicals in aqueous solution. J Phys Chem Ref Data 1985;14:1041– 100
- 35. Flink DH, Tuminello JF, Emptage MH. The inactivation of Fe S cluster containing hydro-lyases by superoxide. J Biol Chem 1993;268:22369-76
- Gardner PR, Fridovich I. Inactivation-reactivation of aconitase in *Escherichia coli*. A sensitive measure of superoxide radical. J Biol Chem 1992;267:8757– 63
- Zhang Y, Marcillat O, Giulivi C, Ernster L, Davies KJ. The oxidative inactivation of mitochondrial electron transport chain components and ATPase. J Biol Chem 1990;265:16330–6
- Beckman JS, Chen J, Ischiropoulos H, Crow JP. Oxidative chemistry of peroxynitrite. Methods Enzym 1994;233:229–40
- 39. White CR, Brock TA, Chang LY, et al. Superoxide and peroxynitrite in atherosclerosis. Proc Natl Acad Sci USA 1994;91:1044–8
- Laurindo FRM, da Luz PL, Uint L, Rocha TF, Jaeger RG, Lopes EA. Evidence for superoxide radical-dependent coronary vasospasm after angioplasty in intact dogs. Circulation 1991;83:1705–15
- Nakazono K, Watanabe N, Matsuno K, Sasaki J, Sato T, Inoue M. Does superoxide underlay the pathogenesis of hypertension? Proc Natl Acad Sci USA 1991;88:10045–8
- 42. Radi R, Beckman JS, Bush KKM, Freeman BA. Peroxynitrite oxidation of sulfhydryls. The cytotoxic potential of superoxide and nitric oxide. J Biol Chem 1990;266:4244–50
- Halliwell B, Hu ML, Louie S, et al. Interaction of nitrogen dioxide with human plasma. Antioxidant depletion and oxidative damage. FEBS Lett 1992;313: 62–6
- 44. Beckman JS, Ischiropoulos H, Zhu L, et al. Kinetics of superoxide dismutase- and iron-catalyzed nitration of phenolics by peroxynitrite. Arch Biochem Biophys 1992;298:438–45
- 45. van der Vliet A, O'Neill CA, Halliwell B, Cross CE, Kaur H. Aromatic hydroxylation and nitration of phenylalanine and tyrosine by peroxynitrite. Evidence

- Bielski BJH, Arudi RL, Sutherland MW. A study of the reactivity of HO₂/O₂⁻ with unsaturated fatty acids. J Biol Chem 1983;258:4759–61
- Aikens J, Dix TA. Perhydroxyl radical (HOO) initiated lipid peroxidation. The role of fatty acid hydroperoxides. J Biol Chem 1991;266:15091–8
- Cochrane CG. Mechanisms of oxidant injury of cells. Mol Aspects Med 1991;12:137–47
- Beauchamp C, Fridovich I. A mechanism for the production of ethylene from methional. The generation of the hydroxyl radical by xanthine oxidase. J Biol Chem 1970;243:4641–6
- 50. Halliwell B, Gutteridge JMC. Role of free radicals and catalytic metal ions in human disease. Methods Enzymol 1990;186:1–85
- Burkitt MJ. ESR spin trapping studies into the nature of the oxidizing species formed in the Fenton reaction: pitfalls associated with the use of 5,5-dimethyl-1-pyrroline-*N*-oxide in the detection of the hydroxyl radical. Free Radical Res Commun 1993;18:43-57
- Condeias LP, Patel KB, Stratford MRL, Wardman P. Free hydroxyl radicals are formed on reaction between the neutrophil-derived species superoxide anion and hypochlorous acid. FEBS Lett 1993;333: 151–3
- Halliwell B, Gutteridge JMC. The antioxidants of human extracellular fluids. Arch Biochem Biophys 1990;200:1–8
- 54. Weinberg ED. Cellular iron metabolism in health and disease. Drug Metab Rev 1990;22:531-79
- Stremmel W, Riedel HD, Niederau C, Strohmeyer G. Pathogenesis of genetic haemochromatosis. Eur J Clin Invest 1993;23:321–9
- Evans PJ, Evans RW, Bomford A, Williams R, Halliwell B. Metal ions catalytic for free radical reactions in the plasma of patients with fulminant hepatic failure. Free Radical Res 1994;20:139–44
- 57. Dizdaroglu M, Nackerdien Z, Chao BC, Gajewski E, Rao G. Chemical nature of in vivo DNA base damage in hydrogen peroxide-treated mammalian cells. Arch Biochem Biophys 1991;285:388–90
- Kyle ME, Nakae D, Sakaida I, Miccadei S, Farber JL. Endocytosis of superoxide dismutase is required in order for the enzyme to protect hepatocytes from the cytotoxicity of hydrogen peroxide. J Biol Chem 1988;263:3784–9
- Biemond P, Van Eijk HG, Swaak AJG, Koster JF. Iron mobilization from ferritin by superoxide derived from stimulated polymorphonuclear leukocytes. Possible mechanism in inflammation diseases. J Clin Invest 1984;73:1576–9
- Bolann BJ, Ulvik RJ. On the limited ability of superoxide to release iron from ferritin. Eur J Biochem 1990;193:899–904
- Breimer L. Repair of DNA damage induced by reactive oxygen species. Free Radical Res Commun 1991;14:159–71
- 62. Stadtman ER, Oliver CN. Metal-catalyzed oxidation

of proteins. Physiological consequences. J Biol Chem 1991;266:2005-8

- Brot N, Weissbach H. Biochemistry of methionine sulfoxide residues in proteins. Biofactors 1991;3: 91-6
- 64. Salo DC, Lin SW, Pacifici RE, Davies KJA. Superoxide dismutase is preferentially degraded by a proteolytic system from red blood cells following oxidative modification by hydrogen peroxide. Free Rad Biol Med 1988;5:335–9
- Burton GW, Traber MG. Vitamin E: antioxidant activity biokinetics and bioavailability. Annu Rev Nutr 1990;10:357–82
- Maiorino M, Chu FF, Ursini F, Davies KJ, Doroshow JH, Esworthy RS. Phospholipid hydroperoxide glutathione peroxidase is the 18-kDa selenoprotein expressed in human tumor cell lines. J Biol Chem 1991;266:7728–32
- Sevanian A, Wratten ML, McLeod LL, Kim E. Lipid peroxidation and phospholipase A2 activity in liposomes composed of unsaturated phospholipids: a structural basis for enzyme activation. Biochim Biophys Acta 1988;961:316–27
- Robberecht W, Sapp P, Viaene MK, et al. Cu/Zn superoxide dismutase activity in familial and sporadic amyotrophic lateral sclerosis. J Neurochem 1994;62:384–7
- Gurney ME, Pu H, Chiu AY, et al. Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. Science 1994;264: 1772–75
- Kagan VE, Serbinova EA, Packer L. Antioxidant effects of ubiquinones in microsomes and mitochondria are mediated by tocopherol recycling. Biochem Biophys Res Commun 1990;169:851–7
- Esterbauer H, Striegl G, Puhl H, Rotheneder M. Continuous monitoring of in vitro oxidation of human low density lipoprotein. Free Radical Res Commun 1989;6:67–75
- 72. Mukai K, Morimoto H, Okauchi Y, Nagaoka S. Kinetic study of reactions between tocopheroxyl radicals and fatty acids. Lipids 1993;28:753–6
- 73. Ernster L, Forsmark P, Nordenbrand K. The mode of action of lipid-soluble antioxidants in biological membranes: relationship between the effects of ubiquinol and vitamin E as inhibitors of lipid peroxidation in submitochondrial particles. Biofactors 1992;3:241–8
- Muller DPR, Goss-Sampson MA. Neurochemical, neurophysiological, and neuropathological studies in vitamin E deficiency. Crit Rev Neurobiol 1990;5: 239–63
- Levine M. New concepts in the biology and biochemistry of ascorbic acid. N Engl J Med 1986;314: 892–902
- Bendich A, Machlin LJ, Scandurra O, Burton GW, Wayner DDM. The antioxidant role of vitamin C. Adv Free Rad Biol Med 1986;2:419–44
- 76a. Halliwell B. How to characterize a biological antioxidant. Free Radical Res Commun 1990;9:1-32
- 77. Nienhuis AW. Vitamin C and iron. N Engl J Med 1981;304:170-1

- Hunt JV, Bottoms MA, Mitchinson MJ. Ascorbic acid oxidation: a potential cause of the elevated severity of atherosclerosis in diabetes mellitus? FEBS Lett 1992;311:161–4
- Stillwell WG, Xu HX, Adkins JA, Wishnock JS, Tannenbaum SR. Analysis of methylated and oxidized purines in urine by capillary gas chromatography– mass spectrometry. Chem Res Toxicol 1989;2: 94–9
- Ames BN. Endogenous oxidative DNA damage, aging, and cancer. Free Radical Res Commun 1989;7: 121–8
- Halliwell B, Dizdaroglu M. The measurement of oxidative damage to DNA by HPLC and GC/MS techniques. Free Radical Res Commun 1992;16:75–87
- Steinberg D, Parthasarathy S, Carew TE, Khoo JC, Witztum JL. Beyond cholesterol. Modifications of low-density lipoprotein that increase atherogenicity. N Engl J Med 1989;320:915–24
- Halliwell B, Chirico S. Lipid peroxidation: its mechanism, measurement and significance. Am J Clin Nutr 1993;57:715S-25S
- Kaur H, Halliwell B. Action of biologically-relevant oxidizing species upon uric acid. Identification of uric acid oxidation products. Chem Biol Interac 1990;73:235–47
- Totter JR. Spontaneous cancer and its possible relationship to oxygen metabolism. Proc Natl Acad Sci USA 1980;77:1763–7
- 86. Harman D. Free radical involvement in ageing. Pathophysiology and therapeutic implications. Drugs Ageing 1993;3:60–80
- Sies H, ed. Oxidative stress, oxidants and antioxidants. London and New York: Academic Press, 1991
- 87a. Golden M. Free radicals and the aetiology of kwashiorkor. Biochemist 1994;June/July:12–15
- Halliwell B, Gutteridge JMC, Cross CE. Free radicals, antioxidants, and human disease: where are we now? J Lab Clin Med 1992;119:598–620
- Grisham MB. Role of reactive oxygen metabolites in inflammatory bowel disease. Curr Opinion Gastroenterol 1993;9:971–80
- Frank L, Iqbal J, Hass M, Massaro D. New "rest period" protocol for inducing tolerance to high O₂ exposure in adult rats. Am J Physiol 1989;257; L226-31
- Iqbal J, Clerch LB, Hass MA, Frank L, Massaro D. Endotoxin increases lung Cu, Zn superoxide dismutase mRNA: O₂ raises enzyme synthesis. Am J Physiol 1989;257:L61–4
- 92. Orrenius S, McConkey DJ, Bellomo G, Nicotera P.

Role of Ca^{2+} in toxic cell killing. Trends Pharmacol Sci 1989;10:281–5

- 93. Halliwell B. Oxidants and human disease: some new concepts. FASEB J 1987;1:358-64
- Merryfield ML, Lardy HA. Ca²⁺ mediated activation of phosphoenolpyruvate carboxykinase occurs via release of Fe²⁺ from rat liver mitochondria. J Biol Chem 1982;257:3628–35
- 95. Gutteridge JMC. Iron promoters of the Fenton reaction and lipid peroxidation can be released from haemoglobin by peroxides. FEBS Lett 1986;201: 291-5
- 96. Puppo A, Halliwell B. Formation of hydroxyl radicals in biological systems. Does myoglobin stimulate hydroxyl radical production from hydrogen peroxide? Free Radical Res Commun 1988;4:415–22
- Gutteridge JMC. Free radicals in disease processes: a compilation of cause and consequence. Free Radical Res Commun 1993;19:141–58
- Halliwell B. Reactive oxygen species and the central nervous system. J Neurochem 1992;59:1609– 23
- 99. Hall ED, Braughler JM. The role of oxygen radicalinduced lipid peroxidation in acute central nervous system trauma. In: Halliwell B, ed. *Oxygen radicals and tissue injury*. Bethesda: FASEB, 1988:92–8
- Esterbauer H, Gebicki J, Puhl H, Jurgens G. The role of lipid peroxidation and antioxidants in oxidative modification of LDL. Free Rad Biol Med 1992;13:341–90
- Hertog MGL, Feskens EJM, Hollman PCH, Katan MB, Kromhout D. Dietary antioxidant flavonoids and the risk of coronary heart disease. The Zutphen elderly study. Lancet 1993;342:1007–11
- 102. Laughton MJ, Halliwell B, Evans PJ, Hoult JRS. Antioxidant and pro-oxidant actions of the plant phenolics quercetin, gossypol and myricetin. Biochem Pharmacol 1989;38:285–65
- Byers T. Vitamin E supplements and coronary heart disease. Nutr Rev 1993;51:333–45
- Krinsky NI. Effects of carotenoids in cellular and animal systems. Am J Clin Nutr 1991;53:238S-46S
- 105. Raven PD, Khouw A, Beltz WF, Parthasarathy S, Witztum JL. Effect of dietary antioxidant combinations in humans. Protection by vitamin E but not by β -carotene. Arteriosclerosis Thromb 1993;13:590–600
- 106. Princen HMG, van Poppel G, Vogelezang C, Buytenhek R, Kok FJ. Supplementation with vitamin E but not β-carotene in vivo protects low density lipoprotein from lipid peroxidation in vitro. Arteriosclerosis Thromb 1992;12:554–62