

# Angiopoietin: A TIE(d) Balance in Tumor Angiogenesis

Winston S.N. Shim,<sup>1,2</sup> Ivy A.W. Ho,<sup>3</sup> and Philip E.H. Wong<sup>1</sup>

<sup>1</sup>Research and Development Unit, National Heart Centre; <sup>2</sup>Department of Surgery, National University of Singapore; and <sup>3</sup>Laboratory of Cancer Gene Therapy, National Cancer Centre, Singapore, Singapore

## Abstract

Angiopoietins (ANG-1 and ANG-2) and their TIE-2 receptor tyrosine kinase have wide-ranging effects on tumor malignancy that includes angiogenesis, inflammation, and vascular extravasation. These multifaceted pathways present a valuable opportunity in developing novel inhibition strategies for cancer treatment. However, the regulatory role of ANG-1 and ANG-2 in tumor angiogenesis remains controversial. There is a complex interplay between complementary yet conflicting roles of both the ANGs in shaping the outcome of angiogenesis. Embryonic vascular development suggests that ANG-1 is crucial in engaging interaction between endothelial and perivascular cells. However, recruitment of perivascular cells by ANG-1 has recently been implicated in its antiangiogenic effect on tumor growth. It is becoming clear that TIE-2 signaling may function in a paracrine and autocrine manner directly on tumor cells because the receptor has been increasingly found in tumor cells. In addition,  $\alpha_5\beta_1$  and  $\alpha_v\beta_5$  integrins were recently recognized as functional receptors for ANG-1 and ANG-2. Therefore, both the ligands may have wide-ranging functions in cellular activities that affect overall tumor development. Collectively, these TIE-2–dependent and TIE-2–independent activities may account for the conflicting findings of ANG-1 and ANG-2 in tumor angiogenesis. These uncertainties have impeded development of a clear strategy to target this important angiogenic pathway. A better understanding of the molecular basis of ANG-1 and ANG-2 activity in the pathophysiologic regulation of angiogenesis may set the stage for novel therapy targeting this pathway. (Mol Cancer Res 2007;5(7):655–65)

## Introduction

Angiogenesis as a rate-limiting step in tumor growth was first mooted more than 3 decades ago (1). Malignant cells transverse such limitations by accumulating mutations that stoke angiogenic response or by sequestering circulating growth inhibitors (2). Intratumoral microvascular density is now recognized as an important and independent prognostic marker for metastasis and overall survival in patients with breast, cervical, colon, lung, renal, ovarian, and esophageal carcinomas (3–9).

Tumor ecosystem comprises malignant cells, endothelial cells, perivascular cells, fibroblasts, inflammatory cells, and their surrounding extracellular matrices. These constituents continuously partake in the evolution of the milieu by expressing a myriad of autocrine and paracrine factors that influence the outcome of the disease. Vascular endothelial growth factor (VEGF) and the angiopoietins (ANG) are among the most important growth factors in the ecosystem. Signaling primarily through their endothelial receptors, these factors are responsible for proliferation, migration, and survival of activated vascular endothelial cells. In addition, these signaling pathways are believed to be responsible for the integrity, maturation, and maintenance of the vascular network. Furthermore, recent findings also suggest that certain types of cancer cells may also be directly responsive to these factors (10–14), although their significance in disease progression remains largely undefined.

ANG-1 is critically important in the formation of vascular networks during developmental angiogenesis (15). Gene transfer of ANG-1 has been shown to promote robust angiogenesis in ischemic tissues (16–18). Surprisingly, ANG-1 has recently been implicated in the inhibition of pathologic vascular expansion via its effect on vessel maturation (19–21). This peculiar idiosyncratic effect of ANG-1 between physiologic and pathologic angiogenesis has profound implication in the development of strategy that targets this pathway for anticancer therapy. Similarly, the context-dependent activation of ANG-2 and inactivation of its cognate TIE-2 receptor also complicate the understanding of this signaling pathway.

Questions aimed at the molecular basis of ANG-1 and ANG-2 action in the angiogenic cycle may help to unravel the conundrum. Is ANG-2 an antagonist or agonist of TIE-2 signaling? Is there a transition phase between proangiogenic and antiangiogenic roles of ANG-1? What are the circumstances that determine the transition between these roles? What role does vessel maturation and vascular stability play in this reversal of function? Does extraendothelial TIE-2 signaling or TIE-2–independent signaling in endothelial cells affect the angiogenic outcome? How does the transition between TIE-2–dependent and TIE-2–independent activity of

Received 2/8/07; revised 4/6/07; accepted 5/7/07.

**Grant support:** National Medical Research Council Singapore grants NMRC0729/2003 and NMRC0730/2003 and Biomedical Research Council Singapore grant BMRC04/1/32/19/355 (W.S.N. Shim) and Singapore Millennium Foundation (I.A.W. Ho).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

**Requests for reprints:** Winston S.N. Shim, Research and Development Unit, National Heart Centre, 17 Third Hospital Avenue, Singapore 168752, Singapore. Phone: 65-6435-0752. E-mail: surssnw@nus.edu.sg

Copyright © 2007 American Association for Cancer Research. doi:10.1158/1541-7786.MCR-07-0072

ANG-1 and ANG-2 affect disease outcome? In this review, we highlight the controversies surrounding this important pathway and attempt to elucidate this Jekyll and Hyde behavior of ANG-1 or ANG-2 to gain insights into the complex tumor ecosystem.

## The ANG Family

The human ANG family comprises the ligands ANG-1, ANG-2, and ANG-4. Their cognate TIE-2 receptor (and a closely related orphan receptor, TIE-1) is mainly expressed in endothelial cells. They lack mitogenic activity toward endothelial cells [although conflicting data are emerging that showed otherwise (22, 23)] but affect distinct aspects of vascular development (24, 25). Transgenic mice lacking ANG-1 or overexpressing ANG-2 have defects attributed to disrupted interaction between endothelial and perivascular cells (15, 25). Mice lacking ANG-2 have defective lymphatic system that can be compensated by ANG-1 (26). This suggests a wide-ranging effect of ANG-1 on both vascular and lymphatic systems. Transgenic mice overexpressing ANG-1 produced enlarged vessels with highly regulated junctional complexes that resulted in leakage-resistant vessels (27). Consistently, hyperpermeable vessels in VEGF-overexpressing mice were restituted by ANG-1, whereby double transgenic mice of ANG-1/VEGF resulted in enhanced angiogenesis with leakage-resistant vessels (28). This suggests a complementary yet contradicting relationship between these important growth factors.

ANGs possess distinct structural domains with their receptor-binding site residing in the fibrinogen-like domain, whereas the coiled-coil region (Fig. 1) multimerizes the former into active multimeric ligands of ANG-1 or ANG-2 (29, 30). Paradoxically, dimeric form of ANG-1 has been found to inactivate TIE-2 receptor (30), and some isoforms of ANG-1 have been reported to negatively regulate TIE-2 activation (31). Interestingly, both ligands were recently reported to function in a TIE-2-independent manner whereby  $\alpha_5\beta_1$  and  $\alpha_v\beta_5$  integrins could act as functional receptors for ANG-1 and ANG-2 (Fig. 2; ref. 32). Interaction between isoforms of ANG-1 and integrin receptors may not be unexpected because differential binding of VEGF isoforms with family of integrins has been reported to induce distinct cellular responses (33). Therefore, activity of ANG-1 and ANG-2 is likely to have broad ramifications because integrins are expressed in multiple cell types.

## ANG-1

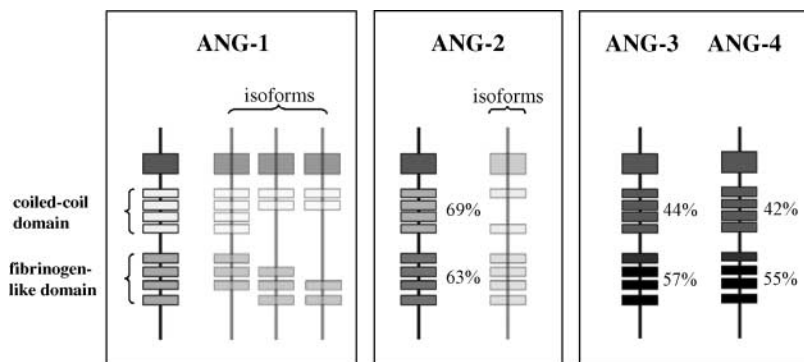
Up-regulation of ANG-1 in high-grade gliomas (34, 35), non-small cell lung carcinoma (36), plasmacytomas (37), and ovarian (38), breast (39, 40), and gastric (41) carcinomas are strongly correlated with tumor malignancy. Furthermore, overexpression of ANG-1 in HeLa, GS9L, U87, U373, and U343 cell lines has been reported to increase tumor growth (see Table 1; refs. 13, 42, 43). Moreover, ANG-1-mobilized, bone marrow-derived endothelial cells have been linked to brain tumor angiogenesis (44) and myeloproliferative disorders (45). Surprisingly, overexpression of ANG-1 in MCF-7 breast cancer cells (19), HT29 colon cancer cells (21, 46), TA3 mammary cancer cells (47), Lewis lung carcinoma (47), and A431 squamous cell carcinoma (20) has been reported to show significant antitumor effect. Its inhibitory effect was linked to recruitment of perivascular cells by ANG-1 that restricts further expansion of tumor vasculature.

## ANG-2

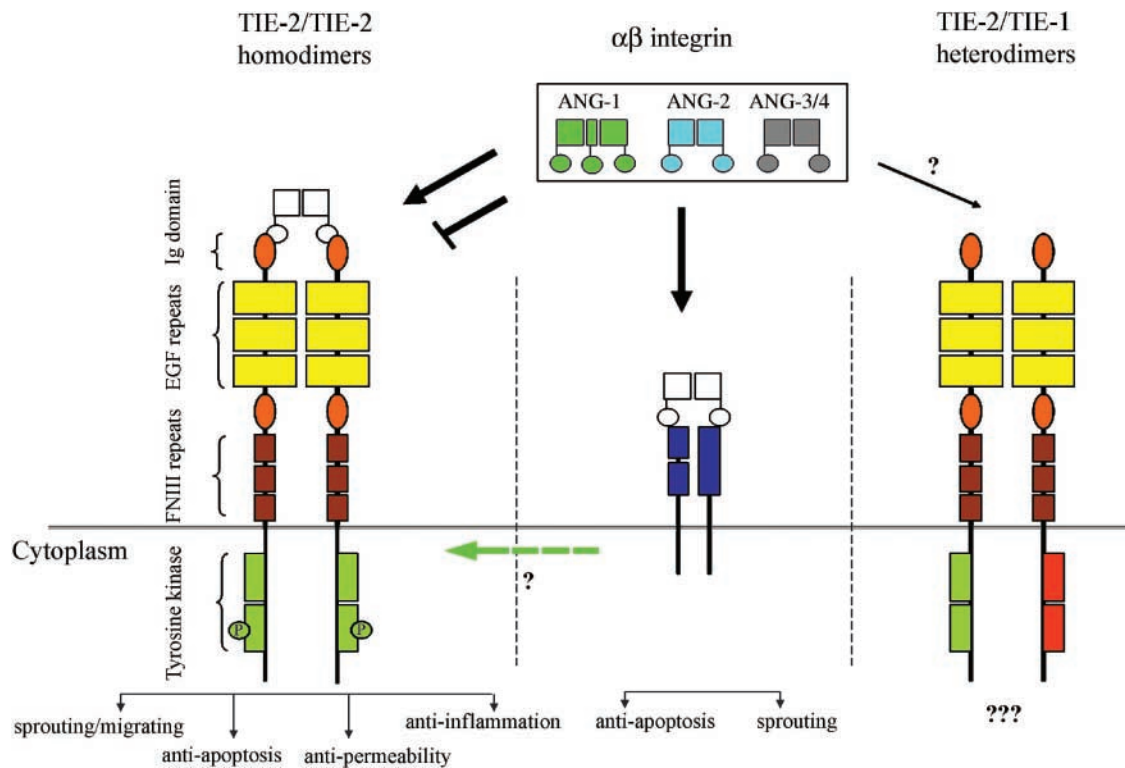
The role of ANG-2 in TIE-2 receptor activation is similarly controversial (see Table 2). Its peculiar context-dependent agonistic and antagonistic relationship with TIE-2 (25, 48-50) has further complicated the understanding of ANG-2 function in vascular development. Embryonic ANG-2 overexpression results in a major disruption of the developing vascular system, suggesting an antagonistic role in angiogenesis (25). Furthermore, it counteracts the angiogenic activity of VEGF and antagonizes the synergistic effect of VEGF with basic fibroblast growth factor in angiogenesis (51, 52). In addition, lung and mammary carcinomas that overexpressed ANG-2 and specific induction of ANG-2 in gliomas were found to retard tumor growth and metastasis (42, 47). In contrast, overexpression of ANG-2 in hepatomas, gliomas, and colorectal and gastric carcinomas was found to enhance angiogenesis and augment tumor malignancy (46, 53-56). Furthermore, strong correlation of ANG-2 with ANG-1 in neuroblastoma (57), gliomas (14, 35), breast and prostate carcinomas (58), hepatocellular carcinoma (59), non-small cell lung carcinomas (60, 61), and gastric adenocarcinoma (62) has been associated with aggressive tumor growth.

## ANG-3/ANG-4

The other two members of ANG family, ANG-3 and ANG-4, are not well studied but are believed to be interspecies



**FIGURE 1.** Structural organization of the ANG family. There are four ANG-1 isoforms with varying regulating activity on TIE-2 receptor (31). ANG-2 (25) and its isoform (125) are believed to be natural antagonists of the TIE-2 pathway. ANG-3 and ANG-4 are believed to be species orthologues in mouse and human, respectively (63), which regulate TIE-2 in a species-specific manner (65). The coiled-coil domain is involved in the multimerization of the ligands and the fibrinogen-like domain functions as the receptor-binding motif. The percentages in each box indicate identity to ANG-1 domains.



**FIGURE 2.** Structural organization of ANG-binding receptors. ANG-1 is known to form trimers and multimers to homodimerize and induce tyrosine phosphorylation of the TIE-2 receptor for intracellular signaling (29, 30). The other ANGs form dimers to bind to TIE-2 receptor (65). ANG-2 acts as antagonistic ligand for TIE-2 in low concentration but is able to activate TIE-2 in high concentration (49). TIE-2/TIE-1 heterodimerization is known to inhibit ANG-2 activation of the receptor (73). ANG-3 and ANG-4 are able to activate TIE-2 receptor in a species-specific manner (65). The integrin  $\alpha_5\beta_1$  and  $\alpha_v\beta_5$  receptors may transduce ANG-1 and ANG-2 signals independent of TIE-2 (32, 76, 77), but they may also work synergistically with TIE-2 receptor (131).

orthologues between mouse and human, respectively (63). The function of ANG-3 and ANG-4 in angiogenesis is equally controversial compared with the more established members of the family. ANG-3 has been reported to act as antagonist that interferes with ANG-1 activation of TIE-2 (63) and Akt in tumor growth (64). However, ANG-3 was recently found to strongly activate mouse TIE-2 receptor, but not its human counterpart, whereas ANG-4 displayed no such species selectivity in TIE-2 activation (65). This may have ramifications in the previously reported results of human ANG-1 and ANG-2 in mouse tumor models whereby the interfering effect of endogenous ANG-3 on TIE-2 binding by the ectopically expressed ligands may not have been properly accounted for.

### Dr. Jekyll and Mr. Hyde in Tumor Angiogenesis

ANG-1 and ANG-2 are known to respond differentially to hypoxia with the latter often being up-regulated in hypoxic/ischemic tissues (66-69). ANG-1 is mainly produced by vascular mural cells, such as smooth muscle cells and pericytes, whereas endothelial cells are the main producers of ANG-2. Therefore, autocrine regulation of TIE-2 activity by ANG-2 may render the receptor less responsive to exogenous stimuli and presents a unique self-modulatory function to endothelial cells during angiogenesis (70). Furthermore, TIE-2 activity is autoinhibited by its COOH terminus (71, 72) and its ligand receptivity toward ANG-2 is reportedly modulated through TIE-1 heterodimerization (73), suggesting a tight regulatory control at the receptor level.

Intracellular signaling pathway of TIE-2 involves multiple cytosolic docking partners [for detail, see review (74)], suggesting that it may be regulated and coordinated in a dose- and spatiotemporal-dependent manner. This is evident by the unique agonistic and antagonistic relationship between ANG-1 and ANG-2 on TIE-2 phosphorylation in endothelial cells but not other cell types (27).

The recent finding of extraendothelial TIE-2 receptor expression further complicates the understanding of ANG/TIE-2 system (see Table 3). It remains to be seen if nonendothelial TIE-2 receptors are functional or functionally similar to the endothelial-specific TIE-2 receptors. Nevertheless, TIE-2 receptors in trophoblasts have been found to mediate cellular migration and proliferation by interacting with ANG-1 and ANG-2, showing their direct effect on non-endothelial cells (75). Similar expression of VEGF receptors in various tumor cells has also been noted, but their implication in tumor angiogenesis remains largely undefined (12). Besides that, TIE-2-independent signaling of ANG-1 and ANG-2 is also increasingly recognized to have important functional roles in cellular behavior. The  $\alpha_5\beta_1$  and  $\alpha_v\beta_5$  integrins have recently been implicated in the differential cell spreading and migration activity of endothelial cells in response to ANG-1 and ANG-2 (32). Moreover, ANG-1 has been found to confer significant survival benefit to myocytes and affect neuronal patterning via  $\beta_1$  integrin signaling (76, 77). Therefore, integrin-expressing tumor cells may respond to ANG-1 and ANG-2 independent of

**TABLE 1. Effect of Stable Ectopic Expression of ANG-1 in Tumor Models**

Tumor type	ANG-1	ANG-2	VEGF	TIE-2	Angiogenesis	Outcomes	Reference
Astrocytomas	↑	ND	=	—	↑	Inducible expression in U87-MG cell line Increase glomeruloid bodies	(135)
Breast cancer	↑	ND	ND	ND	↓	Increase proliferating tumor cells and MVD Stable expression in MCF-7 cell line	(19)
Cervical cancer	↑	ND	=	ND	↑	Retard tumor growth in spite of the presence of FGF-2 Stable expression in HeLa cell line	(13)
Colorectal cancer	↑	+	ND	ND	↓	Increase MVD Decrease tumor cell apoptosis Tumor cell proliferation unaffected	(46)
Colorectal cancer	↑	ND	ND	ND	↓	Increase vessel plasticity with fewer pericytes Stable expression in HT29 cell line	(136)
Glioblastoma	↑	=	=	=	↑	Decrease MVD Reduced tumor cell proliferation Stable transfected KM12L4 cell line	(42)
Glioblastoma	↑	ND	↑	ND	↑	Decrease MVD Decrease tumor cell proliferation and metastasis Decrease ascites formation	(43)
Hepatic colon cancer	↑	ND	ND	ND	↓	Inducible expression in GS9L cell line Highly branched vessels Mature vessels covered with pericytes Increase number of vessels <500 μm <sup>2</sup>	(21)
Squamous cells carcinoma	↑	+	=	↑phos	↓	Increase MVD Stable expression in U87, U373, and U343 cell lines Increase MVD only when VEGF is elevated	(20)
Lung carcinoma	↑	+	=	ND	—	Stable expression in HT29 cell line Higher pericyte coverage in tumor vessels Decrease MVD	(47)
Mammary carcinoma	↑	ND	=	ND	—	Decrease proliferating tumor cells Decrease vascular leakage Stable expression in A431 cell line	(47)
Cervical cancer	↓	ND	=	ND	↓	No change to MVD and VEGF Increase smooth muscle cell coverage of vessels Stable expression in Lewis lung carcinoma cell line	(137)
Gastric carcinoma	↓	ND	ND	ND	↓	No effect on tumor and endothelial cell apoptosis No effect on MVD or vessel maturation Stable expression in TA3 mammary carcinoma cell line	(138)

NOTE: ↑ or ↓ indicates increased or decreased expression levels compared with control tumors; = indicates unchanged expression levels; — indicates no effect on the outcome; + indicates the factor was detectable but levels not quantified against control tumors.  
Abbreviations: MVD, microvascular density; ND, not determined; phos, phosphorylated; FGF, fibroblast growth factor.

the vascular effects of these ligands. Although its implications in tumor development await further clarifications, such relationship between VEGF and integrins has been documented in endothelial cells and, notably, in tumor cells and tumor angiogenesis (78, 79).

In conclusion, the dynamic differential induction of ANG-1 and ANG-2 expression coupled with their paracrine/autocrine receptor-binding activity and the possibility of TIE-1 (73) and TIE-2 (80) cross-modulating their ligand-binding activity indicate a unique self-regulatory mechanism in endothelial responsiveness. Their expression and regulation are expected to have broad implications in the resulting angiogenesis or lack of it (Fig. 3). Such complex interrelationships between TIE-1/TIE-2 and TIE-2/integrin signaling in endothelial and non-endothelial cells may explain the seemingly contradicting outcomes in the ectopic expression of ANG-1 and ANG-2 in various tumor models (Tables 1 and 2).

### Identity Crisis of ANG-1 and ANG-2: Angiogenic or Antiangiogenic?

Some slow growing and poorly angiogenic tumors with low VEGF expression have been shown to exhibit high microvascular permeability, suggesting that threshold levels of VEGF-induced vessel permeability are considerably lower than those needed for inducing angiogenesis (81, 82). Furthermore, microenvironmental concentration rather than the overall dose of VEGF has been found to be important in determining normal and pathologic angiogenic outcome (83). Such parallel observation is yet to be drawn on the diverse roles of ANG-1 in endothelial survival, sprouting, vessel maturation, and vascular permeability. This complex interplay between related, yet at times, conflicting roles of ANG-1 may be crucial in determining the outcome of angiogenesis. Central to this conflict is the dichotomy of functions and processes required for vascular sprouting and vessel maturation.



Vessel sprouts have been shown to loosen vascular integrity and intercellular contact among neighboring endothelial and smooth muscle cells in response to ANG-1 (84). This plastic state enables the endothelium to actively respond to angiogenic factors, such as VEGF, whereas mature vessels that are covered by smooth muscle cells are less responsive to stimulation of VEGF (83, 85). A similar observation has been reported between VEGF and ANG-2 whereby, in the presence of VEGF, ANG-2 promotes vascular sprouts whereas, in its absence, vascular regression accelerates (55, 86). Hence, this window period may enable fine tuning of the neovasculature to adapt to its microenvironment through a regulated process of pruning and remodeling because these plastic vessels are likely to be more susceptible to apoptosis (87, 88). Therefore, the major function of ANG-1 may lie with its antiapoptotic effect on endothelial cells during this plastic phase. It may possibly only have an indirect role in vessel maturation. In fact, conditions favoring vascular maturation and survival in different tissues have distinct consequences on functional outcome. For example, the disparity in the function of ANG-1 is evident in the conflicting conclusions proposed by Du et al. (89) and Zhao et al. (90) on the pathophysiology of pulmonary hypertension. Unregulated recruitment of perivascular cells to the vascular pulmonary networks by ANG-1 or excessive antiapoptosis signal from

ANG-1 on terminal arterioles in the pulmonary vascular bed was separately attributed as the etiology of the disease.

The potent pro-survival effect of ANG-1 alone or in synergy with VEGF has been found to protect endothelial cells from apoptosis (91-96). ANG-1-mediated phosphatidylinositol 3-kinase-dependent activation of Akt and attachment to extracellular matrix are central to the survival of endothelial cells. This antiapoptotic effect is mediated through up-regulation of survivin (97) and suppression of caspase-3, caspase-7, and caspase-9 activity as well as inhibition of second mitochondrial-derived activator of caspase (Smac) release (98, 99). The protective role is evident in radiation, mannitol, and low-density lipoprotein-treated endothelial cells whereby apoptosis was ameliorated by addition of ANG-1 (92, 100, 101). In fact, vascular defects of disrupted endothelial and myocardial layers observed in the ANG-1<sup>-/-</sup> and TIE-2<sup>-/-</sup> mutant mice may be due to impaired survival of endothelium rather than deficiency in vessel maturation as previously thought (102, 103). Indeed, persistent perivascular cell recruitment in vessels composed of TIE-2-deficient endothelial cells that subsequently apoptosed strongly supports this contention (103, 104). Furthermore, overexpression of ANG-1 in the skin and lung failed to show evidence of enhanced recruitment of perivascular cells to the vessels (23, 27, 105). Dilated and pericyte-scarce vessels in

**TABLE 2. Effect of Stable Ectopic Expression of ANG-2 in Tumor Models**

Tumor Type	ANG-1	ANG-2	VEGF	TIE-2	Angiogenesis	Outcomes	Reference
Colorectal	ND	↑	ND	ND	↑	Stable expression in HT29 cell line Increase MVD Enhance tumor cell proliferation	(46)
Gastric	ND	↑	+	ND	↑	Stable expression in MKN-7 cell line Highly vascularized and metastatic tumor Decrease vessel maturation	(56)
Glioma	ND	↑	ND	ND	↑	Stable expression in U87MG cell line Highly invasive with up-regulated MMP-2 Increase angiogenesis	(139)
Glioma	ND	↑	ND	ND	↓	Stable expression in U87 cell line Increase tumor necrosis Decrease vascularization	(140)
Glioblastoma	=	↑	=	↑	↓	Inducible stable expression in GS9L cell line Aberrant vascular cords with aggregated endothelial cells with narrow lumens Less mature vessels with few pericytes Discontinuous basement membrane Decrease MVD Tumor apoptosis unaffected	(42)
Hepatomas	=	↑	ND	ND	↑	Stable expression in HuH-7 cell line Hemorrhagic tumors with hypervascular phenotypes	(53)
Hepatomas	ND	↑	ND	ND	↑	Stable expression in Morris hepatoma cell line Increase tumor perfusion and vascularization Up-regulate Flk-1 expression	(141)
Lung carcinoma	ND	↑	=	ND	↓	Stable expression in Lewis lung carcinoma cell line Increase tumor and endothelial cell apoptosis Decrease metastatic property Decrease vessel maturity	(47)
Mammary carcinoma	ND	↑	=	ND	↓	Stable expression in TA3 mammary carcinoma cell line Increase tumor and endothelial cell apoptosis Decrease metastatic activity Decrease vessel maturity	(47)
Squamous cell carcinoma	+	↑	=	=	—	Stable expression in A431 cell line No effect on MVD or vessel maturation	(20)

NOTE: ↑ or ↓ indicates increased or decreased expression levels compared with control tumors; + indicates the factor was detectable but levels not quantified against control tumors; = indicates unchanged expression levels; — indicates no effect on the outcome.  
Abbreviation: MMP, matrix metalloproteinase.

**TABLE 3. Physiology and Pathology of Extraendothelial Expression of TIE-2 Receptor**

	Findings and Outcomes	Reference
<b>Cancer cell types</b>		
Inflammatory breast cancer cell line	Increase hematogenous metastases and correlated with poor prognosis	(39)
HeLa cervical cell line	Enhance survival of cervical tumor cells	(13)
Neoplastic glial cells	Associated with disease progression and matrix adhesion	(142)
Liver oval cells	Involve in preneoplastic to neoplastic conversion of hepatocytes	(143)
Thyroid tumor cells	Involve in cellular proliferation	(123)
Non-small cell lung carcinoma cells	Unknown function	(60)
Cancerous prostate cells	Unknown function	(124)
Glioma cell lines	Unknown function	(14)
Gastric carcinoma cells	Unknown function	(62)
<b>Normal cell types</b>		
Fetal trophoblasts	Involve in the proliferation, migration, and nitric oxide release	(75)
Ganglion cells	Promote neurite outgrowth when stimulated by ANG-1	(144)
Monocytes and mesenchymal cells	Promote paracrine angiogenic effect and tumor homing	(145)
Nerve cells	Phosphorylated by ANG-1 to prevent apoptosis of neuronal culture through phosphatidylinositol 3-kinase/Akt	(146)
Smooth muscle cells	To synchronize intercellular communication between endothelial and smooth muscle cells	(59)
Synovial cells	Correlate with cellular proliferation and possibly their pathogenesis	(147)
Synoviocytes and stromal fibroblasts	Unknown function	(148)
Synovial lining cells and macrophages	Unknown function	(149)
Choroidal neovascular membranes	Unknown function	(150)
Neuronal and Schwann cells	Unknown function	(151)
Glandular endometrial epithelial cells	Unknown function	(152)
Thyroid and granulosa follicular cells	Unknown function	(153)
Granuloma-associated mesenchymal cells	Unknown function	(154)
Mesenchymal cells and osteoblasts	Unknown function	(155)

venous malformation that are associated with excessive activation of TIE-2 receptor (106) do not support a vessel maturation role for ANG-1. Furthermore, inhibition of TIE-2 function in retinal vasculature failed to affect pericyte recruitment (107). In addition, ANG-1 restored hierarchical structure of vascular network and rescued retinal edema and hemorrhage in the complete absence of smooth muscle cells (108).

The previous observations that ANG-1 restricted tumor growth by promoting vessel integrity via pericyte recruitment (19-21) is difficult to reconcile with the current contradicting findings. It is unclear how vessel maturity may have played a significant role in retarding the tumor angiogenesis. In fact, maturation of vessels and normalization of microcirculation by smooth muscle cell coverage have been linked to a more aggressive tumor growth, possibly due to better nutrient exchange in the previously dysfunction vasculature (42). The reported tumor-inhibiting effect of ANG-1 may be related to its anti-inflammation action (109, 110) because inflammation was recognized as a key trigger for pathologic angiogenesis mediated by VEGF (111).

It is noteworthy that neovascularized tumors exhibit temporal angiogenic phenotype because not all parts of the tumor vessels are concurrently participating in angiogenesis. For example, TIE-2 expression was reported to be restricted to stromal vessels rather than intratumoral vessels in human mammary carcinomas (112). Furthermore, hypoxia-inducing factor-1 modulates the expression of ANG-1 and ANG-2 in a cell type-specific manner, whereby ANG-2 expression was induced in endothelial cells but suppressed in smooth muscle cells, whereas ANG-1 levels were unaffected in both cell types (113). Moreover, only a subset of endothelial cells is responsive to hypoxia induction of ANG-2 (114). Therefore, failure of signal transduction from TIE-2 receptors in different popula-

tions of endothelial cells may account for the observed discrepancies in the action of ANG-1 and ANG-2 (48, 115).

The present evidence suggests that ANG-1 predominantly functions as a survival factor leading to angiogenic sprouting rather than a vessel maturation agent that restricts tumor expansion. There may be a more complex controlling mechanism for vessel maturity whereby ANG-1 may act only indirectly, perhaps, in cooperation with other major mediators, such as platelet-derived growth factor (87), ephrin (116), transforming growth factor- $\beta$ , and sphingosine-1-phosphate (117).

### Angiogenic Cycle Mediated through TIE-2 Pathway

Induction and up-regulation of TIE-2 and ANG-2 expression in endothelial cells are regulated by hypoxia and proinflammatory cytokines, such as tumor necrosis factor- $\alpha$  and interleukin-1 $\beta$  (68, 118-122). Conversely, such stimuli down-regulate the expression of ANG-1 (66, 67), suggesting a delicate inverse relationship between ANG-1 and ANG-2 in the regulation of TIE-2 signaling. Therefore, spatiotemporal changes of these unique relationships among ANG-1, ANG-2, and TIE-2 may be one of the most crucial aspects in determining the outcome of vascular angiogenesis. The initial quiescent endothelium goes through cyclical phases of (a) basal quiescent, (b) plastic, (c) angiogenic, and (d) stable maturing stage (Fig. 4) to complete the angiogenic cycle for neovascularization. The transitions are likely to be influenced by tissue milieu, whereby phase changes may occur depending on the presence of specific growth factors and inhibitors.

In the basal quiescent phase, constitutive expression of ANG-1 from perivascular cells couples with minimal expression of ANG-2 and uniform expression of TIE-2 in the endothelial cells would be expected. Together with basal levels

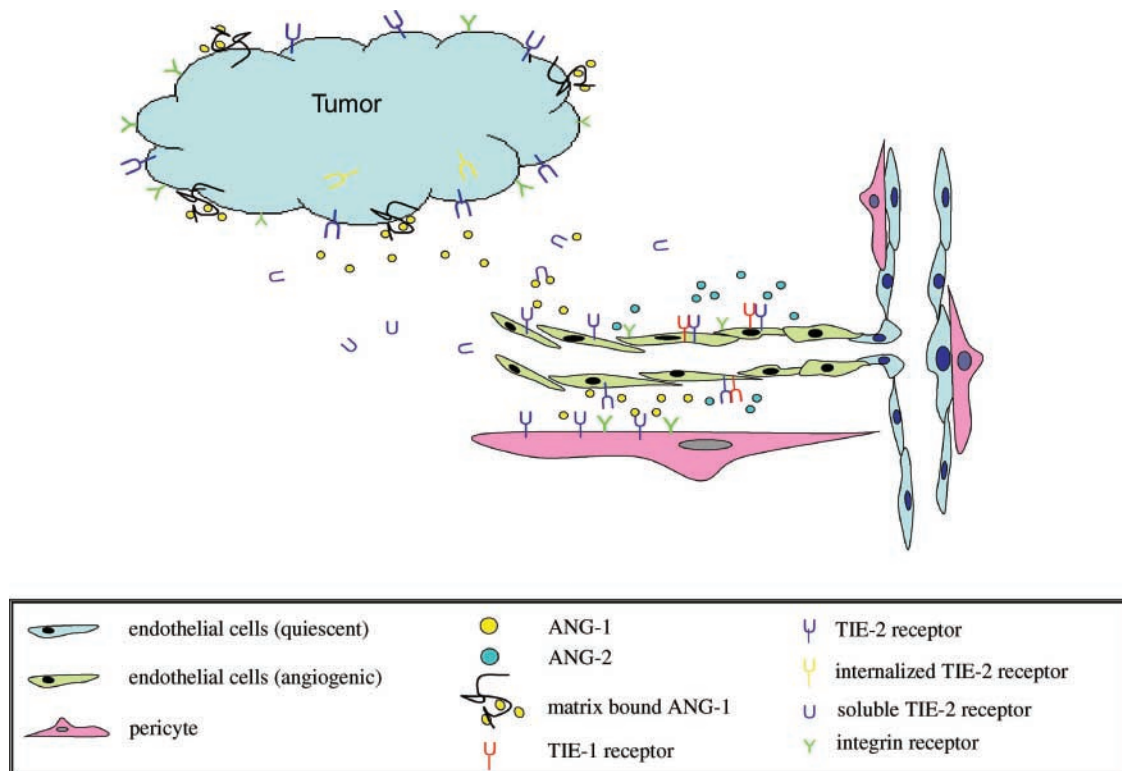
of platelet-derived growth factor and ephrin, this maintains the endothelium in a stable differentiated state by reciprocal interactions between endothelial and perivascular cells. On stimulation, the ratio of ANG-1 to ANG-2 may shift in favor of the latter to promote transient vessel plasticity by dissociation of endothelial cells with perivascular cells. This loosens the tight association between neighboring endothelial cells as well as extracellular matrix during the initial plastic phase. Furthermore, induction of TIE-2 expression during this period may significantly favor increased binding of ANG-2 or increased numbers of unbound, therefore presumably, unphosphorylated TIE-2 receptor, thereby reverting vessels to a more plastic state that are more responsive to angiogenic stimuli.

Up-regulation of ANG-1, ANG-2, and TIE-2 in the active angiogenic phase would promote vessel differentiation by migrating/sprouting and antiapoptotic effect of TIE-2 signaling. There are increasing numbers of tumor cells reported to express TIE-2 receptor (13, 14, 39, 60, 123, 124). However, the significance of TIE-2-expressing tumor cells during this period is largely unknown, but these extraendothelial TIE-2 receptors may serve to sequester the availability of ANG-1 from the vulnerable endothelial cells. Furthermore, it is unclear if the ANG-1-binding integrins on the endothelial and tumor cells may synergize or counteract ANG-1 action through TIE-2 receptor. Nevertheless, the lack of ANG-1 activity in this

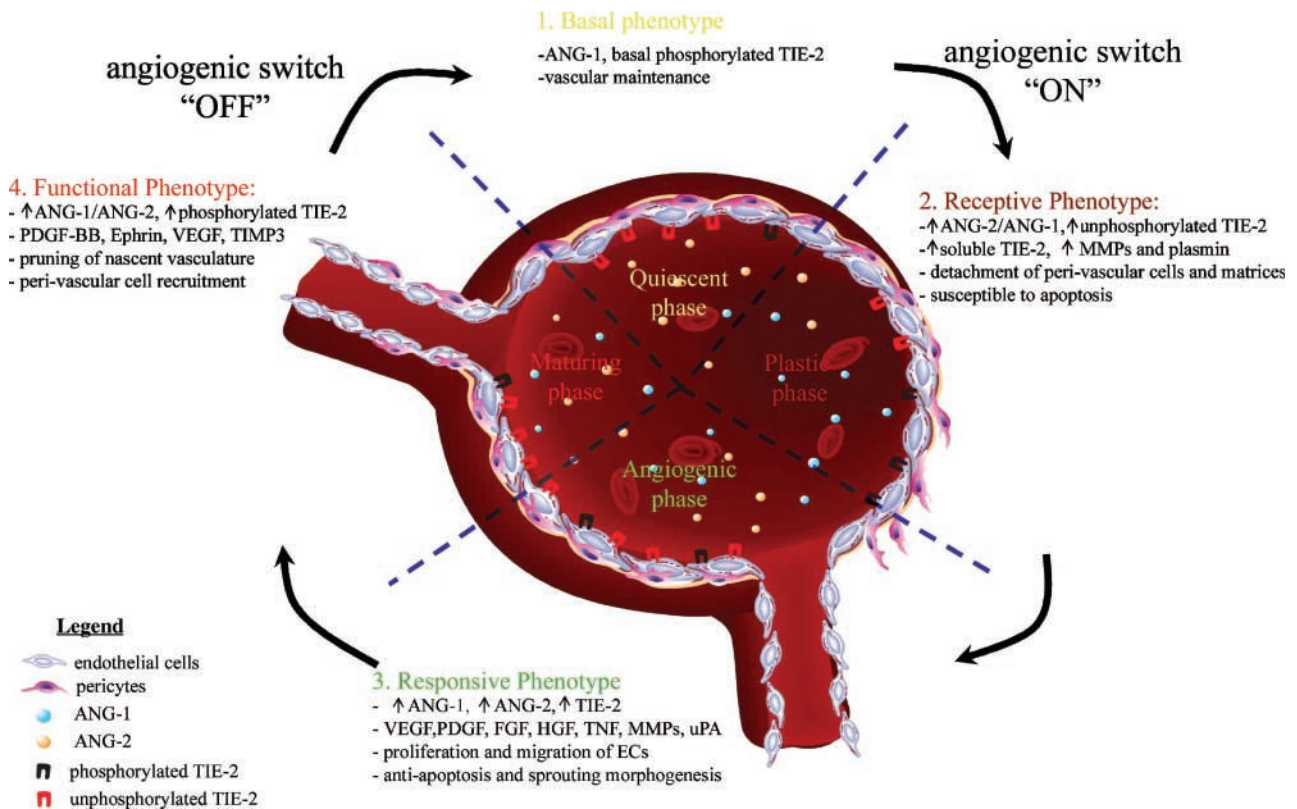
vulnerable state may render the endothelium more susceptible to apoptosis. The loss of this antiapoptotic signal may significantly affect the outcome of tumor angiogenesis. These may be the missing pieces that determine the transition between proangiogenic and antiangiogenic roles of ANG-1 reported in the literature. In the final stage, the cycle reenters quiescent phase after the expression of growth factors returns to basal levels. This tenuous equilibrium between vascular supply and tumor demand may favor a stabilization and maturation of the nascent vasculature. However, this equilibrium is likely to be transient because tumor vasculature is hyperpermeable and lacking in pericyte coverage. This cyclical pathway may explain the governing dynamism in angiogenesis and provide a rational interventional window to strategically target each phase of vascular development in the evolving tumor ecosystem.

### Conclusions

The varied role of TIE-2 signaling pathway in endothelial survival, vessel growth, and vascular maturation may be intrinsic to different types of tumors. However, emerging evidence suggests that additional signaling of ANG-1 and ANG-2 through integrin receptors may be important in their diverse contribution toward tumor growth. The existence of various ligands (ANG-1, ANG-2, and ANG-4), their isoforms



**FIGURE 3.** TIE-2-dependent and TIE-2-independent signaling of ANG-1 and ANG-2 in endothelial and nonendothelial cells. Majority of the ANG-1 is secreted by tumor cells and pericytes, whereas ANG-2 is mainly produced by endothelial cells. The matrix-bound ANG-1 (132) and Weibel-Palade body-stored ANG-2 (133) are likely to act as a readily releasable reservoir in tumor and endothelial cells, respectively. Differential ANG-1 gradient attracts endothelial cell migration toward tumor, whereas ANG-2 maintains a “pericyte-free” endothelium. The internalization of TIE-2 following receptor activation releases the ANG-1 and ANG-2 back into the active pool of ligands (134). Soluble TIE-2 (127) may act as a decoy for ligand binding to regulate the membrane-bound TIE-2, whereas heterodimerization between TIE-1 and TIE-2 may modulate their ligand receptivity (73, 80). Integrin receptors may act as primary and secondary binding partner of the ligands (32, 76, 77).



**FIGURE 4.** Angiogenic cycle of vascular developments. 1, angiogenic switch is triggered by hypoxia, inflammation, or genetic mutations that disrupt the spatiotemporal balance between angiogenic promoters and inhibitors. 2, the up-regulation of ANG-2 in response to the stimuli destabilizes the endothelium by decreasing the phosphorylation status of TIE-2 receptor and resulting in increased vessel plasticity with a more responsive endothelial cells. 3, the activated matrix proteases promote pericyte detachment and facilitate endothelial chemotaxis and sprouting in response to VEGF, ANG-1, fibroblast growth factor (*FGF*), platelet-derived growth factor (*PDGF*), and hepatocyte growth factor (*HGF*). 4, the cyclical supply and demand reaches an equilibrium (even if transiently) after waning effect of the stimuli and tapering ANG-2 expression that enable reinvestment of basement membrane, extracellular matrices, and pericyte recruitment to the nascent vasculature. MMPs, matrix metalloproteinases; TIMP3, tissue inhibitor of metalloproteinase 3; TNF, tumor necrosis factor; uPA, urokinase-type plasminogen activator.

[four ANG-1 isoforms (31) and two ANG-2 isoforms (125)], and soluble forms of TIE-1 and TIE-2 receptors (126, 127) and TIE-2/TIE-1 heterodimerization in modulating receptivity (73) and differential induction of both receptors and ligands (119, 128, 129) compounded the difficulty of precise definition of the role of the ANGs during angiogenesis.

The concept of molecular balance between ANG-1/ANG-2 as a trigger between active angiogenesis and vascular regression is an oversimplification of the inherently complicated process. Together with autocrine or paracrine interactions with its ligands in various cell types, TIE-2 signaling pathway may not be totally restricted to endothelial cells during angiogenesis and the pathway may have wide-ranging functions in other cellular activities. The aberrant vascularization resulted from imbalance between ANG-1 and ANG-2 together with VEGF may explain one aspect of the controversies of TIE-2 signaling in tumor angiogenesis [for detail, see review (130)]. Increasingly, TIE-2-expressing tumor cells and ANG-1-binding integrins may be the emerging puzzles that help in the understanding of the multiple roles of ANG-1 and ANG-2, be it endothelial TIE-2 dependent or independent, in the tumor ecosystem.

It is plausible that an antiangiogenesis approach targeting TIE-2 pathway may be applicable to both endothelial cells as

well as tumor cells. If we could control Dr. Jekyll and Mr. Hyde in the ANGs, a similar strategy targeting the overall tumor ecosystem by controlling the survival pathway of angiogenesis may prove to be more effective in containing malignancy and restricting tumor progression.

## References

- Folkman J. Anti-angiogenesis: new concept for therapy of solid tumors. *Ann Surg* 1972;175:409–16.
- Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature* 2000; 407:249–57.
- Angeletti CA, Lucchi M, Fontanini G, et al. Prognostic significance of tumoral angiogenesis in completely resected late stage lung carcinoma (stage IIIA-N2). Impact of adjuvant therapies in a subset of patients at high risk of recurrence. *Cancer* 1996;78:409–15.
- Tjalma W, Van Marck E, Weyler J, et al. Quantification and prognostic relevance of angiogenic parameters in invasive cervical cancer. *Br J Cancer* 1998; 78:170–4.
- Heimann R, Ferguson D, Gray S, Hellman S. Assessment of intratumoral vascularization (angiogenesis) in breast cancer prognosis. *Breast Cancer Res Treat* 1998;52:147–58.
- Ogawa S, Kaku T, Kobayashi H, et al. Prognostic significance of microvessel density, vascular cuffing and vascular endothelial growth factor expression in ovarian carcinoma: a special review for clear cell adenocarcinoma. *Cancer Lett* 2002;176:111–8.



7. Olewniczak S, Chosia M, Kolodziej B, Kwas A, Kram A, Domagala W. Angiogenesis as determined by computerised image analysis and the risk of early relapse in women with invasive ductal breast carcinoma. *Pol J Pathol* 2003;54:53–9.
8. Chung YC, Hou YC, Chang CN, Hseu TH. Expression and prognostic significance of angiopoietin in colorectal carcinoma. *J Surg Oncol* 2006;94:631–8.
9. Loges S, Clausen H, Reichelt U, et al. Determination of microvessel density by quantitative real-time PCR in esophageal cancer: correlation with histologic methods, angiogenic growth factor expression, and lymph node metastasis. *Clin Cancer Res* 2007;13:76–80.
10. Chen J, De S, Brainard J, Byzova TV. Metastatic properties of prostate cancer cells are controlled by VEGF. *Cell Commun Adhes* 2004;11:1–11.
11. Yigitbasi OG, Younes MN, Doan D, et al. Tumor cell and endothelial cell therapy of oral cancer by dual tyrosine kinase receptor blockade. *Cancer Res* 2004;64:7977–84.
12. Costa C, Soares R, Schmitt F. Angiogenesis: now and then. *APMIS* 2004;112:402–12.
13. Shim WS, Teh M, Bapna A, et al. Angiopoietin 1 promotes tumor angiogenesis and tumor vessel plasticity of human cervical cancer in mice. *Exp Cell Res* 2002;279:299–309.
14. Osada H, Tokunaga T, Hatanaka H, et al. Gene expression of angiogenesis related factors in glioma. *Int J Oncol* 2001;18:305–9.
15. Suri C, Jones PF, Patan S, et al. Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. *Cell* 1996;87:1171–80.
16. Shyu KG, Manor O, Magner M, Yancopoulos GD, Isner JM. Direct intramuscular injection of plasmid DNA encoding angiopoietin-1 but not angiopoietin-2 augments revascularization in the rabbit ischemic hindlimb. *Circulation* 1998;98:2081–7.
17. Chae JK, Kim I, Lim ST, et al. Coadministration of angiopoietin-1 and vascular endothelial growth factor enhances collateral vascularization. *Arterioscler Thromb Vasc Biol* 2000;20:2573–8.
18. Bhardwaj S, Roy H, Karpanen T, et al. Periadventitial angiopoietin-1 gene transfer induces angiogenesis in rabbit carotid arteries. *Gene Ther* 2005;12:388–94.
19. Hayes AJ, Huang WQ, Yu J, et al. Expression and function of angiopoietin-1 in breast cancer. *Br J Cancer* 2000;83:1154–60.
20. Hawighorst T, Skobe M, Streit M, et al. Activation of the tie2 receptor by angiopoietin-1 enhances tumor vessel maturation and impairs squamous cell carcinoma growth. *Am J Pathol* 2002;160:1381–92.
21. Stoeltzing O, Ahmad SA, Liu W, et al. Angiopoietin-1 inhibits vascular permeability, angiogenesis, and growth of hepatic colon cancer tumors. *Cancer Res* 2003;63:3370–7.
22. Kanda S, Miyata Y, Mochizuki Y, Matsuyama T, Kanetake H. Angiopoietin 1 is mitogenic for cultured endothelial cells. *Cancer Res* 2005;65:6820–7.
23. Cho CH, Kim KE, Byun J, et al. Long-term and sustained COMP-Ang1 induces long-lasting vascular enlargement and enhanced blood flow. *Circ Res* 2005;97:86–94.
24. Davis S, Aldrich TH, Jones PF, et al. Isolation of angiopoietin-1, a ligand for the TIE2 receptor, by secretion-trap expression cloning. *Cell* 1996;87:1161–9.
25. Maisonpierre PC, Suri C, Jones PF, et al. Angiopoietin-2, a natural antagonist for Tie2 that disrupts *in vivo* angiogenesis. *Science* 1997;277:55–60.
26. Gale NW, Thurston G, Hackett SF, et al. Angiopoietin-2 is required for postnatal angiogenesis and lymphatic patterning, and only the latter role is rescued by angiopoietin-1. *Dev Cell* 2002;3:411–23.
27. Suri C, McClain J, Thurston G, et al. Increased vascularization in mice overexpressing angiopoietin-1. *Science* 1998;282:468–71.
28. Thurston G, Suri C, Smith K, et al. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science* 1999;286:2511–4.
29. Procopio WN, Pelavin PI, Lee WM, Yeilding NM. Angiopoietin-1 and -2 coiled coil domains mediate distinct homo-oligomerization patterns, but fibrinogen-like domains mediate ligand activity. *J Biol Chem* 1999;274:30196–201.
30. Davis S, Papadopoulos N, Aldrich TH, et al. Angiopoietins have distinct modular domains essential for receptor binding, dimerization and superclustering. *Nat Struct Biol* 2003;10:38–44.
31. Huang YQ, Li JJ, Karparkin S. Identification of a family of alternatively spliced mRNA species of angiopoietin-1. *Blood* 2000;95:1993–9.
32. Carlson TR, Feng Y, Maisonpierre PC, Mrksich M, Morla AO. Direct cell adhesion to the angiopoietins mediated by integrins. *J Biol Chem* 2001;276:26516–25.
33. Hutchings H, Ortega N, Plouet J. Extracellular matrix-bound vascular endothelial growth factor promotes endothelial cell adhesion, migration, and survival through integrin ligation. *FASEB J* 2003;17:1520–2.
34. Stratmann A, Risau W, Plate KH. Cell type-specific expression of angiopoietin-1 and angiopoietin-2 suggests a role in glioblastoma angiogenesis. *Am J Pathol* 1998;153:1459–66.
35. Ding H, Roncari L, Wu X, et al. Expression and hypoxic regulation of angiopoietins in human astrocytomas. *Neuro-oncol* 2001;3:1–10.
36. Takahama M, Tsutsumi M, Tsujiuchi T, et al. Enhanced expression of Tie2, its ligand angiopoietin-1, vascular endothelial growth factor, and CD31 in human non-small cell lung carcinomas. *Clin Cancer Res* 1999;5:2506–10.
37. Nakayama T, Yao L, Tosato G. Mast cell-derived angiopoietin-1 plays a critical role in the growth of plasma cell tumors. *J Clin Invest* 2004;114:1317–25.
38. Martoglio AM, Tom BD, Starkey M, Corps AN, Charnock-Jones DS, Smith SK. Changes in tumorigenesis- and angiogenesis-related gene transcript abundance profiles in ovarian cancer detected by tailored high density cDNA arrays. *Mol Med* 2000;6:750–65.
39. Shirakawa K, Tsuda H, Heike Y, et al. Absence of endothelial cells, central necrosis, and fibrosis are associated with aggressive inflammatory breast cancer. *Cancer Res* 2001;61:445–51.
40. Tangkeangsirisin W, Hayashi J, Serrero G. PC cell-derived growth factor mediates tamoxifen resistance and promotes tumor growth of human breast cancer cells. *Cancer Res* 2004;64:1737–43.
41. Wang J, Wu K, Zhang D, et al. Expressions and clinical significances of angiopoietin-1, -2 and Tie2 in human gastric cancer. *Biochem Biophys Res Commun* 2005;337:386–93.
42. Machein MR, Knedla A, Knoth R, Wagner S, Neuschl E, Plate KH. Angiopoietin-1 promotes tumor angiogenesis in a rat glioma model. *Am J Pathol* 2004;165:1557–70.
43. Zadeh G, Koushan K, Pillo L, Shannon P, Guha A. Role of Ang1 and its interaction with VEGF-A in astrocytomas. *J Neuropathol Exp Neurol* 2004;63:978–89.
44. Udani V, Santarelli J, Yung Y, et al. Differential expression of angiopoietin-1 and angiopoietin-2 may enhance recruitment of bone marrow-derived endothelial precursor cells into brain tumors. *Neurol Res* 2005;27:801–6.
45. Muller A, Lange K, Gaiser T, et al. Expression of angiopoietin-1 and its receptor TEK in hematopoietic cells from patients with myeloid leukemia. *Leuk Res* 2002;26:163–8.
46. Ahmad SA, Liu W, Jung YD, et al. The effects of angiopoietin-1 and -2 on tumor growth and angiogenesis in human colon cancer. *Cancer Res* 2001;61:1255–9.
47. Yu Q, Stamenkovic I. Angiopoietin-2 is implicated in the regulation of tumor angiogenesis. *Am J Pathol* 2001;158:563–70.
48. Witzenbichler B, Maisonpierre PC, Jones P, Yancopoulos GD, Isner JM. Chemotactic properties of angiopoietin-1 and -2, ligands for the endothelial-specific receptor tyrosine kinase Tie2. *J Biol Chem* 1998;273:18514–21.
49. Kim I, Kim JH, Moon SO, Kwak HJ, Kim NG, Koh GY. Angiopoietin-2 at high concentration can enhance endothelial cell survival through the phosphatidylinositol 3'-kinase/Akt signal transduction pathway. *Oncogene* 2000;19:4549–52.
50. Teichert-Kuliszewska K, Maisonpierre PC, Jones N, et al. Biological action of angiopoietin-2 in a fibrin matrix model of angiogenesis is associated with activation of Tie2. *Cardiovasc Res* 2001;49:659–70.
51. Conklin LD, McAninch RE, Schulz D, et al. HIV-based vectors and angiogenesis following rabbit hindlimb ischemia. *J Surg Res* 2005;123:55–66.
52. Ley CD, Olsen MW, Lund EL, Kristjansen PE. Angiogenic synergy of bFGF and VEGF is antagonized by angiopoietin-2 in a modified *in vivo* Matrigel assay. *Microvasc Res* 2004;68:161–8.
53. Tanaka S, Mori M, Sakamoto Y, Makuuchi M, Sugimachi K, Wands JR. Biologic significance of angiopoietin-2 expression in human hepatocellular carcinoma. *J Clin Invest* 1999;103:341–5.
54. Yoshida Y, Oshika Y, Fukushima Y, et al. Expression of angiostatic factors in colorectal cancer. *Int J Oncol* 1999;15:1221–5.
55. Holash J, Maisonpierre PC, Compton D, et al. Vessel cooption, regression, and growth in tumors mediated by angiopoietins and VEGF. *Science* 1999;284:1994–8.
56. Etoh T, Inoue H, Tanaka S, Barnard GF, Kitano S, Mori M. Angiopoietin-2 is related to tumor angiogenesis in gastric carcinoma: possible *in vivo* regulation via induction of proteases. *Cancer Res* 2001;61:2145–53.
57. Eggert A, Ikegaki N, Kwiatkowski J, Zhao H, Brodeur GM, Himelstein BP. High-level expression of angiogenic factors is associated with advanced tumor stage in human neuroblastomas. *Clin Cancer Res* 2000;6:1900–8.
58. Caine GJ, Blann AD, Stonelake PS, Ryan P, Lip GY. Plasma angiopoietin-1, angiopoietin-2 and Tie-2 in breast and prostate cancer: a comparison with VEGF and Flt-1. *Eur J Clin Invest* 2003;33:883–90.

59. Torimura T, Ueno T, Kin M, et al. Overexpression of angiotensin-converting enzyme-2 in hepatocellular carcinoma. *J Hepatol* 2004;40:799–807.
60. Hatanaka H, Abe Y, Naruke M, et al. Significant correlation between interleukin 10 expression and vascularization through angiotensin-converting enzyme-2/TIE2 networks in non-small cell lung cancer. *Clin Cancer Res* 2001;7:1287–92.
61. Tanaka F, Ishikawa S, Yanagihara K, et al. Expression of angiotensin-converting enzyme-2 and its clinical significance in non-small cell lung cancer. *Cancer Res* 2002;62:7124–9.
62. Nakayama T, Yoshizaki A, Kawahara N, et al. Expression of Tie-1 and 2 receptors, and angiotensin-converting enzyme-2 and 4 in gastric carcinoma: immunohistochemical analyses and correlation with clinicopathological factors. *Histopathology* 2004;44:232–9.
63. Valenzuela DM, Griffiths JA, Rojas J, et al. Angiotensin-converting enzyme 3 and 4: diverging gene counterparts in mice and humans. *Proc Natl Acad Sci U S A* 1999;96:1904–9.
64. Xu Y, Liu YJ, Yu Q. Angiotensin-converting enzyme-3 inhibits pulmonary metastasis by inhibiting tumor angiogenesis. *Cancer Res* 2004;64:6119–26.
65. Lee HJ, Cho CH, Hwang SJ, et al. Biological characterization of angiotensin-converting enzyme-3 and angiotensin-converting enzyme-4. *FASEB J* 2004;18:1200–8.
66. Enholm B, Paavonen K, Ristimäki A, et al. Comparison of VEGF, VEGF-B, VEGF-C and Ang-1 mRNA regulation by serum, growth factors, oncoproteins and hypoxia. *Oncogene* 1997;14:2475–83.
67. Ristimäki A, Narko K, Enholm B, Joukov V, Alitalo K. Proinflammatory cytokines regulate expression of the lymphatic endothelial mitogen vascular endothelial growth factor-C. *J Biol Chem* 1998;273:8413–8.
68. Oh H, Takagi H, Suzuma K, Otani A, Matsumura M, Honda Y. Hypoxia and vascular endothelial growth factor selectively up-regulate angiotensin-converting enzyme-2 in bovine microvascular endothelial cells. *J Biol Chem* 1999;274:15732–9.
69. Mandriota SJ, Pepper MS. Regulation of angiotensin-converting enzyme-2 mRNA levels in bovine microvascular endothelial cells by cytokines and hypoxia. *Circ Res* 1998;83:852–9.
70. Scharpfenecker M, Fiedler U, Reiss Y, Augustin HG. The Tie-2 ligand Angiotensin-converting enzyme-2 destabilizes quiescent endothelium through an internal autocrine loop mechanism. *J Cell Sci* 2005;118 (Pt 4):771–80.
71. Niu XL, Peters KG, Kontos CD. Deletion of the carboxyl terminus of Tie2 enhances kinase activity, signaling, and function. Evidence for an autoinhibitory mechanism. *J Biol Chem* 2002;277:31768–73.
72. Shewchuk LM, Hassell AM, Ellis B, et al. Structure of the Tie2 RTK domain: self-inhibition by the nucleotide binding loop, activation loop, and C-terminal tail. *Structure Fold Des* 2000;8:1105–13.
73. Kim KL, Shin IS, Kim JM, et al. Interaction between Tie receptors modulates angiogenic activity of angiotensin-converting enzyme-2 in endothelial progenitor cells. *Cardiovasc Res* 2006;72:394–402.
74. Eklund L, Olsen BR. Tie receptors and their angiotensin-converting enzyme-2 ligands are context-dependent regulators of vascular remodeling. *Exp Cell Res* 2006;312:630–41.
75. Dunk C, Shams M, Nijjar S, et al. Angiotensin-converting enzyme-1 and angiotensin-converting enzyme-2 activate trophoblast Tie-2 to promote growth and migration during placental development. *Am J Pathol* 2000;156:2185–99.
76. Dallabrida SM, Ismail N, Oberle JR, Himes BE, Rupnick MA. Angiotensin-converting enzyme-2 promotes cardiac and skeletal myocyte survival through integrins. *Circ Res* 2005;96:e8–24.
77. Ward NL, Putoczki T, Mearow K, Ivanco TL, Dumont DJ. Vascular-specific growth factor angiotensin-converting enzyme-1 is involved in the organization of neuronal processes. *J Comp Neurol* 2005;482:244–56.
78. Byzova TV, Goldman CK, Pampori N, et al. A mechanism for modulation of cellular responses to VEGF: activation of the integrins. *Mol Cell* 2000;6:851–60.
79. Serini G, Valdembrì D, Bussolino F. Integrins and angiogenesis: a sticky business. *Exp Cell Res* 2006;312:651–8.
80. Saharinen P, Kerkela K, Ekman N, et al. Multiple angiotensin-converting enzyme-2 recombinant proteins activate the Tie1 receptor tyrosine kinase and promote its interaction with Tie2. *J Cell Biol* 2005;169:239–43.
81. Carmeliet P. Basic concepts of (myocardial) angiogenesis: role of vascular endothelial growth factor and angiotensin-converting enzyme-2. *Curr Interv Cardiol Rep* 1999;1:322–35.
82. Graff BA, Bjornæs I, Rofstad EK. Microvascular permeability of human melanoma xenografts to macromolecules: relationships to tumor volumetric growth rate, tumor angiogenesis, and VEGF expression. *Microvasc Res* 2001;61:187–98.
83. Ozawa CR, Banfi A, Glazer NL, et al. Microenvironmental VEGF concentration, not total dose, determines a threshold between normal and aberrant angiogenesis. *J Clin Invest* 2004;113:516–27.
84. Audero E, Cascone I, Zanon I, et al. Expression of angiotensin-converting enzyme-1 in human glioblastomas regulates tumor-induced angiogenesis: *in vivo* and *in vitro* studies. *Arterioscler Thromb Vasc Biol* 2001;21:536–41.
85. Korff T, Kimmina S, Martiny-Baron G, Augustin HG. Blood vessel maturation in a 3-dimensional spheroidal coculture model: direct contact with smooth muscle cells regulates endothelial cell quiescence and abrogates VEGF responsiveness. *FASEB J* 2001;15:447–57.
86. Zagzag D, Amirmovin R, Greco MA, et al. Vascular apoptosis and involution in gliomas precede neovascularization: a novel concept for glioma growth and angiogenesis. *Lab Invest* 2000;80:837–49.
87. Benjamin LE, Hemo I, Keshet E. A plasticity window for blood vessel remodeling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development* 1998;125:1591–8.
88. Grosskreutz CL, Anand-Apte B, Duplax C, et al. Vascular endothelial growth factor-induced migration of vascular smooth muscle cells *in vitro*. *Microvasc Res* 1999;58:128–36.
89. Du L, Sullivan CC, Chu D, et al. Signaling molecules in nonfamilial pulmonary hypertension. *N Engl J Med* 2003;348:500–9.
90. Zhao YD, Campbell AI, Robb M, Ng D, Stewart DJ. Protective role of angiotensin-converting enzyme-1 in experimental pulmonary hypertension. *Circ Res* 2003;92:984–91.
91. Hayes AJ, Huang WQ, Mallah J, Yang D, Lippman ME, Li LY. Angiotensin-converting enzyme-1 and its receptor Tie-2 participate in the regulation of capillary-like tubule formation and survival of endothelial cells. *Microvasc Res* 1999;58:224–37.
92. Kwak HJ, So JN, Lee SJ, Kim I, Koh GY. Angiotensin-converting enzyme-1 is an apoptosis survival factor for endothelial cells. *FEBS Lett* 1999;448:249–53.
93. Papapetropoulos A, Garcia-Cardena G, Dengler TJ, Maisonpierre PC, Yancopoulos GD, Sessa WC. Direct actions of angiotensin-converting enzyme-1 on human endothelium: evidence for network stabilization, cell survival, and interaction with other angiogenic growth factors. *Lab Invest* 1999;79:213–23.
94. Peirce SM, Price RJ, Skalak TC. Spatial and temporal control of angiogenesis and arterIALIZATION using focal applications of VEGF164 and Ang-1. *Am J Physiol Heart Circ Physiol* 2004;286:H918–25.
95. Yamauchi A, Ito Y, Morikawa M, et al. Pre-administration of angiotensin-converting enzyme-1 followed by VEGF induces functional and mature vascular formation in a rabbit ischemic model. *J Gene Med* 2003;5:994–1004.
96. Baffert F, Thurston G, Rochon-Duck M, Le T, Brekken R, McDonald DM. Age-related changes in vascular endothelial growth factor dependency and angiotensin-converting enzyme-1-induced plasticity of adult blood vessels. *Circ Res* 2004;94:984–92.
97. Papapetropoulos A, Fulton D, Mahboubi K, et al. Angiotensin-converting enzyme-1 inhibits endothelial cell apoptosis via the Akt/survivin pathway. *J Biol Chem* 2000;275:9102–5.
98. Dimmeler S, Zeiher AM. Akt takes center stage in angiogenesis signaling. *Circ Res* 2000;86:4–5.
99. Harfouche R, Hassessian HM, Guo Y, et al. Mechanisms which mediate the antiapoptotic effects of angiotensin-converting enzyme-1 on endothelial cells. *Microvasc Res* 2002;64:135–47.
100. Lund EL, Bastholm L, Kristjansen PE. Therapeutic synergy of TNP-470 and ionizing radiation: effects on tumor growth, vessel morphology, and angiogenesis in human glioblastoma multiforme xenografts. *Clin Cancer Res* 2000;6:971–8.
101. Kim I, Moon SO, Han CY, et al. The angiotensin-converting enzyme-2 system in coronary artery endothelium prevents oxidized low-density lipoprotein-induced apoptosis. *Cardiovasc Res* 2001;49:872–81.
102. Jones N, Iljin K, Dumont DJ, Alitalo K. Tie receptors: new modulators of angiogenic and lymphangiogenic responses. *Nat Rev Mol Cell Biol* 2001;2:257–67.
103. Jones N, Voskas D, Master Z, Sarao R, Jones J, Dumont DJ. Rescue of the early vascular defects in Tek/Tie2 null mice reveals an essential survival function. *EMBO Rep* 2001;2:438–45.
104. Puri MC, Partanen J, Rossant J, Bernstein A. Interaction of the TEK and TIE receptor tyrosine kinases during cardiovascular development. *Development* 1999;126:4569–80.
105. Kuroda K, Sapadin A, Shoji T, Fleischmajer R, Lebwohl M. Altered expression of angiotensin-converting enzyme-2 and Tie2 endothelial receptor in psoriasis. *J Invest Dermatol* 2001;116:713–20.
106. Vikkula M, Boon LM, Carraway KL 3rd, et al. Vascular dysmorphogenesis caused by an activating mutation in the receptor tyrosine kinase TIE2. *Cell* 1996;87:1181–90.
107. Hoffmann J, Feng Y, Vom Hagen F, et al. Endothelial survival factors and spatial completion, but not pericyte coverage of retinal capillaries, determine vessel plasticity. *FASEB J* 2005;19:2035–6.

108. Uemura A, Ogawa M, Hirashima M, et al. Recombinant angiopoietin-1 restores higher-order architecture of growing blood vessels in mice in the absence of mural cells. *J Clin Invest* 2002;110:1619–28.
109. Kim I, Moon SO, Park SK, Chae SW, Koh GY. Angiopoietin-1 reduces VEGF-stimulated leukocyte adhesion to endothelial cells by reducing ICAM-1, VCAM-1, and E-selectin expression. *Circ Res* 2001;89:477–9.
110. Hughes DP, Marron MB, Brindle NP. The antiinflammatory endothelial tyrosine kinase Tie2 interacts with a novel nuclear factor- $\kappa$ B inhibitor ABIN-2. *Circ Res* 2003;92:630–6.
111. Ishida S, Usui T, Yamashiro K, et al. VEGF164-mediated inflammation is required for pathological, but not physiological, ischemia-induced retinal neovascularization. *J Exp Med* 2003;198:483–9.
112. Stratmann A, Acker T, Burger AM, Amann K, Risau W, Plate KH. Differential inhibition of tumor angiogenesis by tie2 and vascular endothelial growth factor receptor-2 dominant-negative receptor mutants. *Int J Cancer* 2001;91:273–82.
113. Kelly BD, Hackett SF, Hirota K, et al. Cell type-specific regulation of angiogenic growth factor gene expression and induction of angiogenesis in nonischemic tissue by a constitutively active form of hypoxia-inducible factor 1. *Circ Res* 2003;93:1074–81.
114. Tschedschilsuren G, Aust G, Nieber K, Schilling N, Spänzel-Borowski K. Microvascular endothelial cells differ in basal and hypoxia-regulated expression of angiogenic factors and their receptors. *Microvasc Res* 2002;63:243–51.
115. Boon LM, Brouillard P, Irrthum A, et al. A gene for inherited cutaneous venous anomalies (“glomangiomas”) localizes to chromosome 1p21-22. *Am J Hum Genet* 1999;65:125–33.
116. Gale NW, Baluk P, Pan L, et al. Ephrin-B2 selectively marks arterial vessels and neovascularization sites in the adult, with expression in both endothelial and smooth-muscle cells. *Dev Biol* 2001;230:151–60.
117. Armulik A, Abramsson A, Betsholtz C. Endothelial/pericyte interactions. *Circ Res* 2005;97:512–23.
118. Tian H, McKnight SL, Russell DW. Endothelial PAS domain protein 1 (EPAS1), a transcription factor selectively expressed in endothelial cells. *Genes Dev* 1997;11:72–82.
119. Willam C, Koehne P, Jurgensen JS, et al. Tie2 receptor expression is stimulated by hypoxia and proinflammatory cytokines in human endothelial cells. *Circ Res* 2000;87:370–7.
120. Kim I, Kim JH, Ryu YS, Liu M, Koh GY. Tumor necrosis factor- $\alpha$  upregulates angiopoietin-2 in human umbilical vein endothelial cells. *Biochem Biophys Res Commun* 2000;269:361–5.
121. Ray PS, Sasaki H, Estrada-Hernandez T, Zu L, Maulik N. Effects of hypoxia/reoxygenation on angiogenic factors and their tyrosine kinase receptors in the rat myocardium. *Antioxid Redox Signal* 2001;3:89–102.
122. DeBusk LM, Chen Y, Nishishita T, Chen J, Thomas JW, Lin PC. Tie2 receptor tyrosine kinase, a major mediator of tumor necrosis factor  $\alpha$ -induced angiogenesis in rheumatoid arthritis. *Arthritis Rheum* 2003;48:2461–71.
123. Mitsutake N, Namba H, Takahara K, et al. Tie-2 and angiopoietin-1 expression in human thyroid tumors. *Thyroid* 2002;12:95–9.
124. Wurmbach JH, Hammerer P, Sevinc S, Huland H, Ergun S. The expression of angiopoietins and their receptor Tie-2 in human prostate carcinoma. *Anticancer Res* 2000;20:5217–20.
125. Kim I, Kim JH, Ryu YS, Jung SH, Nah JJ, Koh GY. Characterization and expression of a novel alternatively spliced human angiopoietin-2. *J Biol Chem* 2000;275:18550–6.
126. McCarthy MJ, Burrows R, Bell SC, Christie G, Bell PR, Brindle NP. Potential roles of metalloprotease mediated ectodomain cleavage in signaling by the endothelial receptor tyrosine kinase Tie-1. *Lab Invest* 1999;79:889–95.
127. Quartarone E, Alonci A, Allegra A, et al. Differential levels of soluble angiopoietin-2 and Tie-2 in patients with hematological malignancies. *Eur J Haematol* 2006;77:480–5.
128. Hangai M, Moon YS, Kitaya N, et al. Systemically expressed soluble Tie2 inhibits intraocular neovascularization. *Hum Gene Ther* 2001;12:1311–21.
129. Abdulmalek K, Ashur F, Ezer N, Ye F, Magder S, Hussain SN. Differential expression of Tie-2 receptors and angiopoietins in response to *in vivo* hypoxia in rats. *Am J Physiol Lung Cell Mol Physiol* 2001;281:L582–90.
130. Tait CR, Jones PF. Angiopoietins in tumours: the angiogenic switch. *J Pathol* 2004;204:1–10.
131. Cascone I, Napione L, Maniero F, Serini G, Bussolino F. Stable interaction between  $\alpha_5\beta_1$  integrin and Tie2 tyrosine kinase receptor regulates endothelial cell response to Ang-1. *J Cell Biol* 2005;170:993–1004.
132. Xu Y, Yu Q. Angiopoietin-1, unlike angiopoietin-2, is incorporated into the extracellular matrix via its linker peptide region. *J Biol Chem* 2001;276:34990–8.
133. Fiedler U, Scharpfenecker M, Koidl S, et al. The Tie-2 ligand angiopoietin-2 is stored in and rapidly released upon stimulation from endothelial cell Weibel-Palade bodies. *Blood* 2004;103:4150–6.
134. Bogdanovic E, Nguyen VP, Dumont DJ. Activation of Tie2 by angiopoietin-1 and angiopoietin-2 results in their release and receptor internalization. *J Cell Sci* 2006;119 (Pt 17):3551–60.
135. Zadeh G, Reti R, Koushan K, Baoping Q, Shannon P, Guha A. Regulation of the pathological vasculature of malignant astrocytomas by angiopoietin-1. *Neoplasia* 2005;7:1081–90.
136. Stoeltzing O, Ahmad SA, Liu W, et al. Angiopoietin-1 inhibits tumour growth and ascites formation in a murine model of peritoneal carcinomatosis. *Br J Cancer* 2002;87:1182–7.
137. Shim WS, Teh M, Mack PO, Ge R. Inhibition of angiopoietin-1 expression in tumor cells by an antisense RNA approach inhibited xenograft tumor growth in immunodeficient mice. *Int J Cancer* 2001;94:6–15.
138. Wang J, Wu KC, Zhang DX, Fan DM. Antisense angiopoietin-1 inhibits tumorigenesis and angiogenesis of gastric cancer. *World J Gastroenterol* 2006;12:2450–4.
139. Hu B, Guo P, Fang Q, et al. Angiopoietin-2 induces human glioma invasion through the activation of matrix metalloprotease-2. *Proc Natl Acad Sci U S A* 2003;100:8904–9.
140. Lee OH, Fueyo J, Xu J, et al. Sustained angiopoietin-2 expression disrupts vessel formation and inhibits glioma growth. *Neoplasia* 2006;8:419–28.
141. Kunz P, Hoffend J, Altmann A, et al. Angiopoietin-2 overexpression in murine hepatoma results in increased tumor perfusion and induction of critical angiogenesis-promoting genes. *J Nucl Med* 2006;47:1515–24.
142. Lee OH, Xu J, Fueyo J, et al. Expression of the receptor tyrosine kinase Tie2 in neoplastic glial cells is associated with integrin  $\beta_1$ -dependent adhesion to the extracellular matrix. *Mol Cancer Res* 2006;4:915–26.
143. Kuroda H, Ohtsuru A, Futakuchi M, et al. Distinctive gene expression of receptor-type tyrosine kinase families during rat hepatocarcinogenesis. *Int J Mol Med* 2002;9:473–80.
144. Kosacka J, Figiel M, Engele J, Hilbig H, Majewski M, Spänzel-Borowski K. Angiopoietin-1 promotes neurite outgrowth from dorsal root ganglion cells positive for Tie-2 receptor. *Cell Tissue Res* 2005;320:11–9.
145. De Palma M, Venneri MA, Galli R, et al. Tie2 identifies a hematopoietic lineage of proangiogenic monocytes required for tumor vessel formation and a mesenchymal population of pericyte progenitors. *Cancer Cell* 2005;8:211–26.
146. Valable S, Bellail A, Lesne S, et al. Angiopoietin-1-induced PI3-kinase activation prevents neuronal apoptosis. *FASEB J* 2003;17:443–5.
147. Nakashima M, Uchida T, Tsukazaki T, et al. Expression of tyrosine kinase receptors Tie-1 and Tie-2 in giant cell tumor of the tendon sheath: a possible role in synovial proliferation. *Pathol Res Pract* 2001;197:101–7.
148. Uchida T, Nakashima M, Hirota Y, Miyazaki Y, Tsukazaki T, Shindo H. Immunohistochemical localisation of protein tyrosine kinase receptors Tie-1 and Tie-2 in synovial tissue of rheumatoid arthritis: correlation with angiogenesis and synovial proliferation. *Ann Rheum Dis* 2000;59:607–14.
149. Shahrara S, Volin MV, Connors MA, Haines GK, Koch AE. Differential expression of the angiogenic Tie receptor family in arthritic and normal synovial tissue. *Arthritis Res* 2002;4:201–8.
150. Otani A, Takagi H, Oh H, Koyama S, Matsumura M, Honda Y. Expressions of angiopoietins and Tie2 in human choroidal neovascular membranes. *Invest Ophthalmol Vis Sci* 1999;40:1912–20.
151. Poncet S, Gasc JM, Janzer RC, Meyer S, Juillerat-Jeanneret L. Expression of Tie-2 in human peripheral and autonomic nervous system. *Neuropathol Appl Neurobiol* 2003;29:361–9.
152. Hewett P, Nijjar S, Shams M, Morgan S, Gupta J, Ahmed A. Down-regulation of angiopoietin-1 expression in menorrhagia. *Am J Pathol* 2002;160:773–80.
153. Wulff C, Wiegand SJ, Saunders PT, Scobie GA, Fraser HM. Angiogenesis during follicular development in the primate and its inhibition by treatment with truncated Flt-1-Fc (vascular endothelial growth factor Trap(A40)). *Endocrinology* 2001;142:3244–54.
154. Yuan HT, Yang SP, Woolf AS. Hypoxia up-regulates angiopoietin-2, a Tie-2 ligand, in mouse mesangial cells. *Kidney Int* 2000;58:1912–9.
155. Lewinson D, Maor G, Rozen N, Rabinovich I, Stahl S, Rachmiel A. Expression of vascular antigens by bone cells during bone regeneration in a membranous bone distraction system. *Histochem Cell Biol* 2001;116:381–8.

# Molecular Cancer Research

## Angiopoietin: A TIE(d) Balance in Tumor Angiogenesis

Winston S.N. Shim, Ivy A.W. Ho and Philip E.H. Wong

*Mol Cancer Res* 2007;5:655-665.

**Updated version** Access the most recent version of this article at:  
<http://mcr.aacrjournals.org/content/5/7/655>

**Cited articles** This article cites 155 articles, 67 of which you can access for free at:  
<http://mcr.aacrjournals.org/content/5/7/655.full.html#ref-list-1>

**Citing articles** This article has been cited by 14 HighWire-hosted articles. Access the articles at:  
<http://mcr.aacrjournals.org/content/5/7/655.full.html#related-urls>

**E-mail alerts** [Sign up to receive free email-alerts](#) related to this article or journal.

**Reprints and Subscriptions** To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at [pubs@aacr.org](mailto:pubs@aacr.org).

**Permissions** To request permission to re-use all or part of this article, contact the AACR Publications Department at [permissions@aacr.org](mailto:permissions@aacr.org).