Volume 4 • Issue 7 • November 2001

Everglades Phosphorus Biogeochemistry Land-Use Effects on Amazonian Soil Amazonian Regrowth Forest Biomass South American Ecosystem Functional Types



ISSN 1432 9840 ECOSFJ 4:603-708 (2001)

Springer

Phosphorus Biogeochemistry and the Impact of Phosphorus Enrichment: Why Is the Everglades so Unique?

Gregory B. Noe,¹ Daniel L. Childers,^{1,2} and Ronald D. Jones^{1,2}*

¹Southeast Environmental Research Center, Florida International University, Miami, Florida 33199, USA; and ²Department of Biological Sciences, Florida International University, Miami, Florida 33199, USA

Abstract

The Florida Everglades is extremely oligotrophic and sensitive to small increases in phosphorus (P) concentrations. P enrichment is one of the dominant anthropogenic impacts on the ecosystem and is therefore a main focus of restoration efforts. In this review, we synthesize research on P biogeochemistry and the impact of P enrichment on ecosystem structure and function in the Florida Everglades. There are clear patterns of increased P concentrations and altered structure and processes along nutrient-enrichment gradients in the water, periphyton, soils, macrophytes, and consumers. Periphyton, an assemblage of algae, bacteria, and associated microfauna, is abundant and has a large influence on phosphorus cycling in the Everglades. The oligotrophic Everglades is P-starved, has lower

INTRODUCTION

The Florida Everglades is one of the largest and most unique wetlands in the world. It is also highlyimpacted by hydrologic modification, drainage, exotic species invasions, and nutrient enrichment (Davis and Ogden 1994). In particular, changes in its ecosystem structure and function are a welldocumented result of phosphorus (P) enrichment due to agricultural and urban runoff (reviewed by

Received 23 August 2000; Accepted 23 March 2001.

P concentrations and higher nitrogen-phosphorus (N:P) ratios, and has oxidized to only slightly reduced soil profiles compared to other freshwater wetland ecosystems. Possible general causes and indications of P limitation in the Everglades and other wetlands include geology, hydrology, and dominance of oxidative microbial nutrient cycling. The Everglades may be unique with respect to P biogeochemistry because of the multiple causes of P limitation and the resulting high degree of limitation.

Key words: Everglades; phosphorus; biogeochemistry; wetlands; ecosystem; nutrient cycling; oligotrophy; eutrophy.

Davis 1994). The effects of P enrichment are noteworthy because the components of the Everglades ecosystem appear to be highly responsive to small changes in P concentrations. In this review, we discuss P biogeochemistry in the Everglades and compare less impacted, oligotrophic areas with those that have undergone increased P loading. We will also attempt to identify functional analogies between the Everglades and other wetland ecosystems that are highly limited by P.

Among the dominant characteristics of the Everglades are habitat heterogeneity, large spatial extent, and a distinctive hydrologic regime (McCally

^{*}Corresponding author; e-mail: serc@fiu.edu

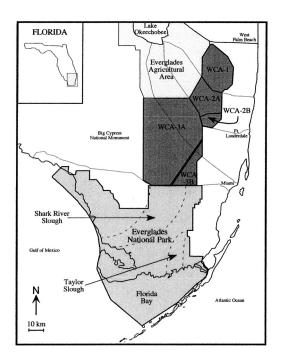


Figure 1. Map of the current Everglades and south Florida.

1999). We will demonstrate that oligotrophy is also characteristic of the Everglades. For the purpose of this review, we limit our definition of the Everglades to the freshwater wetland ecosystem. The predrainage Everglades landscape consisted of a habitat mosaic comprised predominantly of plains and patches of Cladium jamaicense (sawgrass), wet prairies, sloughs, tree islands, marl-forming marshes, and short-hydroperiod peripheral marshes (Davis and others 1994). Prior to efforts to drain the landscape, the freshwater Everglades were hydrologically continuous and covered 1.17 million ha of south Florida (Davis and others 1994). The Everglades ranged from Lake Okeechobee in the north to mangrove forest and Florida Bay in the south, the Gulf of Mexico and cypress forest (including Big Cypress National Monument) in the west, and the Atlantic coastal ridge in the east (Figure 1). Its hydrology was dominated by seasonal patterns of rainfall. The subtropical climate of south Florida is characterized by a dry season (December to May) and a wet season (June to November) with copious amounts of rainfall (mean, 1.4 m y^{-1}) (Duever and others 1994). This resulted in pulsed sheet flow so that water levels in the Northern Everglades were controlled by overflow from Lake Okeechobee and rainfall, whereas in the Southern Everglades hydrology was determined solely by rainfall (Parker 1974). Finally, the Everglades was noted for its

abundance of wildlife, particularly the large numbers of breeding wading birds.

Now, the geographic extent of the Everglades has been reduced, the spatial and temporal patterns of environmental forcing factors (for instance, hydrology and fire) have been altered, and wildlife populations have declined (Davis and Ogden 1994). Construction of an extensive network of nearly 2400 km of canals and dikes was started in the 1880s to drain and compartmentalize the landscape into multiple, discontinuous hydrologic units: the Everglades Agricultural Area (EAA), the Water Conservation Areas (WCAs; WCA-1, -2A, -2B, -3A, and -3B) and Everglades National Park (ENP) (SF-WMD 1999) (Figure 1). Wetlands have been drained for agriculture and urbanization. Today, only 0.62 million ha of the Everglades remain-a roughly 50% loss in spatial extent (Davis and others 1994). Short-hydroperiod wetlands have been preferentially drained, reducing landscape complexity. Animal populations have declined and several species—for example, snail kite (Rostrhamus sociabilis), Florida panther (Felis concolor coryi), wood stork (Mycteria americana), and Cape Sable seaside sparrow (Ammospiza maritima mirabilis)-are now endangered. In addition, the number of nesting wading birds is thought to have decreased 90% since the 1930s (Ogden 1994). Another outcome of ecosystem perturbation has been the successful invasion of many exotic plant and animal species.

Nutrient enrichment, by P in particular, has also had a large impact on the Everglades. Historically, the dominant source of P was rainfall (Davis 1994). Agricultural activities in the EAA (predominantly sugarcane production) and urbanization have increased nutrient concentrations (both nitrogen [N] and P) in waters entering the Everglades. This increase in nutrient loading has been most pronounced in areas that receive water directly from the EAA, such as WCA-2A. Davis (1994) estimated that annual P inputs into the WCAs increased from historic levels of approximately 129 metric tons to current inputs of approximately 376 metric tons as the result of increased P loading from drained agricultural lands. This increase in nutrient loading above natural levels has led to the dramatic expansion of Typha species in areas that receive water from the EAA, Typha species are native to the Everglades but are found only in small stands in oligotrophic areas (Davis and others 1994). Even a small increase in the concentration of total phosphorus (TP) in the water column, from less than 7 to 10–20 μ g L⁻¹, leads to replacement of the extensive cyanobacterial-diatom periphyton mats that characterize the oligotrophic Everglades with filamentous green algae (McCormick and O'Dell 1996; McCormick and Stevenson 1998). We will demonstrate that the oligotrophic Everglades ecosystem is strongly limited by P, as evidenced by the low concentrations of P in the water, soil, periphyton, and macrophytes; highly oxidized soil profiles; adaptations to low P levels; and biotic responses to additions of P.

EVERGLADES RESTORATION

The Everglades Forever Act, passed by the Florida legislature in 1994, requires the development of a numeric water-quality standard for P that will "prevent an imbalance in the natural populations of aquatic flora or fauna . . . and . . . provide a net improvement in the areas already impacted." In addition, the Central and Southern Florida Project Comprehensive Review Study (The Restudy) was authorized by the US Congress in 1992 to study the feasibility of modifying water-control structures and operations to restore the south Florida ecosystem and provide for other water-related needs of the region. The Water Resources Development Act of 2000 authorized the creation of the Comprehensive Everglades Restoration Plan (CERP) as a framework for implementing this hydrologic restoration. The Everglades Forever Act, The Restudy, CERP, and other legislation, regulations, and lawsuits have fostered a new great interest in understanding P biogeochemistry and the impacts of P enrichment on the Everglades. Intrinsic to this understanding is an acknowledgment of the Everglades as a unique wetland ecosystem.

Our Approach

Efforts to protect and restore the Everglades are contingent upon an understanding of why the Everglades are so highly limited by P and how increases in P affect the structure and function of the ecosystem and trigger ecosystem state changes. In this paper, we reviewed the literature on the biogeochemical cycling of P and changes in P cycling in response to enrichment in the Everglades. In addition, we synthesized the published data on P concentrations in the Everglades and identified patterns among ecosystem components and hydrologic units and along nutrient-enrichment gradients. We then utilized this meta-analysis, as well as a summary of the qualitative conclusions from the literature, to develop hypotheses that explain the high degree of P limitation in the oligotrophic Everglades compared to other wetlands. Finally, we developed functional analogies between the Everglades and other P-limited wetlands in an attempt to identify causes and indications of P limitation in wetlands.

Published peer-reviewed literature was surveyed to obtain data on P concentration in water, soils, periphyton, macrophytes, and consumers. Only publications that presented original data and described the methods utilized were included. The South Florida Water Management District (SF-WMD) undertakes an extensive sampling regime and regularly publishes reports on this data. SF-WMD data were not included in this synthesis unless these data were presented in the primary literature. Data from the literature were categorized by (a) hydrologic unit (WCA-1, WCA-2A, WCA-2B, WCA-3A, WCA-3B, ENP-Shark River Slough, and ENP-Taylor Slough) (Figure 1), (b) habitat and position along the nutrient-enrichment gradient (Typha, mixed Typha/Cladium, Cladium, enriched slough, and slough/wet prairie), and (c) ecosystem component (water, soil, periphyton, macrophytes, and fish). Plant community composition is often used as an indicator of nutrient-enrichment gradients in the Everglades. Finally, unpublished studies were included in the nonquantitative review if they reported understudied topics.

Published P concentrations, N:P ratios, and soil P-loading rates were analyzed by meta-analysis. P concentrations and N:P ratios were often estimated from graphs and are reported as ranges if the published values do not include averages. For studies that reported separate data for Typha and Cladium plots in the Typha/Cladium mixed community, data were reported as the range of the monotypic plots. Macrophyte data were weighted by the relative biomass of each species, when reported, in the slough/ wet prairie community. Further details on the interpretation of each publication are listed in the Appendix. In cases where a range of concentrations was reported, the midpoint of the range was used in the meta-analysis. The use of range midpoints is susceptible to skewing by outliers and is most likely in Cladium habitat, where several studies report high soil TP concentrations relative to other Cladium habitat samples within and among studies. These high soil TP plots may be areas of Cladium that have received nutrient-enriched water but have not yet been invaded by Typha.

The matrix of published data was summarized by calculating arithmetic means of nutrient concentrations and geometric means of nutrient ratios and their associated 95% confidence intervals (CI). Published nutrient data in any given hydrologic unit, community, and ecosystem component vary in part due to different analytical techniques and differences in the timing and location of sampling. Consequently, analysis of the published values serves to summarize coarse patterns of P in the Everglades and contrast oligotrophic and enriched areas.

Analysis of variance (ANOVA) was used to test for differences in published nutrient concentration, ratio, and loading data among the four macrophyte communities (*Typha, Typha/Cladium, Cladium,* and slough/wet prairie), where each datum was a single literature value. All ANOVAs were assessed for violations of assumptions with scatterplots; water TP concentration was log-transformed to achieve homoscedasticity and normality of residuals. Differences among habitats for each significant test were assessed with Tukey's HSD multiple comparisons.

GEOLOGY OF THE EVERGLADES

The Everglades began to form about 5000 years BP in a basin underlain by limestone bedrock (Jones 1948; Gleason and Stone 1994). The bedrock is very flat, sloping gently from a high at Lake Okeechobee to a low southward at Florida Bay. The very low relief of the bedrock has variously been estimated to be 2.4 m (Jones 1948), 2.9 m (Light and Dineen 1994), or 4.6 m (Parker 1974) in the 160 km from Lake Okeechobee to Florida Bay. The slope is somewhat exaggerated by deeper soils near Lake Okeechobee. In 1940; the depth of peat ranged from about 2.1 m near the lake to 0.3 m in the Southern Everglades; however, by 1940, significant subsidence of the peat, by roughly 1.5 m, had already occurred in agricultural areas near Lake Okeechobee (Jones 1948). Therefore, the soil surface likely decreased about 6 m in elevation from the shore of Lake Okeechobee to Florida Bay before drainage began. There is very little exposed rock and no terrigenous-clastic sediment input to the Everglades wetlands, although limestone is exposed in some areas of the Southern Everglades. Therefore, P made available from the weathering of mineral rock-the ultimate source of P to ecosystems-is not available to the Everglades.

THE WATER COMPONENT

The hydrology of the Everglades was historically controlled by the subtropical pattern of rainfall, the low relief, and dense vegetation. As a result, historic Everglades hydrology was seasonal in response to rainfall patterns and characterized by a low-velocity sheet flow with a large spatial extent and long periods of inundation (Parker 1974; Fennema and others 1994). In addition, the interaction of surface water with groundwater that had passed through the limestone bedrock resulted in high pH and calcium (Ca) concentrations in surface waters. This interaction with groundwater was greatest in the southern Everglades, where the limestone bedrock is more porous and permeable and is often directly exposed to surface water (Jones 1948). As will be shown later, this carbonate-rich water chemistry has a large impact on the availability of P.

Historically, the primary source of P to the Everglades was from atmospheric deposition rather than the inflowing of surface water. Davis (1994) estimated that wet and dry deposition contributed about 90% of the total P load to the Everglades prior to anthropogenic enrichment. Dry deposition of TP is clearly important to the nutrient budget of the Everglades. However, the estimation of dryfall TP loading is prone to sampling error. In the EAA, 87% of TP deposition was due to dryfall; the remainder came from rainfall, although the rate of TP deposition via dryfall increases near agricultural areas in Florida (Hendry and others 1981). In parts of the Everglades near agriculture, the highest rates of atmospheric dry P loading are likely to occur at the end of the dry season as dust is blown off dry fields that have been fertilized for many years. Calculations of wet deposition depend on the concentration of TP in the rainfall and the amount of rainfall, both of which vary in space and time. The concentration of TP in rainfall in South Florida was estimated to be 10.6 μ g L⁻¹ (Ahn 1999), whereas mean rainfall TP concentrations at meteorological stations within or bordering the Everglades was 7.9 μ g L⁻¹ (data in Ahn 1999). Davis (1994) calculated total (wet and dry) TP deposition to be 0.036 g m^{-2} y⁻¹ using mean annual rainfall rates and an estimate of 29 μ g L⁻¹ in rainfall (pooled wet and dry deposition), which he characterized as an overestimate. Fitz and Sklar (1999) estimated total P deposition to be 0.03 g m⁻² y⁻¹, while Moustafa and others (1996) estimated 0.03 g P m⁻² y⁻¹ in the Kissimmee River basin, approximately 100 km north of the Everglades. Finally, Hendry and others (1981) estimated total P deposition to be 0.017 g $m^{-2} y^{-1}$ at an isolated site in the Florida Keys and $0.096 \text{ g m}^{-2} \text{ y}^{-1}$ near agriculture in the EAA.

Phosphorus in overflow from Lake Okeechobee (which was likely at low concentrations in the past) was probably removed by the custard apple forest (*Annona glabra*) once present at the southern shore of the lake or by the sawgrass plains immediately downstream prior to the draining of wetlands for the EAA and construction of the WCAs. Rudnick and others (1999) reported a high amount of P uptake over a 3-km section of Taylor Slough in the Southern Everglades; TP decreased from a flow-weighted mean of 11.6 to 6.1 μ g L⁻¹. In addition, it

is estimated that more than 90% of P in waters entering the Everglades from the EAA is removed within the WCAs and associated canals before it enters ENP (Rudnick and others 1999).

Today, drainage canals from agricultural fields in the EAA have high concentrations of TP, up to 160 μ g L⁻¹ (Coale and others 1994). Diaz and others (1994) also measured high TP concentrations in EAA canals, 80% of which was SRP. Historically, the annual P load to the entire Everglades from inflowing waters was estimated at 21 metric tons (Davis 1994). In contrast, average annual P load to the WCAs from the EAA was approximately 175 metric tons for the period 1978-91 and 240 metric tons for 1992-96 (Walker 1999). The ENP area has seen smaller changes in total P input (Davis 1994); however, there have been recent increases in P input into areas of ENP near inflow structures. Adjusted for temporal variation in hydrology, TP inputs to Shark River Slough (ENP) increased 5.3% per year and N:P decreased 8.5% per year from 1978 to 1989 (Walker 1991). Due to the implementation of best management practices in the EAA, TP concentrations in the inflow to WCA-2A (SFWMD 2000) and ENP (Walker 1999) decreased in the late 1990s.

Although there is variability in measured TP concentrations in surface water (Table 1), the literature documents a strong trend of increasing P concentration in response to nutrient enrichment (ANOVA: n = 23, P < 0.001, multiple $R^2 = 0.77$). Mean surface-water TP concentrations across the Everglades ranges from 76 to 42 μ g L⁻¹ in *Typha* and Typha/Cladium mix, which is significantly greater than in *Cladium* communities (11 μ g L⁻¹) and unenriched sloughs (10 μ g L⁻¹) (Table 2). The ratio of N:P is an indicator of the relative availability of the two nutrients and can be used cautiously to assess the degree of nutrient limitation by N or P. Phosphorus limitation in freshwater lakes is significantly more frequent than nitrogen limitation when molar N:P ratios are greater than 31, although there is some variation in this threshold among lakes (Downing and McCauley 1992). Molar N:P ratios in surface water change along nutrientenrichment gradients in the Everglades (ANOVA: n = 18, P = 0.001, multiple $R^2 = 0.69$), averaging 94:1, 228:1, 542:1, and 378:1 in Typha, Typha/Cladium mix, Cladium, and slough, respectively (Table 2). In unenriched sloughs of WCA-2A, surface water N:P ratios are above 500:1 (Table 3). In contrast, unenriched sloughs in the southern Everglades typically have N:P ratios of about 275:1 (Table 3). Because TP concentrations are similar in unenriched areas of the northern and southern Everglades (Table 1), the higher N:P ratio in unenriched areas of the northern Everglades must be due to the higher concentrations of N in water from the EAA. The water N:P ratio in the least impacted Everglades is much higher than most oligotrophic lakes (data in Downing and McCauley 1992).

THE PERIPHYTON COMPONENT

The microflora in wetlands are productive and are often a major regulator of nutrient fluxes in freshwater ecosystems (Wetzel 1990). Microbes and algae control the short-term uptake of P in most wetlands (Howard-Williams 1985), although soil and peat accretion determine long-term storage (Richardson and others 1997). Periphyton (a community of algae, bacteria, and microfauna) occurs at the soil surface, attached to macrophytes, and at the water surface. Periphyton standing crop is much greater in the Everglades than in other wetlands (Turner and others 1999); its abundance is visually striking. Everglades periphyton often occurs in association with Utricularia purpurea, a submerged aquatic macrophyte common in wet prairie and slough habitats.

The abundant periphyton of the Everglades reduces water P concentrations and affects P cycling in the water and soil through biotic and abiotic immobilization. Everglades periphyton is efficient at scavenging P from the water column (Scinto 1997; McCormick and others 1998) and has higher uptake rates of P than other ecosystem components (Davis 1982). Unlike macrophytes, the algae and bacteria in periphyton are not restricted to the uptake of dissolved inorganic phosphorus (DIP) but can also utilize dissolved organic phosphorus (DOP) (McCormick and Scinto 1999). In the Everglades, periphyton use DIP and DOP in comparable amounts (Scinto 1997). Scinto (1997) estimated that benthic periphyton alone contributes about 10% of TP accretion to Everglades soils, indicating a role for periphyton in long-term P sequestration. Finally, most P enters the oligotrophic Everglades through atmospheric inputs, and the abundant mats of floating periphyton are likely the primary interceptor of atmospheric inputs of P and therefore the most important regulator of P cycling. The shift to water-column loading of P in enriched areas may have an important impact on the cycling of P.

Calcareous periphyton is characteristic of wet prairies and sloughs in the oligotrophic Everglades (Gleason and Spackman 1974). The precipitation of $CaCO_3$ by periphytic algae can immobilize P and thereby have a large impact on P availability and cycling. In waters with high Ca concentrations, decreased pCO_2 during photosynthetic activity results

		Ecosystem component				
Location	Habitat	Water	Periphyton	Soil	Macrophyte	
EAA	Irrigation canal	100 ¹³ 160 ¹¹				
WCA-1	Typha			1028 ²¹ 1435–2380 ¹⁹		
	Typha/Cladium Cladium			$ 1177^{19} \\ 368^{21} \\ 476-1017^{19} $		
	Slough/wet prairie			413-64219		
WCA-2A	Typha	34 ²⁰ 40–150 ¹⁸ 61 ²⁵ 75 ²³ 101–130 ³²	1900–3390 ⁸ 2500–3750 ¹⁷	$1100 -> 1300^{32}$ $1108 - 1781^{28}$ 1248^{6} 1305^{25} 1338^{12} $1401 - 1611^{7}$ 1474^{23}	1428 ²⁵ 1500 ² 1600 ⁴	
				1475-205919		
				1900 ⁴		
	Typha/Cladium	11 ²⁵ 13–163 ¹⁰ 20–30 ¹⁸ 28–52 ³² 30–65 ²³	500–1750 ¹⁷	500-1100 ³² 529-1022 ²³ 692-1538 ²⁸ 720 ²⁵ 802 ¹² 856-1793 ¹⁹ 924-1292 ⁷ 1350 ⁴	350–550 ²⁹ 500–900 ⁴ 670–680 ²⁵	
	Cladium	5 ²⁵	<10017	$271-547^{23}$	150 ⁴	
		7-20 ²³ 8-11 ¹⁸ 11 ²² 15 ³²	<100-640 ⁸ 132-385 ²²	303^{22} $315-786^{28}$ 432^{6} 441^{25} 473^{12} $495-1201^{7}$ $<500^{32}$ $507-1645^{19}$ 600^{4}	150 ²⁹ 165–197 ²² 200 ² 243 ²⁵	
	Enriched Slough/wet prairie			1496–1538 ²⁸		
	Slough/wet prairie	8 ²⁰ 8–11 ¹⁸ 9 ²⁶ 15 ³²	$30-454^{16} < 100^{17} < 100-640^{8} 200^{26} 385^{22}$	160–435 ²⁶ <500 ³² 530 ²⁸ 586–837 ¹⁹	300-850 ²⁶	
WCA-2B	Typha					
	Typha/Cladium Cladium		67114	321 ¹⁵ 317 ¹⁵	235 ¹⁵ 205 ¹⁵	
	Chuthin			546 ⁶	209	
WCA-3A	Slough/wet prairie Typha Typha/Cladium		160 ¹⁴	220 ¹⁵ 1214 ¹⁹	218 ¹⁵	
	Cladium			351–553 ¹⁹ 548–764 ⁶		
WCA-3B	Slough/wet prairie <i>Typha</i> <i>Typha/Cladium</i>			349–630 ¹⁹		
	<i>Cladium</i> Slough/wet prairie			174–459 ¹	224 ¹	

Table 1. Total Phosphorus Concentration Data for the Everglades

Location	Habitat	Ecosystem component				
		Water	Periphyton	Soil	Macrophyte	
ENP-SRS	Typha			820 ³¹		
				141819		
				1473 ⁵		
	Typha/Cladium					
	Cladium			231-385 ⁵		
				386-1036 ¹⁹		
				611 ³¹		
	Slough/wet prairie	11 ³		334 ³¹		
		15^{30}		322-1063 ¹⁹		
ENP-TS	Typha					
	Typha/Cladium					
	Cladium					
	Slough/wet prairie	7^{3}				
		9 ³⁰				

Table 1.(Continued).

Periphyton, soil, and macrophyte data are presented as $\mu g g^{-1}$ water data are presented as $\mu g L^{-1}$.

References, indicated by superscripts, are listed in the Appendix.

in high pH—as much as 1.2 higher in periphyton mats during the day (Gleason and Spackman 1974). This drop in the partial pressure of soluble CO_2 leads to the crystallization of $CaCO_3$ (Gleason and Spackman 1974) and either the precipitation of Ca-P compounds or the coprecipitation of PO₄ with CaCO₃ (Otsuki and Wetzel 1972; House 1990; Diaz and others 1994). A nighttime decrease in pH at the periphyton–water interface may dissolve CaCO₃ (Gleason and Spackman 1974), thus resulting in increased soluble P concentrations (Diaz and others 1994).

The precipitation of inorganic P bound to Ca may not only occur in association with periphyton. High concentrations of Ca-bound inorganic P have been found in the surface (0-5 cm) soils of areas of the Everglades that received P-enriched water from the EAA but had no periphyton (Qualls and Richardson 1995). Qualls and Richardson (1995) hypothesized that this increase in Ca-bound inorganic P was due to the abiotic precipitation or coprecipitation of Ca phosphates or the adsorption of P to CaCO₃. However, the results of Diaz and others (1994) suggest that the pH and Ca concentrations in the enriched areas that were reported by Qualls and Richardson (1995) were too low for P precipitation to occur. In addition, Ca-bound P in the soils of enriched areas could have been formed by periphyton that existed in the past and was then deposited in the flocculent detrital layer and then accreted into the soil.

Even low levels of P enrichment result in both

the loss of the calcareous periphyton mat (Browder and others 1994; Vymazal and others 1994; McCormick and O'Dell 1996) and changes in algal species composition (Grimshaw and others 1993; Raschke 1993; Vymazal and others 1994; McCormick and O'Dell 1996; McCormick and others 1998). These changes often coincide with short-term increases (Vymazal and others 1994) and long-term decreases (McCormick and others 1998) in periphyton biomass and increases in mass-specific productivity (McCormick and others 1998; McCormick and Scinto 1999). P enrichment also leads to the decreased growth and loss of Utricularia purpurea, a common substrate for periphyton (Craft and others 1995; Vaithiyanathan and Richardson 1999). However, alterations in periphyton composition are also correlated with variations in the degree to which the water column is saturated with CaCO₃. The occurrence of blue-green algal calcareous periphyton is correlated with water in equilibrium with CaCO₃, whereas green algae, particularly desmids, are correlated with water undersaturated with CaCO₃ (Gleason and Spackman 1974). Hydrologic conditions can also influence the saturation and precipitation of CaCO₃ by periphyton (Browder and others 1994). Therefore, changes in the species composition of periphyton could be the result of altered CaCO₃ water chemistry and hydrology as well as increased P.

Vymazal and others (1994) experimentally tested for N vs P limitation of epiphytic periphyton in the

		Everglades			
Component	Bedford and Others 1999	Typha	Typha/Cladium	Cladium	Slough/Wet Prairie
Water TP	_	$76.1 \pm 38.8 (5)^{a}$	$42.3 \pm 36.2 (5)^{a}$	$10.8 \pm 4.8 (5)^{\mathrm{b}}$	$10.4 \pm 2.5 (8)^{a}$
Water N:P	_	$94.1 \pm 52.6 \ (4)^{ab}$	$228.0 \pm 221.1 \ (4)^{bc}$	$542.0 \pm 774.8 (3)^{c}$	$377.6 \pm 164.0 (7)^{c}$
Periphyton TP	_	$2885.0 \pm 3049.4 (2)^{a}$	$898.0 \pm 2884.3 (2)^{b}$	$242.8 \pm 337.1 (3)^{c}$	$242.8 \pm 120.1 \ (6)^{c}$
Periphyton N:P	_	_	86.0	165.0	$151.7 \pm 50.2 (4)$
Soil TP	900 ± 590 (109)	$1402.9 \pm 165.6 (15)^{a}$	$947.3 \pm 230.5 \ (10)^{\mathrm{b}}$	$533.2 \pm 94.0 (20)^{c}$	$467.1 \pm 116.1 (10)^{c}$
Soil TP load	_	$0.60 \pm 0.31 (4)^{a}$	$0.38 \pm 0.98 (2)^{a}$	$0.09 \pm 0.04 (10)^{\mathrm{b}}$	_
Soil N:P	47.1 ± 1.3 (109)	$49.0 \pm 10.3 (10)^{a}$	$77.6 \pm 20.5 \ (6)^{a}$	$144.6 \pm 30.2 (12)^{\mathrm{b}}$	$213.0 \pm 80.1 (4)^{\circ}$
Macrophyte TP	1400 ± 200 (65)	$1509.3 \pm 214.6 (3)^{a}$	$515.0 \pm 346.7 \ (4)^{\mathrm{b}}$	$193.3 \pm 32.7 (7)^{\circ}$	$396.5 \pm 2268.0 (2)^{bc}$
Macrophyte N:P	32.9 ± 3.8 (48)	$16.7 \pm 9.0 (3)^{a}$	$40.2 \pm 21.8 (4)^{\mathrm{b}}$	$76.7 \pm 26.2 (7)^{\circ}$	$62.2 \pm 53.3 (3)^{bc}$

Table 2.	Meta-analysis of	Published Data for the Everglades and Other Wetlar	ıds

Mean \pm 95% confidence interval (CI) is presented with the sample size (number of studies) in parentheses for TP concentration (μ g L⁻¹ or μ g g⁻¹), molar N:P ratio, and annual soil TP load (g m⁻² y⁻¹).

Different letters indicate that a significant difference (lpha < 0.05) exists among Everglades habitats, as determined by Tukey's post hoc tests.

Summary statistics are calculated from the analysis by Bedford and others (1999) of nutrient concentrations in temperate North American wetlands. Macrophyte N:P data from Bedford and others (1999) only include values from peat-based wetlands.

The mean is presented when n = 1; — dashes indicate that data was not found in the literature.

Bolded cells indicate that the 95% CI of a parameter in the Everglades and in temperate North American wetlands (Bedford and others 1999) do not overlap.

Everglades. They found that epiphyton growing on macrophyte stems is predominantly limited by P, although colimitation also occurred (Vymazal and others 1994) at their relatively P-enriched sites in WCA-2B (water $PO_4 = 23-30 \ \mu g \ L^{-1}$) (Craft and others 1995). Periphyton is colimited by N and P in the littoral zone of Lake Okeechobee, which is situated at the northern edge of the Everglades and slightly enriched with phosphorus (water TP = $10-14 \ \mu g \ L^{-1}$) (Havens and others 1999). Combined N and P fertilization results in the loss of the surface periphyton mat, a change in species composition, and increases in chlorophyll a concentrations, carbon uptake, and metabolism (Havens and others 1999). Although Vymazal and others (1994) and Havens and others (1999) found evidence for the colimitation of periphyton by P and N, the quick and large response of periphyton to experimental additions of P in oligotrophic areas suggests that periphyton is predominantly P-limited. The possible existence of periphyton colimitation by N and P is interesting given that the N:P ratio of the water column is much greater, and hence N-rich, than that seen in periphyton (Table 2).

The P content of periphyton mats is strongly correlated with water-column P concentrations (Grimshaw and others 1993; McCormick and O'Dell 1996; McCormick and others 1998) (Table 1) and increases with P enrichment (McCormick and Scinto 1999). TP concentrations in periphyton significantly increase along the eutrophication gradient (ANOVA: n = 13, P < 0.001, multiple $R^2 = 0.97$) from about 250 µg g⁻¹ in unenriched *Cladium*

and slough/wet prairie habitats, to 900 μ g g⁻¹ in slightly enriched sites, to 2900 μ g g⁻¹ in enriched sites (Table 2). Periphyton molar N:P ratios vary (Table 3), but they decrease with enrichment from an average of about 150:1 in unenriched slough and wet prairie communities (Table 2) to 86:1 in mixed *Typha* and *Cladium* habitats (Table 3).

The calcareous floating periphyton mat (metaphyton) in oligotrophic areas blocks most light (Vaithiyanathan and Richardson 1998) and therefore dominates primary productivity in the water column. Experimental removal of metaphyton greatly increases benthic dissolved oxygen (DO) concentrations, suggesting that metaphyton limits primary productivity at the water-soil interface (Vaithiyanathan and Richardson 1998). Therefore, loss of metaphyton in response to P enrichment likely stimulates periphytic and benthic periphyton growth. However, the eventual expansion of Typha following P enrichment is associated with the near elimination of all forms of periphyton. Typha shades sunlight more than *Cladium*, and the conversion of macrophyte communities to stands of Typha decreases the net primary productivity (NPP) of periphyton by 80% (Grimshaw and others 1997). Bacteria within periphyton mats rely on the autotrophic production of algae, and a decrease in photosynthetic productivity leads to decreased heterotrophic activity (Neely and Wetzel 1995). This reduction of both photosynthetic and heterotrophic microbial productivity in response to Typha invasion likely reduces the nutrient assimilation and retention capacities of periphyton and therefore of

		Ecosystem Component				
Location	Habitat	Water	Periphyton	Soil	Macrophyt	
EAA	Irrigation canal	55 ¹³				
WCA-1	Typha			57 ²¹		
	Typha/Cladium			10(2)		
	<i>Cladium</i> Slough/wet prairie			186 ²¹ 186 ²¹		
WCA-2A	Typha	60 ²³		33^4	13 ⁴	
WGA-ZA	Турпи	86 ²⁵		38 ²⁴	13^{1} 18^{2}	
		<111118		4012	20 ²⁵	
		137 ²⁰		44-60 ⁹		
				44-71 ²⁹		
				53 ²⁵		
				57 ⁶	1	
	Typha/Cladium	70–140 ²³ 46–422 ¹⁰		44^4	27 ⁴ 34–43 ²⁹	
		$46-422^{10}$ 166-332 ¹⁸		46–137 ²⁸ 69–133 ²³	34–43 ²⁵ 35–49 ²⁵	
		442^{25}		77^{12}	55-49	
		112		80-82 ⁹		
				86 ²⁵		
	Cladium	180-440 ²³	146-184 ²²	82-201 ²⁸	46-64 ²²	
		$>553^{18}$		91 ⁴	62^{2}	
		929 ²⁵		93–124 ⁹	77–84 ²⁹	
				$133-252^{23}$	82 ²⁵	
				135 ¹² 137 ²⁵	139 ⁴	
				137^{-1} 177^{6}		
	Enriched Slough/wet prairie			51–55 ²⁸		
	Slough/wet prairie	517 ²⁶	126 ²²	164^{28}	46-73 ²⁶	
	0 1	>553 ¹⁸	130-26816	268 ²⁶		
		692 ²⁰	177^{26}			
WCA-2B	Typha					
	Typha/Cladium		86^{14}	6	60 ¹⁵	
	<i>Cladium</i>		12814	1226	82 ¹⁵ 88 ¹⁵	
WCA-3A	Slough/wet prairie <i>Typha</i>		128		88-	
WCA-JA	Typha/Cladium					
	Cladium			93–161 ⁶		
	Slough/wet prairie					
WCA-3B	Турһа					
	Typha/Cladium					
	Cladium					
	Slough/wet prairie			2.25		
ENP-SRS	Typha			38 ⁵ 82 ³¹		
	Typha/Cladium			82		
	Cladium			124 ³¹	6127	
				166–365 ⁵	~ ~	
	Slough/wet prairie	260 ³⁰		252 ³¹	46 ²⁷	
		323 ³				
ENP-TS	Typha					
	Typha/Cladium					
	<i>Cladium</i>	24230				
	Slough/wet prairie	242 ³⁰ 272 ³				
		414				

Table 3. N:P Molar Ratio Data for the Everglades

Data are sorted by ecosystem component, location, and macrophyte habitat. References, indicated by superscripts, are listed in the Appendix. the entire wetland ecosystem (Grimshaw and others 1997).

Periphyton oxygenates the water column in oligotrophic Everglades wetlands (Browder and others 1994; McCormick and others 1997; Vaithiyanathan and Richardson 1998). However, there is conflicting evidence of the impact of P enrichment on water-column DO concentrations. Rader and Richardson (1992) concluded that DO at the soil-water and periphyton-water interface was similar between areas with and without P enrichment, but that midcolumn DO in the dry season was lower in response to P enrichment. Both Swift (unpublished in Browder and others 1994) and Belanger and others (1989) found that DO levels were lower in enriched than in unenriched sites. Finally, McCormick and others (1997) showed that nutrient enrichment was associated with reduced periphyton productivity per unit area, a shift toward increasing community heterotrophy, and protracted periods of oxygen depletion. To summarize, it is likely that P enrichment results in the depletion of water-column DO concentrations.

Finally, periphyton communities provide both food and habitat structure for higher consumers. The highly productive periphyton mats of the Everglades are probably an important base of the food web (Browder and others 1994). However, few studies have documented grazing in periphyton mats. In one of the few published studies on Everglades food webs, Hunt (1952) showed that Florida gar (Lepisosteus platyrhincus) in the man-made Tamiami canal consumed large quantities of mosquito fish (Gambusia holbrooki) and freshwater shrimp (Palaemonetes paludosus). In turn, the algae in the periphyton were a major part of the fish's and shrimp's diet (Hunt 1952). The loss of periphyton mats following P enrichment also has a direct effect on consumer communities.

THE SOILS COMPONENT

In general, phosphate loading in wetland soils is associated with adsorption to minerals, not to organic peat (Richardson 1985). The specific minerals that control P adsorption or precipitation vary depending on wetland pH (Moore and Reddy 1994). In alkaline wetlands, this soil storage is related to calcium carbonate chemistry; whereas in acidic wetlands, aluminum (Al) and iron (Fe) chemistries control P adsorption and precipitation (Richardson 1999). Therefore, P adsorption in the carbonaterich Everglades is likely controlled by pH. In contrast, redox potential (Eh) determines the valence of Fe and Al and consequently controls P adsorption dynamics in low pH wetlands (Patrick and Khalid 1974). The paradox of pH control in the soils of the Everglades and other carbonate-rich systems is that the high carbonate concentration buffers pH changes at the same time that pH changes are responsible for most changes in P from a solid (associated with CaCO₃) to a dissolved state. Thus, minor changes in soil pH at the microscale, such as occurs in periphyton mats, may drive this dynamic in the Everglades.

However, other studies have found that the bulk of PO₄ flux in wetland soils is controlled by soil microbes (Chapin and others 1978; Walbridge 1991; Wetzel 1999). Microbial populations may control this flux directly, by releasing phosphatases to fix P, or indirectly, by lowering the redox potential of soils and causing the release of P from Fe complexes (Wetzel 1999). The size of microbial populations increases, in turn, in response to P additions (Reddy and others 1999). In acid organic wetland soils, 90% of added PO₄ remained available when microbial uptake was inhibited, as compared to only 10% in natural soils (Walbridge 1991). In the Everglades, PO_4 removal in the basic organic soils typical of the ecosystem is controlled by soil microbial processes rather than physical or chemical processes (Amador and others 1992), whereas TP removal is likely controlled by abiotic hydrophobic and ionic interactions with soil particles (Jones and Amador 1992). Organically bound P accounts for 71% of accreted TP in unenriched WCA-2A (Scinto 1997)—a finding that lends support to a biological model of P removal.

Across the Everglades, soil TP concentrations are higher in areas receiving water enriched with nutrients and dominated by Typha than in unenriched areas (ANOVA: n = 55, P < 0.001, multiple $R^2 =$ 0.72) (Table 1). Mean soil TP concentrations are significantly higher in enriched areas with Typha (1403 $\mu g g^{-1}$) than in enriched areas with *Typha* and *Cladium* (947 μ g g⁻¹), which, in turn, are greater than in unenriched Cladium and slough/wet prairie (533 and 467 $\mu g g^{-1}$, respectively) across the Everglades (Table 2). Surface soil TP values are even lower, sometimes less than 100 μ g g⁻¹, in shorthydroperiod marl marshes of the southern Everglades (D. L. Childers unpublished). This pattern of changing soil P concentration along eutrophication gradients is consistent across the Everglades (Doren and others 1997) (Table 1). However, the interior areas of WCA-2A, the location for oligotrophic reference sites in this most frequently studied hydrologic unit, are enriched compared to the southern Everglades (Stober and others 1998). Changes in soil N:P also occur following P enrichment

(ANOVA: n = 32, P < 0.001, multiple $R^2 = 0.75$) (Table 3), decreasing from 213:1 in slough/wet prairie and 145:1 in *Cladium* to 78:1 in mixed *Typha* and *Cladium* and 49:1 in *Typha* habitats (Table 2).

Soil P concentrations in the oligotrophic Everglades and other subtropical/tropical carbonate-rich wetlands are far lower than those found in temperate North American wetlands. In the carbonate-rich wetlands of Belize, soil TP ranges from about 50 to 90 μ g g⁻¹ in *Eleocharis cellulosa* marshes, 90 to 175 μ g g⁻¹ in *Cladium* marshes, and 90 to 290 μ g g⁻¹ in *Typha domingensis* marshes (Rejmánkova and others 1995). In comparison, the mean concentration of TP in surface soils of North American temperate wetlands is 900 \pm 590 μ g g⁻¹ (95% CI) (Bedford and others 1999) (Table 2).

Everglades soil TP in oligotrophic and enriched areas becomes lower with soil depth (Reddy and others 1998), decreasing rapidly below 5 cm (Scinto 1997). Enriched and unenriched soils in WCA-2A have similar TP at lower soil depths (Koch and Reddy 1992; Reddy and others 1993; Craft and Richardson 1993a, 1993b, 1998; Qualls and Richardson 1995; Vaithiyanathan and Richardson 1997). The depth at which TP concentration of unenriched and enriched soil profiles diverge may reflect the extent of peat accumulation following the construction of the water conservation areas and export of TP from drained agricultural land in the EAA (Vaithiyanathan and Richardson 1997; Craft and Richardson 1998). Alternatively, macrophytes are known to influence the vertical profile of TP concentration through the "mining" of P from the lower soil depths.

There is disagreement about whether there are changes in the proportion of inorganic and microbial P in soils along nutrient-enrichment gradients. In WCA-1, Newman and others (1997) and Reddy and others (1998) found that the inorganic P fraction increased in response to P enrichment. In enriched areas of WCA-2A, Vaithiyanathan and Richardson (1997) measured a large increase, Koch and Reddy (1992) found a slight increase, and Qualls and Richardson (1995) and Reddy and others (1998) observed no change in the proportion of TP that was inorganic. The conflicting results of these studies may be a result of spatial and temporal variations in soil P. Any increase in the proportion of inorganic P is most likely due to increased P loading from surface water and the mineralization of organic matter in areas that have been impacted by agricultural drainage (Reddy and others 1998). Phosphorus enrichment may also affect soil microbial P content. The concentration of soil microbial biomass P was found to increase in response to

experimental PO_4 enrichment in WCA-2A (Qualls and Richardson 2000). However, a decreasing proportion of soil TP is found in soil microbial biomass in enriched areas of WCA-1 and WCA-2A (Qualls and Richardson 1995; Reddy and others 1998), suggesting that the relative importance of soil microbial mineralization to the cycling of P is greater in unimpacted oligotrophic areas. In summary, the relative importance of biologically mediated cycling and storage of P decreases in nutrient-enriched areas, where inorganic processes have increased importance.

Microbial processes may no longer be limited by P in impacted areas of the Everglades (Reddy and others 1999). Soil mineralization is limited by P (Miao and DeBusk 1999); however, N is limiting to mineralization in high-TP soils (Amador and Jones 1993, 1995). Koch-Rose and others (1994) observed seasonal variation in soil porewater soluble reactive phosphorus (SRP) at enriched sites but low and stable levels of soil SRP at oligotrophic sites, suggesting that the mineralization of organic matter by soil microbes was limited by P at oligotrophic sites and controlled by water levels and temperature at enriched sites. Increased soil microbial processing (Amador and Jones 1995) and changed soil microbial populations (Drake and others 1996) that result from P enrichment often shift soils from aerobic to anaerobic conditions. Unlike most wetlands, peat and marl soils in the unenriched oligotrophic Everglades are oxidized (>100 mV) (Bachoon and Jones 1992; Colbert 2000), with 0.7–4 mg L^{-1} dissolved oxygen in porewater (Gordon and others 1986). Furthermore, the addition of PO₄ to oligotrophic aerobic (500 mV) Everglades peat soils results in significant decreases in soil redox to anaerobic levels (-200 mV) (Colbert 2000). Drake and others (1996) also found significantly lower redox potentials in a nutrient-enriched site (-302 mV) compared to a less-enriched reference site (-94 mV). This shift to anaerobic conditions following P enrichment increases methanogenesis (Drake and others 1996; Colbert 2000) and may also increase denitrification rates (Koch and Reddy 1992). However, methanogenesis (Bachoon and Jones 1992) and denitrification (Gordon and others 1986; White and Reddy 1999) do not immediately change following the addition of PO₄ to oligotrophic soils.

Many studies have used the depth profile of radioisotopes to estimate the rates of P (Table 4) and peat accumulation. Mean estimates of P accumulation using ¹³⁷Cs or ²¹⁰Pb dating are greater in enriched areas with *Typha* (0.60 g m⁻² y⁻¹) or mixed *Typha* and *Cladium* (0.38 g m⁻² y⁻¹) than in unenriched areas with *Cladium* (0.09 g m⁻² y⁻¹)

Typha		Typha/Cladium		Cladium	
Load	Location	Load	Location	Load	Location
0.39–0.47 ²⁴	WCA-2A	0.25-0.35 ⁹	WCA-2A	$0.01 - 0.02^{24}$	WCA-1
0.46^{6}	WCA-2A	0.28-0.637	WCA-2A	0.02-0.217	WCA-2A
$0.54 - 1.14^{9}$	WCA-2A			0.03 ²²	WCA-2A
$0.56 - 0.78^7$	WCA-2A			$0.03 - 0.09^{24}$	ENP-SRS
				0.06^{6}	WCA-2A
				$0.06 - 0.07^{24}$	WCA-3A
				$0.07 - 0.14^{24}$	WCA-2A
				$0.08 - 0.23^{6}$	WCA-3A
				0.11-0.259	WCA-2A
				0.14^{6}	WCA-2B

Table 4. Annual Soil P-Loading (g $m^{-2} y^{-1}$) Data for the Everglades

(ANOVA: n = 16, P < 0.001, multiple $R^2 = 0.84$) (Table 2). Soils in unenriched areas of the Everglades have lower P accumulation rates, but greater N accumulation rates, than other unenriched freshwater wetlands in the United States (Craft and Richardson 1998). Similar to rates of P accumulation, areas receiving nutrient-enriched water have higher organic soil accretion rates than oligotrophic areas. Organic soil accretion is highest near inflows and lowest near the center of WCA-2A, ranging from 1.13 to 0.27 cm y^{-1} (Reddy and others 1993). At similar locations in WCA-2A, Craft and Richardson (1993a) reported organic soil accretion ranging from 0.40 to 0.16 cm y⁻¹ using 137 Cs activity and 0.66–0.03 cm y⁻¹ with 210 Pb activity (Craft and Richardson 1993b). At enriched sites, organic soil accretion rates were substantially higher after the increase in P loading around 1960 than they were before this period (Craft and Richardson 1998). Organic soil accretion in unenriched areas ranges from 0.068 cm y^{-1} in the interior of WCA-2A (Scinto 1997) to 0.11–0.21 cm y^{-1} across the Everglades (Craft and Richardson 1998).

However, soil is one of the last ecosystem components to show changes after P enrichment begins, or the most difficult component to detect changes due to its high variability. No accumulation of P in soil was observed in the first 4 months after the initiation of P-loading mesocosm experiments in WCA-2A (McCormick and Scinto 1999). After 2 years of adding P to WCA-2B, Craft and others (1995) found statistically significant increased TP concentrations in the surface soils of *Cladium* (approximately 700 vs 1100 μ g g⁻¹) and slough communities (approximately 350 vs 700 μ g g⁻¹) only in the treatments with the highest dosing concentrations of P (4.8 vs 1.2 or 0.6 g $m^{-2} y^{-1}$); no differences were found in the mixed Typha/Cladium community. Following the eventual response of soil P concentration to enrichment (Table 2), the ability of the ecosystem to store P in the soil decreases with increased P loading. Craft and Richardson (1993a) calculated that the efficiency of P removal is lower at enriched sites (87% removal) than at unenriched areas (100% removal) of WCA-2A. Craft and Richardson (1993b) also estimated a 74% efficiency of P removal in the enriched areas of WCA-2A. This divergence between the reported rates of P removal from the same enriched areas of WCA-2A is due to the use of different P-accumulation and P-loading rates in the two Craft and Richardson (1993a, b) calculations.

Despite variation in published values, these estimates of P-removal efficiency suggest that in areas receiving water enriched in P, a front of P-enriched soils will move downstream as the ability of the ecosystem to accumulate P becomes saturated. This hypothesis is supported by studies that have found that soil P-adsorption rates decrease with high P concentrations in Everglades soil (Richardson and Vaithiyanathan 1995; Amador and others 1992). In contrast, soil total nitrogen (TN) concentrations do not vary along the same nutrient-enrichment gradients (Koch and Reddy 1992; Newman and others 1997; Craft and Richardson 1998; Vaithiyanathan and Richardson 1999), although enriched (Typha) areas have higher soil NH₄ concentrations (Koch and Reddy 1992; Newman and others 1997; Vaithiyanathan and Richardson 1997).

Based on an analysis of North American con-

structed wetlands, Richardson and others (1997) proposed the "1-gram rule"—that phosphorus loading into freshwater wetlands in excess of 1 g m^{-2} y⁻¹ results in significant increases in outflow TP concentrations above baseline outputs (>40 μ g L^{-1}). Near large inputs of P into WCA-2A, the threshold TP-loading rate at which water TP concentrations exceed 50 μ g L⁻¹ has been reported to be 0.6 g m⁻² y⁻¹ (Richardson and others 1997). It should be noted that water TP concentrations in oligotrophic areas of the Everglades are close to 10 μ g L⁻¹ (Table 2). Therefore, the loading rate that saturates the ecosystem's capacity to accumulate P and results in increases in outflowing water TP concentrations above background is much lower than 0.6 g m⁻² y⁻¹. By comparison, net atmospheric P loading to the oligotrophic Everglades, and hence the bulk of total loading, is around 0.03 g $m^{-2} y^{-1}$ and soil accumulation averages 0.09 g $m^{-2} y^{-1}$. Finally, such modeling of loading rates and outflowing concentrations does not consider the impact of increased P loading on the structure and function of the wetland.

THE MACROPHYTE COMPONENT

The expansion of Typha has been correlated with the enrichment of soil P concentrations in WCA-2A (Urban and others 1993; Jensen and others 1995; Wu and others 1997), WCA-3B (David 1996), and throughout the entire Everglades system (Doren and others 1997). The soil TP level at which Typha invasion occurs varies from 650 $\mu g g^{-1}$ (Wu and others 1997) to 1200 $\mu g g^{-1}$ (data in Doren and others 1997). Vaithiyanathan and Richardson (1999) reported that Typha domingensis became common at 700–900 μ g TP g⁻¹ and that significant decreases in the frequency of oligotrophic species occurred at soil TP concentrations above 700 μ g g^{-1} . However, shifts in plant communities may begin following small increases in P levels but then become more evident as P levels increase. Typha domingensis is also found in soils enriched with P compared to *Cladium* and *Eleocharis cellulosa* along a nutrient-enrichment gradient in tropical freshwater wetlands in Belize and Mexico (Rejmánkova and others 1996). Among unenriched wetlands in Belize, Rejmánkova and others (1995) measured average soil TP concentrations of about 70 $\mu g g^{-1}$ in Eleocharis cellulosa, 140 μ g g⁻¹ in Cladium, and 180 $\mu g g^{-1}$ in *Typha domingensis* marshes. Although increased soil P could cause Typha expansion, a shift to Typha from oligotrophic plant species can also increase soil P.

The spatial extent of Typha domination in the

Everglades has increased in the recent past. Using satellite imagery of WCA-2A from 1973 to 1991, Jensen and others (1995) estimated that the total area of habitats with Typha increased from 2054 to 26,010 ha, whereas Cladium (including periphyton, open water, and slough) decreased from 41,047 to 26,504 ha. Subsequently, Rutchey and Vilchek (1999) concluded that the use of satellite imagery by Jensen and others (1995) overestimated the area occupied by Typha in WCA-2A but that trends in habitat shifts from 1973 to 1991 were valid. Rutchey and Vilchek (1999) concluded from analyses of aerial photographs that the total area occupied by Typha in WCA-2A increased from 5470 ha in 1991 to 9312 ha in 1995. In 1997, Wu and others utilized aerial imagery to estimate that Typha will be dominant in 50% of WCA-2A by 2003-10 if the driving forces of invasion remain unchanged. Other evidence for vegetation change in WCA-2A includes the distribution of recently (within 6-25 years) deposited macrophyte pollen in relation to soil P concentrations (Jensen and others 1999) and changes in seed banks in areas dominated by Typha (van der Valk and Rosburg 1997).

Everglades vegetation has responded to experimental P enrichment. Craft and others (1995) found that aboveground tissue P concentrations and biomass of emergent vegetation increased only in response to the highest P-addition treatments $(4.8 \text{ g m}^{-2} \text{ y}^{-1} \text{ compared to } 0.6 \text{ or } 1.2 \text{ g m}^{-2} \text{ y}^{-1})$ in WCA-2B; Cladium root biomass increased in the medium P-addition treatment. Macrophyte biomass responses were observed only in the 2nd (final) year of fertilization, and species composition did not differ among nutrient-addition treatments (Craft and others 1995), suggesting that there is a lag between an increase in P and changes in vegetation. Newman and others (1996) found that the aboveground biomass of Typha domingensis increased in response to elevated water TP concentration (100 vs 50 μ g L⁻¹) over 2 years in ex situ mesocosms. However, Cladium and Eleocharis interstincta did not respond to increased P concentrations (Newman and others 1996). No significant differences in aboveground biomass were detected among the three species at the lower nutrient level after 2 years (Newman and others 1996). Finally, Daoust (1998) added P to Cladium and wet prairie habitats in ENP. In this study, Cladium responded to P enrichment by increasing below ground biomass in the 1st year and adding aboveground biomass in the 2nd year (Daoust 1998). In contrast, Eleocharis sp. in the wet prairie did not respond until the 2nd year, when it increased stem turnover rates but did

not change aboveground or belowground biomass (Daoust 1998).

Changes in P loading to the Everglades are concomitant with altered hydrology. Urban and others (1993) found that increased inundation was correlated with increases in *Typha* density through time in WCA-2A, although water depth is not related to distance along the WCA-2A transects (Urban and others 1993; McCormick and others 1996) and the spatial distribution of Typha was related to soil TP. In addition, Newman and others (1998) concluded that the proliferation of Typha in two small northern Everglades remnants with elevated soil nutrients was a result of flooding and fire. However, Wu and others (1997) found that Typha invasion rates in WCA-2A escalated with increases in water depth only where soil TP concentrations were low. The mesocosm experiment of Newman and others (1996) demonstrates that the aboveground biomass of Typha domingensis, and not Cladium or Eleocharis, increases in response to increased water depth, but only at lower nutrient levels. Finally, David (1996) used an analysis of vegetation change in WCA-3A to conclude that improved hydroperiod without water quality improvements may be inadequate for Everglades restoration because it will likely result in monotypic stands of Typha domingensis. Therefore, it is unlikely that Typha expansion would occur without P enrichment.

Macrophytes in oligotrophic Everglades marshes have very low tissue P concentrations (Table 1). Previous work has identified and reviewed the low nutrient concentrations in Everglades plants compared to other wetlands (Steward and Ornes 1975; Miao and DeBusk 1999). Total P concentrations in live aboveground macrophyte tissues increase in enriched habitats (ANOVA: n = 16, P < 0.001, multiple $R^2 = 0.94$). Mean TP concentration in *Cladium* vegetation is roughly 200 and 420 μ g g⁻¹, respectively, in sloughs and wet prairies, whereas average Typha tissue concentrations are much higher, around 1500 μ g g⁻¹ (Table 2). Bedford and others (1999) report an average live tissue TP concentration of 1400 $\mu g g^{-1}$ (± 200 95% CI) for temperate North American wetland plant species; average TP concentrations in oligotrophic bogs (700 μ g g⁻¹) and poor fens (600 μ g g⁻¹) are also greater than in Cladium vegetation (Table 2). Plant tissue TP concentrations are also higher in temperate peatlands than in oligotrophic Everglades-roughly 750 $\mu g g^{-1}$ in bogs and 1100 $\mu g g^{-1}$ in fens (Aerts and others 1999). Mean tissue TP concentrations are lower in the oligotrophic Everglades than along a broad nutrient gradient in Central America, where Cladium, Eleocharis cellulosa, and Typha domingensis TP concentrations are 1170, 1210, and 1890 μ g g⁻¹, respectively (Rejmánkova and others 1996). However, plant tissue TP concentrations in the oligotrophic marshes of Belize average 200 μ g g⁻¹ in *Cladium*, 500 μ g g⁻¹ in *Eleocharis cellulosa*, and 1400 μ g g⁻¹ in *Typha domingensis* (Rejmánkova and others 1995).

The N:P ratio is useful for evaluating nutrient economy in plant communities (Koerselman and Meuleman 1996). Koerselman and Meuleman (1996) suggested that N-limitation occurs at molar N:P ratios below 31:1, whereas plant communities are limited by P at N:P ratios above 35:1. The N:P ratio in live aboveground tissues decreases in response to enrichment (ANOVA: n = 17, P <0.001, multiple $R^2 = 0.81$). Cladium in the oligotrophic Everglades has very high molar N:P ratios, ranging from 46:1 to 139:1 (Table 3) and averaging 77:1 in the literature (Table 2), suggesting extreme P limitation. Cladium N:P ratios in the Everglades are much greater than vegetation N:P values in peat-based temperate wetland ecosystems (Table 2) (Bedford and others 1999). The N:P ratio of aboveground and belowground Typha tissues increases from an enriched (average for all tissue = 15:1) to an unenriched site (average for all tissue =33:1) along a eutrophication gradient in WCA-2A (Koch and Reddy 1992). In addition, the average N:P ratio of all Cladium tissues increases from 20:1 to 139:1 along the same gradient (Koch and Reddy 1992). Miao and Sklar (1998) found similar trends in Typha and Cladium N:P ratios. Typha may be N-limited in enriched areas, as suggested by the literature's average N:P ratio of 17:1 (Table 2). In contrast, *Cladium* N:P ratios (average = 77:1) (Table 2) are much higher than the P-limitation threshold suggested by Koerselman and Meuleman (1996), and *Cladium* shoot and root growth are responsive to the addition of P but not N (Steward and Ornes 1983). However, observations of N:P ratios of multiple species in an oligotrophic area of ENP suggest that not all species are necessarily P-limited (Daoust and Childers 1999). This assertion relies on the generality of the community-level N:P threshold for P limitation for individual species in communities, which has not been confirmed (Koerselman and Meuleman 1996). To summarize, Everglades macrophytes are in general highly limited by P in oligotrophic areas but may become N-limited in areas enriched with P. The degree of P limitation of macrophyte growth in the oligotrophic Everglades appears to be greater than in other wetland ecosystems.

The low P concentrations in the live tissue of vegetation in the oligotrophic Everglades and effi-

cient resorption of P from senescing leaves (Richardson and others 1999) result in low P concentrations in macrophyte detritus (Davis 1991). TP concentrations in the standing dead leaves of Cla*dium* are less than 100 $\mu g g^{-1}$ in interior, lessenriched areas of WCA-2A (DeBusk and Reddy 1998; Qualls and Richardson 2000). The nutrient content of detritus is an important determinant of decomposition rates (Webster and Benfield 1986; Enriquez and others 1993), and the litter of Everglades macrophytes has low decomposition (Amador and Jones 1993) and P-mineralization rates (Amador and Jones 1995). Low decomposition rates in long-hydroperiod areas of the Everglades are evidenced by a thick peat layer. In the Everglades, Cladium has decreased P nutrient use efficiency at high soil P (Richardson and others 1999). In addition, Typha has higher P in aboveground (Miao and Sklar 1998) and all tissues (Koch and Reddy 1992). Typha in highly enriched areas (De-Busk and Reddy 1998), as well as in oligotrophic areas (Qualls and Richardson 2000), also has higher TP concentrations in standing dead leaves than Cladium in oligotrophic areas of the Everglades, indicating that Typha is less proficient at P resorption (Sensu Killingbeck 1996) than Cladium. The increase in tissue nutrient levels and a shift to species with higher tissue nutrient concentrations in response to P enrichment in the Everglades results in altered rates of detrital decomposition (Davis 1991; DeBusk and Reddy 1998; Qualls and Richardson 2000). Thus, P enrichment may initiate a positive feedback cycle following the conversion of vegetation to

Dead macrophyte tissue is an important sink for P in the Everglades. At intermediate to highly enriched water-column P concentrations, P is immobilized in macrophyte detritus and P content increases during decomposition, whereas the P content of detritus does not change at low watercolumn P concentrations (Davis 1991; Qualls and Richardson 2000). The P immobilization rate for macrophyte detritus (0.07 g m⁻² y⁻¹) (Davis 1991) is similar to estimates of total soil P load in unenriched areas (Table 2). However, the detritus immobilization rate of P in enriched areas (1.42 g m⁻² y^{-1}) (Davis 1991) is greater than total soil loading estimates (Table 2). This difference in macrophyte detrital immobilization rates compared to total soil loading rates in enriched areas (Table 4) suggests that some detrital P is eventually exported downstream to less impacted marshes instead of accreted to soil following P enrichment.

Typha.

THE CONSUMER COMPONENT

Due in part to the oligotrophy of the ecosystem, the oligotrophic Everglades has very low fish and aquatic invertebrate biomass relative to other freshwater wetlands (Turner and others 1999). The Everglades faunal community is very responsive to P enrichment, suggesting food web limitations by P. Rader and Richardson (1992, 1994) observed an increase in the number of species, including species not found in oligotrophic marshes, and an increase in the density of most invertebrate and small fish species in sweep nets and soil cores from nutrientenriched sloughs of WCA-2A. However, they (1994) found that the density of the freshwater shrimp (*Palaemonetes paludosus*) decreased drastically in P-enriched sloughs, from 54.5 to 1.2 m⁻³.

Furthermore, in a comparison of litterbag colonization in eutrophic (Typha) and nonimpacted areas (Cladium) of WCA-2A, Urban (unpublished in Davis 1994) found a near absence of snails, an absence of isopods, a doubling of annelids, and in general fewer taxa in eutrophic sites. There is no difference in total macroinvertebrate standing crop between Typha communities and Cladium or wet prairie (nonenriched) communities in WCA-2A and ENP, although the total fish biomass is greater in enriched areas (Turner and others 1999). Turner and others (1999) hypothesized that a trophic cascade was operating and that the fish (primarily carnivorous) were reducing the invertebrate biomass in the nutrient-enriched areas. Finally, Richardson and others (1999) documented increased herbivory of Cladium leaves in P-enriched areas compared to less enriched areas of WCA-2A. This increase in herbivory occurs in conjunction with a decrease in herbivore-deterring phenolic secondary compounds and an increase in TP concentration in Cladium leaves (Richardson and others 1999).

There are no published data on P concentrations in Everglades consumers. A preliminary estimate of average TP concentrations is 38,500 $\mu g g^{-1}$ (dry mass) in three dominant fish species of the Everglades (C. Stevenson unpublished). In contrast, mean TP concentrations in fish from Laurentian oligotrophic lakes is 14,900 $\mu g g^{-1}$ (Sterner and George 2000). A review of whole-fish P concentrations by Sterner and George (2000) indicated that typical TP concentrations in fish range from roughly 20,000 to 40,000 $\mu g g^{-1}$. These preliminary data suggest that fish in the Everglades may have high TP concentrations relative to those in other ecosystems. The fact that fish in extremely P-limited Everglades marshes may potentially be sequestering, or bioaccumulating, more P than fish in other ecosystems is particularly intriguing and warrants further investigation.

Palaemonetes paludosus is a dominant invertebrate (61% of total invertebrate biomass) (Turner and others 1999) and an important part of the Everglades food web (Kushlan and Kushlan 1980); fish consume large numbers of them (Hunt 1952). The diet of P. paludosus comes primarily from periphyton mats (Hunt 1952) and from a periphyton/macrophyte detritus mix (Browder and others 1994). Therefore, the loss of the calcareous periphyton-Utricularia assemblage and the concurrent decrease in P. paludosus density in nutrient-enriched areas should also affect higher trophic levels. An interpretation of data presented in Rader and Richardson (1994), and similar to the trophic evaluation of Rader (1994), indicates that the proportion of individuals that were detritivorous increased from about 33% in unenriched areas to 42% in enriched areas, whereas the proportion that was herbivorous decreased from about 44% to 25% with nutrient enrichment. Therefore, a shift from a grazer-based to a detritivore-based consumer community has occurred in enriched areas of WCA-2A. Finally, the shift in periphyton composition toward less palatable species and an alteration in the structure of the periphyton mat may both decrease the food value of periphyton and the habitat value of the mat, resulting in changes to primary and secondary productivity in the Everglades (Browder and others 1994). These cascading effects of periphyton loss on ecosystem dynamics need further study.

The number of nesting wading birds in the southern and central Everglades is thought to have declined 90% from the period of early drainage (around the 1930s) to the 1980s in response to altered hydrology (Ogden 1994). Historically, P deposition by wading birds was locally important at colonial nesting sites on tree islands. This relocation and concentration of P has become less important as the number of wading birds nesting in the Everglades has declined; P deposition by wading birds at nesting sites may have been as high as 331 g m^{-2} y^{-1} in 1934, but it was only 0.90 g m⁻² y⁻¹ in 1987 (Frederick and Powell 1994). This historic estimate of P deposition by wading birds is four orders of magnitude greater than atmospheric deposition rates (approximately 0.03 g $m^{-2} y^{-1}$), whereas the recent loading rate is only 30 times greater than atmospheric sources of P. However, avian P deposition has been very localized because of the patchy distribution of colonial nesting sites in the landscape. In addition to declines in the magnitude of deposition, the spatial location of most P deposition by wading birds has shifted from the mangrove zone at the estuarine fringe of Florida Bay to the water conservation areas, especially WCA-3, due to the shift in nesting sites (Frederick and Powell 1994). The large decline in the rate of P deposition and the different locations of these birds has undoubtedly affected the cycling of P at both the land-scape and the local scales.

Research Gaps

Our review of the literature, in the context of other freshwater wetland ecosystems, revealed several significant gaps in our understanding of P biogeochemistry and the effects of P enrichment on the Everglades and other oligotrophic ecosystems. First, most research on P in the Everglades has focused on WCA-2A (Table 1), the most enriched hydrologic unit of the Everglades. Thus, there is a dearth of data on P in the other WCAs and ENP. Furthermore, the way in which periphyton facilitates the removal of P by adsorption to CaCO₃ or precipitation with Ca has not been adequately quantified. Although gradients in soil TP concentrations along nutrient and macrophyte habitat gradients are well documented, the order in which ecosystem components respond to and are changed by P additions is not clear. Nor have consumer trophic dynamics and P relations been adequately studied. In addition, the role of the flocculent detrital layer in P biogeochemistry needs further evaluation. P cycling in different ecosystem components needs further study. Finally, more controlled experimental additions of P should be done to separate the effects of concurrent N and P additions (such as discharge from EAA) from P fertilization alone, so that we can assess the effects of low levels of P enrichment and document the events that occur when an Everglades marsh is first exposed to P additions. This research will help to alleviate controversy by identifying TP concentrations in water entering the Everglades that can "prevent an imbalance in the natural populations of aquatic flora or fauna" (Everglades Forever Act). Filling in these gaps in our understanding of P will improve efforts to manage and restore the Everglades.

ECOSYSTEM FUNCTIONAL ANALOGIES

There are several similarities between the Everglades and other P-limited wetland ecosystems, including their hydrogeomorphology, the microbial dominance of P cycling, and their soils. For example, the wetlands of the Yucatan Peninsula in Belize and Mexico have a flat limestone bedrock and a climate similar to the Everglades (Rejmánkova and others 1995) and an analogous ecosystem structure and function. Freshwater wetlands with a plant community structure similar to the Everglades also occur on the Zapata Peninsula of Cuba, an area with similar hydrology and flat karstic limestone bedrock (Borhidi and others 1983). Phosphorus is also very limiting in spring-fed calcareous wetlands in England (Boyer and Wheeler 1989). Wetlands receiving water from a carbonate-dominated landscape are likely to be P-limited due to the biologically mediated or abiotic removal of P by precipitation with Ca or adsorption to CaCO₃ (see, for example, Boyer and Wheeler 1989).

Hydrology may also influence P limitation in wetlands. The large flat bedrock of the Everglades results in a hydrology that is primarily driven by precipitation; similarly, ombrotrophic temperate bogs have lower leaf P concentrations than temperate fens in which the hydrology is controlled by precipitation, surface water, and groundwater (Aerts and others 1999). In contrast, leaf N concentrations do not differ among fens and bogs (Aerts and others 1999). In other words, the conservative nature of P cycling necessitates the presence of an external supply of P. A low level of allochthonous P inputs to ecosystems will likely result in P limitation.

Another characteristic that the Everglades shares with other P-limited wetlands is the microbial control of P uptake and cycling. In a Michigan fen, Richardson and Marshall (1986) demonstrated that microbes control initial P uptake and cycling in surface waters. In addition, microbial processes strongly influence the supply of available P in Plimited pocosin wetlands in North Carolina (Walbridge 1991). As in the Everglades, periphyton mats are also abundant in the P-limited freshwater wetlands of the Yucatan Peninsula (Rejmánkova and other 1995). Periphyton has also been shown to dominate P uptake and cycling in the oligotrophic littoral zone of a coastal lake (Howard-Williams and Allanson 1981). In arctic wetlands, soil microbes immobilize P after fertilization and control the availability of P to plants (Jonasson and others 1996). When microbial populations crash in the winter, P is released in arctic wetlands (Chapin and others 1978). In the subtropical Everglades, there are no prolonged freezes to lyse microbial cells; therefore, microbes are likely to be strong competitors for P throughout the year.

However, it should be noted that the microbial control of P cycling in the Everglades and other P-limited wetlands could be a response to ecosystem P limitation rather than the cause of P limitation. Microbes may dominate nutrient cycling in oligotrophic wetlands because their high surface area-to-volume ratio makes them more competitive assimilators of available P. The importance of periphyton to the primary production and P cycling of many areas of the Everglades may be due to its competitive uptake of P and a position in the water column that allows it to capture scarce P inputs.

Finally, soils with a high organic content are also more likely to be limited by P. Peatlands, including bogs and fens, are often P-limited, but marshes with mineral-rich soils are generally N-limited (Bedford and others 1999). N:P ratios in soils and the live tissues of plants in temperate wetlands are, on average, significantly higher in peat than in mineral soils (Bedford and others 1999). This is probably due to the low mineral content of organic soils and the resulting lack of imported P from minerals. Soils low in P, as well as N, are also more likely to accumulate organic matter in the soil due to their low decomposition rates. In general, wetlands are more likely to be P-limited when inputs of P are restricted.

CONCLUSIONS

The oligotrophic Everglades is extremely P-limited due to a suite of factors. This sensitivity to P enrichment is ultimately due to the low-relief limestone bedrock and large spatial extent of the Everglades, as well as the conservative nature of P cycling. Other wetland ecosystems share some of the same causes of P limitation, but we found none that included all of these factors. Very little exogenous P is available to the Everglades from mineral rock weathering or allochthonous sediments, and water flow rates are low. Thus, the primary natural source of P to the oligotrophic Everglades is atmospheric deposition, which supplies low levels of P. In addition, high Ca concentrations and pH remove P from the water column by the precipitation of Ca-P minerals and the adsorption of PO₄ onto CaCO₃. As a consequence of the low supply and availability of P to the ecosystem, water-column TP concentrations in oligotrophic areas average about 10 μ g L⁻¹. Periphyton is very abundant compared to other wetlands and rapidly removes available P from the water column, while Cladium is able to grow at low P levels. The biomass of fish and aquatic invertebrates is also smaller than in other wetlands. In addition, the soil profile is oxidized to slightly reduced, even in peat soils. Finally, the uniqueness of the Everglades is evident in the very low P concentrations in plants and very high N:P ratios in its soils and plants, as compared to other wetlands (see Table 2). Therefore, the P biogeochemistry of the Everglades is likely unique.

Phosphorus enrichment modifies the structure and function of the Everglades ecosystem. Increased P loading increases the concentration of P in most components (water, periphyton, soil, and macrophytes), alters biogeochemical processes, eliminates calcareous periphyton mats, deoxygenates soils, stimulates the invasion of Typha, and increases the abundance of fish. Some of these changes may be beneficial, such as the potential benefit to wading birds from increased fish abundance. However, at the same time, fish may be less available to wading birds in areas where the vegetation structure has been altered by thick stands of Typha. In general, P enrichment leads to a distinctly different ecosystem than the historic oligotrophic Everglades.

We sought to identify the causes and indications of P limitation in wetlands. The functional similarities between the Everglades and other P-limited wetlands include their hydrogeomorphology, the microbial dominance of P cycling, and their soils. Not surprisingly, wetlands in general tend to be limited by P when supply rates are low or removal rates are high. Two common causes of low P supply to wetlands are a hydrology that is predominantly controlled by precipitation and their location in a watershed with low content or availability of mineral-bound P in bedrock and soils. Finally, common mechanisms for P removal in wetlands include location in a calcareous landscape and microbial dominance of P cycling. In a limestone-dominated watershed, P availability can be limited by CaCO₃ adsorption or Ca-P precipitation. Microbes often dominate the cycling of P in P-limited wetland ecosystems; however, this may be a response to, rather than a cause of, limitation.

Our review of Everglades P cycling and our analysis of this system in the context of other nutrientpoor freshwater wetland ecosystems has led us to consider the relationship between oligotrophy and P limitation in a more general way. To that end, we hypothesize that most oligotrophic freshwater wetland ecosystems are P-limited. The biota of oligotrophic ecosystems can offset N limitation by the fixation of atmospheric N2 (Redfield 1958; Schindler 1977; Howarth and others 1988; Short and others 1990; Vitousek and Howarth 1991; Levine and Schindler 1992), whereas P cycling is conservative. Similarly, oligotrophic freshwater lakes are predominantly limited by P (Downing and McCauley 1992) and there is a trend toward increasing N limitation with eutrophication (Smith 1998). Therefore, N-limited oligotrophic wetlands should be rare, and the status of nutrient limitation in wetlands is likely determined by P loading. We hope that these hypotheses on the general causes of P limitation within and among wetland ecosystems will stimulate more research and synthesis on the nature of P biogeochemistry.

ACKNOWLEDGMENTS

We thank Joy Zedler and the *Ecosystems* editors for inviting this review. Many colleagues from the South Florida Wetland Ecosystems Lab, Southeast Environmental Research Center (SERC), and Department of Biological Sciences at Florida International University provided valuable insights on phosphorus in the Everglades and made useful comments on the manuscript. This review was made possible through funding to the Southeast Environmental Research Center from the United States Department of the Interior, Everglades National Park, Arthur R. Marshall Loxahatchee National Wildlife Refuge, and the South Florida Water Management District. This is SERC publication series contribution no. 144.

REFERENCES

- Aerts R, Verhoeven JTA, Whigham DF 1999. Plant-mediated controls on nutrient cycling in temperate fens and bogs. Ecology 80:2170–81.
- Ahn H. 1999. Statistical modeling of total phosphorus concentrations measured in south Florida rainfall. Ecol Model 116: 33–44.
- Amador JA, Jones RD. 1995. Carbon mineralization in pristine and phosphorus-enriched peat soils of the Florida Everglades. Soil Sci 159:129–41.
- Amador JA, Jones RD. 1993. Nutrient limitations on microbial respiration in peat soils with different total phosphorus content. Soil Biol Biochem 25:793–801.
- Amador JA, Richany GH, Jones RD. 1992. Factors affecting phosphate uptake by peat soils of the Florida Everglades. Soil Sci 153:463–70.
- Bachoon D, Jones RD. 1992. Potential rates of methanogenesis in sawgrass marshes with peat and marl soils in the Everglades. Soil Biol Biochem 24:21–7.
- Bedford BL, Walbridge MR, Aldous A. 1999. Patterns in nutrient availability and plant diversity of temperate North American wetlands. Ecology 80:2151–69.
- Belanger TV, Scheidt DJ, Platko JR II. 1989. Effects of nutrient enrichment on the Florida Everglades. Lake Reservoir Manage 5:101–11.
- Borhidi, A, Muñiz O, Del Risco E. 1983. Plant communities of Cuba, I. Fresh and salt water, swamp and coastal vegetation. Acta Botanica Hungarica 29:337–76.
- Boyer MLH, Wheeler BD. 1989. Vegetation patterns in springfed calcareous fens: calcite precipitation and constraints on fertility. J Ecol 77:597–609.
- Browder JA, Gleason PJ, Swift DR. 1994. Periphyton in the Everglades: spatial variation, environmental correlates, and ecological implications. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration Delray Beach (FL): St. Lucie Press. 379–418.
- Chapin FS III, Barsdate RJ, Barèl D. 1978. Phosphorus cycling in

Alaskan coastal tundra: a hypothesis for the regulation of nutrient cycling. Oikos 31:189–99.

- Coale FJ, Izuno FT, Bottcher AB. 1994. Phosphorus drainage water from sugarcane in the Everglades Agricultural Area as affected by drainage rate. J Environ Quality 23:121–26.
- Colbert GL. 2000. The effects of phosphorus and carbon additions on anaerobic microbial activity in peat soils of the Florida Everglades [thesis]. Miami: Florida International University.
- Craft CB, Richardson CJ. 1993a. Peat accretion and N, P, and organic C accumulation in nutrient-enriched and unenriched Everglades peatlands. Ecol Appl 3:446–58.
- Craft CB, Richardson CJ. 1993b. Peat accretion and phosphorus accumulation along a eutrophication gradient in the northern Everglades. Biogeochemistry 22:133–56.
- Craft CB, Richardson CJ. 1998. Recent and long-term organic soil accretion and nutrient accumulation in the Everglades. Soil Science Soc of Am J 62:834–43.
- Craft CB, Vymazal J, Richardson CJ. 1995. Response of Everglades plant communities to nitrogen and phosphorous additions. Wetlands 15:258–71.
- Daoust RJ. 1998. Investigating how phosphorus controls structure and function in two Everglades wetland plant communities [thesis]. Miami: Florida International University.
- Daoust RJ, Childers DL. 1999. Controls on emergent macrophyte composition, abundance, and productivity in freshwater Everglades wetland communities. Wetlands 19:262–75.
- David PG. 1996. Changes in plant communities relative to hydrologic conditions in the Florida Everglades. Wetlands 16:15– 23.
- Davis SM. 1991. Growth, decomposition, and nutrient retention in *Cladium jamaicense* Crantz and *Typha domingensis* Pers. in the Florida Everglades. Aquat Bot 40:203–24.
- Davis SM. 1982. Patterns of radiophosphorus accumulation in the Everglades after its introduction into surface water. West Palm Beach (FL): South Florida Water Management District.
- Davis SM. 1994. Phosphorous inputs and vegetation sensitivity in the Everglades. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 357–78.
- Davis SM, Gunderson LH, Park WA, Richardson JR, and Mattson JE. 1994. Landscape dimension, composition, and function in a changing Everglades ecosystem. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 419–44.
- Davis SM, Ogden JC, editors. 1994. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press,
- DeBusk WF, Reddy KR. 1998. Turnover of detrital organic carbon in a nutrient-impacted Everglades marsh. Soil Sci Soc Am J 62:1460–8.
- DeBusk WF, Reddy KR, Koch MS, Wang Y. 1994. Spatial distributions of soil nutrients in a northern Everglades marsh: Water Conservation Area 2A. Soil Sci Soc Am J 58:543–52.
- Diaz OA, Reddy KR, and Moore PA. Jr. 1994. Solubility of inorganic phosphorus in stream water as influenced by pH and calcium concentration. Water Res 28:1755–63.
- Doren RF, Armentano TV, Whiteaker LD, and Jones RD. 1997. Marsh vegetation patterns and soil phosphorus gradients in the Everglades ecosystem. Aqua Bot 56:145–63.
- Downing JA, McCauley E. 1992. The nitrogen:phosphorus relationship in lakes. Limnol Oceanogr 37:936–45.
- Drake HL, Aumen NG, Kuhner C, Wagner C, Grie Bhammer A,

Schmittroth M. 1996. Anaerobic microflora of Everglades sediments: effects of nutrients on population profiles and activities. Appl Environ Microbiol 62:486–93.

- Duever MJ, Meeder JF, Meeder LC, McCollum JM. 1994. The climate of south Florida and its role in shaping the Everglades ecosystem. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 225–48.
- Enriquez S, Duarte CM, Sand-Jensen K. 1993. Patterns in decomposition rates among photosynthetic organisms: the importance of detritus C:N:P content. Oecologia 94:457–71.
- Fennema RJ, Neidrauer CJ, Johnson RA, MacVicar TK, and Perkins WA. 1994. A computer model to simulate natural Everglades hydrology. In: Davis SM, and Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 249–90.
- Fitz HC, Sklar FH. 1999. Ecosystem analysis of phosphorus impacts and altered hydrology in the Everglades: a landscape modeling approach. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 585–620.
- Frederick PC, Powell GVN. 1994. Nutrient transport by wading birds in the Everglades. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 571–84.
- Gleason PJ, Spackman WS Jr. 1974. Calcareous periphyton and water chemistry in the Everglades. In: Gleason PJ, editor. Environments of South Florida: present and past. Miami: Miami Geological Society, p 146–81.
- Gleason PJ, Stone P. 1994. Age, origin, and landscape evolution of the Everglades peatland. In: Davis SM, and Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 149–97.
- Gordon AS, Cooper WJ, Scheidt DJ. 1986. Denitrification in marl and peat sediments in the Florida Everglades. Appl Environ Microbiol 52:987–91.
- Grimshaw HJ, Rosen M, Swift DR, Rodberg K, Noel JM. 1993. Marsh phosphorous concentrations, phosphorous content and species composition of Everglades periphyton communities. Arch Hydrobiol 128:257–76.
- Grimshaw HJ, Wetzel RG, Brandenburg M, Segerblom K, Wenkert LJ, Marsh GA, Charnetzky W, Haky JE, Carraher C. 1997. Shading of periphyton communities by wetland emergent macrophytes: decoupling of algal photosynthesis from microbial nutrient retention. Arch Hydrobiol 139:17–27.
- Havens KE, East TL, Rodusky AJ, Sharfstein B. 1999. Littoral periphyton responses to nitrogen and phosphorus: an experimental study in a subtropical lake. Aquat Bot 63:267–90.
- Hendry CD, Brezonik PL, Edgerton ES. 1981. Atmospheric deposition of nitrogen and phosphorus in Florida. In: Eisenreich SJ, editor. Atmospheric pollutants in natural waters. Ann Arbor (MI): Ann Arbor Science. p 199–215.
- House WA. 1990. The prediction of phosphate coprecipitation with calcite in freshwaters. Water Res 24:1017–23.
- Howard-Williams C. 1985. Cycling and retention of nitrogen and phosphorus in wetlands: a theoretical and applied perspective. Freshwater Biol 15:391–431.
- Howard-Williams C, Allanson BR. 1981. Phosphorus cycling in a dense *Potamogeton pectinatus* L. bed. Oecologia 49:56–66.
- Howarth R, Marino WR, Lane J, Cole JJ. 1988. Nitrogen fixation in freshwater, estuarine, and marine ecosystems. 1. Rates and importance. Limnol Oceanog 33:669–87.

- Hunt BP. 1952. Food relationships between Florida spotted gar and other organisms in the Tamiami Canal, Dade County, Florida. Trans Am Fish Soc 82:13–33.
- Jensen JE, Cooper SR, and Richardson CJ. 1999. Calibration of modern pollen along a nutrient gradient in Everglades Water Conservation Area–2A. Wetlands 19:675–88.
- Jensen JR, Rutchey K, Koch MS, Narumalani S. 1995. Inland wetland change detection in the Everglades Water Conservation Area 2A using a time series of normalized remotely sensed data. Photogram Eng Remote Sens 61:199–209.
- Jonasson S, Michelsen A, Schmidt IK, Nielsen EV, Callaghan TV. 1996. Microbial biomass C, N, and P in two arctic soils and responses to addition of NPK fertilizer and sugar: implications for plant nutrient uptake. Oecologia 106:507–15.
- Jones LA. 1948. Soils, geology, and water control in the Everglades region. Bulletin no. 442. Gainesville (FL): University of Florida Agricultural Experimental Station.
- Jones RD, Amador JA. 1992. Removal of total phosphorus and phosphate by peat soils of the Florida Everglades. Can J Fish Aquat Sci 49:577–83.
- Killingbeck KT. 1996. Nutrients in senesced leaves: keys to the search for potential resorption and resorption proficiency. Ecology 77:1716–27.
- Koch MS, Reddy KR. 1992. Distribution of soil and plant nutrients along a trophic gradient in the Florida Everglades. Soil Sci Soc Am J 56:1492–99.
- Koch-Rose MS, Reddy KR, Chanton JP. 1994. Factors controlling seasonal nutrient profiles in a subtropical peatland of the Florida Everglades. J Environ Qual 23:526–33.
- Koerselman W, Meuleman AFM. 1996. The vegetation N:P ratio: a new tool to detect the nature of nutrient limitation. J Appl Ecol 33:1441–50.
- Kushlan JA, Kushlan MS. 1980. Population fluctuations of the prawn, *Palaemonetes paludosus*, in the Everglades. Am Midland Nat 103:401–3.
- Levine SN, Schindler DW. 1992. Modification of the N:P ratio in lakes by in situ processes. Limnol Oceanogr 37:917–35.
- Light SS, Dineen JW. 1994. Water control in the Everglades: a historical perspective. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 47–84.
- McCally D. 1999. The Everglades: an environmental history. Gainesville (FL): University Press of Florida,
- McCormick PV, Chimney MJ, Swift DR. 1997. Diel oxygen profiles and water column community metabolism in the Florida Everglades, U.S.A. Arch Hydrobiol 140:117–29.
- McCormick PV, O'Dell MB. 1996. Quantifying periphyton responses to phosphorus in the Florida Everglades: a synopticexperimental approach. J North Am Benthol Soc 15:450–68.
- McCormick PV, Rawlik PS, Lurding K, Smith EP, Sklar FH. 1996. Periphyton–water quality relationships along a nutrient gradient in the northern Florida Everglades. J North Am Benthol Soc 15:433–49.
- McCormick PV, Scinto LJ. 1999. Influence of phosphorus loading on wetlands periphyton assemblages: a case study from the Everglades. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 301–20.
- McCormick PV, Shuford RBE III, Backus JG, Kennedy WC. 1998. Spatial and seasonal patterns of periphyton biomass and productivity in the northern Everglades, Florida, USA. Hydrobiologia 362:185–208.

- McCormick PV, Stevenson RJ. 1998. Periphyton as a tool for ecological assessment and management in the Florida Everglades. J Phycol 34:726–33.
- Miao SL, DeBusk WF. 1999. Effects of phosphorous enrichment on structure and function of sawgrass and cattail communities in the Everglades. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 275–99.
- Miao SL, Sklar FH. 1998. Biomass and nutrient allocation of sawgrass and cattail along a nutrient gradient in the Florida Everglades. Wetlands Ecol Manage 5:245–63.
- Moore PA Jr, Reddy KR. 1994. Role of Eh and pH on phosphorus geochemistry in sediments of Lake Okeechobee, Florida. J Environ Qual 23:955–64.
- Moustafa MZ, Chimney MJ, Fontaine TD, Shih G, Davis S. 1996. The response of a freshwater wetland to long-term "low level" nutrient loads—marsh efficiency. Ecol Eng 7:15–33.
- Neely RK, Wetzel RG. 1995. Simultaneous use of 14C and 3H to determine autotrophic production and bacterial protein production in periphyton. Microb Ecol 30:227–73.
- Newman S, Grace JB, Koebel JW. 1996. Effects of nutrients and hydroperiod on *Typha, Cladium,* and *Eleocharis*: implications for Everglades restoration. Ecol Appl 6:774–83.
- Newman S, Reddy KR, DeBusk WF, Wang Y, Shih G, Fisher MM. 1997. Spatial distribution of soil nutrients in a northern Everglades marsh: Water Conservation Area 1. Soil Sci Soc Am J 61:1275–83.
- Newman S, Schuette J, Grace JB, Rutchey K, Fontaine T, Reddy KR, Pietrucha M. 1998. Factors influencing cattail abundance in the northern Everglades. Aquat Bot 60:265–80.
- Ogden JC. 1994. A comparison of wading bird nesting colony dynamics (1931–1946 and 1974–1989) as an indication of ecosystem conditions in the southern Everglades. In: Davis SM, Ogden JC, editors. Everglades: the ecosystem and its restoration. Delray Beach (FL): St. Lucie Press, p 533–70.
- Otsuki A, Wetzel RG. 1972. Coprecipitation of phosphate with carbonates in a marl lake. Limnol Oceanog 17:763–7.
- Parker GG. 1974. Hydrology of the pre-drainage system of the Everglades in southern Florida. In: Gleason PJ, editor. Environments of South Florida: present and past. Miami: FL Miami Geological Society, p 18–27.
- Patrick WH Jr, Khalid RA. 1974. Phosphate release and sorption by soils and sediments: effect of aerobic and anaerobic conditions. Science 186:53–5.
- Qualls RG, Richardson CJ. 1995. Forms of soil phosphorus along a nutrient enrichment gradient in the northern Everglades. Soil Sci 160:183–98.
- Qualls RG, Richardson CJ. 2000. Phosphorus enrichment affects litter decomposition, immobilization, and soil microbial phosphorus in wetland mesocosms. Soil Sci Soc Am J 64:799–808.
- Rader RB. 1994. Macroinvertebrates of the northern Everglades: species composition and trophic structure. Florida Sci 57:22–33.
- Rader RB, Richardson CJ. 1992. The effects of nutrient enrichment on algae and macroinvertebrates in the Everglades: a review. Wetlands 12:121–35.
- Rader RB, Richardson CJ. 1994. Response of macroinvertebrates and small fish to nutrient enrichment in the northern Everglades. Wetlands 14:134–46.
- Raschke RL. 1993. Diatom (Bacillariphyta) community response to phosphorus in the Everglades National Park, USA. Phycologia 32:48–58.
- Reddy KR, DeLaune RD, DeBusk WF, Koch MS. 1993. Long-

term nutrient accumulation rates in the Everglades. Soil Sci Soc Am J 57:1147–55.

- Reddy KR, Wang Y, DeBusk WF, Fisher MM, Newman S. 1998. Forms of soil phosphorus in selected hydrologic units of the Florida Everglades. Soil Sci Soc Am J 62:1134–47.
- Reddy KR, White JR, Wright A, Chua T. 1999. Influence of phosphorus loading on microbial processes in the soil and water column of wetlands. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 249–73.
- Redfield AC. 1958. The biological control of the chemical factors in the environment. Am Sci 46:205–21.
- Rejmánkova E, Pope KO, Pohl MD, Rey-Benayas JM. 1995. Freshwater wetland plant communities of northern Belize: implications for paleoecological studies of Maya wetland agriculture. Biotropica 27:28–36.
- Rejmánkova E, Pope KO, Post R, Maltby E. 1996. Herbaceous wetland of the Yucatan Peninsula: communities at extreme ends of environmental gradients. Int Rev Ges Hydrobiol 81: 223–52.
- Richardson CJ. 1985. Mechanisms controlling phosphorous retention capacity in freshwater wetlands. Science 228:1424–6.
- Richardson CJ. 1999. The role of wetlands in storage, release, and cycling of phosphorus on the landscape: a 25-year retrospective. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 47–68.
- Richardson CJ, Ferrel GM, Vaithiyanathan P. 1999. Nutrient effects on stand structure, resorption efficiency, and secondary compounds in Everglades sawgrass. Ecology 80:2182–92.
- Richardson CJ, Marshall PE. 1986. Processes controlling movement, storage, and export of phosphorus in a fen peatland. Ecol Monogr 56:279–302.
- Richardson CJ, Qian S, Craft CB, Qualls RG. 1997. Predictive models for phosphorus retention in wetlands. Wetlands Ecol Manage 4:159–75.
- Richardson CJ, Vaithiyanathan P. 1995. Phosphorus sorption characteristics of Everglades soils along a eutrophication gradient. Soil Sci Soc Am J 59:1782–8.
- Rudnick DT, Chen Z, Childers DL, Boyer JN, Fontaine TD III. 1999. Phosphorus and nitrogen inputs to Florida Bay: the importance of the Everglades watershed. Estuaries 22:398–416.
- Rutchey K, Vilchek L. 1999. Air photointerpretation and satellite imagery analysis techniques for mapping cattail coverage in a northern Everglades impoundment. Photogram Eng Remote Sens 65:185–91.
- Schindler DW. 1977. Evolution of phosphorus limitation in lakes. Science 195:260–2.
- Scinto LJ. 1997. Phosphorus cycling in a periphyton-dominated freshwater wetland [dissertation]. Gainesville (FL): University of Florida,
- Short FT, Dennison WC, Capone DG. 1990. Phosphorus-limited growth of the tropical seagrass *Syringodium filiforme* in carbonate sediments. Mar Ecol Prog Ser 62:169–74.
- Smith VH. 1998. Cultural eutrophication of inland, estuarine, and coastal waters. In: Pace ML, Groffman PM, editors. Successes, limitations, and frontiers in ecosystem science. New York: Springer-Verlag, p 7–49.
- [SFWMD] South Florida Water Management District. 2000. Everglades consolidated report. West Palm Beach (FL): South Florida Water Management District.
- [SFWMD] South Florida Water Management District. 1999. Ev-

erglades interim report. West Palm Beach (FL): South Florida Water Management District.

- Sterner RW, George NB. 2000. Carbon, nitrogen, and phosphorus stoichiometry of cyprinid fishes. Ecology 81:127–40.
- Steward KK, Ornes WH. 1975. The autecology of sawgrass in the Florida Everglades. Ecology 56:162–71.
- Steward KK, Ornes WH. 1983. Mineral nutrition of sawgrass (*Cladium jamaicense* Crantz) in relation to nutrient supply. Aquat Bot 16:349–59.
- Stober J, Scheidt D, Jones R, Thornton K, Gandy L, Stevens D, Trexler J, Rathbun S. 1998. South Florida ecosystem assessment. Athens (GA): United States Environmental Protection Agency.
- Turner AM, Trexler JC, Jordan CF, Slack SJ, Geddes P, Chick JH, Loftus WF. 1999. Targeting ecosystem features for conservation: standing crops in the Florida Everglades. Conserv Biol 13:898–911.
- Urban NH, Davis SM, Aumen NG. 1993. Fluctuations in sawgrass and cattail densities in Everglades Water Conservation Area 2A under varying nutrient, hydrologic, and fire regimes. Aquat Bot 46:202–23.
- Vaithiyanathan P, Richardson CJ. 1998. Biogeochemical characteristics of the Everglades sloughs. J Environ Qual 27:1439–50.
- Vaithiyanathan P, Richardson CJ. 1999. Macrophyte species changes in the Everglades: examination along a eutrophication gradient. J Environ Qual 28:1347–58.
- Vaithiyanathan P, Richardson CJ. 1997. Nutrient profiles in the Everglades: examination along the eutrophication gradient. Sci Total Environ 205:81–95.
- van der Valk AG, Rosburg TR. 1997. Seed bank composition along a phosphorous gradient in the northern Florida Everglades. Wetlands 17:228–36.
- Vitousek PM, Howarth RW. 1991. Nitrogen limitation on land and in the sea: how can it occur? Biogeochemistry 13:87–115.
- Vymazal J, Craft CB, Richardson CJ. 1994. Periphyton response to nitrogen and phosphorus additions in Florida Everglades. Algolog Studies 73:75–97.
- Vymazal J, Richardson CJ. 1995. Species composition, biomass, and nutrient content of periphyton in the Florida Everglades. J Phycol 31:343–54.
- Walbridge MR. 1991. Phosphorus availability in acid organic soils of the lower North Carolina coastal plain. Ecology 72: 2083–100.
- Walker WW. 1991. Water quality trends at inflows to Everglades National Park. Water Resources Bull 27:59–72.
- Walker WW Jr. 1999. Long-term water quality trends in the Everglades. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 447–66.
- Webster JR, Benfield EF. 1986. Vascular plant breakdown in freshwater ecosystems. Ann Rev Ecol System 17:567–94.
- Wetzel RG. 1990. Land–water interfaces: metabolic and limnological regulators. Verh Int Verein Limnol 24:6–24.
- Wetzel RG. 1999. Organic phosphorus mineralization in soils and sediments. In: Reddy KR, O'Connor GA, Schelske CL, editors. Phosphorous biogeochemistry in subtropical ecosystems. Boca Raton (FL): Lewis Publishers, p 225–48.
- White JR, Reddy KR. 1999. Influence of nitrate and phosphorus loading on denitrifying enzyme activity in Everglades wetland soils. Soil Sci Soc Am J 63:1945–54.
- Wu Y, Sklar FH, Rutchey K. 1997. Analysis and simulations of fragmentation patterns in the Everglades. Ecol Appl 7:268–76.

Appendix. Survey of Literature Included in the Meta-analysis

No.	Study	Soil Depth	Date	Notes on Interpretation and Usage
1	Steward and Ornes 1975	NR	1971	
2	Davis 1991	n/a	1975–80	Only live aboveground tissues included. <i>Cladium</i> = lowest SRP, <i>Typha</i> = highest SRP.
3	Walker 1991	n/a	1977-89	Median concentration.
4	Koch and Reddy 1992	5 cm	1990	<i>Typha/Cladium</i> reported as range. Macrophyte data is an average of leaves and roots.
5	Amador and Jones 1993	10 cm	NR	<i>Typha</i> = "high." <i>Cladium</i> = "low and intermediate."
6	Craft and Richardson 1993a	1964 or 10 cm	1989	Soil sampled down to peak ¹³⁷ Cs depth (accumulation since 1964), or if no accumulation, 10 cm.
7	Craft and Richardson 1993b	1964	1990	Soil sampled down to peak ¹³⁷ Cs depth (1964). <i>Typha</i> = St. 1, <i>Typha/Cladium</i> = 2–3, <i>Cladium</i> = 4–6.
8	Grimshaw and others 1993	n/a	1978–79	<i>Typha</i> = $B1-2$, <i>Cladium</i> and slough/wet prairie = $B5-10$
9	Reddy and others 1993	1964	1990	Soil TP load and N:P data sampled down to peak ¹³⁷ Cs depth (1964).
10	Urban and others 1993	n/a	1986–91	All sites classified as Typha/Cladium.
11	Coale and others 1994	n/a	1988-90	Only data from typical (fast) drainage practice included.
12	DeBusk and others 1994	10 cm	1990 ND	
13	Diaz and others 1994	n/a	NR	Control plate only
14	Vymazal and others 1994 Craft and others 1995	n/a	1990	Control plots only.
15	Craft and others 1995	30 cm	1990	Control plots only. Macrophyte aboveground biomass is an average weighted by species biomass. Slough macrophyte average includes <i>Utricularia</i> with associated periphyton. Soil depth = 15 cm in slough/ wet prairie.
16	Vymazal and Richardson 1995	n/a	1991–92	Range is for detached and attached periphyton, through time.
17	McCormick and O'Dell 1996	n/a	1995	Water data presented in #20. E1, E2, F2 = <i>Typha</i> . E3, F3, E4, F4 = <i>Typha/Cladium</i> . Other sites = <i>Cladium</i> and slough/wet prairie.
18	McCormick and others 1996	n/a	1994–95	Site classification identical to #19.
19	Doren and others 1997	25 cm	NR	<i>Typha/Cladium</i> = both <i>Typha</i> and <i>Cladium</i> have more than 25% frequency.
20	Grimshaw and others 1997	n/a	1994–95	"Unenriched slough" in Table 1 described as slough and <i>Cladium</i> = slough/wet prairie.
21	Newman and others 1997	10 cm	1991	Interior plots = <i>Cladium</i> and slough/wet prairie
22	Scinto 1997	9 cm	1993–95	Soil TP averaged over surface 9 cm.
23	Vaithiyanathan and Richardson 1997	2.5 cm	1993–95	Sites less than 2 km downstream = <i>Typha</i> ; sites betweer 2 and 6 km = <i>Typha/Cladium</i> ; sites more than 6 km = <i>Cladium</i> .
24	Craft and Richardson 1998	n/a	NR	Accretion data includes both ¹³⁷ Cs (1964) and ²¹⁰ Pb estimates (1962).
25	Miao and Sklar 1998	NR	1994–95	<i>Typha/Cladium</i> reported as range. Macrophyte data are an average of leaf, root, shoot base, and rhizome, weighted by relative biomass.
26	Vaithiyanathan and Richardson 1998	5 cm	1993–96	Range in macrophyte concentration is the range of species means (<i>Nymphaea</i> = average of petiole and leaf).
27	Daoust and Childers 1999	n/a	1995–96	<i>Cladium</i> datum is an average of wet and dry seasons. Average was weighted by species-relative biomass.
28	Jensen and others 1999	5 cm	1996	<i>Typha/Cladium</i> = both species denser than "nearby" or "sparse."
29	Richardson and others 1999	n/a	1990–96	Only <i>Cladium</i> data published for <i>Typha/Cladium</i> habitat. Soil data already published, not included.
30	Rudnick and others 1999	n/a	1984–95	Flow-weighted average concentration.
31	Turner and others 1999	10 cm	1996	WCA-2A soils data not used, reported in #12.
32	Vaithiyanathan and Richardson 1999	5 cm	1993–95	<i>Typha</i> = P-enrichment category 4–5. <i>Typha/Cladium</i> = $1-3$, <i>Cladium</i> and slough/wet prairie = 0 .

NR, not reported; n/a, not applicable