Key words: *hepatitis B virus/nucleotide sequence/divergence* 

## **The Complete Nucleotide Sequence of the Genome of a Hepatitis B Virus Isolated from a Naturally Infected Chimpanzee**

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## **SUMMARY**

The complete nucleotide sequence of a strain of hepatitis B virus, originally isolated from a naturally infected chimpanzee, has been determined. Interesting features of the sequence include the presence of an in-phase stop codon in the 'pre-core' region of the core antigen open reading frame. The sequence shows approximately  $10\%$  nucleotide divergence from all of the other hepatitis B virus sequences previously published and the possibility that this divergence is the result of passage through chimpanzees is discussed.

The occurrence of naturally acquired hepatitis B virus (HBV) infection in the colony of chimpanzees of the Zoological Society of London has been reported previously (Zuckerman *et al.,* 1978). One of the animals, chimpanzee K, was persistently infected with a high titre of hepatitis B surface antigen (HBsAg), and was also positive for both hepatitis B e antigen (HBeAg) and circulating virus. The subtype of the surface antigen was *adw.* The mother of this animal died from natural causes in February, 1973, and we have been unable to test her serum. However, a female sibling was also HBsAg-positive and it is possible that both became infected perinatally. We describe here the cloning and sequencing of the genome of this virus.

Virus was pelleted from the plasma of chimpanzee K and further purified by sedimentation through sucrose. Virus DNA was purified by standard methods following completion of the plus strand by the endogenous DNA polymerase. Because the DNA was found to lack the single *EcoRI* site common to most strains of HBV and a convenient single-cut enzyme was not readily identifiable, it was cloned as two separate *PstI* fragments of 1758 and 1424 nucleotides (nt) into pBR328. The *PstI* fragments were later subcloned into M 13mp 19 so that clones containing both orientations of the fragments were obtained. DNA sequencing was performed using the chain termination method (Sanger *et al.,* 1977). A nested set of templates was obtained from each clone by digestion with exonuclease III according to the method of Henikoff (1984). Any gaps in the sequence were filled by subcloning and sequencing specific restriction enzyme fragments, so that a complete sequence was obtained for both strands of the genome.

The nucleotide sequence of this strain of hepatitis B virus is shown in Fig. 1. The sequence reveals the four major open reading frames of HBV (surface, core, polymerase and X genes as indicated) and is numbered from the ATG of the HBcAg open reading frame (Fig. 2). The *PstI*  sites used in the cloning process are located at nucleotide positions 1305 and 3063. We believe that the two individual sequences are contiguous because the junctions result in intact genes (for HBsAg/P and X respectively), the sequences across the junctions are homologous to **other**  published sequences and the total number of nucleotides (3182) is in agreement with that found for other subtypes of HBV.

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A L E 8 P E 8 C S P N H I A L R Q A I L C W G E L M T L A S W V G N N L E D P A GCCT T AGAGTCI C CAGAGCAT TGT TCAC{3T AACCAIACAGCACT TAGGCAAGCTATA{3IGT GCIGGGGIGAGT f AATGACT C TGGCCTCCTGGGTGGGCAA I AA I I T GGAAGAICCAGCA 130 140 150 16{3 17{3 180 190 200 210 220 230 240

S R E Q V V N Y V N T N M G L K I R Q L L W F H I S C L T F G R E T V L E Y L V<br>TCCAGGGAACAAGTAGTTAATTATGTCAATACCAATATGGGTTTAAAGATCAGACAATTATTGTGGTTTCATATTTCCTGTCTTACTTTTGGAAGAGAAACTGTCCTTGAGTATTTGGTG 250 260 270 280 290 300 310 320 330 340 350 360

Polymerase M P L S Y Q H F R K L L L L D D E A G P L £ E E L S F G V W I R T P P A Y R P P N A P I L S f L P E f T V V R R R G R S P R R R f I CT T 1 T GCAGT C TGCA T I CGCACT C CCCCCGCI T ATAGACCACCAAA I GC{3CCI A TCT T ATCAACACT f CCCGAAACTA{3T G IT GI TAGACGACGAGGCACGTCCCCIAGAAGAAGAACT 370 38{3 390 400 410 42{3 450 440 450 460 470 480

P R L A D [ G L N R R V A E D L N L Q L P N V S I P W l H K V G N F T G L Y S 5 P S P R R R R S Q S P R R R R S Q 5 P A S Q C \* Cone C••TCGCCTC•CA•A{3GAA••TCTCAATCG•CGCGICG•AGAAGATCTCAATCTC{3AGCTTCC{3AA•GTTAGTATTCCT fGGACTCAIAAGGTGGGAAATTITACTGGGCITTATICTTC 490 500 510 520 530 540 550 560 570 580 590 600

T L P V F N P N W Q T P S F P D I H L H Q D I I N K C E Q F V G P L T V N E K R<br>TACTCTACCTGTCTTTAACCCTAACTGCCAAACTCCTTCTTTCCCTGATATTCATTTGCACCAAGATATTATTAACAAGTGTGAACAATTTGTGGGCCCTCTTAACGGTTAATGAAAAAAG 610 62{3 630 640 650 660 67{3 680 690 700 710 728

R L K L S M P A R F Y P N S T K Y L P L E K G I K P Y Y P D N V V N H Y F Q T R<br>AAGAITGAAGTTAAGTATGCCTGCCAGATTTTATCCAAATTCTACCAAGTATTTGCCTCTAGAGAAGGCTATAAAACCCTATTATCCAGATAATGTAGTTAATCATTACT 730 740 750 76{3 770 780 79{3 80{3 810 820 830 848

H Y L H T k w Q A G I L Y K R 0 T 7 R 5 A 8 F C G S P Y S W E Q £ L Q H G A E 5 N G Q N = Sur face = M G Q N<br>ACACTATTITACATACCAGGCAGAAACCATTITATATAAGAGAGAAACCAGAGGCGCTTCATTCTGGGTCACCATATTCTTGGGAACAAGAGCTACAGC<u>ATG</u>GGGCAGAATC

850 860 870 880 890 900 91{3 920 *950* 940 950 960

FH Q Q S A G IFSR A P V G S S I Q S K H Q Q S R L G L Q P Q K Q L L A R G N<br>L S T S N P L G F F P E H Q L D P A F K A N T N N P D W D F N P A K D Y W A L L A R G N<br>TTTCCACCAATCCGCGGATTTTTTCCCGAGCACCAGTTGGATCCAGCATTCGAACCAA 97{3 980 990 1000, 1010 1020 1 {33{3 104{3 1050 1060 1070 108{3

E G R S W S V R 5 R V H P T T W R S F G V E P S S S G H T N N F A S K S A S C L<br>T K V G A G A F G P G F T P P H G G L L G L S P Q A Q G I L T T L P A A P P P A G L L T T L P A N P P P A G L L<br>CCAAGGTAGEGEGEGEGEGEGEGEGEGEGE

H Q S V V R K A A Y P T F S T T K R H S 5 8 G H A V E L H N I S S S S A G S Q 5 S T N R 0 S G R Q P f P L S P P L R D T H P Q A M Q W N S T T F H Q A L Q 0 P R C CAC{3AAICGCCAG T CACGAAGGCAGCCTACCCCACT T TCTC{3A{3CACf AAGACACACTCA T CCTCAGGCCATGCA{3TGCAACTCCACAACAT T T CAT{3AAGCTCTG{3AGGAT CCCAGAG 121{3 1220 1230 1240 1250 1260 1270 1280 1290 1300 131{3 1320

K G P V F S C W W L Q F R N I E P C S E Y C L S H L V S L L D D W G P C T E H G<br>V R G L Y F P A G G S S S S C T L N P V P N T A 5 H I S S V F S T T G D P A P MAGGCCITATITICTICCIGGCA COLOCAGI<br>TAAGGCCITGTATITICTICCIGGCIGCCIC

E H H I R I P R T P A R V T G G V F L V 0 K N P H N T A E S R L V V D F S Q F 5 E N I T 5 G F L G P L L V L Q A G F F L L T K I L T I P Q S L D S W W T S L N F A GAACATCACATCA••ATT{3CTAGGACCCCT•CTCG•G•IACAGGCGGGGTITTTCTTGTTGACAAAAA•CC•CACAATACCGCA{3AGTC•AGACTCGTGGTGGACTTCTCTCAAT••TC 145{3 146{3 1470 1480 1490 15{30 151 {3 1520 1530 1540 1550 1560

R G S T R V P W P K F A V P N L Q P L T N L L S S N L S W L S L D V S A A F Y H<br>L G G A P V C L G Q N S Q S P T S N H S P T S C P P I G C Y R W M C L R R F I<br>TAGGGGGACACCOCTGTICACCA AATICOCCACACCOCCCACCACCOCCOCCACCOCCOCCOC

LPLHPAAMPHLLVGSSGLSRYVARLSSNSRILDHQHGTMQ<br>IFLFLFILQLCITATIONS(ITTLLVLLDYQGINIAAGGATATIONS)<br>IFLFICTIONS(CONTAGTEDITIONS) 1790 GMLPHONS (ITTLEIN) 1790 GMLPGSSTINIA<br>1690 1700 1710 1710 1720 1730 1740 1750 1760 1770 1780 1790

NLHDSCSRNLFDSLMLLYKTF{3RKLHLYSHPIIMGFRKIP KTCTTPAQGTSLIPSC{3CTKPSBGNCTCIPIPSSWAFAKF AAACITG•A••A{3T{3CT•{3TCAAG•AACC••T•T•AT•CCC•CATG•T•CT•TA•AAAACCITC•GA{3••AAA•TG•ACCT•TA•TC•CA•C{3CA•CA•{3A••••CTITC•CAAAA•TCC 1810 182{3 1830 1840 1850 186{3 1870 1880 1890 1980 1910 1920

MGVGLSPFLLAQFYSAICSVVRRAFPHCLAFSYMDDVVLG LWOWASVRFSWLSLLAPFVQWFACLSPTVWLLAIWMMWYW TATGG~GTG~G{3CTCAGT~CG~TTCTCC~GCT~A~TTTACTA~CG{3CA~TTGI~CA~TGG~T~GCAG~CT~T{3CC{3CA~G~T~CT~TTAGCTATAT~GATGATGTGGTATTGGG 1930 1940 1950 1960 197{3 1980 *199{3* 2000 2010 2020 2030 2040

AKS VQHHE SLYTA VTNFLLSLGIHLNPNKTKRWGYSLHFM<br>GPNLYNILSPFIPI PLLSPFIPI (PIFTCLWVYI \* Surface<br>GGCCAAAFCTETAGAACETTGAGETCCTITATACCCTGTIACCAHTTECTITICGTITECGTITATACETTAGAACEAAACEAAACECTGGGGTIATTCCCTACATTTCAT<br>2050 2050 2060 2070

GYVIGSWGTLPQEHIVQKIKN{3FRKLPVNRPIDWKVCQRI GGGT•ATGTAATTGG{3AGT•GGG••ACA•TACCA•AAGAACATATTGTACAAAAAATCAAAAATTGTT•CAGAAAA{3••CCTGT•AACAGACCTA•AGA•TGGAAAGTATGTCAAAGAAI 2170 2180 2190 22{330 221{3 2220 2230 224{3 2250 2260 2270 2280

VCLLGFAApFTQ{3GYPALMPLYACIQAKQAFTFSPTYKAF TGTGGGTCT~TTGGGATTTGCT~CCCC~TTTACGCAATGTGGTTATCCT~CGTTAATGCCA~TATGCATGTATACAA~CAAAACA~GC~TCACT~TCTCGCCAAC~ATAAGGCCT~ 2290 2300 2310 2320 2330 2340 2350 2360 2370 2380 2390 2400

LSQQYSTLYPVARQRSGLCQVFADATPTGWOLVMGHQRMR TCTAAGTCAACAA•MTCGA•C•T•TACCCCGTTGCCC•GCAACGGTCCGGTCTGTGCCAAG•{3•TTGCTGACGCAACCC{3CACTGGCTGGGGC•TG•TCATG•GCCATCAGC•CMGCG 2410 2420 2430 2440 2450 2460 2470 2480 2490 2500 2510 2520

GIFVAPLPIHTAQLLAACTCCGAT.<br>TGGAACCTTTGTGGCTCCTCTGCCGATCCATACTGCGGAACTCCTAGCAGCTTGTTTTGCTCGCAGCCGGTCTGGAGAAACTTATCGGAACTGACAATTCTGTCGTCCTCCCGGAA 2530 2540 2550 2560 2570 2580 2590 2600 2610 2620 2630 2640 Y T S F P W L L G C A A N W I L R G T S F V Y V P S A L N P A D D P S R G R L G<br>X Gene M A A R L C C Q L D T S R D V L C L R P V G A E S C G R P F S G P L R<br>ATATACATELTITICCATEGECING CONTEGERATE CONTEGES CONTECTING CONTEGE LYRPLIRLLFQPTTGRTSLYAVSPSVPSHLPVRVHFASPL ALPPSHPSALPTDYGAHLSLRGLPVCAFSSAGPCALRFTS •C•CIACCGCCCICTCATCCGTCTGCT•TTCCAACCGACTACGGGGCG•A•CT••CTITACGCGG•C•CCCCG•CTG•GCCTTCTCATCTGCCGG•CCGTGTGCACTTCGCTI•ACCTCT 2770 2780 2790 2800 2810 2820 2830 2840 2850 2860 2870 2880 H V A W R P P \* Polymerase<br>A R C M E T T V N A P R N L P K V L H K R T L G L S A M S T T K I E T Y F K D C<br>DCACGTIGCAIGGAGACCIGCAGGACCICCCCAGCGAGAGAGACCICCCATAAGAGGACCACTITICGACTITICAGCAATGICAGCACCACCAACGATIGCATACTICAAACAC VFKDWEELGEEIRLKVFVLGGCRHKLVCTPAPCNFFISA\* ~TAIITAAGGACTGGGAGGAGCTGGGGGAGGAGA~TAGG~AAAGGTCTT~QTATTAGGA~GCTG~AGGCA~AAAT~GGTC~G~ACACCAGCACCA~GCAAC~TT~TCACCT~TGCCIAG 3010 3020 3030 3040 3050 3060 3070 3080 3090 3100 3110 3120 X Gene<br>TCATCTCATGTTCATGTCCTACTGTTCAAGCCTCCAAGTTGTGCCTTGGGTGCCTTTAGGGC<br>3130 3140 3150 3160 3170 3170 31780

Fig. 1. The complete nucleotide sequence of the LSH strain of HBV. Position 1 is the start of the core region. The predicted amino acid sequences of the four major open reading frames are shown above the nucleotide sequence. The following sequences are underlined: the stop codon in the pre-core region (nt 3177), the start of pre-S1 (nt 948), pre-S2 (nt 1272) and the major sAg gene (nt 1437) and the direct repeats DR1 (nt 3106 to 3116) and DR2 (nt 2872 to 2882).

The sequence includes most of the expected features of the HBV genome. The 11 bp direct repeats, which are believed to be central to the replication strategy of the virus (for review, see Ganem & Varmus, 1987), are located at positions 2872 and 3106. The enhancer element (TGTTTGCT; Shaul & Ben-Levy, 1987) is located at position 2463. A consensus sequence which may function as a specific glucocorticoid receptor site (CAANNTGTYCT; Tur-Kaspa *et al.,* 1986) is located at nucleotide position 1636. The extended TATA box (TATATAA) which is presumed to be the pre-S1 promoter (Cattaneo *et al.,* 1987) is located at position 876. A motif with homology to the simian virus 40 late promoter, which is presumed to be the promoter for the major 2-1 kb surface antigen transcript (Cattaneo *et al.,* 1983), is located around position 1225. A signal believed to be involved in the termination of all HBV transcripts (TATAAA) is located at position 16 and a second signal which may be involved in the cleavage/polyadenylation reaction (Hart *et al.,* 1985; Renan, 1987) is located at position 43 to 56.

The amino acid sequences deduced for the four main open reading frames are also shown in Fig. 1 and a schematic diagram of the genome organization forms Fig. 2. The *adw* subtype may be confirmed from the sequence of the major HBsAg protein, lysine at amino acid position 122 (nt 1801) confirms the d subtype and lysine at amino acid position 160 (nt 1915) the w subtype (Okamoto *et al.,* 1987). A six base pair insertion (which would occur between nt 455 and 456 in our clone) is present in both the *adw-1* (Valenzuela *et al.,* 1980) and *adw-2* (Ono *et al.,* 1983) sequences but not in this sequence, despite the fact that the *adw-2* sequence is otherwise conserved for around 40 base pairs either side of this position. The region encoding the pre-S1 domain of HBsAg displays considerable size variation and the sequences of this clone and others reported in the literature are compared in Fig. 3. It may be seen that the size of the deletion in this clone is identical to that of the *ayw* sequence reported by Galibert *et al.* (1979) and the amino-terminal amino acid sequence of pre-S1 most closely resembles that of the *ayw* subtype. Comparison of overall nucleotide homology with other strains (Table 1) shows 9.4 to  $10\%$ nucleotide differences compared to other published HBV sequences. These other sequences themselves fall into three groups : *adw-1* and *adw-2; ayw, ayw-2* and *adyw* [the *adyw* sequence was derived from a mixed clone bank (Burrell *et al.,* 1979) and has the characteristics of the *ayw* subtype according to the definition of Okamoto *et al.* (1987)]; and *adr, adr-4* and *ayr* (references for these sequences are given in Table 1).

Some of the effects of these changes on the amino acid sequences of the HBV proteins are noteworthy. Near to the carboxyl terminus of the core protein the sequence Pro-Ala (nt 535)



Fig. 2. A diagrammatic representation of the genome of the LSH strain of HBV. The inner circle shows the numbering system (clockwise, with the start of the HBcAg open reading frame as position 1) and the two *Pstl* sites used in the original cloning. The locations of the four major open reading frames are shown outside this circle, the broken lines preceding the HBcAg gene denote the position of the pre-core region with the in-phase stop codon.



Fig. 3. Comparison of the LSH sequence nt 942 to 962 with other HBV strains. Sequences are taken from Galibert *et al.* (1979; *ayw ),* Ono *et al.* (1983; *adw-2 ),* Valenzuela *et al.* (1980; *adw-1* ), Fujiyama *et al.*  (1983; *adr)* and Okamoto *et al.* (1986; *ayr).* 

Table 1. Percentage variability between the nucleotide sequence of the LSH strain of HBV and *other strains\** 

	$adw-1$	$adw-2$	avw	$a$ yw-2	adyw	adr	$adr-4$	ayr
adw-LSH	$9-8$	9.5	9.5	$9-4$	9.7	9.6	10.0	$10-0$
ayr	9.7	9.6	10-2	$10-3$	10.9	$2-4$	2.2	
$adr-4$	$9-4$	10.0	$10-5$	$10-7$	$11-4$	2.0		
adr	8.9	9.9	10-6	11.5	12.0			
adyw	11-0	$10-6$	$3-6$	$4-4$				
$a$ <i>yw</i> -2	11-0	10-1	$2 - 7$					
avw	10.8	9.9						
$adw-2$	1.8							

\* Comparisons are with the sequences of Valenzuela *et al.* (1980; *adw-1),* Ono *et al.* (1983; *adw-2* and adr), Galibert *et al.* (1979, *ayw ),* Bichko *et ul.* (1985, *ayw-2 ),* Fujiyama *et al.* (1983, *adr-4 ),* Okamoto *et al.* (1986, *ayr ),* and Will *et al.* (1982, *adyw).* 

replaces the Arg-Glu found in all other published sequences. These changes also affect the amino acid sequence of the polymerase. A protease-like sequence near the amino terminus of the core protein which may be involved in the generation of HBeAg (Miller, 1987) is conserved in the LSH strain (nt 79 to 111). The sequence of the major surface antigen is highly conserved when compared to other published sequences. As mentioned above, the pre-S1 amino acid sequence is found to be highly variable between different HBV strains and a number of unique changes occur in this region of the LSH strain. If the receptor-binding site for the hepatocyte is located in the pre-S1 region (Neurath *et al.,* 1986) it is conceivable that some of these changes may represent adaptation of the virus to the chimpanzee host. However, the pre-S2 region is also diverged in our strain and, taken together, these changes affect the amino acid sequence of the polymerase more than that of the pre-S proteins. The middle third of the polymerase is most diverged, in our strain, but the region that shows homology to other viral RNA-dependent DNA polymerases (Toh *et al.,* 1983) is highly conserved. A region of the X protein near the amino terminus (nt 2757 to 2803) which is somewhat variable among HBV strains is less conserved in the LSH sequence. The X protein appears to be a transcriptional *trans-activator* (Twu & Schloemer, 1987) and, again, it is possible that changes in this protein may be the result of adaptation to a chimpanzee host.

Atypically, the LSH sequence reveals an in-phase stop codon in the 'pre-core' region of the core antigen open reading frame (position 3177). In order to try to eliminate the possibility that we had cloned a defective virus we obtained a second clone of the 1424 nt *PstI* fragment from a different plasma sample from the same animal (taken over 12 months later) and sequenced it from the *PstI* site through the stop codon. The presence of the stop codon was confirmed and the sequence of that strand found to be identical to that in Fig. 1 over 250 nt.

The presence of this in-phase stop codon is particularly noteworthy. A cloned HBV genome with a stop codon in the pre-core region was found in The Netherlands to be non-infectious for chimpanzees (Will *et al.,* 1985). The clone, however, had other anomalies including a six base pair duplication at the end of the X gene which may interfere with its replication and render it defective. Chang *et al.* (1987) have introduced a frameshift mutation into the pre-core region of an avian hepadnavirus (duck HBV) and have shown that the resultant clone, when inoculated intrahepatically into newborn ducklings, gives rise to an apparently normal, productive infection. If both the pre-core and core start codons are present on a transcript of this gene, translation starts preferentially at the pre-core AUG (Weimer *et al.,* 1987). However, core antigen without the pre-core domain is the major product *in vivo* and this may reflect heterogeneity of the 5' ends of these transcripts, with the most abundant family of transcripts initiated in the pre-core region itself. The pre-core region resembles a signal sequence and it has been argued that it may be responsible for the secretion of HBeAg (Ou *et al.,* 1986; Roossinck *et al.,* 1986; Uy *et al.,* 1986) though, in this study, the chimpanzee was highly positive for HBeAg (using a monoclonal antibody-based assay; Sorin-Diagnostics, Italy). We have used site-directed mutagenesis to remove this stop codon from our clone (M. Donati, A. J. Wolstenholme & T. J. Harrison, unpublished observations) and plan to investigate the secretion of HBeAg by wildtype and mutant virus in eukaryotic expression systems. Although we cannot rule out the possibility that we have sequenced the genomes of two defective viruses, it appears likely that expression of the pre-core region is not required for infectivity in chimpanzees.

The sequence of this clone shows approximately  $10\%$  nucleotide differences compared to other published HBV sequences (Table 1) including the other clone containing a pre-core stop codon (Will *et al.,* 1982). It has been suggested that the chimpanzee responsible for infecting the colony may itself have been infected naturally in Africa (Zuckerman *et al.,* 1978) and this may be the first reported complete sequence of an African isolate of the virus. Alternatively, the sequence divergence may in part represent adaptation of the human virus to a chimpanzee host as discussed above. A sequence of the hepatitis delta virus cloned from an infected human (Makino *et al.*, 1987) was found to be  $11\%$  divergent from that of virus cloned from infected chimpanzees (Wang *et al.,* 1986; Kos *et al.,* 1986) and Makino *et al.* suggested adaptation to the chimpanzee host as one possible explanation of this divergence. Since HBV replicates via an RNA intermediate (Summers & Mason, 1982) it may have the high mutation rate typical of **RNA viruses. It is now clear that HBsAg subtype specificity depends upon relatively minor changes in protein sequence (Okamoto** *et al.,* **1987) and divergence of HBV nucleotide sequence may be otherwise largely independent of these specificities.** 

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## REFERENCES

- BICHKO, V., PUSHKO, P., DREILINA, D., PUMPEN, P. & GREN, E. (1985). Subtype *ayw* variant of hepatitis B virus. DNA primary structure analysis. *FEBS Letters* 185, 208-212.
- BURRELL, C. J., MACKAY, P., GREENAWAY, P. J., HOFSCHNEIDER, P. H. & MURRAY, K. (1979). Expression in *E. coli* of hepatitis virus DNA sequence cloned in plasmid pBR322. *Nature, London* 279, 43-47.
- CATTANEO, R., WILL, H., HERNANDEZ, N. & SCHALLER, H. (1983). Signals regulating hepatitis B surface antigen transcription. *Nature, London 305,* 336-338.
- CATTANEO, R., SPRENGEL, R., WILL, H. & SCHALLER, H. (1987). Molecular biology of hepatitis B. In *Primers in Developmental Biology,* vol. 2: *Molecular Genetics of Mammalian Cells,* pp. 271-295. Edited by G. M. Malacinski. London: Collier Macmillan.
- CHANG, C., ENDERS, G., SPRENGEL, R., PETERS, N., VARMUS, H. E. & GANEM, D. (1987). Expression of the precore region of an avian hepatitis B is not required for viral replication. *Journal of Virology* 61, 3322-3325.
- FUJIYAMA, A., MIYANCHARA, A., NOZAKI, C., YONEYAMA, J., OHTOMO, N. & MATSUBARA, K. (1983). Cloning and structural analysis of hepatitis B virus DNAs, subtype *adr. Nucleic Acids Research* 11, 4601-4610.
- GALIBERT, F., MANDART, E., FITOUSSI, F., TIOLLAIS, P. & CHARNAY, P. (1979). Nucleotide sequence of the hepatitis B virus genome (subtype *ayw)* cloned in *E. coli. Nature, London* 281, 646-650.
- GANEM, D. & VARMUS, H. E. (1987). The molecular biology of hepatitis B viruses. *Annual Review of Biochemistry 56,*  651-693.
- HART, R. P., McDEVITT, M. A., ALI, H. & NEVINS, J. R. (1985). Definition of essential sequences and functional equivalence of elements downstream of the adenovirus E2A and the early simian virus 40 polyadenylation sites. *Molecular and Cellular Biology* 5, 2975-2983.
- HENIKOFF, S. (1984). Unidirectional digestion with exonuclease III creates targeted break points for DNA sequencing. *Gene 28,* 351-359.
- KOS, A., DIJKEMA, R., ARNBERG, A. C., VAN DER MEIDE, P. H. & SCHELLEKENS, H. (1986). The hepatitis delta ( $\delta$ ) virus possesses a circular RNA. *Nature, London* 323, 558-560.
- MAKINO, S., CHANG, M-F., SHIEH, C=K., KAMAHORA, T., VANNIER, D. M., GOVINDARAJAN, S. & LAI, M. M. C. (1987). Molecular cloning and sequencing of a human hepatitis delta (6) virus RNA. *Nature, London* 329, 343-346.
- MILLER, R. H. (1987). Proteolytic self-cleavage of hepatitis B virus core protein may generate serum e antigen. *Science* 236, 722-725.
- NEURATH, A. R., KENT, S. B. H., STRICK, N. & PARKER, K. (1986). Identification and chemical synthesis of a host cell receptor binding site on hepatitis B virus. *Cell 46,* 429-436.
- OKAMOTO, H., IMAI, M., SHIMOZAKI, M.. HOSHI, Y., IIZUKA, H., GOTANDA, T., TSUDA, F., MIYAKAWA, Y. & MAYUMI, M. (1986). Nucleotide sequence of a cloned hepatitis B virus genome, subtype *ayr:* comparison with genomes of the other three serotypes. *Journal of General Virology* 67, 2305-2314.
- OKAMOTO, H., IMAI, M., TSUDA, F., TANAKA, T., MIYAKAWA, Y. & MAYUMI, M. (1987). Point mutation in the S gene of hepatitis B virus for a d/y or w/r subtypic change in two blood donors carrying a surface antigen of compound subtype *adyr* or *adwr. Journal of Virology* 61, 3030-3034.
- ONO, Y., ONDA, H., SASADA, R., IGARISHI, K., SUGINO, Y. & NISHIOKA, K. (1983). The complete nucleotide sequences of the cloned hepatitis B virus DNA: subtype *adr* and *adw. Nucleic Acids Research* 11, 1747-1757,
- ou, J. H., LAUB, O. & RUTTER, W. J. (1986). Hepatitis B virus gene function: the precore targets the core antigen to cellular membranes and causes the secretion of the E antigen. *Proceedings of the National Academy of Sciences*, *U.S.A.* 83, 1578-1582.

RENAN, M. J. (1987). Conserved 12-bp element downstream from mRNA polyadenylation sites. *Gene 60,* 245-254.

- ROOSSINCK, M. J., JAMEEL, S., LOUKIN, S. H. & SIDDIQUI, A. (1986). Expression of hepatitis B viral core region in mammalian cells. *Molecular and Cellular Biology* 6, 1393-1400.
- SANGER, F., NICKLEN, S. & COULSON, A. R. (1977). DNA sequencing with chain-terminating inhibitors. *Proceedings of the National Academy of Sciences, U.S.A.* 74, 5463-5467.
- **SHAUL, Y. &** BEN-LEVY, R. (1987). Multiple nuclear proteins in liver cells are bound to hepatitis B virus enhancer element and its upstream sequences. *EMBO Journal* 6, 1913-1920.
- SUMMERS, J. & MASON, W. S. (1982). Replication of the genome of a hepatitis B-like virus by reverse transcription of an RNA intermediate. *Cell* 29, 403-415.
- TOH, H., HAYASHIDA, H. & MIYATA, T. (1983). Sequence homology between retroviral reverse transcriptase and putative polymerases of hepatitis B virus and cauliflower mosaic virus. *Nature, London* 305; 827-829.
- TUR-KASPA, R., BURK, R. D., SHAUL, Y. & SHAFRITZ, D. A. (1986). Hepatitis B virus DNA contains a glucocorticoidresponsive element. *Proceedings of the National Academy of Sciences, U.S.A.* 83, 1627-1631.

- TWU, R-J. & SCHLOEMER, R. H. (1987). Transcriptional *trans-activating* function of hepatitis B virus. *Journal of Virology* 61, 3448-3453.
- UY, A., BRUSS, V., GERLICH, W. H., KOCHEL, H. G. & THOMSSEN, R. (1986). Precore sequence of hepatitis B virus inducing e antigen and membrane association of the viral core protein. *Virology* 155, 89-96.
- VALENZUELA, P., QUIROGA, M., ZALDIVAR, J., GRAY, P. & RUTTER, W. J. (1980). The nucleotide sequence of the hepatitis B viral genome and the identification of the major viral genes. In *Animal Virus Genetics,* pp. 57-70. Edited by B. Fields, R. Jaenisch & C. F. Fox. New York & London: Academic Press.
- WANG, K-S., CHOO, Q-L., WEINER, A. J., OU, J-H., NAJARIAN, R. C., THAYER, R. M., MULLENBACH, G. T., DENNISTON, K. J., GERIN, J. L. & HOUGHTON, M. (1986). Structure, sequence and expression of the hepatitis delta  $(\delta)$  viral genome. *Nature, Lo~tdon* 323, 508-514.
- WEIMER, T., SALFIELD, J. & WILL, H. (1987). Expression of the hepatitis B virus core gene *in vitro* and *in vivo. Journal of Virology* 61, 3109-3113.
- WILL, H., KUHN, C., CATrANEO, R. & SCHALLER, H. (1982). Structure and function of the hepatitis B virus genome. In *Primary and Tertiary Structure of Nucleic Acids and Cancer Research,* vol. 12, pp. 237-247. Edited by M. Miwa, S. Nishimura, A. Rich, D. G. Soell & T. Sugimura. Tokyo: Japan Scientific Society Press.
- WILL, H., CATTANEO, R., DARAI, G., DEINHARDT, F., SCHELLEKENS, H. & SCHALLER, H. (1985). Infectious hepatitis B virus from cloned DNA of known nucleotide sequence. *Proceedings of the National Academy of Sciences~ U.S.A.* 82, 891-895.
- ZUCKERMAN, A. J., THORNTON, A., HOWARD, C. R., TSIQUAYE, K. N., JONES, D. M. & BRAMBELL, M. R. (1978). Hepatitis B outbreak among chimpanzees at the London Zoo. *Lancet* ii, 652-654.

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