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Claudin Proteins in Human Cancer: Promising New Targets for Diagnosis and Therapy

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Abstract

The tight junction proteins claudins are abnormally regulated in several human cancers. In particular, claudin-3 and claudin-4 are frequently overexpressed in several neoplasias, including ovarian, breast, pancreatic, and prostate cancers. Although the exact roles of these proteins in tumorigenesis are still being uncovered, it is clear that they represent promising targets for cancer detection, diagnosis, and therapy. (Cancer Res 2005; 65(21): 9603-6)

Claudins Are Tight Junction Proteins

Tight junctions, together with adherens junctions and desmosomes, form the apical junctional complex in epithelial and endothelial cellular sheets. Adherens junctions and desmosomes are responsible for the mechanical adhesion between adjacent cells, whereas tight junctions are essential for the tight sealing of the cellular sheets, thus controlling paracellular ion flux and therefore maintaining tissue homeostasis (1). Tight junctions also play a crucial role in the maintenance of cell polarity by forming a fence that prevents lateral diffusion of membrane proteins and lipids, thereby maintaining the differential composition of the apical and basolateral domains. Finally, because of the ability of tight junction proteins to recruit signaling proteins (2), tight junctions have also been hypothesized to be involved in the regulation of proliferation, differentiation, and other cellular functions.

When observed by electron microscopy, tight junctions form multiple strands that seem to provide the structural basis for adhesion between adjacent cells (1). Tight junctions are composed of three major integral membrane proteins, occludin, claudins, and junctional adhesion molecules. Although the exact roles of these proteins are not completely clear, it seems that the claudins form the backbone of the tight junction strands. The claudin family of proteins is comprised of 23 members of closely related transmembrane proteins (see Fig. 1). Although the expression pattern of claudins is tissue specific, most tissues express multiple claudins, which can interact in both homotypic and heterotypic fashion to form the tight junction strands. It is believed that the exact combination of claudin proteins within a given tissue can determine the selectivity and strength of the tight junctions. Claudins are polymerized together within a given cell and can interact with the claudin of the adjacent cells to form an adhesive structure.

The high degree of cellular organization typically observed in normal differentiated tissues is often lost in cancer. Tumor cells

frequently exhibit abnormal tight junction function as well as decreased differentiation and cell polarity (3, 4). Loss of tight junction integrity may be particularly important in allowing the diffusion of nutrients and other factors necessary for the survival and growth of the tumor cells (5). In addition, decreased polarity and differentiation may be important for the metastatic phenotype, where individual cells must leave the primary site and enter the blood vessels to reach distant sites (6). Finally, the destruction of functional tight junctions in cancer may have a role in growth control. For example, in *Drosophila*, mutations in many tumor suppressor genes lead to loss of cell polarity and overproliferation of the epithelia (7). Based on the similarity between the vertebrate and *Drosophila* epithelia, mammalian cells are likely to require cytoarchitectural cues for cell growth control as well.

Claudin Expression in Cancer

The expression of occludin and claudins, the two major transmembrane proteins that contribute to formation of tight junctions, has been found to be altered in several cancers. An early study in the field showed that occludin was often down-regulated in gastrointestinal tumors (8). Similarly, other studies have shown that claudins are down-regulated in various cancers. For example, claudin-1 has been found to be reduced in breast cancer (9, 10) as well as in colon cancer (11). Claudin-7 has also been found down-regulated in invasive breast cancer (12) and in head and neck cancer (13). These reports of decreased tight junction protein expression in cancer are consistent with the generally accepted idea that tumorigenesis is accompanied by a disruption of tight junctions, a process that may play an important role in the loss of cohesion, invasiveness, and lack of differentiation observed in cancer cells. In addition to the down-regulation of protein levels, phosphorylation of tight junction proteins, including claudins, may affect tight junction function in cancer (14). For example, phosphorylation of claudin-1 by mitogen activated protein kinases (15) and protein kinase C (16), as well as phosphorylation of claudin-5 by cyclic AMP-dependent protein kinase (17, 18) have been reported. Also, WNK4 kinase has been shown to phosphorylate claudin-3 and claudin-4, and decrease tight junction function (19). Interestingly, phosphorylation of claudin-3 and claudin-4 in ovarian cancer cells has been shown to disrupt tight junctions (20).

Paradoxically, other studies have shown that certain claudin proteins are up-regulated in cancer. In fact, the overwhelming majority of the studies published thus far report an over-expression of claudins in various cancers (see Table 1). One of the first studies reporting this fact was a serial analysis of gene expression (SAGE) study of ovarian cancer showing that *CLDN3* and *CLDN4* (encoding claudin-3 and claudin-4, respectively) were among the most highly up-regulated genes in this cancer (21). Several additional reports have since confirmed the high

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expression of these two claudins in ovarian cancer (22–25). In addition, claudin-3 and claudin-4 have also been reported to be expressed in other cancers, such as breast (26), prostate (27), and pancreatic (28–32) cancers. Other claudins are differentially expressed in a number of human neoplasms and these data are summarized in Table 1.

Roles of Claudin in Cancer

As mentioned above, the loss of claudins and other tight junction proteins in cancer has been interpreted as a mechanism for the loss of cell adhesion and an important step in the progression of cancer to metastasis. Consistent with this hypothesis, a recent study showed that expression of claudin-4 in pancreatic cancer cells reduces invasiveness of these cells (33). In addition, claudin-1 reexpression in cancer cells can lead to increased apoptosis in three-dimensional cultures (34). On the other hand, as discussed previously, many claudins, such as claudin-3 and claudin-4, are typically up-regulated in many cancers (Table 1), suggesting that these proteins may have a positive effect on tumorigenesis. Recent work has shown that, at least in the case of ovarian cells, expression of claudin-3 and claudin-4 may lead to an increase in invasion, motility, and cell survival (35), all characteristics important for metastasis. Consistent with these *in vitro* findings is a report that claudin-4 expression in pancreatic

intraductal papillary mucinous neoplasms was associated with a more invasive phenotype (31). Similarly, expression of claudin-3 and claudin-4 was observed in advanced ovarian cancer but not in ovarian cystadenomas (22). Therefore, the functions of claudins may be highly tissue specific and may depend on the exact molecular circuitry of the cell.

Claudins as Diagnosis Markers and Therapeutic Targets

Because of the high specificity of claudin expression patterns in cancer, it has been suggested that claudins may represent useful molecular markers for many different cancers. For example, a set of four markers, including claudin-3, was found to be sufficient to accurately identify all 158 ovarian cancers tested, including eight early-stage serous cancers (24). In addition, claudin expression may be used as a prognostic indicator because low claudin-1 expression has been shown to be associated with a poor prognosis in stage II colon cancer (11). Claudin-10 expression has also been shown to be an independent prognostic factor for hepatocellular carcinoma recurrence after curative hepatectomy (36).

Interestingly, claudin-3 and claudin-4 are receptors for the *Clostridium perfringens* enterotoxin (CPE; ref. 37). CPE is a single

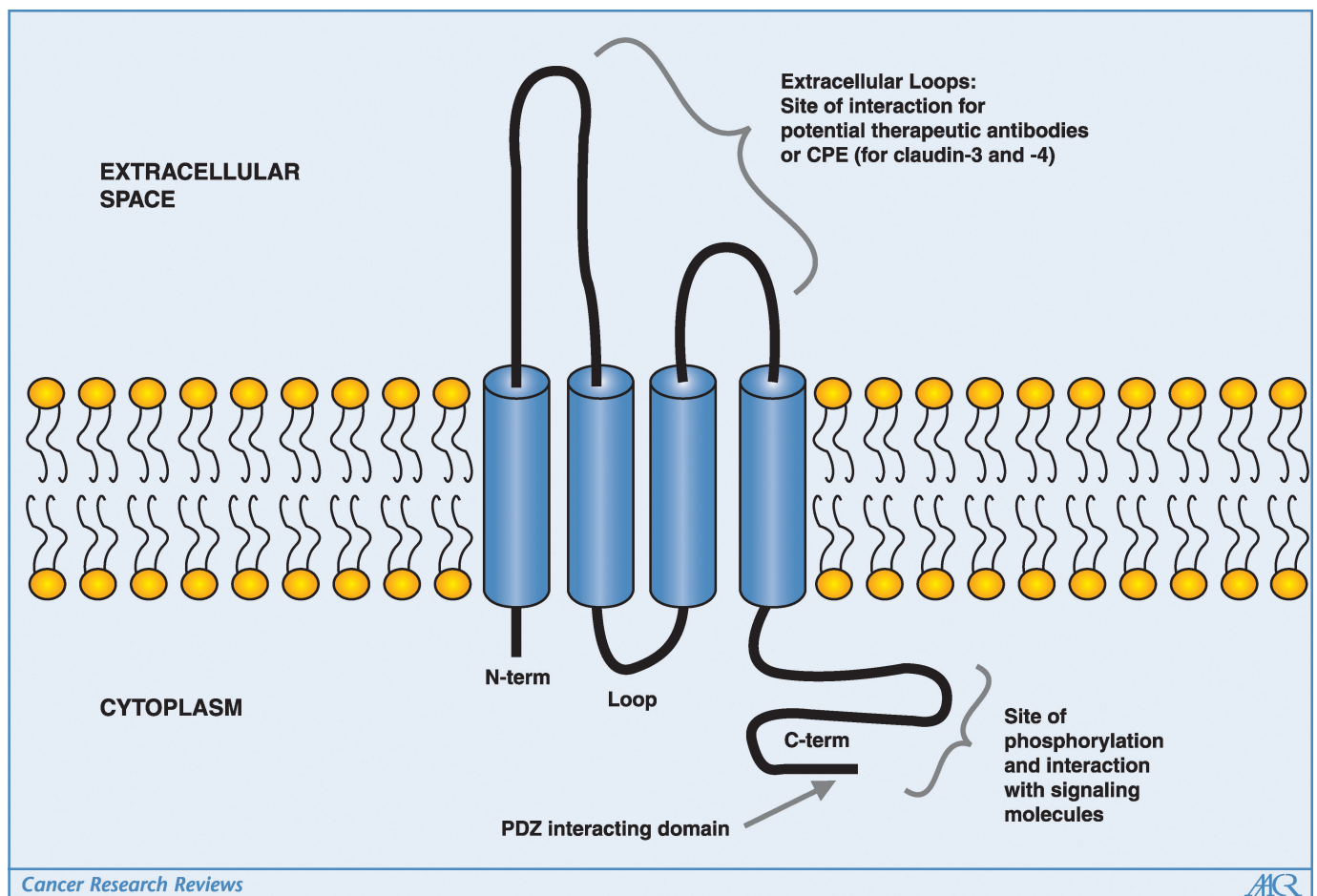


Figure 1. Structure of claudins. Claudin proteins are a family of proteins containing four transmembrane domains. The extracellular loops represent promising targets for therapy using monoclonal antibodies or, in the case of claudin-3 and claudin-4, the CPE. The COOH-terminal region of claudins is phosphorylated, contains a PDZ-binding domain, and has been implicated in signal transduction.

Table 1. Claudin expression in cancer

Cancer	Claudin gene	Expression	References
Breast	<i>CLDN1</i>	Down	(9)
	<i>CLDN7</i>	Down	(12)
	<i>CLDN1, CLDN3, CLDN4</i>	Variable	(10)
	<i>CLDN3</i>	Up	(26)
	<i>CLDN4</i>	Up	(26)
Breast and Paget's disease	<i>CLDN2, CLDN3, CLDN4, CLDN5</i>	Variable	(44)
Colon	<i>CLDN1</i>	Variable	(11)
Gastric	<i>CLDN4</i>	Down	(45)
Hepatocellular carcinoma	<i>CLDN10</i>	Up	(36)
Hepatocellular carcinoma (mouse model)	<i>Cldn7</i>	Up	(46)
Head and neck squamous cell carcinoma	<i>CLDN7</i>	Down	(13)
Squamous cell carcinoma	<i>CLDN1</i>	Up	(47)
	<i>CLDN4</i>	Up	(47)
	<i>CLDN3</i>	Up	(21, 24, 25, 48)
Ovarian	<i>CLDN4</i>	Up	(21, 23, 25)
	<i>CLDN16</i>	Up	(49)
	<i>CLDN4</i>	Up	(28–30, 32)
Pancreatic	<i>CLDN4</i>	Up	(31)
Pancreatic (intraductal papillary mucinous neoplasms)	<i>CLDN4</i>	Up	(31)
	<i>CLDN3</i>	Up	(27)
Prostate	<i>CLDN4</i>	Up	(27)
	<i>CLDN10</i>	Up	(50)
Thyroid papillary cancer	<i>CLDN10</i>	Up	(50)

polypeptide of 35 kDa, which, upon binding to its receptors, causes cytolysis through its effects on membrane permeability. High expression of claudin-3 and claudin-4 in multiple cancers may thus represent a unique opportunity for innovative therapy using CPE (38). Prostate adenocarcinoma cells expressing claudin-3 and claudin-4 have indeed been shown to be sensitive to CPE-mediated cytolysis (27). Specificity was evident as prostate cancer cells lacking claudin-3 and claudin-4 were unaffected by CPE treatment. Similar experiments established that breast (26), ovarian (39), and pancreatic (29) cancer cells are also sensitive to CPE treatment, provided that they express claudin-3 and/or claudin-4, as these cancers often do. Interestingly, human tumors grown as xenografts in immunocompromised mice could also successfully be treated using CPE, again on the condition of claudin-3 or claudin-4 expression (26, 29, 39). Importantly, these studies showed that no significant toxicity was encountered in mice upon intratumoral CPE treatment. However, claudin-3 and/or claudin-4 are expressed in several normal human tissues, including the gut, the lungs, and the kidneys (27). This expression pattern may represent a problem in the use of CPE for systemic cancer therapy, and it remains to be seen whether this approach will be useful in the clinic. Clearly, approaches that would involve regional application of CPE would be preferable. In addition, it has been suggested that a nontoxic, but claudin-specific, COOH-terminal CPE fragment (C-CPE; ref. 40) could be delivered locally to certain normal tissues and prevent CPE toxicity. Other potential problems with the use of CPE in tumor treatment include the occasional lack of surface claudin expression (22), the possibility of an immune response against CPE in treated patients as well as the penetration of CPE into the tumor mass. Additional studies will be required to clearly ascertain these issues.

The C-CPE fragment represents another potential opportunity for the treatment of claudin-3- and claudin-4-expressing tumors. Indeed, C-CPE could be used as a specific carrier for cytotoxic agents and, therefore, provide selective drug delivery. Addition-

ally, it has been suggested that, because C-CPE can destroy tight junctions (41), this peptide may be useful in combination therapy with conventional chemotherapeutic by increasing drug delivery to the interior of tumors. However, it seems that claudin-3 and claudin-4 expression is not necessarily associated with the formation of functional tight junctions in tumors and this approach may not be generally viable (22). Because claudins are transmembrane proteins and typically have two relatively large extracellular loops (see Fig. 1; ref. 42), these proteins may also offer promising targets for antibody-based therapy. Antibodies that specifically recognize different extracellular loops have been produced and shown to specifically bind claudins on the surface of the cell, providing a proof of principle for the approach (42).

The advent of gene expression profiling techniques has allowed the unbiased identification of genes that are differentially expressed in cancer. Although tight junction proteins have been studied for their role in tumorigenesis for many years, SAGE studies of breast (43) and ovarian (21) cancers allowed for the first time the identification of specific claudin family members as potential biomarkers for these cancers. Subsequent array analyses have confirmed these findings and also identified claudins as proteins frequently altered in cancer (see Table 1). These findings are important because the unusual expression patterns of claudins suggest utility for detection, diagnosis, and treatment of drug-resistant cancers. Although clinical trials will be required to establish this potential, basic research on claudins is likely to remain valuable for providing important insights into normal and neoplastic cellular physiology.

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References

1. Tsukita S, Furuse M, Itoh M. Multifunctional strands in tight junctions. *Nat Rev Mol Cell Biol* 2001;2:285-93.
2. Mitic LL, Anderson JM. Molecular architecture of tight junctions. *Annu Rev Physiol* 1998;60:121-42.
3. Weinstein RS, Merk FB, Alroy J. The structure and function of intercellular junctions in cancer. *Adv Cancer Res* 1976;23:23-89.
4. Soler AP, Miller RD, Laughlin KV, Carp NZ, Klurfeld DM, Mullin JM. Increased tight junctional permeability is associated with the development of colon cancer. *Carcinogenesis* 1999;20:1425-31.
5. Mullin JM. Potential interplay between luminal growth factors and increased tight junction permeability in epithelial carcinogenesis. *J Exp Zool* 1997;279:484-9.
6. Martin TA, Jiang WG. Tight junctions and their role in cancer metastasis. *Histol Histopathol* 2001;16:1183-95.
7. Wodarz A. Tumor suppressors: linking cell polarity and growth control. *Curr Biol* 2000;10:R624-6.
8. Kimura Y, Shiozaki H, Hirao M, et al. Expression of occludin, tight-junction-associated protein, in human digestive tract. *Am J Pathol* 1997;151:45-54.
9. Kramer F, White K, Kubbies M, Swisshelm K, Weber BH. Genomic organization of claudin-1 and its assessment in hereditary and sporadic breast cancer. *Hum Genet* 2000;107:249-56.
10. Tokes AM, Kulka J, Paku S, et al. Claudin-1, -3 and -4 proteins and mRNA expression in benign and malignant breast lesions: a research study. *Breast Cancer Res* 2005;7:R296-305.
11. Resnick MB, Konkin T, Routhier J, Sabo E, Pricolo VE. Claudin-1 is a strong prognostic indicator in stage II colonic cancer: a tissue microarray study. *Mod Pathol* 2005;18:511-8.
12. Kominsky SL, Argani P, Korz D, et al. Loss of the tight junction protein claudin-7 correlates with histological grade in both ductal carcinoma *in situ* and invasive ductal carcinoma of the breast. *Oncogene* 2003;22:2021-33.
13. Al Moustafa AE, Alaoui-Jamali MA, Batist G, et al. Identification of genes associated with head and neck carcinogenesis by cDNA microarray comparison between matched primary normal epithelial and squamous carcinoma cells. *Oncogene* 2002;21:2634-40.
14. Li D, Mrsny RJ. Oncogenic Raf-1 disrupts epithelial tight junctions via downregulation of occludin. *J Cell Biol* 2000;148:791-800.
15. Fujibe M, Chiba H, Kojima T, et al. Thr²⁰³ of claudin-1, a putative phosphorylation site for MAP kinase, is required to promote the barrier function of tight junctions. *Exp Cell Res* 2004;295:36-47.
16. Nunbhakdi-Craig V, Machleidt T, Ogris E, Bellotto D, White CL III, Sontag E. Protein phosphatase 2A associates with and regulates atypical PKC and the epithelial tight junction complex. *J Cell Biol* 2002;158:967-78.
17. Ishizaki T, Chiba H, Kojima T, et al. Cyclic AMP induces phosphorylation of claudin-5 immunoprecipitates and expression of claudin-5 gene in blood-brain-barrier endothelial cells via protein kinase A-dependent and -independent pathways. *Exp Cell Res* 2003;290:275-88.
18. Soma T, Chiba H, Kato-Mori Y, et al. Thr(207) of claudin-5 is involved in size-selective loosening of the endothelial barrier by cyclic AMP. *Exp Cell Res* 2004;300:202-12.
19. Yamauchi K, Rai T, Kobayashi K, et al. Disease-causing mutant WNK4 increases paracellular chloride permeability and phosphorylates claudins. *Proc Natl Acad Sci U S A* 2004;101:4690-4.
20. D'Souza T, Agarwal R, Morin PJ. Phosphorylation of claudin-3 at threonine 192 by pka regulates tight junction barrier function in ovarian cancer cells. *J Biol Chem* 2005;280:26233-40.
21. Hough CD, Sherman-Baust CA, Pizer ES, et al. Large-scale serial analysis of gene expression reveals genes differentially expressed in ovarian cancer. *Cancer Res* 2000;60:6281-7.
22. Rangel LBA, Agarwal R, D'Souza T, et al. Tight junction proteins claudin-3 and claudin-4 are frequently overexpressed in ovarian cancer but not in ovarian cystadenomas. *Clin Cancer Res* 2003;9:2567-75.
23. Hibbs K, Skubitz KM, Pambuccian SE, et al. Differential gene expression in ovarian carcinoma: identification of potential biomarkers. *Am J Pathol* 2004;165:397-414.
24. Lu KH, Patterson AP, Wang L, et al. Selection of potential markers for epithelial ovarian cancer with gene expression arrays and recursive descent partition analysis. *Clin Cancer Res* 2004;10:3291-300.
25. Santin AD, Zhan F, Bellone S, et al. Gene expression profiles in primary ovarian serous papillary tumors and normal ovarian epithelium: identification of candidate molecular markers for ovarian cancer diagnosis and therapy. *Int J Cancer* 2004;112:14-25.
26. Kominsky SL, Vali M, Korz D, et al. *Clostridium perfringens* enterotoxin elicits rapid and specific cytolysis of breast carcinoma cells mediated through tight junction proteins claudin 3 and 4. *Am J Pathol* 2004;164:1627-33.
27. Long H, Crean CD, Lee WH, Cummings OW, Gabig TG. Expression of *Clostridium perfringens* enterotoxin receptors claudin-3 and claudin-4 in prostate cancer epithelium. *Cancer Res* 2001;61:7878-81.
28. Gress TM, Muller-Pillasch F, Geng M, et al. A pancreatic cancer-specific expression profile. *Oncogene* 1996;13:1819-30.
29. Michl P, Buchholz M, Rolke M, et al. Claudin-4: a new target for pancreatic cancer treatment using *Clostridium perfringens* enterotoxin. *Gastroenterology* 2001;121:678-84.
30. Nichols LS, Ashfaq R, Iacobuzio-Donahue CA. Claudin 4 protein expression in primary and metastatic pancreatic cancer: support for use as a therapeutic target. *Am J Clin Pathol* 2004;121:226-30.
31. Sato N, Fukushima N, Maitra A, et al. Gene expression profiling identifies genes associated with invasive intraductal papillary mucinous neoplasms of the pancreas. *Am J Pathol* 2004;164:903-14.
32. Terris B, Blaveri E, Crnogorac-Jurcovic T, et al. Characterization of gene expression profiles in intraductal papillary-mucinous tumors of the pancreas. *Am J Pathol* 2002;160:1745-54.
33. Michl P, Barth C, Buchholz M, et al. Claudin-4 expression decreases invasiveness and metastatic potential of pancreatic cancer. *Cancer Res* 2003;63:6265-71.
34. Hoevel T, Macek R, Swisshelm K, Kubbies M. Reexpression of the TJ protein CLDN1 induces apoptosis in breast tumor spheroids. *Int J Cancer* 2004;108:374-83.
35. Agarwal R, D'Souza T, Morin PJ. Claudin-3 and claudin-4 expression in ovarian epithelial cells enhances invasion and is associated with increased matrix metalloproteinase-2 activity. *Cancer Res* 2005;65:7378-85.
36. Cheung ST, Leung KL, Ip YC, et al. Claudin-10 expression level is associated with recurrence of primary hepatocellular carcinoma. *Clin Cancer Res* 2005;11:551-6.
37. Katahira J, Sugiyama H, Inoue N, Horiguchi Y, Matsuda M, Sugimoto N. *Clostridium perfringens* enterotoxin utilizes two structurally related membrane proteins as functional receptors *in vivo*. *J Biol Chem* 1997;272:26652-8.
38. Michl P, Gress TM. Bacteria and bacterial toxins as therapeutic agents for solid tumors. *Curr Cancer Drug Targets* 2004;4:689-702.
39. Santin AD, Cane S, Bellone S, et al. Treatment of chemotherapy-resistant human ovarian cancer xenografts in C.B-17/SCID mice by intraperitoneal administration of *Clostridium perfringens* enterotoxin. *Cancer Res* 2005;65:4334-42.
40. Hanna PC, Wnek AP, McClane BA. Molecular cloning of the 3' half of the *Clostridium perfringens* enterotoxin gene and demonstration that this region encodes receptor-binding activity. *J Bacteriol* 1989;171:6815-20.
41. Sonoda N, Furuse M, Sasaki H, et al. *Clostridium perfringens* enterotoxin fragment removes specific claudins from tight junction strands: evidence for direct involvement of claudins in tight junction barrier. *J Cell Biol* 1999;147:195-204.
42. Offner S, Hekele A, Teichmann U, et al. Epithelial tight junction proteins as potential antibody targets for pancreatic cancer therapy. *Cancer Immunol Immunother* 2005;54:431-45.
43. Nacht M, Ferguson AT, Zhang W, et al. Combining serial analysis of gene expression and array technologies to identify genes differentially expressed in breast cancer. *Cancer Res* 1999;59:5464-70.
44. Soini Y. Claudins 2, 3, 4, and 5 in Paget's disease and breast carcinoma. *Hum Pathol* 2004;35:1531-6.
45. Lee SK, Moon J, Park SW, Song SY, Chung JB, Kang JK. Loss of the tight junction protein claudin 4 correlates with histological growth-pattern and differentiation in advanced gastric adenocarcinoma. *Oncol Rep* 2005;13:193-9.
46. Borlak J, Meier T, Halter R, Spänel R, Spänel-Borowski K. Epidermal growth factor-induced hepatocellular carcinoma: gene expression profiles in precursor lesions, early stage and solitary tumours. *Oncogene* 2005;24:1809-19.
47. Morita K, Tsukita S, Miyachi Y. Tight junction-associated proteins (occludin, ZO-1, claudin-1, claudin-4) in squamous cell carcinoma and Bowen's disease. *Br J Dermatol* 2004;151:328-34.
48. Heinzelmann-Schwarz VA, Gardiner-Garden M, Henshall SM, et al. Overexpression of the cell adhesion molecules DDR1, claudin 3, and Ep-CAM in metaplastic ovarian epithelium and ovarian cancer. *Clin Cancer Res* 2004;10:4427-36.
49. Rangel LBA, Sherman-Baust CA, Werny RP, Schwartz DR, Cho KR, Morin PJ. Characterization of novel human ovarian cancer-specific transcripts (HOSTs) identified by serial analysis of gene expression. *Oncogene* 2003;22:7225-32.
50. Aldred MA, Huang Y, Liyanarachchi S, et al. Papillary and follicular thyroid carcinomas show distinctly different microarray expression profiles and can be distinguished by a minimum of five genes. *J Clin Oncol* 2004;22:3531-9.