

# Recent Developments on Endothelin Antagonists as Immunomodulatory Drugs - from Infection to Transplantation Medicine

Philipp C. Nett<sup>\*†</sup>, Mauro M. Teixeira<sup>#</sup>, Daniel Candinas<sup>†</sup> and Matthias Barton<sup>\*.1</sup>

*\*Department of Medicine, Internal Medical I, Medical Policlinic, University Hospital Zürich, Switzerland; †Department of Visceral and Transplantation Surgery, University Hospital Bern, Switzerland; #Departamento de Bioquímica e Imunologia, Instituto Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil*

Received: September 18, 2006; Accepted: September 28, 2006; Revised: September 30, 2006

**Abstract:** Endothelin, a potent endogenous vasoconstrictor and mitogen that acts through the ET<sub>A</sub> and ET<sub>B</sub> receptors, has been not only implicated in the regulation of cardiovascular homeostasis but also in inflammatory responses, including that induced by infection and solid organ transplantation. Changes in capillary perfusion and leukocyte recruitment are important features of inflammation. The concentrations of ET are elevated in many forms of inflammation and are especially high in sepsis. The rise in plasma levels of ET during early stages of inflammation may initially have some positive homeostatic effects that might help to maintain vascular tone and blood pressure. However, high levels of ET compromise the appropriate matching of flow to tissue needs and contribute to the pathophysiology of microcirculatory derangements. Attempts at regulating the effects of ET by the use of pharmacological antagonists are complicated by important interactions between the ET<sub>A</sub> and ET<sub>B</sub> receptors. This review highlights findings of recent studies and patents in this area showing that the ET system, apart from being a marker of vascular and tissue injury, is directly involved in the pathophysiology of these disease processes as an immunomodulatory mediator.

**Keywords:** Bacteria, disease, drug therapy, human, physiology, vasoconstriction, rejection, trypanosoma.

## INTRODUCTION

In response to several infectious and non-infectious stimuli, monocytes and macrophages release a number of mediators including cytokines which are involved in the inflammatory response [1]. These cytokines lead to further expression of mediators and co-stimulatory molecules which feed back into the inflammatory cascade [1, 2]. Endothelin is among the mediators released during inflammatory activation and together with different proinflammatory molecules, such as interleukin (IL)-1 and tumor necrosis factor (TNF)- $\alpha$ , may play a role in the cascade of events leading to tissue inflammation [3, 4]. Endothelin (ET) was initially characterized as a potent smooth-muscle spasmogen [5, 6] and is indeed a potent endogenous vasoconstrictor and mitogen [7]. Endothelin has been reported to have numerous biologic properties within the cardiovascular, respiratory, renal, endocrine, gastrointestinal, and neurologic systems [8-10] and is produced by several cell types, including endothelial cells, leukocytes, macrophages and monocytes [10-13].

ET-1, the predominant of three endothelin isoforms [14], is a product of endopeptidasic cleavage of prepro-ET-1 to big ET-1 which is later transformed by ET-converting enzyme (ECE)-1 and/or ECE-2 into its active form (Fig. 1) [9, 15]. Regulation of ET production primarily occurs at the transcriptional level which is achieved by controlling the activity of the promoter and the stability of messenger RNA [16]. In a variety of cells, ET expression is controlled by autocrine and paracrine mechanisms [17] and induced by

physiochemical factors such as blood flow, pulsatile stretch, sheer stress, hypoxia, pH or by cytokines, hormones and vasoactive agents [9, 15, 18-21] which also change the biological responsiveness to ET [15, 22].

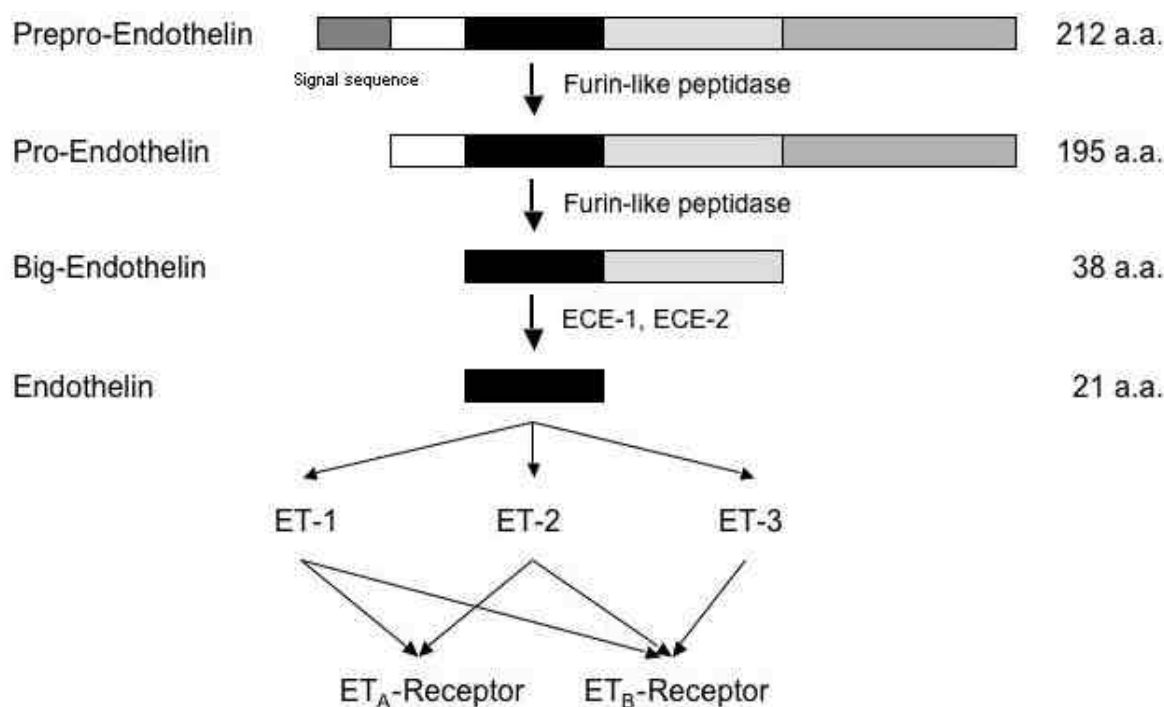
In mammals, two ET receptors have been identified: ET receptor A (ET<sub>A</sub>) and B (ET<sub>B</sub>), which are members of the G-protein-coupled superfamily [8, 9, 15]. ET<sub>A</sub> receptors have selective affinity for ET-1 and ET-2, whereas ET<sub>B</sub> receptors have similar affinity for all ET isoforms (Fig. 1) [15, 23]. Both receptors stimulate phospholipase C which leads to increased formation of diacylglycerol and inositol-1,4,5-triphosphate which activates protein kinase C pathway and increases intracellular Ca<sup>2+</sup>, respectively [24]. ET<sub>A</sub> receptors are the predominant subtype mediating vasoconstriction and cells in humans whereas ET<sub>B</sub> receptors are mainly expressed in vascular endothelial cells, mediating vasodilatation via release of nitric oxide (NO) and dilator prostanoids and thereby inhibiting cell proliferation and inflammation [8, 9, 23].

In this review, we summarize the current understanding of the mechanisms and signal transduction pathways triggered by ET in inflammatory and immunomodulatory processes and discuss the findings of previous studies evaluating the use of selective and non-selective ET receptor antagonists.

## VASCULAR EFFECTS OF ENDOTHELIN

Experimental studies using molecular and pharmacological inhibition of the ET system have demonstrated that ET-1 takes part in normal cardiovascular homeostasis [23, 25, 26]. Thus, ET-1 plays a major role in the functional and structural changes observed in arterial and pulmonary hypertension, glomerulosclerosis, atherosclerosis, mainly

<sup>1</sup>Address correspondence to this author at the Department of Medicine, Internal Medicine I, Medical Policlinic, University Hospital Zürich, Rämistrasse 100, CH-8091 Zürich, Switzerland; Tel: +41-1-255 5663; Fax: +41-1-255 8747; E-mail: barton@usz.ch



**Fig. (1).** Molecular components of the endothelin system. Processing of precursor peptides by furin-like peptidase results in formation of big-ET-1, big-ET-2 and big-ET-3. These 38-amino acids (a.a.) peptides are further processed by ECE-1 and ECE-2, chymases and non-ECE metalloprotease into vasoactive ET which activates tissue ET<sub>A</sub> and/or ET<sub>B</sub> receptors. a.a., amino acids; ECE, endothelin-converting enzyme; ET, endothelin.

through pressure-independent mechanisms [15]. Under experimentally induced pathological conditions (e.g. heart failure), the expression of ET-1 and its receptors in cardiomyocytes is increased, and treatment with ET receptor antagonists improves survival and cardiac function [15].

ET<sub>A</sub> receptors are the predominant ET vasoconstrictor receptors in arteries. ET<sub>A</sub> and ET<sub>B</sub> receptors on smooth muscle mediate contraction, cell proliferation, and hypertrophy [27]. Vasoconstrictor ET<sub>B</sub> receptors are present in the veins [28] and pulmonary vessels [29-31] in larger numbers than in arteries, although ET<sub>A</sub> still predominate over ET<sub>B</sub> receptors in these vessels. ET<sub>B</sub> receptors are also localized on endothelial cells and act through the production of NO and prostacyclin to exert vasodilator and antiinflammatory effects [26]. Whether vasoconstriction or vasodilatation is the most important effect of ET-1 under normal conditions may depend on the vascular bed [32]. It has been reported that in coronary arteries there are few endothelial vasodilator ET<sub>B</sub> receptors [33]. As a result, ET-1 acts on coronary vessels mainly as a vasoconstrictor. In other vascular beds, however, ET-1 may even function as a vasodilator under physiological conditions [34, 35].

#### ENDOTHELIN IN INFLAMMATORY AND IMMUNOMODULATORY PROCESSES

Increasing evidence supports a role for ET as a pro-inflammatory cytokine and fibrotic factor which is released in inflammatory and immune reactions [36, 37]. ET-1 acts as a proinflammatory peptide via vascular and non-vascular related mechanisms: ET-1 induces the release of pro-

inflammatory cytokines [13, 36], provokes local ischemia [38] or alters the epithelial permeability allowing antigen translocation [39].

Thus, ET-1 has been implicated in many diseases characterized by inflammation and/or fibrotic remodeling, such as atherosclerosis [40], ischemia-reperfusion injury [41], alveolitis [42], dermatitis [43], Crohn's disease and ulcerative colitis [38, 44, 45], systemic sclerosis [46, 47], dermatomyositis/polymyositis [48], systemic lupus erythematoses [49], rheumatoid arthritis [50], liver cirrhosis [51], and glomerulosclerosis [52, 53] (Table 1).

Both ET<sub>A</sub> and ET<sub>B</sub> receptors appear to be involved in these vascular and non-vascular related processes [54-56]. In patients with Crohn's disease and ulcerative colitis, the plasma levels of ET-1 are significantly higher than in healthy controls and the density of ET positive cells as well as the expression of ET<sub>A</sub> and ET<sub>B</sub> receptors is increased [8, 38]. ET-1 is involved in many cell signaling pathways that include Ca<sup>2+</sup> mobilization and activation of proinflammatory cytokines, extracellular signal-regulated kinases (ERK)1/2, and cyclin D1 [10, 57]. ET-1 also increases vascular permeability [58] as well as time- and dose-dependent entrapment of circulating platelets [59].

Recently, the downstream intracellular signal transduction pathways of ET-1 have been in part identified. In this regard, a key role of mitogen-activated protein kinases (MAPK) has been suggested, which operate via phosphorylation cascades responsible for regulation of several substrates, mainly including transcription factors implicated in inflammation, development, cell proliferation and

**Table 1. Sites of Endothelin and Endothelin Receptors Expression During Inflammatory and Immune Reactions**

Tissue	Cell type	Source
<b>Brain</b>	Neurons	[7-8, 71, 96]
	Astrocytes	[7-8, 71]
	Microglia	[7-8, 71]
<b>Peripheral nerve</b>		[7-8]
<b>Blood</b>	Macrophages	[7-8, 11, 14, 161]
	Monocytes	[7-8, 12]
	Leukocytes	[7-8, 14, 57, 89, 92]
<b>Vascular system</b>	Endothelial cells	[6-8, 14-19, 39, 48, 63, 70, 76, 79, 83, 97, 142]
	Vascular smooth muscle cells	[7-8, 14, 74]
<b>Heart</b>	Cardiomyocytes	[7-8, 10, 14, 64, 78, 83, 85, 106, 136, 165-66, 179]
<b>Liver</b>	Kupfer cells	[7-8, 14, 149, 165]
	Hepatocytes	[7-8, 14, 50, 147, 163-65, 169]
<b>Renal</b>	Peritubular cells	[7-8, 14, 20, 51, 163-4]
	Mesangial cells	[7-8, 52]
<b>Pancreas</b>	Exocrine	[7-8, 14, 164]
	Endocrine	[7-8, 117]
<b>Gastro-intestinal</b>	Epithelial cells	[7-8, 14, 37, 38, 43, 44, 81, 139, 144]
<b>Lung</b>	Epithelial cells	[3, 7-8, 14, 41, 57, 102, 157, 165, 169]
	Endothelial cells	[3, 7-8, 14, 157]
	Vascular smooth muscle cells	[3, 7-8, 14, 102]
<b>Prostate</b>		[7-8, 14]

apoptosis [60, 61]. Three major MAPK subgroups are currently known, activated by dual phosphorylation on tyrosine and threonine residues and named c-Jun N-terminal kinases (JNK), p38, and ERK1/2, respectively [62]. In particular, ERK activation is frequently required for cell growth and differentiation induced by various stimuli, also including ET-1 [63, 64]. Important downstream targets of the MAPKs and ET-1 pathways are cell cycle regulatory molecules such as cyclins, cyclin-dependent kinases and cyclin-dependent kinase inhibitors [65]. The cyclins are a target of both ET-1 and the MAPK pathway [66].

#### **EVIDENCE THAT ENDOTHELIN TRIGGERS MEDIATORS OF INFLAMMATION AND CELL ADHESION MOLECULES**

ET-1 is known to stimulate neutrophils to release elastase [67, 68] and to stimulate monocytes to produce various cytokines, including IL-1 [36, 69], IL-6 [70, 71], IL-8 [72], IL-10 [73], TNF- $\alpha$  [74], tumor growth factor (TGF)- $\beta$  [42], granulocyte-macrophage colony-stimulating factor (GCSF) [75], monocyte chemoattractant protein (MCP)-1 [76, 77] and nuclear factor (NF)- $\kappa$ B [13, 78] promoting the inflammatory cascade (Table 2). Overexpression of ET-1

stimulates expression of TNF- $\alpha$ , interferon (IFN)- $\gamma$ , IL-1 and IL-6 [79]. Conversely, TNF- $\alpha$  and IL-1 modulate the expression of ET-1 in the inflammatory cascade [80, 81], which may further amplify its deleterious effects. Akin to ET-1, many of these mediators increase expression of cell adhesion molecules [82, 83], which induce the adhesion of circulating leucocytes to endothelial cells, an initial step in the events leading to cellular infiltrate in an inflamed tissue. Lopez-Farre *et al.* [84] demonstrated that ET-1 enhances neutrophil accumulation, suggesting a direct link between ET-1 and the inflammatory process. In cultured endothelial cells, ET-1 promotes neutrophil aggregation [85] and stimulates surface expression of CD11b/CD1 on human neutrophils augmenting their adhesion to endothelial cells [84] and myocytes [86]. Activated endothelial cells are an important source of cytokines, adhesion molecules and growth factors enhancing inflammatory cells to migrate and accumulate in the extravascular tissue [87, 88]. In addition to the neutrophil-activating properties of ET-1 it also acts as a chemoattractant for monocytes [89], activator of mast cells [90-92] and inductor of significant eosinophil migration mediated by an increase in the local levels of eotaxin and IL-5 [93].

**Table 2. Stimulatory Effects of Endothelin During Inflammatory and Immunomodulatory Reactions**

Target/cell type	Stimulatory effects	Source
Neutrophils, endothelial cells	Elastase	[66, 67]
Pulmonary mast cells	Eotaxin	[91]
Monocytes	E-Selectine	[95]
Macrophages	GCSF	[74]
Monocytes	ICAM-1	[95]
Monocytes	IFN-	[78]
Monocytes, endothelial cells	IL-1	[24, 68, 78]
Pulmonary mast cells	IL-5	[91]
Monocytes, endothelial cells	IL-6	[69, 70, 78]
Endothelial cells	IL-8	[71]
Endothelial cells	IL-10	[72]
Fibroblasts	MCP-1	[75, 76]
Macrophages, endothelial cells	NF- B	[12, 77]
-	TGF-	[41]
Neutrophils, monocytes	TNF-	[73, 78]
Endothelial cells	Angiogenesis	[7-8, 99]
Neutrophils, monocytes, macrophages	Cell adhesion	[7-8, 83-85]
Monocytes	Chemotaxis	[88-91]

Macrophage, monocytes and polymorphonuclear leukocytes also secrete ET-1 [12, 13, 94]. In addition to its ability to prime leukocytes for chemotaxis [95], ET-1 stimulates also the expression of adhesion molecules which are key players in the leukocyte-endothelial cell interaction [96]. Hayasaki *et al.* [86] documented that ET-1 induces intracellular adhesion molecule (ICAM)-1 expression on cultured cardiomyocytes and endothelial cells. Similarly, Zouki *et al.* [97] reported that ET-1 increased the expression of E-selectin and ICAM-1 on cultured human endothelial cells in a concentration-dependent fashion. The production of MCP-1, a potent chemoattractant for monocytes, can be also induced by ET-1 through ET<sub>A</sub> receptor activation in endothelial cells. This effect is augmented by the proinflammatory cytokines TNF- and IL-1 [98].

Previous studies demonstrated that ET-1 and ET-3, acting through the ET<sub>B</sub> receptor, have dose-dependent stimulatory, proliferative and migratory effects on endothelial cells [99]. While ET-1 and ET-2 are reported to be equipotent in promoting DNA synthesis, ET-3 is less active [100]. During the formation of new blood vessels, endothelial cells are stimulated to migrate, proliferate and invade surrounding tissue to form capillaries [101]. ET-1, similar to VEGF, induces these angiogenic effects and, in concert with VEGF, displays a potent additive effect on the different stages of

the angiogenic process. In this scenario, ET-1 signaling is mediated mainly by the ET<sub>B</sub> receptor [102]. Recent studies emphasize a key inflammatory role of the endothelial cells, either by overexpression of inflammatory mediators or by stimulating formation of new blood vessels, in the disease process leading to the systemic organ involvement [103]. ET released after activation and/or damage of endothelial cells, might play an important role inflammatory processes and vasculopathy in autoimmune diseases [48]. Increased plasma ET levels have been found in various autoimmune diseases such as systemic lupus erythematoses [49], systemic sclerosis [47] or rheumatoid arthritis [50]. Moreover, raised ET serum levels have been implicated in the pathophysiology of both fibrotic and vascular manifestations of systemic sclerosis [47].

ET-1 exerts mitogenic activity on smooth muscle cells, myocytes, and fibroblasts [15, 104, 105]. In epithelial cells, ET-1 is known to induce gene expression and release of fibronectin [106] which is an important extracellular matrix component, as well as being chemoattractant factor for fibroblasts [107]. There is increasing evidence that ET-1 can function as a profibrotic cytokine by stimulating fibroblast chemotaxis and proliferation [108-110] and procollagen production [111, 112]. ET-1 may act in concert with several other profibrogenic molecules, including TGF- [113], platelet-derived growth factor (PDGF) [114], epidermal growth factor (EGF) [115], fibroblast growth factor (FGF) [116], insulin-like growth factor (IGF)-1 [117, 118], IL-11 [119] and insulin [64, 120, 121] to potentiate cellular transformation or replication. In particular, the synergistic interactions involving these mediators are crucial for implementing the tissue repair response leading to fibroblast proliferation, myofibroblast differentiation and collagen synthesis [117]. Within this context, a pivotal role is mediated by MAPKs that participate in the cross-talk between ET-1 and the other fibrogenic factors at several levels. Furthermore, rat cardiac fibroblasts have been shown to synthesize ET-1 [122], which induces collagen synthesis via both ET<sub>A</sub> and ET<sub>B</sub> receptors [123] as well as induces matrix metalloproteinase-2 expression [124].

The role of ET-1 in the pathogenesis of infectious diseases has only recently received attention. For example, ET-1 has been implicated in the etiology of vascular compromise and multi-organ dysfunction in the setting of septic shock [125, 126] and neuronal injury due to *Streptococcus pneumoniae* meningitis [127]. Investigators have reported that circulating ET levels increase significantly in septic individuals which may be a beneficial effect in maintaining the blood pressure and organ perfusion during the early phase of septic shock [22, 87] and which correlates with mortality [128, 129]. Recent evidence also suggests that ET-1 plays a significant role in vascular dysfunction and organ failure associated with sepsis and septic shock [126]. It has been shown that endotoxin administration results in upregulation of the ET system [130, 131] causing monocytes to produce proinflammatory cytokines such as TNF- [36]. Endotoxin stimulates the production of TNF- in monocytes and macrophages [13, 132]. Indeed, several reports have illustrated that antagonism of ET in septic animals improves metabolic acidosis as well as coronary, renal, splanchnic, pulmonary and intestinal perfusion [133-135]. In particular,

release of ET-1 is stimulated by endotoxin [136, 137]. Moreover, patients with *Rickettsia conorii*-induced vasculitis exhibit increased plasma levels of ET-1 [138]. *Rickettsia* share some important similarities with *Trypanosoma cruzi* (Chagas' disease) as both can reside within the endothelial cell cytoplasm and cause perturbations of the host cell [139]. ET is expressed during *Trypanosoma cruzi* infection in mice [65], and high plasma levels of immunoreactive ET were found in patients with severe chagasic cardiomyopathy [140].

## EFFECTS OF ENDOTHELIN RECEPTOR BLOCKADE ON INFLAMMATORY AND IMMUNE PROCESSES

### Cell Adhesion and Migration

Activated leukocyte adhesion is the initial and rate-limiting step in the development of inflammatory cellular infiltrate. Both ET<sub>A</sub> and ET<sub>B</sub> receptors are involved in the ET-induced activation of leukocyte adhesion [86, 97, 141]. Based on these observations, selective and non-selective ET receptor antagonists might be an effective treatment as immunomodulatory agents blocking the recruitment of leukocytes by reduction of leukocyte adhesion and preventing the ET-1 dependent progression of the inflammatory cascade.

Anthoni *et al.* [142] reported that bosentan, a non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist, prevents the adhesion of leukocytes in colonic submucosal venules and reduces inflammation in a mouse model of inflammatory bowel disease. A comparable effect was also observed with the non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist LU420627. Prophylactic oral administration of bosentan reduces inflammation and myeloperoxidase activity in colonic tissue in trinitrobenzene sulphamate-induced colitis in rats [143, 144]. The mechanisms of bosentan-induced reduction of leukocyte firm adherence remain speculative, but may depend on the blockade of ET-1 induced up-regulation of cell adhesion molecules on the endothelium and leukocytes [84, 86, 97, 145-147]. In particular, the finding that anti-inflammatory effects of ET-1 blockade are mediated by vascular cell adhesion molecule (VCAM)-1 in isolated endothelial cells suggests that these cell adhesion molecules may play a role in the attenuation of sticking and the elevation of rolling velocity [146]. This is supported by the observation that treatment with an anti-VCAM-1 oligonucleotide exerts a more potent protective effect on leucocyte endothelial adhesion when compared to bosentan [148], which reduces the infection-associated increase in nitrate/nitrite (NO<sub>x</sub>) serum concentration [149]

Previous studies using cardiac myocytes have shown that the selective ET<sub>B</sub> receptor antagonist BQ-788 inhibits neutrophil-endothelial cell adhesion and ICAM-1 expression on endothelial cells [86], whereas the selective ET<sub>A</sub> receptor antagonist S-0139 promotes neutrophil-cardiac myocytes adhesion and ICAM-1 production. This may be related to the differences in ET<sub>A</sub> and ET<sub>B</sub> receptor density on various cell types. For example, ET<sub>A</sub> represents 90% of the receptors on the surface of cardiac myocytes, while the ET<sub>B</sub> receptor is the dominant receptor on endothelial cells and macrophages which also predominantly express the ET<sub>B</sub> receptor [150,

151]. However, different groups have reported varying results. Zouki *et al.* [97] reported that the ET<sub>A</sub>-selective antagonist FR139317 markedly attenuated the ET-1 stimulated neutrophil adherence, suggesting a role of the ET<sub>A</sub> receptor in neutrophil-endothelial attachment. This discrepancy of the results may be due to the use of different antagonists. It also remains possible that short- versus long-term exposure of endothelial cell to ET-1 may affect endothelial adhesiveness differently. The finding that the ET<sub>A</sub> receptor-selective antagonist BQ610, but not the ET<sub>B</sub> receptor-selective antagonist BQ788, decreased ET-1 induced MCP-1 expression implicates a role for the ET<sub>A</sub> receptor in ET-1-mediated inflammation [98]. Recently, it has been shown that aryl-alkane-sulfonamides and their derivatives may ameliorate ET-1-mediated inflammation [152, 153] as well as an ET<sub>A</sub> selective N-(5-isoxazolyl) benzene-sulfonamide receptor antagonist that can be modified in the 4-position with aryl and substituted aryl groups to generate a ET<sub>B</sub> selective receptor antagonist [154, 155].

The mechanisms responsible for increased production of TNF- $\alpha$  by ET-1 and inhibition of TNF- $\alpha$  production by ET-1 receptor antagonism remain unclear. It has been suggested that ET-1 may play a potential proinflammatory role in pathological conditions [156]. However, there have recently been published a number of studies investigating this question. Quehenberger *et al.* [157], for example, demonstrated that the human ET-1 gene has NF- $\kappa$ B binding site and reported that transcription of the ET-1 gene is controlled by NF- $\kappa$ B. This group clearly demonstrated that binding of advanced glycation end products (AGE's) on the surface of erythrocytes, which occurs during oxidative stress, results in a NF- $\kappa$ B-dependent ET-1 induction *in vitro* [157]. It is likely that inhibition of TNF- $\alpha$  production by ET receptor antagonism is mediated by the NF- $\kappa$ B pathway. The effects of NF- $\kappa$ B in inflammation are well known, e.g., induction of synthesis of several proinflammatory cytokines including IL-1 and IL-6 [158]. Browatzki *et al.* [159] recently demonstrated that ET<sub>A</sub> receptor activation induces IL-6 synthesis via NF- $\kappa$ B activation in human vascular smooth muscle cells, extending recent observations that ET-1 is a potent stimulus for activation of NF- $\kappa$ B in human hepatic stellate cells [160]. These mechanisms may be responsible for the synthesis of most of proinflammatory cytokines including the autocrine activation of ET-1 synthesis.

### Allergic Inflammation

ET-1 has been reported to induce eosinophil migration mediated by an increase in the local levels of eotaxin and IL-5 [93, 161]. This observation explains previous reports showing the accumulation of eosinophils rather than neutrophils, as well as an increase in immunoreactive ET in the broncho-alveolar fluid in experimental airway inflammation [162-164], and in a murine model of allergic pleurisy [165, 166]. Treatment of these animals with a selective ET receptor antagonist significantly decreased the production of proinflammatory mediators [42] and eosinophilia [161].

In this context recent patents propose to use a novel class of indole-based ECE inhibitors to diminish the production of

ET-1 to inhibit bronchoconstriction and pulmonary vasoconstriction in newborn mammals [167, 168].

### **Atherosclerosis and Fibrosis**

Inflammation of the vessel wall is a characteristic feature of atherosclerosis. ET-1 contributes to the pathogenesis of atherosclerosis and its chronic complications [40]. Browatzki *et al.* [78] investigated the effect of ET-1 on the proinflammatory transcription factor NF- $\kappa$ B in human monocytes which are the major source of inflammatory mediators in atheroma and are located in rupture-prone plaque areas. In these monocytes, ET-1 caused NF- $\kappa$ B activation which was blocked by inhibition of the I- $\kappa$ B-degrading proteasome complex as well as the ET<sub>A</sub> receptor antagonist BQ123 but not by the ET<sub>B</sub> receptor antagonist BQ788. ET-1 stimulated expression of the proinflammatory molecule CD40 but not of the cytokine IL-6 in a NF- $\kappa$ B-dependent manner. The data demonstrate the ability of ET-1 to differently activate inflammatory pathways in human monocytes [78].

Recently, Ammarguella *et al.* [169, 170] reported that myocardial inflammation and fibrosis in hearts of deoxycorticosterone acetate hypertensive rats were, at least in part, mediated by ET-1. In this model, the selective ET<sub>A</sub> receptor antagonist A-127722 prevented cardiac fibrosis in hearts by normalizing the levels of procollagen I and III and TGF- $\beta$ . A-127722 also abrogated the activation of the inflammatory mediator NF- $\kappa$ B and the expression of platelet-endothelial cell adhesion molecule (PECAM)-1 and VCAM-1 [169, 170]. Hoher *et al.* [171] also demonstrated that ET-1 plays an important role in the development of cardiac fibrosis in a two-kidney, one-clip rat model of renovascular hypertension. In this model, the use of the ET<sub>B</sub>-selective antagonist IRL1038, but not the ET<sub>A</sub>-selective antagonist BQ-123, attenuated cardiac fibrosis.

While benzo-1,3-dioxolyl- and ET benzofuranyl-substituted pyrrolidine derivatives could be useful for the treatment of congestive heart failure [172-175], recent patents reveal that pyrrolidine and piperidine derivatives act as selective ET<sub>A</sub> and ET<sub>B</sub> receptor antagonists and that novel alkanesulfonamides may have beneficial effects in atherosclerosis [176, 177].

### **Transplantation**

In organ transplantation, several studies confirmed the involvement of ET-1 in ischemia-reperfusion injury in heart, lung, liver, and kidney allografts [178, 179]. Treatment with the selective ET<sub>A</sub> receptor antagonist BSF208075 has also protective effects on the microcirculation after ischemia/reperfusion. Uhlmann *et al.* [180, 181] were able to show that BSF208075 not only affects the expression of vasoactive genes, but also decrease gene expression of proinflammatory cytokines such as TNF- $\alpha$ , IL-1 and IL-6 *in vitro*. Ischemia/reperfusion injury promotes a microcirculation-associated inflammatory response, involving release of deleterious mediators, such as reactive oxygen species, TNF- $\alpha$ , IL-1, the upregulation of leucocytic and endothelial adhesion molecules such as selectins,  $\alpha$ -integrins, intracellular adhesion molecule-1, and the interaction of platelets and leucocyte with the microvascular endothelium [180-182]. Based on these results selective antagonism of ET<sub>A</sub> receptors may have anti-inflammatory potential through suppression of the

mRNA of the genes of proinflammatory cytokines such as TNF- $\alpha$ , IL-1 and IL-6. Several other studies demonstrated also a pathogenetic involvement of ET-1 in ischemia/reperfusion injury of heart, lung, liver and kidney allografts [178, 183].

Elevated levels of ET-1 have been associated with inflammation and immune responses [15, 184, 185] as well as with transplant-associated diseases [179]. High ET-1 expression in early posttransplant biopsy specimens was related to poor long-term allograft function following renal transplantation [186], and combined endothelial and myocardial protection could be achieved by ET antagonism enhancing transplant allograft preservation [187]. Multiple chemokines, cytokines, and proinflammatory mediators are involved in the immune response following transplantation [188]. ET-1 accumulates during cold storage of allografts and can be detected in the effluent preservation solution [189]. In addition, ET-1 is very likely to play a pivotal role in the development of chronic allograft dysfunction [190] which represents one of the major causes of late allograft loss [191]. Increased expression of components of the ET system has been reported in areas of neointimal proliferation, a hallmark of chronic allograft vasculopathy [187, 189].

There is growing evidence that hemodynamic effects of calcineurin inhibitors (CNI) may be caused by an altered expression of ET-1 [37, 192]. Enhanced levels of ET-1 lead to constriction of smooth muscle cells and cell proliferation, as could be shown in cell culture experiments [193, 194]. The plasma levels of big ET-1 are dependent on CNI plasma levels 1 year after successful heart transplantation in patients [195]. Previous studies using ET<sub>A</sub> receptor antagonism have used concomitant immunosuppressive therapy [15]; thus, the potential immunomodulatory actions of ET-1 and the actual contribution of ET-1 to chronic rejection were masked because of the immunosuppressive agents. In a recent study in the absence of immunosuppression, we found that treatment with the ET<sub>A</sub> receptor antagonist darusentan following cardiac transplantation in the absence of immunosuppression changed circulating levels of IL-1 and TNF-tissue and gene expression of different interleukins in an organ-specific fashion independent of graft atherosclerosis [190]. These findings are compatible with a direct immunomodulatory role for ET<sub>A</sub> receptors during allograft rejection [190, 196]. Braun *et al.* [197] reported that ET<sub>A</sub> receptor blockade with LU135252 prevented long-term deleterious changes on renal allograft function and morphology independently from systemic blood pressure and did not lower proteinuria, strongly arguing against any effect on glomerular hemodynamics and upregulation of renal ET-1 synthesis in a rat model of renal allograft rejection in which a role for the ET<sub>A</sub> receptor has been demonstrated [198].

In this context a recent patent reports that tripeptide derivatives [199, 200], as well as pyrimidine sulfamides derivatives and benzo-fused heterocycles could show significant benefit in the treatment of ischemia and proliferative disorders associated with ET-1 [176, 201].

### **Infection**

Ono *et al.* [202] have reported that myocardial and ET-1 plasma levels increased in mice with encephalomyocarditis

virus-induced myocarditis. The observation that ET-1 levels were higher in the myocardium than those in plasma suggests that the heart is major source of endogenous ET-1 in myocarditis. The non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist bosentan improved survival in these mice and reduced cellular infiltration and myocardial necrosis [202]. In addition, Pandey *et al.* [203] reported that another non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonist, SB217242, improved cardiac contractility and relaxation in these mice as well. ET-1 contributes to the pathogenesis of murine viral myocarditis and treatment with the non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonists bosentan or SB217242 significantly improved myocarditis [202, 203]. In another study, Albertini *et al.* [128] reported that ET<sub>A</sub> receptor blockade in rats completely prevented endotoxin-induced mortality and attenuated serum indices of myocardial, renal, liver and lung injury. Previous studies on experimental septic shock models evaluated the effects of ET<sub>A</sub> receptor antagonists on injury and dysfunction of a single organ and showed that ET-1 has strong cardiodepressant effects in sepsis [135, 204].

We have recently reported that ET-1 contributes to the pathogenesis of chagasic cardiomyopathy (*Trypanosoma cruzi*) showing that interventions inhibiting the synthesis of ET-1 and/or neutral endopeptidase might have either deleterious or protective effects on myocardial structure and function in chagasic cardiomyopathy [137, 149, 184, 205]. *Trypanosoma cruzi* infection induces ET-1 release from endothelial cells [206], and the released ET-1 contracts vascular smooth muscle via activation of ET<sub>A</sub> receptors [207]. The mechanism by which ET-1 controls *Trypanosoma cruzi* infection are not investigated yet, however, ET-1 acting mainly on ET<sub>A</sub> receptors can activate macrophages to release cytokines such as TNF- $\alpha$  [55]. As TNF- $\alpha$ -induced NO production by macrophages plays an important role in the acute phase of the infection [208], it is possible that the ability of ET-1 to release TNF- $\alpha$  may account for the effects observed. In this regard, we have shown that inflammatory mediators such as platelet-activating factors and chemokines, which like ET-1 act on G-protein-coupled receptors, activate macrophages to produce NO and kill *Trypanosoma cruzi* [209].

## CURRENT AND FUTURE DEVELOPMENTS

Endothelin (ET), a naturally occurring polypeptide with a broad range of activities including vasospastic, proinflammatory and profibrotic properties, has been implicated in inflammatory and immunomodulatory processes. These are characterized by vascular damage, inflammatory infiltrates and progressive fibrosis of the skin and internal organs. Plasma concentrations of ET-1 may be elevated in certain inflammatory reactions and may favor the inflammatory cascade by facilitating the recruitment of leukocytes, the expression of cell adhesion molecules and cytokines. ET-1 also induces fibrosis by stimulating fibrotic factors, collagen synthesis, and fibroblast proliferation. Although there is growing evidence to suggest that ET-1 may act as a proinflammatory, profibrotic, and mitogenic factor, it is not clear whether activation of the ET system under these conditions is adaptive or pathogenic. While the relative importance of the receptors mediating these effects remains uncertain, both ET<sub>A</sub> and ET<sub>B</sub> receptors appear to be

involved. In this context, the ET represents a novel target for immunomodulation. Recent studies with selective ET<sub>A</sub>, ET<sub>B</sub> or non-selective ET<sub>A</sub>/ET<sub>B</sub> receptor antagonists indicate that ET-receptors blockade, indeed, holds the potential to markedly improve consequences of local or systemic inflammatory processes improving symptoms and prognosis of the underlying disease. Clinical trials – in part under way [210, 211, 212] are now required to test this hypothesis in patients.

## ACKNOWLEDGEMENTS

Supported by the Swiss National Foundation (SNF 3200-058426.99, 3232-058421.99, 81ZH-064227 and 32-108258/1) the Hanne-Liebermann-Stiftung, the Hartmann-Müller-Stiftung, the Olga-Mayenfisch-Stiftung, and a grant-in-aid by the Department of Clinical Research, University of Bern.

## REFERENCES

- [1] Luster AD. Chemokines--chemotactic cytokines that mediate inflammation. *N Engl J Med* 1998; 338(7): 436-45.
- [2] Keane MP, Strieter RM. Chemokine signaling in inflammation. *Crit Care Med* 2000; 28(4 Suppl): N13-26.
- [3] Saleh D, Furukawa K, Tsao MS, *et al.* Elevated expression of endothelin-1 and endothelin-converting enzyme-1 in idiopathic pulmonary fibrosis: possible involvement of proinflammatory cytokines. *Am J Respir Cell Mol Biol* 1997; 16(2): 187-93.
- [4] Graido-Gonzalez E, Doherty JC, Bergreen EW, Organ G, Telfer M, McMillen MA. Plasma endothelin-1, cytokine, and prostaglandin E2 levels in sickle cell disease and acute vaso-occlusive sickle crisis. *Blood* 1998; 92(7): 2551-5.
- [5] Hickey KA, Rubanyi G, Paul RJ, Highsmith RF. Characterization of a coronary vasoconstrictor produced by cultured endothelial cells. *Am J Physiol* 1985; 248(5 Pt 1): C550-6.
- [6] Gillespie MN, Owasoyo JO, McMurtry IF, O'Brien RF. Sustained coronary vasoconstriction provoked by a peptidergic substance released from endothelial cells in culture. *J Pharmacol Exp Ther* 1986; 236(2): 339-43.
- [7] Yanagisawa M, Kurihara H, Kimura S, *et al.* A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature* 1988; 332(6163): 411-5.
- [8] Rubanyi GM, Botelho LH. Endothelins. *FASEB J* 1991; 5(12): 2713-20.
- [9] Levin ER. Endothelins. *N Engl J Med* 1995; 333(6): 356-63.
- [10] Kedzierski RM, Yanagisawa M. Endothelin system: the double-edged sword in health and disease. *Annu Rev Pharmacol Toxicol* 2001; 41: 851-76.
- [11] Gray MO, Long CS, Kalinyak JE, Li HT, Karlner JS. Angiotensin II stimulates cardiac myocyte hypertrophy via paracrine release of TGF-beta 1 and endothelin-1 from fibroblasts. *Cardiovasc Res* 1998; 40(2): 352-63.
- [12] Ehrenreich H, Anderson RW, Fox CH, *et al.* Endothelins, peptides with potent vasoactive properties, are produced by human macrophages. *J Exp Med* 1990; 172(6): 1741-8.
- [13] Cunningham ME, Huribal M, Bala RJ, McMillen MA. Endothelin-1 and endothelin-4 stimulate monocyte production of cytokines. *Crit Care Med* 1997; 25(6): 958-64.
- [14] Inoue A, Yanagisawa M, Kimura S, *et al.* The human endothelin family: three structurally and pharmacologically distinct isopeptides predicted by three separate genes. *Proc Natl Acad Sci USA* 1989; 86(8): 2863-7.
- [15] Luscher TF, Barton M. Endothelins and endothelin receptor antagonists: therapeutic considerations for a novel class of cardiovascular drugs. *Circulation* 2000; 102(19): 2434-40.
- [16] Bloch KD, Friedrich SP, Lee ME, Eddy RL, Shows TB, Quertermous T. Structural organization and chromosomal assignment of the gene encoding endothelin. *J Biol Chem* 1989; 264(18): 10851-7.
- [17] Inoue A, Yanagisawa M, Takuwa Y, Mitsui Y, Kobayashi M, Masaki T. The human preendothelin-1 gene. Complete nucleotide sequence and regulation of expression. *J Biol Chem* 1989; 264(25): 14954-9.

- [18] Rakugi H, Tabuchi Y, Nakamaru M, *et al.* Evidence for endothelin-1 release from resistance vessels of rats in response to hypoxia. *Biochem Biophys Res Commun* 1990; 169(3): 973-7.
- [19] Malek A, Izumo S. Physiological fluid shear stress causes downregulation of endothelin-1 mRNA in bovine aortic endothelium. *Am J Physiol* 1992; 263(2 Pt 1): C389-96.
- [20] Macarthur H, Warner TD, Wood EG, Corder R, Vane JR. Endothelin-1 release from endothelial cells in culture is elevated both acutely and chronically by short periods of mechanical stretch. *Biochem Biophys Res Commun* 1994; 200(1): 395-400.
- [21] Wesson DE, Simoni J, Green DF. Reduced extracellular pH increases endothelin-1 secretion by human renal micro-vascular endothelial cells. *J Clin Invest* 1998; 101(3): 578-83.
- [22] Battistini B, Forget MA, Laight D. Potential roles for endothelins in systemic inflammatory response syndrome with a particular relationship to cytokines. *Shock* 1996; 5(3): 167-83.
- [23] Barton M, Luscher TF. Endothelin antagonists for hypertension and renal disease. *Curr Opin Nephrol Hypertens* 1999; 8(5): 549-56.
- [24] Simonson MS, Dunn MJ. Cellular signaling by peptides of the endothelin gene family. *FASEB J* 1990; 4(12): 2989-3000.
- [25] Benigni A, Remuzzi G. Endothelin antagonists. *Lancet* 1999; 353(9147): 133-8.
- [26] Schiffrin EL. Endothelin: role in hypertension. *Biol Res* 1998; 31(3): 199-208.
- [27] de Nucci G, Thomas R, D'Orleans-Juste P, *et al.* Pressor effects of circulating endothelin are limited by its removal in the pulmonary circulation and by the release of prostacyclin and endothelium-derived relaxing factor. *Proc Natl Acad Sci USA* 1988; 85(24): 9797-800.
- [28] Moreland S, McMullen DM, Delaney CL, Lee VG, Hunt JT. Venous smooth muscle contains vasoconstrictor ETB-like receptors. *Biochem Biophys Res Commun* 1992; 184(1): 100-6.
- [29] Russell FD, Davenport AP. Characterization of endothelin receptors in the human pulmonary vasculature using bosentan, SB209670, and 97-139. *J Cardiovasc Pharmacol* 1995; 26 Suppl 3: S346-7.
- [30] Sato K, Oka M, Hasunuma K, Ohnishi M, Kira S. Effects of separate and combined ETA and ETB blockade on ET-1-induced constriction in perfused rat lungs. *Am J Physiol* 1995; 269(5 Pt 1): L668-72.
- [31] Davie N, Haleen SJ, Upton PD, *et al.* ET<sub>A</sub> and ET<sub>B</sub> receptors modulate the proliferation of human pulmonary artery smooth muscle cells. *Am J Respir Crit Care Med* 2002; 165(3): 398-405.
- [32] Russell FD, Skepper JN, Davenport AP. Detection of endothelin receptors in human coronary artery vascular smooth muscle cells but not endothelial cells by using electron microscope autoradiography. *J Cardiovasc Pharmacol* 1997; 29(6): 820-6.
- [33] Bacon CR, Davenport AP. Endothelin receptors in human coronary artery and aorta. *Br J Pharmacol* 1996; 117(5): 986-92.
- [34] Verhaar MC, Strachan FE, Newby DE, *et al.* Endothelin-A receptor antagonist-mediated vasodilatation is attenuated by inhibition of nitric oxide synthesis and by endothelin-B receptor blockade. *Circulation* 1998; 97(8): 752-6.
- [35] Goddard J, Johnston NR, Hand MF, *et al.* Endothelin-A receptor antagonism reduces blood pressure and increases renal blood flow in hypertensive patients with chronic renal failure: a comparison of selective and combined endothelin receptor blockade. *Circulation* 2004; 109(9): 1186-93.
- [36] McMillen MA, Sumpio BE. Endothelins: polyfunctional cytokines. *J Am Coll Surg* 1995; 180(5): 621-37.
- [37] Solini A, Santini E, Ferrannini E. Enhanced angiotensin II-mediated effects in fibroblasts of patients with familial hypercholesterolemia. *J Hypertens* 2005; 23(2): 367-74.
- [38] Murch SH, Braegger CP, Sessa WC, MacDonald TT. High endothelin-1 immunoreactivity in Crohn's disease and ulcerative colitis. *Lancet* 1992; 339(8790): 381-5.
- [39] Oktar BK, Coskun T, Bozkurt A, *et al.* Endothelin-1-induced PMN infiltration and mucosal dysfunction in the rat small intestine. *Am J Physiol Gastrointest Liver Physiol* 2000; 279(3): G483-91.
- [40] Barton M, Haudenschild CC, d'Uscio LV, Shaw S, Munter K, Luscher TF. Endothelin ETA receptor blockade restores NO-mediated endothelial function and inhibits atherosclerosis in apolipoprotein E-deficient mice. *Proc Natl Acad Sci USA* 1998; 95(24): 14367-72.
- [41] Bajory Z, Hutter J, Krombach F, Messmer K. The role of endothelin-1 in ischemia-reperfusion induced acute inflammation of the bladder in rats. *J Urol* 2002; 168(3): 1222-5.
- [42] Finsnes F, Lyberg T, Christensen G, Skjonsberg OH. Effect of endothelin antagonism on the production of cytokines in eosinophilic airway inflammation. *Am J Physiol Lung Cell Mol Physiol* 2001; 280(4): L659-65.
- [43] Griswold DE, Douglas SA, Martin LD, *et al.* Endothelin B receptor modulates inflammatory pain and cutaneous inflammation. *Mol Pharmacol* 1999; 56(4): 807-12.
- [44] Letizia C, Boirivant M, De Toma G, *et al.* Plasma levels of endothelin-1 in patients with Crohn's disease and ulcerative colitis. *Ital J Gastroenterol Hepatol* 1998; 30(3): 266-9.
- [45] MacCarthy PA, Grocott-Mason R, Prendergast BD, Shah AM. Contrasting inotropic effects of endogenous endothelin in the normal and failing human heart: studies with an intracoronary ET<sub>A</sub> receptor antagonist. *Circulation* 2000; 101(2): 142-7.
- [46] Becvar R, Stork J, Pesakova V, *et al.* Clinical correlations of potential activity markers in systemic sclerosis. *Ann N Y Acad Sci* 2005; 1051: 404-12.
- [47] Vancheswaran R, Magoulas T, Efrat G, *et al.* Circulating endothelin-1 levels in systemic sclerosis subsets--a marker of fibrosis or vascular dysfunction? *J Rheumatol* 1994; 21(10): 1838-44.
- [48] Mayes MD. Endothelin and endothelin receptor antagonists in systemic rheumatic disease. *Arthritis Rheum* 2003; 48(5): 1190-9.
- [49] Yoshio T, Masuyama J, Mimori A, Takeda A, Minota S, Kano S. Endothelin-1 release from cultured endothelial cells induced by sera from patients with systemic lupus erythematosus. *Ann Rheum Dis* 1995; 54(5): 361-5.
- [50] Haq A, El-Ramahi K, Al-Dalaan A, Al-Sedairy ST. Serum and synovial fluid concentrations of endothelin-1 in patients with rheumatoid arthritis. *J Med* 1999; 30(1-2): 51-60.
- [51] Rockey DC, Chung JJ. Endothelin antagonism in experimental hepatic fibrosis. Implications for endothelin in the pathogenesis of wound healing. *J Clin Invest* 1996; 98(6): 1381-8.
- [52] Barton M, Vos I, Shaw S, *et al.* Dysfunctional renal nitric oxide synthase as a determinant of salt-sensitive hypertension: mechanisms of renal artery endothelial dysfunction and role of endothelin for vascular hypertrophy and glomerulosclerosis. *J Am Soc Nephrol* 2000; 11(5): 835-45.
- [53] Ortmann J, Amann K, Brandes RP, *et al.* Role of podocytes for reversal of glomerulosclerosis and proteinuria in the aging kidney after endothelin inhibition. *Hypertension* 2004; 44(6): 974-81.
- [54] Teder P, Noble PW. A cytokine reborn? Endothelin-1 in pulmonary inflammation and fibrosis. *Am J Respir Cell Mol Biol* 2000; 23(1): 7-10.
- [55] Barton M. Endothelial dysfunction and atherosclerosis: endothelin receptor antagonists as novel therapeutics. *Curr Hypertens Rep* 2000; 2(1): 84-91.
- [56] Traupe T, Ortmann J, Munter K, Barton M. Endothelial therapy of atherosclerosis and its risk factors. *Curr Vasc Pharmacol* 2003; 1(2): 111-21.
- [57] Suzuki E, Nagata D, Kakoki M, *et al.* Molecular mechanisms of endothelin-1-induced cell-cycle progression: involvement of extracellular signal-regulated kinase, protein kinase C, and phosphatidylinositol 3-kinase at distinct points. *Circ Res* 1999; 84(5): 611-9.
- [58] Helset E, Kjaeve J, Hauge A. Endothelin-1-induced increases in microvascular permeability in isolated, perfused rat lungs requires leukocytes and plasma. *Circ Shock* 1993; 39(1): 15-20.
- [59] Helset E, Lindal S, Olsen R, Myklebust R, Jorgensen L. Endothelin-1 causes sequential trapping of platelets and neutrophils in pulmonary microcirculation in rats. *Am J Physiol* 1996; 271(4 Pt 1): L538-46.
- [60] Widmann C, Gibson S, Jarpe MB, Johnson GL. Mitogen-activated protein kinase: conservation of a three-kinase module from yeast to human. *Physiol Rev* 1999; 79(1): 143-80.
- [61] Chang L, Karin M. Mammalian MAP kinase signalling cascades. *Nature* 2001; 410(6824): 37-40.
- [62] MacCorkle RA, Tan TH. Mitogen-activated protein kinases in cell-cycle control. *Cell Biochem Biophys* 2005; 43(3): 451-61.
- [63] Whelchel A, Evans J, Posada J. Inhibition of ERK activation attenuates endothelin-stimulated airway smooth muscle cell proliferation. *Am J Respir Cell Mol Biol* 1997; 16(5): 589-96.



- [64] Nett PC, Ortmann J, Celeiro J, *et al.* Transcriptional regulation of vascular bone morphogenetic protein by endothelin receptors in early autoimmune diabetes mellitus. *Life Sci* 2006; 78(19): 2213-8.
- [65] Petkova SB, Tanowitz HB, Magazine HI, *et al.* Myocardial expression of endothelin-1 in murine *Trypanosoma cruzi* infection. *Cardiovasc Pathol* 2000; 9(5): 257-65.
- [66] Petkova SB, Ashton A, Bouzahzah B, Huang H, Pestell RG, Tanowitz HB. Cell cycle molecules and diseases of the cardiovascular system. *Front Biosci* 2000; 5: D452-60.
- [67] Hafstrom I, Ringertz B, Lundeberg T, Palmblad J. The effect of endothelin, neuropeptide Y, calcitonin gene-related peptide and substance P on neutrophil functions. *Acta Physiol Scand* 1993; 148(3): 341-6.
- [68] Halim A, Kanayama N, el Maradny E, Maehara K, Terao T. Activated neutrophil by endothelin-1 caused tissue damage in human umbilical cord. *Thromb Res* 1995; 77(4): 321-7.
- [69] Fabricio AS, Rae GA, Zampronio AR, D'Orleans-Juste P, Souza GE. Central endothelin ET<sub>B</sub> receptors mediate IL-1-dependent fever induced by preformed pyrogenic factor and corticotropin-releasing factor in the rat. *Am J Physiol Regul Integr Comp Physiol* 2006; 290(1): R164-71.
- [70] Saito Y, Nakao K, Mukoyama M, Imura H. Increased plasma endothelin level in patients with essential hypertension. *N Engl J Med* 1990; 322(3): 205.
- [71] Stankova J, D'Orleans-Juste P, Rola-Pleszczynski M. ET-1 induces IL-6 gene expression in human umbilical vein endothelial cells: synergistic effect of IL-1. *Am J Physiol* 1996; 271(4 Pt 1): C1073-8.
- [72] Zidovetzki R, Chen P, Chen M, Hofman FM. Endothelin-1-induced interleukin-8 production in human brain-derived endothelial cells is mediated by the protein kinase C and protein tyrosine kinase pathways. *Blood* 1999; 94(4): 1291-9.
- [73] Huribal M, Kumar R, Cunningham ME, Sumpio BE, McMillen MA. Endothelin-stimulated monocyte supernatants enhance neutrophil superoxide production. *Shock* 1994; 1(3): 184-7.
- [74] Ruetten H, Thiemermann C. Endothelin-1 stimulates the biosynthesis of tumour necrosis factor in macrophages: ET-receptors, signal transduction and inhibition by dexamethasone. *J Physiol Pharmacol* 1997; 48(4): 675-88.
- [75] Waters CE, Shi-Wen X, Denton CP, Abraham DJ, Pearson JD. Signaling pathways regulating intercellular adhesion molecule 1 expression by endothelin 1: comparison with interleukin-1beta in normal and scleroderma dermal fibroblasts. *Arthritis Rheum* 2006; 54(2): 649-60.
- [76] Luft FC. Proinflammatory effects of angiotensin II and endothelin: targets for progression of cardiovascular and renal diseases. *Curr Opin Nephrol Hypertens* 2002; 11(1): 59-66.
- [77] Duerrschmidt N, Wippich N, Goettsch W, Broemme HJ, Morawietz H. Endothelin-1 induces NAD(P)H oxidase in human endothelial cells. *Biochem Biophys Res Commun* 2000; 269(3): 713-7.
- [78] Browatzki M, Pfeiffer CA, Schmidt J, Kranzhofer R. Endothelin-1 induces CD40 but not IL-6 in human monocytes via the proinflammatory transcription factor NF-kappaB. *Eur J Med Res* 2005; 10(5): 197-201.
- [79] Yang LL, Gros R, Kabir MG, *et al.* Conditional cardiac overexpression of endothelin-1 induces inflammation and dilated cardiomyopathy in mice. *Circulation* 2004; 109(2): 255-61.
- [80] Maemura K, Kurihara H, Morita T, Oh-hashii Y, Yazaki Y. Production of endothelin-1 in vascular endothelial cells is regulated by factors associated with vascular injury. *Gerontology* 1992; 38 Suppl 1: 29-35.
- [81] Klemm P, Warner TD, Hohlfeld T, Corder R, Vane JR. Endothelin 1 mediates *ex vivo* coronary vasoconstriction caused by exogenous and endogenous cytokines. *Proc Natl Acad Sci USA* 1995; 92(7): 2691-5.
- [82] Arndt H, Bolanowski MA, Granger DN. Role of interleukin 8 on leucocyte-endothelial cell adhesion in intestinal inflammation. *Gut* 1996; 38(6): 911-5.
- [83] Panes J, Granger DN. Leukocyte-endothelial cell interactions: molecular mechanisms and implications in gastrointestinal disease. *Gastroenterology* 1998; 114(5): 1066-90.
- [84] Lopez Farre A, Riesco A, Espinosa G, *et al.* Effect of endothelin-1 on neutrophil adhesion to endothelial cells and perfused heart. *Circulation* 1993; 88(3): 1166-71.
- [85] Gomez-Garre D, Guerra M, Gonzalez E, *et al.* Aggregation of human polymorphonuclear leukocytes by endothelin: role of platelet-activating factor. *Eur J Pharmacol* 1992; 224(2-3): 167-72.
- [86] Hayasaki Y, Nakajima M, Kitano Y, Iwasaki T, Shimamura T, Iwaki K. ICAM-1 expression on cardiac myocytes and aortic endothelial cells via their specific endothelin receptor subtype. *Biochem Biophys Res Commun* 1996; 229(3): 817-24.
- [87] Krishnaswamy G, Kelley J, Yerra L, Smith JK, Chi DS. Human endothelium as a source of multifunctional cytokines: molecular regulation and possible role in human disease. *J Interferon Cytokine Res* 1999; 19(2): 91-104.
- [88] Majka M, Janowska-Wieczorek A, Ratajczak J, *et al.* Numerous growth factors, cytokines, and chemokines are secreted by human CD34(+) cells, myeloblasts, erythroblasts, and megakaryoblasts and regulate normal hematopoiesis in an autocrine/paracrine manner. *Blood* 2001; 97(10): 3075-85.
- [89] Achmad TH, Rao GS. Chemotaxis of human blood monocytes toward endothelin-1 and the influence of calcium channel blockers. *Biochem Biophys Res Commun* 1992; 189(2): 994-1000.
- [90] Sessa WC, Kaw S, Hecker M, Vane JR. The biosynthesis of endothelin-1 by human polymorphonuclear leukocytes. *Biochem Biophys Res Commun* 1991; 174(2): 613-8.
- [91] Uchida Y, Ninomiya H, Sakamoto T, *et al.* ET-1 released histamine from guinea pig pulmonary but not peritoneal mast cells. *Biochem Biophys Res Commun* 1992; 189(2): 1196-201.
- [92] Maurer M, Wedemeyer J, Metz M, *et al.* Mast cells promote homeostasis by limiting endothelin-1-induced toxicity. *Nature* 2004; 432(7016): 512-6.
- [93] Cui P, Sharmin S, Okumura Y, Yamada H, Yano M, Mizuno D, *et al.* Endothelin-1 peptides and IL-5 synergistically increase the expression of IL-13 in eosinophils. *Biochem Biophys Res Commun* 2004; 315(4): 782-7.
- [94] Sessa WC, Kaw S, Zembowicz A, Anggard E, Hecker M, Vane JR. Human polymorphonuclear leukocytes generate and degrade endothelin-1 by two distinct neutral proteases. *J Cardiovasc Pharmacol* 1991; 17 Suppl 7: S34-8.
- [95] Langenfeld MR, Nakhla S, Death AK, Jessup W, Celermajer DS. Endothelin-1 plus oxidized low-density lipoprotein, but neither alone, increase human monocyte adhesion to endothelial cells. *Clin Sci (Lond)* 2001; 101(6): 731-8.
- [96] Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 1994; 76(2): 301-14.
- [97] Zouki C, Baron C, Fournier A, Filep JG. Endothelin-1 enhances neutrophil adhesion to human coronary artery endothelial cells: role of ET(A) receptors and platelet-activating factor. *Br J Pharmacol* 1999; 127(4): 969-79.
- [98] Chen P, Shibata M, Zidovetzki R, Fisher M, Zlokovic BV, Hofman FM. Endothelin-1 and monocyte chemoattractant protein-1 modulation in ischemia and human brain-derived endothelial cell cultures. *J Neuroimmunol* 2001; 116(1): 62-73.
- [99] Noiri E, Hu Y, Bahou WF, Keese CR, Giaever I, Goligorsky MS. Permissive role of nitric oxide in endothelin-induced migration of endothelial cells. *J Biol Chem* 1997; 272(3): 1747-52.
- [100] Battistini B, Chailier P, D'Orleans-Juste P, Briere N, Sirois P. Growth regulatory properties of endothelins. *Peptides* 1993; 14(2): 385-99.
- [101] Distler O, Neidhart M, Gay RE, Gay S. The molecular control of angiogenesis. *Int Rev Immunol* 2002; 21(1): 33-49.
- [102] Salani D, Tarabozetti G, Rosano L, *et al.* Endothelin-1 induces an angiogenic phenotype in cultured endothelial cells and stimulates neovascularization *in vivo*. *Am J Pathol* 2000; 157(5): 1703-11.
- [103] Libby P, Sukhova G, Lee RT, Galis ZS. Cytokines regulate vascular functions related to stability of the atherosclerotic plaque. *J Cardiovasc Pharmacol* 1995; 25 Suppl 2: S9-12.
- [104] Matsuya M, Sasaki H, Aoto H, *et al.* Cell adhesion kinase beta forms a complex with a new member, Hic-5, of proteins localized at focal adhesions. *J Biol Chem* 1998; 273(2): 1003-14.
- [105] Gallelli L, Pelaia G, D'Agostino B, *et al.* Endothelin-1 induces proliferation of human lung fibroblasts and IL-11 secretion through an ET<sub>A</sub> receptor-dependent activation of MAP kinases. *J Cell Biochem* 2005; 96(4): 858-68.
- [106] Marini M, Carpi S, Bellini A, Patalano F, Mattoli S. Endothelin-1 induces increased fibronectin expression in human bronchial

- epithelial cells. *Biochem Biophys Res Commun* 1996; 220(3): 896-9.
- [107] Malemud CJ. Matrix metalloproteinases (MMPs) in health and disease: an overview. *Front Biosci* 2006; 11: 1696-701.
- [108] Shahar I, Fireman E, Topilsky M, *et al.* Effect of endothelin-1 on alpha-smooth muscle actin expression and on alveolar fibroblasts proliferation in interstitial lung diseases. *Int J Immunopharmacol* 1999; 21(11): 759-75.
- [109] Gray MJ, Zadoks RN, Fortes ED, *et al.* Listeria monocytogenes isolates from foods and humans form distinct but overlapping populations. *Appl Environ Microbiol* 2004; 70(10): 5833-41.
- [110] Hafizi S, Wharton J, Chester AH, Yacoub MH. Profibrotic effects of endothelin-1 via the ETA receptor in cultured human cardiac fibroblasts. *Cell Physiol Biochem* 2004; 14(4-6): 285-92.
- [111] Kahaleh MB. Endothelin, an endothelial-dependent vasoconstrictor in scleroderma. Enhanced production and profibrotic action. *Arthritis Rheum* 1991; 34(8): 978-83.
- [112] Dawes KE, Cambrey AD, Campa JS, *et al.* Changes in collagen metabolism in response to endothelin-1: evidence for fibroblast heterogeneity. *Int J Biochem Cell Biol* 1996; 28(2): 229-38.
- [113] Shimajo N, Jesmin S, Zaedi S, *et al.* Eicosapentaenoic acid prevents endothelin-1-induced cardio-myocyte hypertrophy *in vitro* through the suppression of TGF- $\beta$ 1 and phosphorylated JNK. *Am J Physiol Heart Circ Physiol* 2006.
- [114] Yahiaoui L, Villeneuve A, Valderrama-Carvajal H, Burke F, Fixman ED. Endothelin-1 regulates proliferative responses, both alone and synergistically with PDGF, in rat tracheal smooth muscle cells. *Cell Physiol Biochem* 2006; 17(1-2): 37-46.
- [115] Beaucage P, Moreau P. EGF Receptor Transactivation in Angiotensin II and Endothelin Control of Vascular Protein Synthesis *in vivo*. *J Cardiovasc Pharmacol* 2004; 44 Suppl 1: S20-3.
- [116] Li P, Oparil S, Sun JZ, Thompson JA, Chen YF. Fibroblast growth factor mediates hypoxia-induced endothelin-1 receptor expression in lung artery smooth muscle cells. *J Appl Physiol* 2003; 95(2): 643-51; discussion 863.
- [117] Zhang S, Smartt H, Holgate ST, Roche WR. Growth factors secreted by bronchial epithelial cells control myofibroblast proliferation: an *in vitro* co-culture model of airway remodeling in asthma. *Lab Invest* 1999; 79(4): 395-405.
- [118] Holgate ST, Davies DE, Lackie PM, Wilson SJ, Puddicombe SM, Lordan JL. Epithelial-mesenchymal interactions in the pathogenesis of asthma. *J Allergy Clin Immunol* 2000; 105(2 Pt 1): 193-204.
- [119] Vignola AM, Mirabella F, Costanzo G, *et al.* Airway remodeling in asthma. *Chest* 2003; 123(3 Suppl): 417S-22S.
- [120] Ortmann J, Traupe T, Nett P, *et al.* Upregulation of Vascular ET<sub>B</sub> Receptor Gene Expression after Chronic ET<sub>A</sub> Receptor Blockade in Prediabetic NOD Mice. *J Cardiovasc Pharmacol* 2004; 44: S105-S108.
- [121] Ortmann J, Nett PC, Celeiro J, *et al.* Endothelin inhibition delays onset of hyperglycemia and associated vascular injury in type I diabetes: evidence for endothelin release by pancreatic islet beta-cells. *Biochem Biophys Res Commun* 2005; 334(2): 689-95.
- [122] King KL, Winer J, Phillips DM, Quach J, Williams PM, Mather JP. Phenylephrine, endothelin, prostaglandin F<sub>2alpha</sub> and leukemia inhibitory factor induce different cardiac hypertrophy phenotypes *in vitro*. *Endocrine* 1998; 9(1): 45-55.
- [123] Guarda E, Katwa LC, Myers PR, Tyagi SC, Weber KT. Effects of endothelins on collagen turnover in cardiac fibroblasts. *Cardiovasc Res* 1993; 27(12): 2130-4.
- [124] Bergman MR, Cheng S, Honbo N, Piacentini L, Karliner JS, Lovett DH. A functional activating protein 1 (AP-1) site regulates matrix metalloproteinase 2 (MMP-2) transcription by cardiac cells through interactions with JunB-Fra1 and JunB-FosB heterodimers. *Biochem J* 2003; 369(Pt 3): 485-96.
- [125] Iskit AB, Senel I, Sokmensuer C, Guc MO. Endothelin receptor antagonist bosentan improves survival in a murine caecal ligation and puncture model of septic shock. *Eur J Pharmacol* 2004; 506(1): 83-8.
- [126] Mitaka C. Septic shock and endothelin. *Nippon Rinsho* 2004; 62 Suppl 9: 661-4.
- [127] Pfister LA, Tureen JH, Shaw S, *et al.* Endothelin inhibition improves cerebral blood flow and is neuroprotective in pneumococcal meningitis. *Ann Neurol* 2000; 47(3): 329-35.
- [128] Albertini M, Clement MG, Hussain SN. Role of endothelin ETA receptors in sepsis-induced mortality, vascular leakage, and tissue injury in rats. *Eur J Pharmacol* 2003; 474(1): 129-35.
- [129] Forni M, Mazzola S, Ribeiro LA, *et al.* Expression of endothelin-1 system in a pig model of endotoxic shock. *Regul Pept* 2005; 131(1-3): 89-96.
- [130] Pittet JF, Morel DR, Hemsén A, *et al.* Elevated plasma endothelin-1 concentrations are associated with the severity of illness in patients with sepsis. *Ann Surg* 1991; 213(3): 261-4.
- [131] Gandhi CR, Stephenson K, Olson MS. A comparative study of endothelin- and platelet-activating-factor-mediated signal transduction and prostaglandin synthesis in rat Kupffer cells. *Biochem J* 1992; 281 (Pt 2): 485-92.
- [132] McMillen MA, Huribal M, Kumar R, Sumpio BE. Endothelin-stimulated human monocytes produce prostaglandin E2 but not leukotriene B4. *J Surg Res* 1993; 54(4): 331-5.
- [133] Weitzberg E, Lundberg JM, Rudehill A. Elevated plasma levels of endothelin in patients with sepsis syndrome. *Circ Shock* 1991; 33(4): 222-7.
- [134] Wanecek M, Oldner A, Rudehill A, Sollevi A, Alving K, Weitzberg E. Cardiopulmonary dysfunction during porcine endotoxin shock is effectively counteracted by the endothelin receptor antagonist bosentan. *Shock* 1997; 7(5): 364-70.
- [135] Wanecek M, Weitzberg E, Alving K, Rudehill A, Oldner A. Effects of the endothelin receptor antagonist bosentan on cardiac performance during porcine endotoxin shock. *Acta Anaesthesiol Scand* 2001; 45(10): 1262-70.
- [136] Sugiura M, Inagami T, Kon V. Endotoxin stimulates endothelin-release *in vivo* and *in vitro* as determined by radioimmunoassay. *Biochem Biophys Res Commun* 1989; 161(3): 1220-7.
- [137] Camargos ER, Machado CR, Teixeira Jr AL, *et al.* Role of endothelin during experimental Trypanosoma cruzi infection in rats. *Clin Sci (Lond)* 2002; 103 Suppl 48: 64S-67S.
- [138] Davi G, Giammarresi C, Vigneri S, *et al.* Demonstration of Rickettsia Conorii-induced coagulative and platelet activation *in vivo* in patients with Mediterranean spotted fever. *Thromb Haemost* 1995; 74(2): 631-4.
- [139] Shi RJ, Simpson-Haidaris PJ, Lerner NB, Marder VJ, Silverman DJ, Sporn LA. Transcriptional regulation of endothelial cell tissue factor expression during Rickettsia rickettsii infection: involvement of the transcription factor NF-kappaB. *Infect Immun* 1998; 66(3): 1070-5.
- [140] Salomone OA, Caeiro TF, Madoery RJ, *et al.* High plasma immunoreactive endothelin levels in patients with Chagas' cardiomyopathy. *Am J Cardiol* 2001; 87(10): 1217-20; A7.
- [141] Boros M, Massberg S, Baranyi L, Okada H, Messmer K. Endothelin 1 induces leukocyte adhesion in submucosal venules of the rat small intestine. *Gastroenterology* 1998; 114(1): 103-14.
- [142] Anthoni C, Mennigen RB, Rijcken EJ, *et al.* Bosentan, an endothelin receptor antagonist, reduces leukocyte adhesion and inflammation in a murine model of inflammatory bowel disease. *Int J Colorectal Dis* 2005; 1-10.
- [143] Hogaboam CM, Muller MJ, Collins SM, Hunt RH. An orally active non-selective endothelin receptor antagonist, bosentan, markedly reduces injury in a rat model of colitis. *Eur J Pharmacol* 1996; 309(3): 261-9.
- [144] Gulluoglu BM, Kurtel H, Gulluoglu MG, *et al.* Role of endothelins in trinitro-benzene sulfonic acid-induced colitis in rats. *Digestion* 1999; 60(5): 484-92.
- [145] Sanz MJ, Johnston B, Issekutz A, Kubes P. Endothelin-1 causes P-selectin-dependent leukocyte rolling and adhesion within rat mesenteric microvessels. *Am J Physiol* 1999; 277(5 Pt 2): H1823-30.
- [146] Ishizuka T, Takamizawa-Matsumoto M, Suzuki K, Kurita A. Endothelin-1 enhances vascular cell adhesion molecule-1 expression in tumor necrosis factor alpha-stimulated vascular endothelial cells. *Eur J Pharmacol* 1999; 369(2): 237-45.
- [147] McCarron RM, Wang L, Stanimirovic DB, Spatz M. Endothelin induction of adhesion molecule expression on human brain microvascular endothelial cells. *Neurosci Lett* 1993; 156(1-2): 31-4.
- [148] Rijcken E, Kriegelstein CF, Anthoni C, *et al.* ICAM-1 and VCAM-1 antisense oligonucleotides attenuate *in vivo* leukocyte adherence and inflammation in rat inflammatory bowel disease. *Gut* 2002; 51(4): 529-35.

- [149] Rachid MA, Camargos ER, Barcellos L, *et al.* Blockade of endothelin ET<sub>A</sub>/ET<sub>B</sub> receptors favors a role for endothelin during acute *Trypanosoma cruzi* infection in rats. *Microbes Infect* 2006.
- [150] Babaei S, Picard P, Ravandi A, *et al.* Blockade of endothelin receptors markedly reduces atherosclerosis in LDL receptor deficient mice: role of endothelin in macrophage foam cell formation. *Cardiovasc Res* 2000; 48(1): 158-67.
- [151] Sakurai-Yamashita Y, Yamashita K, Yoshida A, *et al.* Rat peritoneal macrophages express endothelin ET<sub>B</sub> but not endothelin ET<sub>A</sub> receptors. *Eur J Pharmacol* 1997; 338(2): 199-203.
- [152] Boss C, Bolli MH, Weller T, Fischli W, Clozel M. Bis-sulfonamides as endothelin receptor antagonists. *Bioorg Med Chem Lett* 2003; 13(5): 951-4.
- [153] Weller, T., Bolli, M., Boss, C., Clozel, M., Fischli, W.: US20067091201 (2006).
- [154] Chan MF, Kois A, Verner EJ, *et al.* The discovery and structure-activity relationships of nonpeptide, low molecular weight antagonists selective for the endothelin ET<sub>B</sub> receptor. *Bioorg Med Chem* 1998; 6(12): 2301-16.
- \*[155] Chan, M.F., Raju, B.G., Kois, A., Verner, E.J., Wu, C., Castillo, R.S., Yalamoori, V., Balaji, V.N., Ramnarayan, K.: US6541498 (2003).
- [156] Parissis JT, Korovesis S, Giazitoglou E, Kalivas P, Katritsis D. Plasma profiles of peripheral monocyte-related inflammatory markers in patients with arterial hypertension. Correlations with plasma endothelin-1. *Int J Cardiol* 2002; 83(1): 13-21.
- [157] Quehenberger P, Bierhaus A, Fasching P, *et al.* Endothelin 1 transcription is controlled by nuclear factor-kappaB in AGE-stimulated cultured endothelial cells. *Diabetes* 2000; 49(9): 1561-70.
- [158] Gaddipati JP, Sundar SV, Calemine J, Seth P, Sidhu GS, Maheshwari RK. Differential regulation of cytokines and transcription factors in liver by curcumin following hemorrhage/resuscitation. *Shock* 2003; 19(2): 150-6.
- [159] Browatzki M, Schmidt J, Kubler W, Kranzhofer R. Endothelin-1 induces interleukin-6 release via activation of the transcription factor NF-kappaB in human vascular smooth muscle cells. *Basic Res Cardiol* 2000; 95(2): 98-105.
- [160] Gallois C, Habib A, Tao J, *et al.* Role of NF-kappaB in the antiproliferative effect of endothelin-1 and tumor necrosis factor-alpha in human hepatic stellate cells. Involvement of cyclooxygenase-2. *J Biol Chem* 1998; 273(36): 23183-90.
- [161] Sharmin S, Shiota M, Murata E, *et al.* A novel bioactive 31-amino acid ET-1 peptide stimulates eosinophil recruitment and increases the levels of eotaxin and IL-5. *Inflamm Res* 2002; 51(4): 195-200.
- [162] Finsnes F, Skjonsberg OH, Tonnessen T, Naess O, Lyberg T, Christensen G. Endothelin production and effects of endothelin antagonism during experimental airway inflammation. *Am J Respir Crit Care Med* 1997; 155(4): 1404-12.
- [163] Finsnes F, Christensen G, Lyberg T, Sejersted OM, Skjonsberg OH. Increased synthesis and release of endothelin-1 during the initial phase of airway inflammation. *Am J Respir Crit Care Med* 1998; 158(5 Pt 1): 1600-6.
- [164] Finsnes F, Skjonsberg OH, Lyberg T, Christensen G. Endothelin-1 production is associated with eosinophilic rather than neutrophilic airway inflammation. *Eur Respir J* 2000; 15(4): 743-50.
- [165] Sampaio AL, Rae GA, Henriques MG. Participation of endogenous endothelins in delayed eosinophil and neutrophil recruitment in mouse pleurisy. *Inflamm Res* 2000; 49(4): 170-6.
- [166] Sampaio AL, Rae GA, Henriques MM. Role of endothelins on lymphocyte accumulation in allergic pleurisy. *J Leukoc Biol* 2000; 67(2): 189-95.
- [167] Brands M, Erguden JK, Hashimoto K, *et al.* Novel, selective indole-based ECE inhibitors: lead optimization via solid-phase and classical synthesis. *Bioorg Med Chem Lett* 2005; 15(19): 4201-5.
- \*[168] Weigand, S., Frey, R., Stasch, J.P.: WO06037491A1 (2006).
- [169] Ammarguella F, Larouche I, Schiffrin EL. Myocardial fibrosis in DOCA-salt hypertensive rats: effect of endothelin ET<sub>A</sub> receptor antagonism. *Circulation* 2001; 103(2): 319-24.
- [170] Ammarguella FZ, Gannon PO, Amiri F, Schiffrin EL. Fibrosis, matrix metalloproteinases, and inflammation in the heart of DOCA-salt hypertensive rats: role of ET<sub>A</sub> receptors. *Hypertension* 2002; 39(2 Pt 2): 679-84.
- [171] Hocher B, George I, Rebstock J, *et al.* Endothelin system-dependent cardiac remodeling in renovascular hypertension. *Hypertension* 1999; 33(3): 816-22.
- [172] Liu G, Kozmina NS, Winn M, *et al.* Design, synthesis, and activity of a series of pyrrolidine-3-carboxylic acid-based, highly specific, orally active ET<sub>B</sub> antagonists containing a diphenylmethylamine acetamide side chain. *J Med Chem* 1999; 42(18): 3679-89.
- [173] Jae HS, Winn M, von Geldern TW, *et al.* Pyrrolidine-3-carboxylic acids as endothelin antagonists. 5. Highly selective, potent, and orally active ET<sub>A</sub> antagonists. *J Med Chem* 2001; 44(23): 3978-84.
- [174] Winn, M., Boyd, S.A., Hutchins, C.W., Jae, H.S., Tasker, A.S., Vongeldern, T.W., Kester, J.A., Sorensen, B.K.: CA2195677C (2005).
- [175] Winn, M., Boyd, S.A., Hutchins, C.W., Jae, H.S., Tasker, A.S., Von Geldern, T.W., Kester, J.A., Sorensen, B.K., Szczepankiewicz, B.G., Henry, K.J. Jr., Liu, G., Wittenberger, S.J., King, S.A.: EP0885215B1 (2006).
- [176] Bolli MH, Marfurt J, Grisostomi C, *et al.* Novel benzo[1,4] diazepin-2-one derivative as endothelin receptor. *J Med Chem* 2004; 47(11): 2776-95.
- [177] Actelion Pharmaceuticals Ltd.: IL0162534A0 (2005).
- [178] Behrend M. The endothelin receptor antagonist TAK-044 in the treatment of reperfusion injury in organ transplantation. *Expert Opin Investig Drugs* 1999; 8(7): 1079-91.
- [179] Guttman RD, Forbes RD, Zheng SX. The pathophysiology of chronic rejection. *Am J Med Sci* 1997; 313(5): 302-4.
- [180] Uhlmann D, Gabel G, Ludwig S, *et al.* Effects of ET<sub>A</sub> receptor antagonism on proinflammatory gene expression and microcirculation following hepatic ischemia/reperfusion. *Microcirculation* 2005; 12(5): 405-19.
- [181] Uhlmann D, Gaebel G, Armann B, *et al.* Attenuation of proinflammatory gene expression and microcirculatory disturbances by endothelin A receptor blockade after orthotopic liver transplantation in pigs. *Surgery* 2006; 139(1): 61-72.
- [182] Witzigmann H, Ludwig S, Armann B, *et al.* Endothelin(A) receptor blockade reduces ischemia/reperfusion injury in pig pancreas transplantation. *Ann Surg* 2003; 238(2): 264-74.
- [183] Hansen A, Bekeredjian R, Filusch A, *et al.* Cardioprotective Effects of the Novel Selective Endothelin-A Receptor Antagonist BSF 461314 in Ischemia-Reperfusion Injury. *J Am Soc Echocardiogr* 2005; 18(11): 1213-20.
- [184] Camargos ER, Rocha LL, Rachid MA, *et al.* Protective role of ETA endothelin receptors during the acute phase of *Trypanosoma cruzi* infection in rats. *Microbes Infect* 2004; 6(7): 650-6.
- [185] Sampaio AL, Rae GA, Henriques MG. Effects of endothelin ETA receptor antagonism on granulocyte and lymphocyte accumulation in LPS-induced inflammation. *J Leukoc Biol* 2004; 76(1): 210-6.
- [186] Takeda M, Ishida T, Ishimura T, Fujisawa M. Correlation between endothelin expression in early post-transplant biopsy specimens and long-term allograft function in living-related renal transplantation. *Clin Transplant* 2006; 20(1): 26-9.
- [187] Fedak PW, Rao V, Verma S, *et al.* Combined endothelial and myocardial protection by endothelin antagonism enhances transplant allograft preservation. *J Thorac Cardiovasc Surg* 2005; 129(2): 407-15.
- [188] Mueller TF, Ma C, Lederer JA, Perkins DL. Differentiation of stress, metabolism, communication, and defense responses following transplantation. *J Leukoc Biol* 2003; 73(3): 379-90.
- [189] Gottmann U, van der Woude FJ, Braun C. Endothelin receptor antagonists: a new therapeutic option for improving the outcome after solid organ transplantation? *Curr Vasc Pharmacol* 2003; 1(3): 281-99.
- [190] Lattmann T, Hein M, Horber S, *et al.* Activation of pro-inflammatory and anti-inflammatory cytokines in host organs during chronic allograft rejection: role of endothelin receptor signaling. *Am J Transplant* 2005; 5(5): 1042-9.
- [191] Nett PC, Heisey DM, Fernandez LA, Sollinger HW, Pirsch JD. Association of cytomegalovirus disease and acute rejection with graft loss in kidney transplantation. *Transplantation* 2004; 78(7): 1036-41.

- [192] Busauschina A, Schnuelle P, van der Woude FJ. Cyclosporine nephrotoxicity. *Transplant Proc* 2004; 36(2 Suppl): 229S-233S.
- [193] Bunchman TE, Brookshire CA. Cyclosporine-induced synthesis of endothelin by cultured human endothelial cells. *J Clin Invest* 1991; 88(1): 310-4.
- [194] Komuro I, Kurihara H, Sugiyama T, Yoshizumi M, Takaku F, Yazaki Y. Endothelin stimulates c-fos and c-myc expression and proliferation of vascular smooth muscle cells. *FEBS Lett* 1988; 238(2): 249-52.
- [195] Kocik M, Malek I, Glagolicova A, Pirk J. The effect of cyclosporin A on the level of big endothelin in patients one year after orthotopic heart transplantation. *Transpl Int* 2004; 17(2): 65-70.
- [196] Adams DH, Russell ME, Hancock WW, Sayegh MH, Wyner LR, Karnovsky MJ. Chronic rejection in experimental cardiac transplantation: studies in the Lewis-F344 model. *Immunol Rev* 1993; 134: 5-19.
- [197] Braun C, Conzelmann T, Vetter S, *et al.* Treatment with a combined endothelin A/B-receptor antagonist does not prevent chronic renal allograft rejection in rats. *J Cardiovasc Pharmacol* 2000; 36(4): 428-37.
- [198] Knoll T, Oltersdorf J, Gottmann U, *et al.* Influence of acute selective endothelin-receptor-A blockade on renal hemodynamics in a rat model of chronic allograft rejection. *Transpl Int* 2003; 16(6): 425-9.
- \*[199] Liu, K., Yu, W., Liang, Y., Wang, H., Zhao, Y., Ding, Z.: US6953780 (2005).
- [200] Yu W, Liang Y, Liu K, Zhao Y, Fei G, Wang H. The chemical syntheses and bioactivities of novel peptide-based endothelin antagonists. *J Pept Res* 2002; 59(3): 134-8.
- [201] Bolli MH, Boss C, Clozel M, Fischli W, Hess P, Weller T. The use of sulfonamide pyrimidines incorporating an unsaturated side chain as endothelin receptor antagonists. *Bioorg Med Chem Lett* 2003; 13(5): 955-9.
- [202] Ono K, Matsumori A, Shioi T, Furukawa Y, Sasayama S. Contribution of endothelin-1 to myocardial injury in a murine model of myocarditis: acute effects of bosentan, an endothelin receptor antagonist. *Circulation* 1999; 100(17): 1823-9.
- [203] Pandey AS, Stewart DJ, Cernacek P, Dawood F, Wen WH, Liu P. Chronic endothelin-1 blockade preserves myocardial contractility in dilated cardiomyopathy. *J Cardiovasc Pharmacol* 1998; 31 Suppl 1: S306-8.
- [204] Wanecek M, Oldner A, Sundin P, Alving K, Weitzberg E, Rudehill A. Effects on haemodynamics by selective endothelin ET<sub>B</sub> receptor and combined endothelin ET<sub>A</sub>/ET<sub>B</sub> receptor antagonism during endotoxin shock. *Eur J Pharmacol* 1999; 386(2-3): 235-45.
- [205] Jelicks LA, Chandra M, Shirani J, *et al.* Cardioprotective effects of phosphoramidon on myocardial structure and function in murine Chagas' disease. *Int J Parasitol* 2002; 32(12): 1497-506.
- [206] Wittner M, Christ GJ, Huang H, *et al.* Trypanosoma cruzi induces endothelin release from endothelial cells. *J Infect Dis* 1995; 171(2): 493-7.
- [207] Tanowitz HB, Wittner M, Morris SA, *et al.* The putative mechanistic basis for the modulatory role of endothelin-1 in the altered vascular tone induced by Trypanosoma cruzi. *Endothelium* 1999; 6(3): 217-30.
- [208] Brener Z, Gazzinelli RT. Immunological control of Trypanosoma cruzi infection and pathogenesis of Chagas' disease. *Int Arch Allergy Immunol* 1997; 114(2): 103-10.
- [209] Aliberti JC, Machado FS, Souto JT, *et al.* beta-Chemokines enhance parasite uptake and promote nitric oxide-dependent microbistatic activity in murine inflammatory macrophages infected with Trypanosoma cruzi. *Infect Immun* 1999; 67(9): 4819-26.
- [210] Korn JH, Mayes M, Matucci Cerinic M, *et al.* Digital ulcers in systemic sclerosis: prevention by treatment with bosentan, an oral endothelin receptor antagonist. *Arthritis Rheum* 2004; 50(12): 3985-93.
- [211] Ligeiro D, Fonseca JE, Abade O, *et al.* Influence of Human Leukocyte Antigen-DRB1 on the susceptibility to rheumatoid arthritis and on the production of anti-cyclic citrullinated peptide antibodies in a Portuguese population. *Ann Rheum Dis* 2006.
- [212] Barton M, Mullins JJ, Bailey MA, Kretzler M. Role of endothelin receptors for renal protection and survival in hypertension: Waiting for clinical trials. *Hypertension* 2006; 48: 834-7.