

#### Review

# Mitochondria and vascular pathology

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#### Abstract:

Functional and structural changes in mitochondria are caused by the opening of the mitochondrial permeability transition pore (PTP) and by the mitochondrial generation of reactive oxygen species (ROS). These two processes are linked in a vicious cycle that has been extensively documented in ischemia/reperfusion injuries of the heart, and the same processes likely contribute to vascular pathology. For instance, the opening of the PTP causes cell death in isolated endothelial and vascular smooth muscle cells. Indeed, atherosclerosis is exacerbated when mitochondrial antioxidant defenses are hampered, but a decrease in mitochondrial ROS formation reduces atherogenesis.

Determining the exact location of ROS generation in mitochondria is a relevant and still unanswered question. The respiratory chain is generally believed to be a main site of ROS formation. However, several other mitochondrial components likely contribute to ROS generation. Recent reports highlight the relevance of monoamine oxidases (MAO) and p66<sup>Shc</sup>. For example, the absence of p66<sup>Shc</sup> in hypercholesterolemic mice has been reported to reduce the occurrence of foam cells and early atherogenic lesions. On the other hand, MAO inhibition has been shown to reduce oxidative stress in many cell types eliciting significant protection from myocardial ischemia. In conclusion, evidence will be presented to demonstrate that (i) mitochondria are major sites of ROS formation; (ii) an increase in mitochondrial ROS formation and/or a decrease in mitochondrial antioxidant defenses exacerbate atherosclerosis; and (iii) mitochondrial dysfunction is likely a relevant mechanism underlying several risk factors (i.e., diabetes, hyperlipidemia, hypertension) associated with atherosclerosis.

Key words: oxidative stress, mitochondria, p66<sup>Shc</sup>, monoamine oxidase

**Abbreviations:** apoE – apolipoprotein E, MAO – monoamine oxidase, PTP – mitochondrial permeability transition pore, ROS – reactive oxygen species, SOD – superoxide dismutase

#### Mitochondrial mechanisms of cell injury

The energy-linked processes and metabolic activities occurring in mitochondria are required to maintain

cell viability. This uncomplicated conception is paradoxically counterbalanced by the well-established notion that mitochondria play a significant role in cell death [8, 45, 58]. In addition to the profound imbalance between ATP synthesis and utilization that occurs as a consequence of mitochondrial dysfunction [31, 71], the impairment of ionic homeostasis [12, 34, 43, 61, 69] and the formation of reactive oxygen species (ROS) [15, 63, 77] represent two additional processes through which mitochondria accelerate, or

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even determine, the evolution of cell injury toward necrosis or apoptosis [8, 32, 45, 47, 58].

A large body of evidence supports the concept that ROS are formed within mitochondria under physiological and pathological conditions [3, 33, 40, 56, 77]. The superoxide anion  $(O_2 \cdot \overline{\ })$ , formed by Complex I and III, is rapidly transformed into hydrogen peroxide  $(H_2O_2)$  by a family of metalloenzymes, the superoxide dismutases (SOD) [38]. Particularly relevant in this process is the mitochondrial form of SOD (MnSOD or SOD-2). Widespread organ damage associated with severe mitochondrial dysfunction has been observed in mice lacking SOD-2 [52]. Recent work has demonstrated that CuZnSOD (SOD-1), commonly referred to as the cytosolic isoform, is also present in the mitochondrial intermembrane space [60].

The major link between mitochondria and vascular derangements is oxidative stress [4, 26, 35, 42, 50]. In particular, the development of atherosclerosis appears to depend on the mitochondrial metabolism of ROS. In fact, when mitochondrial antioxidant defenses are hampered, atherosclerosis is exacerbated, whereas a decrease in mitochondrial ROS formation reduces atherogenesis [50]. For instance, the absence of SOD-2 increases mtDNA damage and accelerates atherosclerosis in apoE knockout mice [59]. These findings are consistent with results obtained in ischemia/reperfusion experiments, which show that SOD-2 overexpression elicits cardioprotection [19], whereas a heterozygous deficiency of this enzyme impairs postischemic recovery of the heart [1].

In addition to the damage to all of cell's components, oxidative stress increases the occurrence of cell death, especially by apoptosis, which greatly contributes to the progress of atherosclerotic lesions. In this respect, an important consequence of ROS accumulation is increased susceptibility to opening of the mitochondrial permeability transition pore (PTP) [7]. PTP opening is especially sensitive to oxidative stress, since it is favored by decreases in NADPH(H<sup>+</sup>)/NADP<sup>+</sup> and -SH/-S-S ratios [23, 24]. Recent evidence suggests that PTP opening and ROS formation are linked in a vicious cycle. In addition to being a likely consequence of oxidative stress, PTP opening has been shown to increase mitochondrial ROS formation in cardiac myocytes [41].

Despite the large body of evidence that relates PTP opening to cell death, especially in the case of an ischemia/reperfusion injury to the heart [29], definite proof of PTP involvement in atherogenesis is lacking.

While data obtained *in vitro* indicate that PTP opening causes cell death in isolated endothelial and vascular smooth muscle cells [26, 28], no information on the effect of pharmacological or genetic inhibition of the PTP on atherogenesis *in vivo* is available.

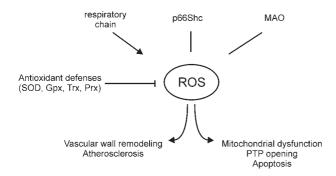
The mitochondrial formation of ROS may be modulated by NO· [67, 72] as a consequence of the inhibition of cytochrome oxidase [6, 13, 20, 44, 77]. This reversible process can be transformed into irreversible damage to the respiratory chain when NO-formation is sustained. Indeed, the reaction of NO-with  $O_2$ · generates peroxynitrite, which can cause the irreversible nitration of proteins [5]. Interestingly, a proteomic study showed that one-third of the proteins nitrated during an inflammatory challenge were mitochondrial in origin [2].

It must be noted that ROS are also produced within mitochondria at sites other than the inner mitochondrial membrane [30, 33], by proteins such as monoamine oxidase (MAO) and p66Shc. These additional mitochondrial processes produce significant amounts of ROS. For instance, in brain mitochondria MAO activity results in steady state concentrations of H<sub>2</sub>O<sub>2</sub> that are 48-fold higher than those originating from the respiratory chain in the presence of antimycin A [15]. Therefore, the generation of ROS, especially H<sub>2</sub>O<sub>2</sub>, by mitochondria is not just an unfortunate side effect of respiration, but can also be catalyzed by specific enzymes, such as MAO and p66Shc. It is tempting to speculate that, under physiological conditions, the inner mitochondrial membrane scavenges ROS produced at other mitochondrial or cellular sites. In such a scenario, the increase in ROS formation detected under pathological conditions might result, at least in part, from a dysfunction of the inner mitochondrial membrane's scavenging abilities.

This review will focus on the relevant contributions of MAO and p66<sup>Shc</sup> to both mitochondrial ROS formation and cell injury, as summarized in Figure 1.

# Oxidative stress, vascular pathology and p66<sup>Shc</sup>

The Shc family of proteins gets its name from the abbreviation of the <u>Src homology</u> 2 domain and a <u>collagen-homology</u> region [55]. Two members of this adaptor protein family, p52<sup>Shc</sup> and p46<sup>Shc</sup>, are phos-



**Fig. 1.** Mitochondrial sources of reactive oxygen species (ROS). If not efficiently neutralized by intrinsic antioxidant systems, ROS produced by the respiratory chain, p66 shc and MAO in mitochondria can trigger secondary signaling pathways that lead to vascular wall remodeling and atherosclerosis. Alternatively, excess ROS production at these sites can also induce mitochondrial dysfunction resulting in PTP opening and apoptosis. Gpx – glutathione peroxidase, MAO – monoamine oxidase, Prx – peroxiedoxin, PTP – mitochondrial permeability transition pore, SOD – superoxide dismutase, Trx – thioredoxin

phorylated in response to mitotic signals and result in Ras activation. A third member of the family, p66<sup>Shc</sup>, which is not involved in Ras activation, is composed of the entire p52<sup>Shc</sup>/p46<sup>Shc</sup> sequence and an additional amino-terminal proline-rich region, named CH2, which contains a serine phosphorylation site implicated in oxidative stress signaling. Indeed, p66<sup>Shc</sup> has been shown to play a significant role in a wide range of pathological conditions related to oxidative stress [22, 55]. p66<sup>Shc</sup> garnered a great deal of attention when it was demonstrated that its deletion resulted in a 30% increase in lifespan [54]. Subsequently, studies carried out in a wide range of experimental models established quite clearly that ROS formation is reduced in cells lacking p66Shc, and that systemic and intracellular markers of oxidative stress are diminished in  $p66^{Shc-/-}$  mice [39, 62, 76]. The relationship of p66<sup>Shc</sup> to ROS formation was elucidated by showing that it partially localizes within mitochondria, where it catalyzes electron transfer from cytochrome c to oxygen [39]. More recent work suggests that PKC β phosphorylation of p66<sup>Shc</sup> on Ser<sup>36</sup> could cause its translocation to mitochondria [65]. It seems that the increase in mitochondrial ROS formation caused by p66Shc amplifies the PKC β signaling triggered by an initial oxidative stress.

The link between p66<sup>Shc</sup> and vascular pathology was originally highlighted by studying the effects of hypercholesterolemia [57]. When compared to wild type littermates, hypercholesterolemic p66<sup>Shc-/-</sup> mice displayed reduced levels of isoprostane and oxidized

LDL, a decreased number of foam cells, and a reduction in the extent of both apoptosis and early atherogenic lesions. These initial findings prompted several additional studies, especially in the field of diabetic vasculopathy. In studies of streptozotocin-induced diabetes, p66Shc-/- mice showed glycemic levels similar to wild type mice, but were protected against glomerulopathy as shown by the preservation of renal structure and function, and by a marked reduction in oxidative stress [53]. Using the same experimental model, p66<sup>Shc-/-</sup> mice were found to have unimpaired acetylcholine-induced vasorelaxation, due to the unchanged availability of NO [16]. In addition, the lack of p66Shc was shown to protect against diabetic cardiomyopathy by preventing the senescence of cardiac progenitor cells, which hampers cardiac and vascular cell turnover [70].

The findings obtained in mice lacking p66<sup>Shc</sup> prove beyond any doubt that mitochondrial ROS formation is not just an accidental by-product of the respiratory chain, and that most intracellular oxidative stress originates in mitochondria. However, at present the translation of these concepts into clinical practice is limited by the lack of drugs that prevent ROS formation by p66<sup>Shc</sup>.

### Monoamine oxidase

#### Structural features and biochemical role

Monoamine oxidase, a flavoenzyme located within the outer mitochondrial membrane, is responsible for the oxidative deamination of neurotransmitters and dietary amines. It exists in two isoforms, MAO-A and B, which differ in substrate specificity and inhibitor sensitivity [37]. In both isoforms, FAD is covalently bound to a cysteine residue [36]. MAO-A and B are anchored to the outer membrane of mitochondria through a C-terminal  $\alpha$ -helix segment that protrudes from the basal face of the structure. The analysis of crystal diffraction data indicates that MAO-B crystallizes as a dimer; with each monomer containing a C-terminal membrane bound domain, a FAD binding domain and a substrate-binding domain [11]. In contrast to human MAO-B, human MAO-A crystallizes as a monomer [27]. Analysis of residue side chains in both active sites shows substrates to have less freedom of rotation in the MAO-B site than in MAO-A. The structural basis for this difference can be partially attributed to the conformational differences in the region from residue 200 to 215 that constitutes the "cavity shaping loop" in both isoforms. This loop has a more extended conformation in MAO-A and a more compact conformation in MAO-B.

In peripheral tissues, the MAO isoforms are involved in the oxidative catabolism of amines from the blood, and in preventing dietary amines from entering circulation. In the central and peripheral nervous system, intraneuronal MAO-A and -B protect neurons from exogenous amines, terminate the actions of amine neurotransmitters and regulate the contents of intracellular amine stores [79].

MAO-A, which preferentially catalyzes the oxidative deamination of norepinephrine (NE) and serotonin (5-HT), is inhibited by low concentrations of clorgyline. In contrast, MAO-B, which has a higher affinity for phenylethylamine and benzylamine, is inhibited by selegiline [79]. Both isoforms catalyze the deamination of dopamine, tyramine, octopamine and tryptamine, and are inhibited by pargyline. MAO catalyzes the following reaction:

$$RCH_2NR'R'' + O_2 + H_2O \rightarrow RCHO + NHR'R' + H_2O_2$$

Kinetic studies have shown that amine binding to the enzyme precedes oxygen binding [75]. In the first step, the reduction of the FAD cofactor yields an aldehyde intermediate and an ammonium, while, in a second step, the prosthetic group is oxidixed with the concomitant production of hydrogen peroxide.

$$RCH_2NH_2 + MAO \rightarrow RCHO + NH_4^+ + MAO$$
-reduced  
 $MAO$ -reduced  $+ O_2 \rightarrow MAO + H_2O_2$ 

The aldehyde intermediates are rapidly metabolized to the corresponding carboxylic acids by the action of aldehyde dehydrogenase (ALDH). A failure of this latter enzyme might increase the deleterious aspects of MAO activity by generating potentially harmful aldehyde compounds, and magnifying the damage done by MAO-induced H<sub>2</sub>O<sub>2</sub> formation. Indeed, a decrease in ALDH activity appears to be involved in both oxidative stress and nitrate tolerance [25, 78], whereas increased ALDH activity has been reported to result in decreased injury to ischemic hearts [18].

The main physiological role of MAO is the degradation of endogenous monoamine neurotransmitters and dietary amines, such as tyramine, which may

cause hypertensive crises if not properly catabolized [79]. Similarly, MAO-B in microvessels and at the blood-brain barrier has a protective function acting as a metabolic barrier, and preventing the entrance of xenobiotic and potentially toxic neurotransmitters.

The deletion of the MAO-A and MAO-B genes has proven their important role in neurotransmitter metabolism and behavior. MAO-A knockout mice have elevated brain levels of serotonin, norepinephrine and, to a lesser extent, dopamine [17], whereas only 2-phenylethylamine levels are increased in MAO-B knockout mice [79]. Both MAO-A and -B knockout mice show an increased response to stress, similar to that observed after the administration of non-selective MAO inhibitors. However, these studies, and the deletion of both MAO-A and MAO-B, in a rare form of human Norrie disease, indicate that MAO is not essential for survival [48]. Gene deletion has shown that MAO-A activity is important during development. A compulsive aggressive behavior results from a lack of MAO-A function in humans [14] and mice [73]. This effect, which might reflect the importance of serotonin during development, can be mimicked by administering the MAO-A inhibitor clorgyline during the early postnatal period.

The distribution of MAO in the brain has been well studied. MAO-A is prevalently found in noradrenergic neurons, whereas MAO-B has been detected in serotoninergic and histaminergic neurons and in glial cells [49]. With respect to peripheral tissues, MAO-A has been found in placenta, liver, intestine and thyroid gland, while platelets, liver and kidney contain mainly MAO-B. Human cardiomyocytes contain both enzymes, although MAO-A is the predominant isoform [74].

## The therapeutic potential of MAO inhibitors

MAO's roles, in terminating the action of neurotransmitters in the central and peripheral nervous system and in the oxidation of dietary amines in extraneuronal tissues, have been extensively studied. However, less attention has been paid to the products of MAO activity. Monoamine catabolism results in the formation of aldehydes, ammonium cations and H<sub>2</sub>O<sub>2</sub>.

MAOs are involved in numerous pathologies. The important role of MAOs in neuronal and psychiatric disorders is demonstrated by the beneficial effects elicited by MAO inhibitors. The therapeutic potential of MAO inhibition was realized in the early 1950s, when iproniazid, an antituberculosis treatment, was

shown to improve mood while reducing MAO activity [66, 68]. A wide spectrum of MAO inhibitors are available today and these are proving to have therapeutic value in several pathologies, including affective disorders, neurodegenerative diseases, stroke and aging [68, 79]. MAO inhibitors are distinguished on the basis of their specificity for each isoform and the nature of their binding to the enzyme [68, 79]. They can be classified into three groups: (i) irreversible and non-selective inhibitors, such as phenelzine and tranylcypromine; (ii) irreversible and selective inhibitors, such as selegiline for MAO-B and clorgyline for MAO-A; and (iii) reversible and selective inhibitors, such as moclobemide for MAO-A and lazabemide for MAO-B.

MAO-B seems to be involved in the loss of dopaminergic neurons that occurs in Parkinson's disease, most likely due to increased dopamine catabolism, which results in an elevated production of the reactive oxygen species responsible for the oxidative damage in nigrostriatal neurons. Indeed, MAO-B inhibition has been shown to afford neuroprotection [79]. Furthermore, an increase in brain MAO-B activity is also associated with diseases such as Alzheimer's and Huntington's disease. Depression, panic attacks and personality disorders are also associated with changes in dopaminergic, noradrenergic and serotoninergic neurotransmission, which are regulated by both isoforms of MAO [79].

In addition to being implicated in neurodegenerative diseases, the MAO isoforms, especially MAO-A, have been shown to play a prominent role in myocardial injury caused by post-ischemic reperfusion [9]. Preliminary evidence also suggests that MAO-A contributes to the maladaptive evolution from myocardial hypertrophy to failure [46]. In particular, MAO-A has been demonstrated to be an important source of ROS in the receptor-independent apoptotic effects of serotonin in isolated cardiomyocytes and in post ischemic myocardial injury [9, 10]. In fact, MAO-dependent increases in ROS production appear to be relevant in serotonin-induced myocyte hypertrophy in vitro [10]. In addition, MAO-A can promote apoptosis through ROS-dependent sphingosine kinase inhibition that results in the accumulation of ceramide [64]. With respect to vasculature, MAO-A-mediated ROS production has been shown to induce mitogenic signaling in smooth muscle cells by a process that may involve the activation of the metalloproteinase MMP-2, which likely contributes to vascular wall remodeling [21]. Interestingly, MAO-A activity has been reported to increase with aging [51]. It is tempting to speculate that the resulting increase in H<sub>2</sub>O<sub>2</sub> formation might contribute to aging-associated pathologies, such as congestive heart failure and vascular pathologies.

Considering MAO's important role as a source of  $H_2O_2$  in both the brain and in the heart following post-I/R cardiac injury, MAO inhibition will likely represent an important tool for both the study and the treatment of vascular pathologies that share oxidative stress as a common denominator.

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#### References:

- Asimakis GK, Lick S, Patterson C: Postischemic recovery of contractile function is impaired in SOD2<sup>+/-</sup> but not SOD1<sup>+/-</sup> mouse hearts. Circulation, 2002, 105, 981–986.
- Aulak KS, Miyagi M, Yan L, West KA, Massillon D, Crabb JW, Stuehr DJ: Proteomic method identifies proteins nitrated in vivo during inflammatory challenge. Proc Natl Acad Sci USA, 2001, 98, 12056–12061.
- Balaban RS, Nemoto S, Finkel T: Mitochondria, oxidants, and aging. Cell, 2005, 120, 483

  –495.
- 4. Bauersachs J, Widder JD: Endothelial dysfunction in heart failure. Pharmacol Rep, 2008, 60, 119–126.
- 5. Beckman JS, Koppenol WH: Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. Am J Physiol, 1996, 271, C1424–C1437.
- Beltran B, Mathur A, Duchen MR, Erusalimsky JD, Moncada S: The effect of nitric oxide on cell respiration: A key to understanding its role in cell survival or death. Proc Natl Acad Sci USA, 2000, 97, 14602–14607.
- Bernardi P, Krauskopf A, Basso E, Petronilli V, Blachly-Dyson E, Di Lisa F, Forte MA: The mitochondrial permeability transition from in vitro artifact to disease target. FEBS J, 2006, 273, 2077–2099.
- 8. Bernardi P, Petronilli V, Di Lisa F, Forte M: A mitochondrial perspective on cell death. Trends Biochem Sci, 2001, 26, 112–117.
- Bianchi P, Kunduzova O, Masini E, Cambon C, Bani D, Raimondi L, Seguelas MH et al.: Oxidative stress by monoamine oxidase mediates receptor-independent cardiomyocyte apoptosis by serotonin and postischemic myocardial injury. Circulation, 2005, 112, 3297–3305.
- Bianchi P, Pimentel DR, Murphy MP, Colucci WS, Parini A: A new hypertrophic mechanism of serotonin in cardiac myocytes: receptor-independent ROS generation. FASEB J, 2005, 19, 641–643.

- Binda C, Newton-Vinson P, Hubálek F, Edmondson DE, Mattevi A: Structure of human monoamine oxidase B, a drug target for the treatment of neurological disorders. Nat Struct Biol, 2002, 9, 22–26.
- Brookes PS, Yoon Y, Robotham JL, Anders MW, Sheu SS: Calcium, ATP, and ROS: a mitochondrial love-hate triangle. Am J Physiol Cell Physiol, 2004, 287, C817–C833.
- Brown GC, Cooper CE: Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. FEBS Lett, 1994, 356, 295–298.
- Brunner HG, Nelen M, Breakefield XO, Ropers HH, van Oost BA: Abnormal behavior associated with a point mutation in the structural gene for monoamine oxidase A. Science, 1993, 262, 578–580.
- 15. Cadenas E, Davies KJ: Mitochondrial free radical generation, oxidative stress, and aging. Free Radic Biol Med, 2000, 29, 222–230.
- Camici GG, Schiavoni M, Francia P, Bachschmid M, Martin-Padura I, Hersberger M, Tanner FC et al.: Genetic deletion of p66<sub>Shc</sub> adaptor protein prevents hyperglycemia-induced endothelial dysfunction and oxidative stress. Proc Natl Acad Sci USA, 2007, 104, 5217–5222.
- Cases O, Seif I, Grimsby J, Gaspar P, Chen K, Pournin S, Muller U et al.: Aggressive behavior and altered amounts of brain serotonin and norepinephrine in mice lacking MAOA. Science, 1995, 268, 1763–1766.
- 18. Chen CH, Budas GR, Churchill EN, Disatnik MH, Hurley TD, Mochly-Rosen D: Activation of aldehyde dehydrogenase-2 reduces ischemic damage to the heart. Science, 2008, 321, 1493–1495.
- Chen Z, Siu B, Ho YS, Vincent R, Chua CC, Hamdy RC, Chua BH: Overexpression of MnSOD protects against myocardial ischemia/reperfusion injury in transgenic mice. J Mol Cell Cardiol, 1998, 30, 2281–2289.
- Cleeter MW, Cooper JM, Darley-Usmar VM, Moncada S, Schapira AH: Reversible inhibition of cytochrome c oxidase, the terminal enzyme of the mitochondrial respiratory chain, by nitric oxide. Implications for neurodegenerative diseases. FEBS Lett, 1994, 345, 50–54.
- Coatrieux C, Sanson M, Negre-Salvayre A, Parini A, Hannun Y, Itohara S, Salvayre R, Auge N: MAO-Ainduced mitogenic signaling is mediated by reactive oxygen species, MMP-2, and the sphingolipid pathway. Free Radic Biol Med, 2007, 43, 80–89.
- Cosentino F, Francia P, Camici GG, Pelicci PG, Luscher TF, Volpe M: Final common molecular pathways of aging and cardiovascular disease: role of the p66Shc protein. Arterioscler Thromb Vasc Biol, 2008, 28, 622–628.
- 23. Costantini P, Chernyak BV, Petronilli V, Bernardi P: Modulation of the mitochondrial permeability transition pore by pyridine nucleotides and dithiol oxidation at two separate sites. J Biol Chem, 1996, 271, 6746–6751.
- Costantini P, Chernyak BV, Petronilli V, Bernardi P: Selective inhibition of the mitochondrial permeability transition pore at the oxidation-reduction sensitive dithiol by monobromobimane. FEBS Lett, 1995, 362, 239–242.

- Daiber A, Wenzel P, Oelze M, Schuhmacher S, Jansen T, Munzel T: Mitochondrial aldehyde dehydrogenase (ALDH-2)-Maker of and marker for nitrate tolerance in response to nitroglycerin treatment. Chem Biol Interact, 2009, 178, 40–47.
- Davidson SM, Duchen MR: Endothelial mitochondria: contributing to vascular function and disease. Circ Res, 2007, 100, 1128–1141.
- 27. De Colibus L, Li M, Binda C, Lustig A, Edmondson DE, Mattevi A: Three-dimensional structure of human monoamine oxidase A (MAO A): relation to the structures of rat MAO A and human MAO B. Proc Natl Acad Sci USA, 2005, 102, 12684–12689.
- Detaille D, Guigas B, Chauvin C, Batandier C, Fontaine E, Wiernsperger N, Leverve X: Metformin prevents high-glucose-induced endothelial cell death through a mitochondrial permeability transition-dependent process. Diabetes, 2005, 54, 2179–2187.
- 29. Di Lisa F, Bernardi P: Mitochondria and ischemia-reperfusion injury of the heart: Fixing a hole. Cardiovasc Res, 2006, 70, 191–199.
- 30. Di Lisa F, Bernardi P: Mitochondrial function and myocardial aging. A critical analysis of the role of permeability transition. Cardiovasc Res, 2005, 66, 222–232.
- Di Lisa F, Blank PS, Colonna R, Gambassi G, Silverman HS, Stern MD, Hansford RG: Mitochondrial membrane potential in single living adult rat cardiac myocytes exposed to anoxia or metabolic inhibition. J Physiol, 1995, 486, 1–13.
- 32. Di Lisa F, Menabò R, Canton M, Petronilli V: The role of mitochondria in the salvage and the injury of the ischemic myocardium. Biochim Biophys Acta, 1998, 1366, 69–78.
- 33. Droge W: Free radicals in the physiological control of cell function. Physiol Rev, 2002, 82, 47–95.
- 34. Duchen MR: Mitochondria and calcium: from cell signalling to cell death. J Physiol, 2000, 529, 57–68.
- Dworakowski R, Alom-Ruiz SP, Shah AM: NADPH oxidase-derived reactive oxygen species in the regulation of endothelial phenotype. Pharmacol Rep, 2008, 60, 21–28.
- 36. Edmondson DE, Binda C, Mattevi A: The FAD binding sites of human monoamine oxidases A and B. Neurotoxicology, 2004, 25, 63–72.
- 37. Edmondson DE, Mattevi A, Binda C, Li M, Hubalek F: Structure and mechanism of monoamine oxidase. Curr Med Chem, 2004, 11, 1983–1994.
- Fridovich I: Superoxide radical and superoxide dismutases. Annu Rev Biochem, 1995, 64, 97–112.
- Giorgio M, Migliaccio E, Orsini F, Paolucci D, Moroni M, Contursi C, Pelliccia G et al.: Electron transfer between cytochrome c and p66Shc generates reactive oxygen species that trigger mitochondrial apoptosis. Cell, 2005, 122, 221–233.
- 40. Głąb M, Łojek A, Wrzosek A, Dołowy K, Szewczyk A: Endothelial mitochondria as a possible target for potassium channel modulators. Pharmacol Rep, 2006, 58, Suppl, 89–95.
- 41. Gordon LI, Burke MA, Singh AT, Prachand S, Lieberman ED, Sun L, Naik TJ et al.: Blockade of the erbB2 receptor induces cardiomyocyte death through mitochon-

- drial and reactive oxygen species-dependent pathways. J Biol Chem, 2009, 284, 2080-2087.
- 42. Gutierrez J, Ballinger SW, Darley-Usmar VM, Landar A: Free radicals, mitochondria, and oxidized lipids: the emerging role in signal transduction in vascular cells. Circ Res. 2006, 99, 924–932.
- 43. Hajnóczky G, Pacher P, Lin X: Spatio-temporal organization of the mitochondrial phase of apoptosis. IUBMB Life, 2001, 52, 237–245.
- 44. Haynes V, Elfering SL, Squires RJ, Traaseth N, Solien J, Ettl A, Giulivi C: Mitochondrial nitric-oxide synthase: role in pathophysiology. IUBMB Life, 2003, 55, 599–603.
- 45. Hengartner MO: The biochemistry of apoptosis. Nature, 2000, 407, 770–776.
- 46. Kaludercic N, Feng N, Nagayama T, Bedja D, Carpi A, Vecoli C, Cormaci G et al.: Monoamine oxidase A is upregulated in cardiac hypertrophy and is a major determinant of the transition from compensation to failure. Circ Res, 2007, 101(11), 7 (abstract).
- 47. Lemasters JJ, Qian T, Bradham CA, Brenner DA, Cascio WE, Trost LC, Nishimura Y et al.: Mitochondrial dysfunction in the pathogenesis of necrotic and apoptotic cell death. J Bioenerg Biomembr, 1999, 31, 305–319.
- 48. Lenders JW, Eisenhofer G, Abeling NG, Berger W, Murphy DL, Konings CH, Wagemakers LM et al.: Specific genetic deficiencies of the A and B isoenzymes of monoamine oxidase are characterized by distinct neurochemical and clinical phenotypes. J Clin Invest, 1996, 97, 1010–1019.
- Levitt P, Pintar JE, Breakefield XO: Immunocytochemical demonstration of monoamine oxidase B in brain astrocytes and serotonergic neurons. Proc Natl Acad Sci USA, 1982, 79, 6385–6389.
- 50. Madamanchi NR, Runge MS: Mitochondrial dysfunction in atherosclerosis. Circ Res, 2007, 100, 460–473.
- Maurel A, Hernandez C, Kunduzova O, Bompart G, Cambon C, Parini A, Frances B: Age-dependent increase in hydrogen peroxide production by cardiac monoamine oxidase A in rats. Am J Physiol Heart Circ Physiol, 2003, 284, H1460–H1467.
- 52. Melov S, Coskun PE, Wallace DC: Mouse models of mitochondrial disease, oxidative stress, and senescence. Mutat Res, 1999, 434, 233–242.
- 53. Menini S, Amadio L, Oddi G, Ricci C, Pesce C, Pugliese F, Giorgio M et al.: Deletion of p66Shc longevity gene protects against experimental diabetic glomerulopathy by preventing diabetes-induced oxidative stress. Diabetes, 2006, 55, 1642–1650.
- 54. Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, Lanfrancone L, Pelicci PG: The p66shc adaptor protein controls oxidative stress response and life span in mammals. Nature, 1999, 402, 309–313.
- Migliaccio E, Giorgio M, Pelicci PG: Apoptosis and aging: role of p66Shc redox protein. Antioxid Redox Signal, 2006, 8, 600–608.
- 56. Murphy MP: How mitochondria produce reactive oxygen species. Biochem J, 2009, 417, 105–111.
- 57. Napoli C, Martin-Padura I, de Nigris F, Giorgio M, Mansueto G, Somma P, Condorelli M et al.: Deletion of the p66Shc longevity gene reduces systemic and tissue oxidative stress, vascular cell apoptosis, and early

- atherogenesis in mice fed a high-fat diet. Proc Natl Acad Sci USA, 2003, 100, 2112–2116.
- 58. Newmeyer DD, Ferguson-Miller S: Mitochondria: releasing power for life and unleashing the machineries of death. Cell, 2003, 112, 481–490.
- Ohashi M, Runge MS, Faraci FM, Heistad DD: MnSOD deficiency increases endothelial dysfunction in ApoEdeficient mice. Arterioscler Thromb Vasc Biol, 2006, 26, 2331–2336.
- Okado-Matsumoto A, Fridovich I: Subcellular distribution of superoxide dismutases (SOD) in rat liver: Cu,Zn-SOD in mitochondria. J Biol Chem, 2001, 276, 38388–38393.
- 61. Orrenius S, Zhivotovsky B, Nicotera P: Regulation of cell death: the calcium-apoptosis link. Nat Rev Mol Cell Biol, 2003, 4, 552–565.
- 62. Orsini F, Migliaccio E, Moroni M, Contursi C, Raker VA, Piccini D, Martin-Padura I et al.: The life span determinant p66Shc localizes to mitochondria where it associates with mitochondrial heat shock protein 70 and regulates trans-membrane potential. J Biol Chem, 2004, 279, 25689–25695.
- Papa S, Skulachev VP: Reactive oxygen species, mitochondria, apoptosis and aging. Mol Cell Biochem, 1997, 174, 305–319.
- 64. Pchejetski D, Kunduzova O, Dayon A, Calise D, Seguelas MH, Leducq N, Seif I et al.: Oxidative stress-dependent sphingosine kinase-1 inhibition mediates monoamine oxidase A-associated cardiac cell apoptosis. Circ Res, 2007, 100, 41–49.
- 65. Pinton P, Rimessi A, Marchi S, Orsini F, Migliaccio E, Giorgio M, Contursi C et al.: Protein kinase C beta and prolyl isomerase 1 regulate mitochondrial effects of the life-span determinant p66Shc. Science, 2007, 315, 659–663.
- 66. Pletscher A: The discovery of antidepressants: a winding path. Experientia, 1991, 47, 4–8.
- 67. Poderoso JJ, Carreras MC, Lisdero C, Riobo N, Schopfer F, Boveris A: Nitric oxide inhibits electron transfer and increases superoxide radical production in rat heart mitochondria and submitochondrial particles. Arch Biochem Biophys, 1996, 328, 85–92.
- Riederer P, Lachenmayer L, Laux G: Clinical applications of MAO-inhibitors. Curr Med Chem, 2004, 11, 2033–2043.
- 69. Rizzuto R, Pinton P, Ferrari D, Chami M, Szabadkai G, Magalhaes PJ, Di Virgilio F, Pozzan T: Calcium and apoptosis: facts and hypotheses. Oncogene, 2003, 22, 8619–8627.
- 70. Rota M, LeCapitaine N, Hosoda T, Boni A, De Angelis A, Padin-Iruegas ME, Esposito G et al.: Diabetes promotes cardiac stem cell aging and heart failure, which are prevented by deletion of the p66shc gene. Circ Res, 2006, 99, 42–52.
- Rouslin W, Erickson JL, Solaro RJ: Effects of oligomycin and acidosis on rates of ATP depletion in ischemic heart muscle. Am J Physiol Heart Circ Physiol, 1986, 250, H503–H508.
- 72. Sarkela TM, Berthiaume J, Elfering S, Gybina AA, Giulivi C: The modulation of oxygen radical production

- by nitric oxide in mitochondria. J Biol Chem, 2001, 276, 6945–6949.
- Shih JC: Cloning, after cloning, knock-out mice, and physiological functions of MAO A and B. Neurotoxicology, 2004, 25, 21–30.
- Sivasubramaniam SD, Finch CC, Rodriguez MJ, Mahy N, Billett EE: A comparative study of the expression of monoamine oxidase-A and -B mRNA and protein in non-CNS human tissues. Cell Tissue Res, 2003, 313, 291–300.
- 75. Tipton KF, Boyce S, O'Sullivan J, Davey GP, Healy J: Monoamine oxidases: certainties and uncertainties. Curr Med Chem, 2004, 11, 1965–1982.
- 76. Trinei M, Giorgio M, Cicalese A, Barozzi S, Ventura A, Migliaccio E, Milia E et al.: A p53-p66Shc signalling pathway controls intracellular redox status, levels of

- oxidation-damaged DNA and oxidative stress-induced apoptosis. Oncogene, 2002, 21, 3872–3878.
- 77. Turrens JF: Mitochondrial formation of reactive oxygen species. J Physiol, 2003, 552, 335–344.
- Wenzel P, Muller J, Zurmeyer S, Schuhmacher S, Schulz E, Oelze M, Pautz A et al.: ALDH-2 deficiency increases cardiovascular oxidative stress—evidence for indirect antioxidative properties. Biochem Biophys Res Commun, 2008, 367, 137–143.
- 79. Youdim MB, Edmondson D, Tipton KF: The therapeutic potential of monoamine oxidase inhibitors. Nat Rev Neurosci, 2006, 7, 295–309.

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