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Molecular evolution of the prostate cancer susceptibility locus *RNASEL*: Evidence for positive selection

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Abstract

Recent research implicates viral infection as a factor that may contribute to the risk of prostate cancer. Allelic variation at the *RNASEL* locus is associated with the risk of infection by a newly discovered retrovirus called XMRV, and with hereditary risk of prostate cancer. This evidence suggests that the *RNASEL* locus has undergone antagonistic coevolution with the retrovirus over evolutionary time. If this is the case, then both the *RNASEL* locus and the retrovirus should show evidence of positive selection. Here we use molecular-evolutionary methods to investigate the prediction that the *RNASEL* locus will exhibit evidence of positive selection. We find evidence that positive selection has acted on this locus over evolutionary time. We further find, using a Bayesian estimation procedure, that Asp541Glu, which was found to be associated with prostate cancer risk in Caucasians in a recent meta-analysis, shows an elevated probability of positive selection. Previous studies provide evidence for rapid evolution of the infection-mediating *gag* gene in the XMRV retrovirus. Taken together, these results suggest that antagonistic coevolution may have occurred between a specific host locus involved in immune defense (*RNASEL*) and a viral pathogen. In turn, genetic variation associated with this apparent coevolution may influence susceptibility to prostate cancer.

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Keywords: XMRV; Prostate cancer; Virus; RNASEL; Coevolution; Positive selection

1. Introduction

A substantial body of evidence links specific parasitic infections to cancer risk and progression in a variety of tissues (e.g. breast cancer: Pogo and Holland, 1997; colon cancer: Laghi et al., 1999). The mechanisms that cause associations between infection and cancer are still being elucidated. In some cases, the infection and the associated interactions between parasites and the immune system can cause chronic tissue damage and repeated tissue repair that increase the risk of cancer. In other cases, viruses may activate mechanisms that prevent apoptosis, or increase cell replication rates, to enhance their own replication (Barber, 2001). These adaptations on the part of viruses can have the secondary effect of promoting the development and survival of cancer cell lineages, again increasing the risk of cancer (Persing and Prendergast, 1999; Barber, 2001; Ewald, 2000).

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Recently, evidence has been found that implicates viral 44 infection in the risk of prostate cancer (reviewed by 45 Wagenlehner et al., 2007) mediated by RNASEL, the long 46 sought-after Hereditary Prostate Cancer 1 gene (Carpten et al., 47 2002; reviewed by Silverman, 2007). Research involving the 48 hybridization of RNA from prostate tumors of individuals 49 bearing different genotypes at amino acid position 462 of the 50 RNASEL locus to a microarray representing conserved 51 sequences from a wide range of viruses identified a novel 52 gammaretrovirus (XMRV) related to xenotropic murine 53 leukemia viruses (Urisman et al., 2006). There were two 54 alleles involved, arginine (R) and glutamine (Q), with the 55 former being common relative to the latter. A high proportion 56 (40%) of the tumors from individuals homozygous for the 57 susceptible version of the 462 allele (QQ) harbored the genome 58 of XMRV, whereas few of the tumors (2%) from heterozygous 59 or homozygous wildtype (RR) genotypes showed any evidence 60 for the presence of the retrovirus. These findings indicate that 61 the XMRV virus interacts with prostate cells and may increase 62 the risk of cancer in individuals genetically susceptible due to 63 their genotype at the RNASEL locus. 64

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Urisman et al. (2006) sequenced multiple copies of several 65 different genes in the XMRV viral genome (gag, gag-pro-pol, and env) and compared them, using phylogenetic analysis, to other MuLVs. The XMRV sequences formed a strongly supported clade, which was the sister clade to an MuLV clade-containing viral strains NFS-Th-1 and NZB-9-1. Analysis of the gag leader sequence of the gag gene of XMRV showed high levels of sequence variation consistent with the 72 possibility of positive selection (Urisman et al., 2006), and 73 studies of this region in closely related MuLVs implicate it as an 74 important mediator of infectivity and virulence (Corbin et al., 75 1994; Fujisawa et al., 1998). 76

The evidence linking genotypic variation at the RNASEL 77 locus with susceptibility to viral infection suggests that this 78 gene may be subject to antagonistic coevolution with XMRV. 79 such that different genetic strains of the parasite are evolving 80 over time to exploit the host, and host loci are evolving in 81 response. This type of antagonistic coevolution should leave a 82 molecular evolutionary signature in both the host and parasite 83 genomes across evolutionary time, as for diverse other genes 84 involved in host-parasite evolutionary dynamics (Yang and 85 Bielawski, 2000). Specifically, this type of coevolution is 86 expected to leave a signal of positive (diversifying) selection, 87 where new protein variants are selected at such a high rate that 88 the rate of non-synonymous substitution exceeds the rate of 89 synonymous substitution, controlled for opportunity (Hughes, 90 91 1999; Yang and Bielawski, 2000).

In this paper, we use methods from molecular evolutionary 92 genetics designed to test for the signal of diversifying selection 93 across species in a phylogenetic context. We use these methods 94 to search for the signal of diversifying selection in the RNASEL 95 gene, to test the prediction that this gene has been involved in 96 97 antagonistic coevolution with pathogens.

RNASEL codes for the RNase L protein, a constitutively 98 expressed latent endoribonclease that mediates the IFN-99 inducible 2-5A system (Li and Tai, 2006). It functions (in part) 100 as a tumor suppressor, acting to induce apoptosis of cancer cells 101 (Malathi et al., 2004), and is also associated the activity of 102 interferons (Zhou et al., 1997). RNase L appears to implement 103 antiviral effects through several different mechanisms, including 104 cleavage of viral RNA, prevention of viral protein synthesis 105 through destruction of rRNA, induction of apoptosis and 106 107 induction of other antiviral genes (Silverman, 2007).

An association of RNASEL with hereditary prostate cancer 108 has been found in a variety of studies (e.g. Carpten et al., 2002; 109 Rennert et al., 2002), although other studies have found no 110 association (e.g. Downing et al., 2003). A recent meta-analysis 111 supported a role for polymorphism in the RNASEL gene in the 112 113 development of prostate cancer (particularly the Asp541Glu polymorphism), but there was substantially heterogeneity 114 among studies (Li and Tai, 2006). In this paper, we analyze 115 sequence variation in the DNA sequences of this gene across 116 multiple species, to determine if RNASEL shows signs of 117 positive selection over evolutionary time. This analysis 118 provides a preliminary test of the hypothesis that the molecular 119 evolution of the RNASEL gene has been driven in part by host-120 viral antagonistic coevolution. 121

2. Methods

The program CODEML in the PAML computer program package (Yang, 1997) implements maximum-likelihood-based methods that test for a statistical excess of non-synonymous nucleotide changes (leading to amino acid substitutions), over synonymous changes, in sequence data of protein coding genes.

DNA sequences for the RNASEL genes from different mammals were obtained from GenBank. First, the mouse or human sequence was obtained by searching the peptide database. This protein was then used for a translated BLAST search (tblastn). Genes with the same name with an e score less than $1 \times e^{-20}$ were assumed to be orthologous. We identified eight sequences appropriate for analysis: mouse (NM011882), rat (NM182673), dog (XM547430), cow (XR027398), macaque (NM001042433), human (NM021133), orangutan (CR858878) and chimpanzee (XM524990). DNA sequences from GenBank were saved in Fasta format and analyzed with the programs Clustal X (Thompson et al., 1997) and RevTrans (Wernersson and Pedersen, 2003). Sequences were imported into RevTrans in Fasta format and translated. The amino acid sequences were aligned in Clustal X using the multiple alignment parameters 'Gap opening penalty = 15.00, Gap extension penalty = 35.00'. The Gonnet matrix was used to characterize amino acid similarities. Several different gap opening and extension penalties were tried to optimize the alignment. The alignment was also inspected by eye and further adjusted. The final peptide alignment was then used in RevTrans as a "guide alignment". The DNA sequences were translated, aligned to match the guide alignment, and reverse translated to give the final DNA sequence alignment.

The aligned sequences were imported into the phylogenetic analysis program PAUP 4.0b10 (Swofford, 2003). An unweighted heuristic search with TBR and 10 random addition-order replicates was used to identify the most parsimonious tree, which was used as a base topology to calculate the following parameters via maximum likelihood: base frequencies, transition/transversion ratio, gamma parameter (alpha), and proportion of invariant sites. These estimates were then used to parameterize a maximum likelihood analysis of the phylogenetic relationships of the sequences involved. A heuristic search was used, with TBR and 10 random additionorder replicates. The tree obtained agreed with previous estimates of the relationships among the taxa involved based on multiple datasets (Springer et al., 2004). The tree from PAUP was imported into TreeView (Page, 2000) and edited to insure q1 that it was an unrooted tree appropriate for analysis with PAML (Yang, 1997).

Analyses of dN/dS ratios (ω) were carried out with the CODEML program in PAML 3.15 (Yang, 1997), using the sequence alignments from Clustal X and RevTrans, and the tree from PAUP. We used the N-sites model, which uses codons as sites and allows ω to vary among sites (Nielsen and Yang, 1998). We also used a lineage-specific model to investigate selection on the human lineage specifically (Yang, 1998). The phylogenetic trees obtained from the PAUP analyses were used to produce a treefile for preliminary analyses in PAML using 121

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the one-ratio model (M0). The estimates of branch lengths produced by this model were then incorporated into the treefiles, and the tree plus branch lengths files were used as the main treefiles for all subsequent analyses in PAML (Yang, 1997).

Simulation studies indicate that the power and accuracy of the log-likelihood ratio tests used to investigate the significance of evidence for positive selection increases with the number and diversity of sequences used (Anisomova et al., 2001, 2002). These simulation studies also indicate that the tests are robust if the treelength in the analyses is greater than one substitution per codon. This criterion was met for our dataset.

The following models were used to analyze the dataset for 190 each gene region: M0 (single rate model), M1 (neutral model), 191 M2 (basic selection model). M7 (continuous distribution 192 model), and M8 (continuous distribution plus selection model). 193 Codon frequencies were estimated from the average nucleotide 194 frequencies at the three-codon positions for all runs. Log-195 likelihood ratio tests were used to test for significant differences 196 in the fit of the models incorporating selection relative to their 197 (nested) counterparts that did not allow positive selection 198 (Nielsen and Yang, 1998). We focused on comparing model 1 199 (neutral model) results to model 2 (selection) results, and model 200 7 (continuous distribution) to model 8 (continuous plus 201 selection), as recommended by Yang et al. (2000). We also 202 compared model 8 to model 8 with the value of omega fixed at 203 204 one, which has been used as a stringent test in some recent analyses (e.g. Clark and Swanson, 2005). These tests provide a 205 useful series of metrics for interpreting the significance of the 206 results. The order of stringency of the LRTs is as follows: M7 207 versus M8 < M1 versus M2 < M8 versus M8 fixed omega. We 208 used the Bayes empirical Bayes method implemented in the 209 most recent version of PAML to estimate posterior probabilities 210 of selection on each codon. This method allows assessment of 211 the probabilities that specific codon sites in the gene in question 212 are under positive selection. 213

To check that the method had not converged on a local 214 maximum (leaving a global maximum undetected) we carried 215 out several runs for each set of models, using three initial values 216 (0.5, 2 and 10) for kappa (transition/transversion ratio) and 217 omega (0.1, 1 and 10). The final likelihoods were compared and 218 the highest likelihood taken as the best estimate. We did not 219 detect any case where the initial analysis was trapped on a local 220 maximum. 221

3. Results

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Our analysis showed strong evidence for positive selection 223 on the RNASEL gene, with an omega of 2.92 at 7% of the codon 224 sites for the discrete approximation model (M2), and 2.32 (14%) 225 of sites) for the continuous approximation plus selection model 226 (M8). The log-likelihood ratio tests (LRT's) were highly 227 significant (P < 0.01) in every comparison (M2 vs. M1, M8 vs. 228 M7, and M8 vs. M8 with omega fixed at one). Using the Bayes 229 empirical Bayes method (Yang et al., 2005), we estimated 230 significant posterior probabilities of positive selection at 10 231 codon sites under the continuous approximation plus selection 232

(M8) model (the first number lists the codon, the letter denotes 233 the common amino acid at that site, and the fraction in 234 parentheses is the estimate of the probability that the site is 235 under positive selection): 40R (0.96), 47K (0.95), 102Q (0.96), 236 178K (0.96), 302E (0.96), 322Y (0.99), 323H (0.98), 618C 237 (1.0), 647N (0.98), 654H (0.99). A number of other sites also 238 showed high posterior probabilities of being under positive 239 selection under both models: posterior probabilities of positive 240 selection higher than 75% were estimated at 35 other sites. 241 Thus, these sites showed an elevated posterior probability of 242 being under positive selection, relative to the random 243 expectation (Yang et al., 2005). 244

One site showing polymorphism associated with prostate 245 cancer risk (Li and Tai, 2006), Asp541Glu, also showed an 246 elevated probability of positive selection in our analysis 247 $(\omega = 2.19, \text{ posterior probability} = 82\%)$. This polymorphism 248 does not reduce the RNase L activity (Xiang et al., 2003), but 249 has been associated with risk of prostate cancer in some studies. 250 In a study of a Japanese population sample, homozygotes for 251 Asp541 showed increased risk for prostate cancer (Nakazato 252 et al., 2003). In contrast, homozygotes for Glu541 were found 253 to be at higher risk in a European-American Caucasian sample Q2 254 by Noonan-Wheeler et al. (2006). Shook et al. (2007) found an 255 elevated risk for Glu541 homozygotes in Hispanic Caucasians, 256 but no association was found in non-Hispanic Caucasians or 257 African Americans. Other studies have also found no 258 association between Asp541Glu and the risk of prostate cancer 259 (e.g. Maier et al., 2005). Hence there is variation across studies, 260 populations and ethnic groups in the relationship between 261 Asp541Glu and prostate cancer risk, and in the specific 262 genotypes that confer susceptibility. A recent meta-analysis 263 comparing multiple studies concluded that there is significant 264 evidence across studies for an effect of polymorphism at this 265 locus on prostate cancer (Li and Tai, 2006). However, the study 266 also demonstrated significant heterogeneity among studies of 267 different populations for the association of this polymorphism 268 with both sporadic and hereditary prostate cancer risk. We note 269 here that the associations found could be due to spurious results, 270 such as Type I errors. 271

We also carried out a lineage specific analysis of positive 272 selection to determine if there was any evidence of positive 273 selection on the human lineage in particular. Lineage-specific 274 analyses have reduced power relative to codon-specific 275 analyses because they average estimation of selection across 276 the entire gene. The ML model indicated a relatively high ratio 277 of non-synonymous to synonymous substitutions on the 278 RNASEL locus along the human lineage, with an average ω 279 of 0.92. Although the ratio does not exceed one (and hence does 280 not exceed the threshold for positive selection), it appears 281 elevated relative to the average ratio of 0.51 across the rest of 282 the tree. 283

4. Discussion

The molecular evolutionary analyses reported here provide285evidence that positive selection has acted on the *RNASEL* locus,286a gene that shows variation associated with susceptibility to287

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infection by XMRV and with predisposition to prostate cancer. 288 This inference is also supported by the results of Bustamante 289 et al. (2005), who found highly significant evidence of positive 290 selection (p = 0.004) on *RNASEL* using comparisons of human 291 intraspecific polymorphism with human-chimp divergence in a 292 survey of over 11,000 genes. Such positive selection is found 293 most commonly in the context of parasite-host antagonistic 294 coevolution, and such interactions provide some of the best 295 evidence for positive selection via antagonistic interaction in 296 297 many taxa (Hughes, 1999). For RNASEL, the available evidence suggests that its connection with prostate cancer may have 298 evolved, at least in part, in the context of resistance to infection 299 by the XMRV virus. Previous research suggesting the action of 300 positive selection on XMRV (Urisman et al., 2006) is also 301 consistent with this interpretation. 302

Immune system surveillance of incipient tumors (Dunn 303 et al., 2004), tradeoffs between immune function and 304 testosterone production (e.g. Verhulst et al., 1999; Muehlenbein 305 and Bribiescas, 2005), or microbe-induced inflammatory 306 reactions favoring the development of cancer (Clevers, 2004) 307 all provide potential explanations for a connection between loci 308 associated with both immunity to specific pathogens and risk of 309 cancer. In the case of *RNASEL*, the ability of the gene to induce 310 apoptosis, and the ability of the virus to inhibit apoptosis, may 311 be an important locus of interaction. 312

Conflicting results have been found concerning the roles of 313 314 specific variants in RNASEL in both hereditary and sporadic prostate cancer (Li and Tai, 2006). For example, the 315 homozygous Gln462 genotype in the Arg462Gln polymorph-316 ism is associated with increased risk for sporadic (non-317 hereditary) cases of prostate cancer in Caucasians (Casey et al., 318 2002), whereas the reverse association has been found for 319 hereditary prostate cancer in Asian populations (e.g. Nakazato 320 et al., 2003). Li and Tai (2006) also found significant 321 322 heterogeneity across all studies for the effect of Arg462Gln on hereditary prostate cancer in their meta-analysis. The same 323 situation occurs for polymorphism associated with prostate 324 cancer at Asp541Glu, as described in Section 3 (see above). Li 325 and Tai (2006) argued that the effect of particular mutations 326 might be specific to particular ethnic groups in some cases. 327 Antagonistic coevolution can generate polymorphism in the 328 virus for exploitation of specific host genotypes and 329 polymorphism in the host for resistance to specific viral 330 genotypes (Frank 2002). Hence, variation in which allele is 331 associated with risk of prostate cancer (in association with viral 332 infection) in different populations or ethnic groups is not 333 unexpected in this context. Indeed, variation in patterns of 334 335 positive selection among ethnic groups is common in analyses of the human HapMap data (Voight et al., 2006), and genes 336 associated with immune function are particularly likely to show 337 this kind of variation. 338

The discovery that Arg541Glu shows a high posterior probability of positive selection, and also shows an association with prostate cancer (Li and Tai, 2006), indicates that this site may have been an important locus of viral-host coevolution, and consequent risk of prostate cancer. That is, variants at this position may show variation in susceptibility to the virus, and consequent variation in the risk of prostate cancer associated with prolonged viral infection. The specific variants that confer susceptibility apparently differ among populations and ethnic groups, consistent with independent coevolutionary trajectories in different populations.

Our study also revealed a number of other sites that show high posterior probabilities of positive selection. These findings suggest that variation at these sites has been associated with antagonistic coevolution with parasites over the course of mammalian evolution. Indeed, evidence from the evolutionary theory of host–pathogen interactions indicates that natural selection will continually favor new variants that enhance parasite virulence and host resistance, and this can lead to the evolution of multiple variants at multiple sites over evolutionary time (Frank, 2002). Investigation of linkages between these selected sites, and prostate cancer risk or progression, may yield novel insights into the evolutionary-genetic basis of this disease.

Recent research suggests that other loci may also mediate susceptibility to cancer in the context of interactions between the immune system and cancer. For example, positive selection has been documented for the antimicrobial beta-defensin genes of some mammals (Maxwell et al., 2003; Luenser and Ludwig, 2005), some of which exhibit aberrant expression in prostate cancer (Donald et al., 2003), and for the alpha-defensins (Lynn et al., 2004; Patil et al., 2004), which are highly overexpressed in colon cancer (Nam et al., 2005).

Uncited references

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Crespi and Summers (2006), Hassel et al. (1993), Page (1996), Rokman et al. (2002), Simard et al. (2003), Walsh (2003) and Wiklund et al. (2004).

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