Effects of Sesame Oil Against After the Onset of Acetaminophen-Induced Acute Hepatic Injury in Rats

Victor Raj Mohan Chandrasekaran, PhD¹; Se-Ping Chien, MS²; Dur-Zong Hsu, DVM, PhD¹; Yu-Chung Chang, MD, PhD³; and Ming-Yie Liu, PhD^{1,4}

Financial disclosure: This study was supported by grants from the National Science Council, Taiwan.

Background: Acetaminophen (APAP) is a safe and effective analgesic and antipyretic when used at therapeutic levels. However, an acute or cumulative overdose can cause severe liver injury with the potential to progress to liver failure in humans and experimental animals. Much attention has been paid to the development of an antioxidant that protects against APAP-induced acute hepatic injury. Hence, we aimed to investigate the effect of sesame oil against after the onset of acute hepatic injury in APAPoverdosed rats. *Methods:* Male Wistar rats were first given 2 oral doses (1,000 mg/kg each) of APAP (at 0 and 24 hours) and then 1 oral dose of sesame oil (8 mL/kg at 24 hours). *Results:* After 48 hours, APAP increased aspartate and alanine aminotransferase levels in the rats' serum and centrilobular necrosis in liver tissue.

cetaminophen (APAP, N-[4-hydroxyphenyl]acetamide) is widely used as an analgesic and antipyretic.¹ APAP has a good safety profile,^{2,3} but an APAP overdose may lead to severe hepatic necrosis and fatal hepatic failure.⁴ APAP is also the most common substance in self-poisoning in the United Kingdom and United States.^{5,6} At therapeutic doses, it is conjugated with either a sulfate or a glucuronide and is safely excreted.⁷ However, when taken in toxic doses, APAP is metabolized by the cytochrome P450 system to form highly electrophilic N-acetylp-benzoquinone imine (NAPQI).⁸⁻¹⁰ Further, glutathione (GSH), the main component of the endogenous sulfhydryl Journal of Parenteral and Enteral Nutrition Volume 34 Number 5 September 2010 567-573 © 2010 American Society for Parenteral and Enteral Nutrition 10.1177/0148607110362584 http://jpen.sagepub.com hosted at http://online.sagepub.com

In addition, APAP significantly decreased the rats' glutathione levels and mitochondrial aconitase activity, but increased superoxide anion, hydroxyl radical, and lipid peroxidation levels. Oral sesame oil (8 mL/kg, given at 24 hours) reversed all APAP-altered parameters and protected the rats against APAP-induced acute liver injury. *Conclusion:* We hypothesize that sesame oil acts as a useful agent that maintains intracellular glutathione levels and inhibits reactive oxygen species, thereby protecting rats against after the onset of APAP-induced acute oxidative liver injury. (JPEN J Parenter Enteral Nutr. 2010;34:567-573)

Keywords: acetaminophen; glutathione; aconitase; reactive oxygen species; sesame oil

pool, is a reducing agent and antioxidant important for scavenging free radicals and reducing oxidative stress.¹¹ When excessive amounts of APAP are ingested, there is more NAPQI in the liver, which depletes GSH and causes hepatic damage.¹²

Sesame oil (SO), derived from the plant species *Sesamum indicum* L., is an herbaceous annual in the family *Pedaliaceae*.¹³ SO increases the hepatic detoxification of chemicals and reduces the incidence of chemically induced oxidative stress after the onset of endotoxication.¹⁴ Prophylactic use of SO attenuates APAP-induced reactive oxygen species and lipid peroxidation, thereby preventing hepatic injury.¹⁵ However, the effect of SO against hepatic injury after the onset of APAP poisoning is unclear. Therefore, we investigated the effect of SO against after the onset of acute hepatic injury in APAP-overdosed rats.

Materials and Methods

Chemicals

APAP, SO, and polyethylene glycol (PEG) were obtained from Sigma-Aldrich Co. (St Louis, MO). APAP was prepared in a suspension using 40% PEG.¹⁵

From ¹Department of Environmental and Occupational Health, National Cheng Kung University Medical College, Tainan; ²Department of Living Science, Tainan University of Technology, Tainan; ³Department of Surgery, National Cheng Kung University Medical College, Tainan; ⁴Sustainable Environment Research Centre, National Cheng Kung University, Tainan, Taiwan.

Received for publication March 19, 2009; accepted for publication October 21, 2009.

Address for correspondence to: Ming-Yie Liu, PhD, Department of Environmental and Occupational Health, National Cheng Kung University Medical College, 138 Sheng-Li Road, Tainan 70428, Taiwan; e-mail: myliu@mail.ncku.edu.tw.

Animals

Ten-week-old male albino Wistar rats (n = 30; 250–300 g), purchased from our Institutional Laboratory Animal Center were used in this study. They were given pellet feed (Richmond Standard; PMI Feeds, Inc, St Louis, MO) and water ad libitum. The animal facility had a 12-hour light/ dark cycle and central air-conditioning (25°C, 70% humidity) throughout the experimental period. The animal care and experimental protocols were in accordance with nationally approved guidelines.

Experimental Design

Food was withheld from the rats for at least 12 hours before each experiment, and the rats were pair-fed after 3 hours of APAP and SO treatments. All experimental treatments were given orally. The doses of APAP and SO were selected based on our previous study.¹⁵ The rats were divided into 5 groups (n = 6 per group): Group I, healthy, untreated controls (HC group); Group II (SO group) received SO (8 mL/kg) at 24 hours; Group III (PEG group) received PEG (3 mL/kg) at 0 hours and 24 hours; Group IV (APAP group), positive controls, received APAP (1,000 mg/kg) at 0 hours and 24 hours; and Group V (ASO group) first received APAP (1,000 mg/kg) at 0 hours and 24 hours and then SO (8 mL/kg) at 24 hours. After 48 hours, we collected rat serum to assess the levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT), and we harvested a small piece of liver tissue from each rat for histopathologic examination. We also assessed the levels of GSH, superoxide anion, hydroxyl radical, and lipid peroxidation in the liver tissue. In addition, we also measured aconitase activity in isolated mitochondria.

Blood Collection

Blood was collected in serum separation tubes from a femoral vein via venipuncture while the rats were under mild ether anesthesia. The tubes were left to clot at room temperature for 30 minutes and then centrifuged at 1,000 g at 4° C for 10 minutes.

Assessment of Hepatic Injury

We assessed hepatic injury by measuring the levels of AST and ALT in serum using a biochemistry analyzer (Fujifilm Dri-Chem 3500s; Fujifilm, Kanagawa, Japan). Hepatic injury was further confirmed using histologic studies. A small piece of liver tissue from each rat was cut and placed in 4% phosphate-buffered formalin. The tissue pieces were dehydrated using a graded percentage of alcohol and then fixed in paraffin wax for 1 hour to form blocks. The blocks were trimmed and cut into 4-µm-thick sections, stained with hematoxylin and eosin, and then mounted using Depex Polystyrene dissolved in xylene mountant. The permanently mounted sections of liver tissue were examined under a microscope (Eclipse E 600; Nikon Instech Co Ltd, Kawasaki, Japan; 100× magnification) to assess hepatic injury.¹⁶ Histopathologic score categories indicated the type of injury: necrosis, sinusoidal dilatation, and lymphocytic infiltration. The scoring system was scaled from 1 to 4 (1 = no abnormalities, 2 =mildly abnormal, 3 =moderately abnormal, and 4 =markedly abnormal).¹⁷

Determination of GSH Levels

A 10% liver-tissue homogenate (1 g in 10 mL of ice-cold 10% trichloroacetic acid) was used to determine GSH levels. In brief, tissue samples were homogenized and centrifuged (3,000 rpm for 10 minutes), and then 500 μ L supernatant was added to 2 mL 0.3 M Na₂HPO₄ 2H₂O solution. Next, 200 μ L dithiobis(2-nitrobenzoic acid) (1% sodium citrate, 0.4 mg/mL) was added, and the absorbance was immediately measured at 412 nm.¹⁸

Determination of Superoxide Anion and Hydroxyl Radical Levels in Liver Tissue

Liver tissue was homogenized (1:10; w/v) in Tris-sucrose buffer (0.24 M sucrose in 20 mM Tris-HCl buffer containing 1 mM ethylene-diaminetetraacetic acid [EDTA; pH 7.4]). The homogenates were centrifuged at 400 g at 4°C for 30 minutes. Superoxide anion and hydroxyl radical levels were measured using a high-performance chemiluminescence (CL) analyzer (CLA-2100; Tohoku Electronic Industrial Co Ltd, Rifu, Japan). In brief, 400 µL supernatant was mixed with 200 µL phosphatebuffered saline in a stainless dish, and then the background CL count was read for 60 seconds. After adjusting the baseline count, 100 μ L lucigenin and indoxyl β -D-glucuronide (17 mM dissolved in phosphate-buffered saline, to determine superoxide anion and hydroxyl radical, respectively) were injected into the machine, and CL was counted for another 15 minutes at 10-second intervals. The data were analyzed using chemiluminescence analyzer data acquisition software CLA-DAS (Tohoku Electronic Industrial Co.).¹⁴

Isolation of Mitochondria From Liver Tissue

Liver tissue was washed once with ice-cold isolation buffer (10 mL Tris-MOPS [0.1 M; pH 7.4], 20 mL sucrose [1 M], and 1 mL Ethylene glycol-bis(2-aminoethylether)-N,N,N,N-tetraacetic acid-Tris buffer [0.1 M; pH 7.4]) and cut into small pieces; the buffer was discarded. Five milliliters of fresh isolation buffer was then added, and the mixture was homogenized. The homogenates were centrifuged at 600 g at 4°C for 10 minutes, and then the supernatants were centrifuged at 7,000 g at 4°C for 10 minutes, after which the supernatants were discarded.



Figure 1. Effects of sesame oil against after the onset of acetaminophen (APAP)-induced hepatic injury. The rats were divided into 5 groups of 6. Group I (HC): healthy controls; Group II (SO): given oral sesame oil (8 mL/kg) at 24 hours; Group III (PEG): given oral polyethylene glycol (3 mL/kg) at 0 and 24 hours; Group IV (APAP): positive controls, given oral APAP (1,000 mg/kg) at 0 and 24 hours; and Group V (ASO): first given oral APAP (1,000 mg/kg) at 0 and 24 hours. Aspartate transaminase (AST) and alanine transaminase (ALT) levels were assessed in rat serum 48 hours later. Data are given as mean \pm standard error. Different letters (a,b, and c) indicate a significant difference between groups. (P < .05; one-way analysis of variance, and then the Tukey multiple-comparison test).

The pellets were washed once with isolation buffer, and the above centrifugation steps were repeated twice. After the 3-step centrifugation, the supernatants were discarded and the pellets were suspended in 1 mL isolation buffer and used for further analysis.¹⁹

Determination of Mitochondrial Aconitase Activity

Mitochondrial aconitase activity was determined using a commercial kit (Bioxytech Aconitase-340; Oxis Research, Foster City, CA) based on the molar extinction coefficient of nicotinamide adenine dinucleotide phosphate (6.220×10^{-3}) and the temperature coefficient (2.4435), and then read on a spectrophotometer (DU 640B; Beckman, Fullerton, CA) at 340 nm.

Measuring Lipid Peroxidation in Liver Tissue

Liver tissue was homogenized in Tris-HCl buffer (20 mM; pH 7.4) and then centrifuged at 2,500 g at 4°C for 10 minutes. Two hundred microliters of supernatant were analyzed for malondialdehyde (MDA) levels using a kit (Bioxytech MDA-586; Oxis Research) and then read on a spectrophotometer (Beckman) at 586 nm.

Statistical Analysis

The data were analyzed using one-way analysis of variance (ANOVA) and the Tukey multiple-comparison test to evaluate the significance between the treatment groups. Results are given as mean \pm standard error. A *P* value < .05 was considered statistically significant.

Results

Effects of Sesame Oil After the Onset of APAPinduced Hepatic Injury

To examine the effects of SO after the onset of APAPinduced liver toxicity, we measured AST and ALT levels in serum. APAP significantly increased AST and ALT levels compared with those in the HC, SO, and PEG groups (all P < .001). SO significantly protected against after the onset of APAP-induced liver toxicity by preventing the rise of AST and ALT levels at 48 hours (Figure 1).

Effects of Sesame Oil on Liver Histopathology After the Onset of APAP-induced Liver Damage

We analyzed the histopathologic features of liver tissue to find out the effects of SO against after the onset of APAPinduced liver toxicity. APAP-treated liver tissue showed centrilobular necrosis, lymphocytic infiltration, and sinusoidal dilatation (Figure 2A). This necrotic effect was minimal in the ASO group compared with that in the APAP group. APAP significantly altered the normal architecture of the liver compared with that in the HC, SO, and PEG groups (P < .001; Figure 2B). Neither SO alone



Figure 2. Effects of sesame oil against after the onset of acetaminophen (APAP)-induced hepatic histopathology. The rats were divided into 5 groups of 6. Group I (HC): healthy controls; Group II (SO): given oral sesame oil (8 mL/kg) at 24 hours; Group III (PEG): given oral polyethylene glycol (3 mL/kg) at 0 and 24 hours; Group IV (APAP): positive controls, given oral APAP (1,000 mg/kg) at 0 and 24 hours; and Group V (ASO): first given oral APAP (1,000 mg/kg) at 0 and 24 hours, and then oral sesame oil (8 mL/kg) at 24 hours. Hepatic histopathologic changes (A) and their respective scores (B) were observed 48 hours after the initial treatment. The arrow in the APAP image indicates the necrotic region (hematoxylin and eosin stain; 100× magnification). Data are given as mean \pm standard error. Different letters (a,b, and c) indicate a significant difference between groups. (*P* < .05; one-way analysis of variance and then the Tukey multiple-comparison test).

nor PEG alone altered the normal architecture of liver tissue.

Effects of Sesame Oil Against After the Onset of APAP-induced GSH Levels, Superoxide Anion Generation, and Mitochondrial Aconitase Activity

APAP significantly (P < .001) decreased GSH (Figure 3A) levels and increased superoxide anion generation (Figure 3B) in liver tissue compared with that in the HC, SO, PEG, and ASO groups (all P < .001). In addition, APAP significantly (P < .001) decreased aconitase activity

(Figure 3C) in rat liver mitochondria compared with that in the HC, SO, PEG, and ASO groups (all P < .001). We found no differences in those levels among the HC, SO, PEG, and ASO groups.

Effects of Sesame Oil Against After the Onset of APAP-induced Hydroxyl Radical and Lipid Peroxidation Levels

To study the effect of SO against after the onset of APAP-induced hydroxyl radical generation and lipid peroxidation, we measured hydroxyl radical and MDA levels in liver tissue. Hydroxyl radical and MDA levels



Figure 3. Effects of sesame oil on the levels of glutathione (GSH), superoxide anion generation in liver tissue, and mitochondrial aconitase activity after the onset of acetaminophen-induced hepatic injury. The rats were divided into 5 groups of 6. Group I (HC): healthy controls; Group II (SO): given oral sesame oil (8 mL/kg) at 24 hours; Group III (PEG): given oral polyethylene glycol (3 mL/kg) at 0 and 24 hours; Group IV (APAP): positive controls, given oral APAP (1,000 mg/kg) at 0 and 24 hours; and Group V (ASO): first given oral APAP (1,000 mg/kg) at 0 and 24 hours, and then oral sesame oil (8 mL/kg) at 24 hours. Forty-eight hours after the initial treatment, we assessed GSH and superoxide anion levels in liver tissue. In addition, aconitase activity was estimated in rat liver mitochondria. Data are given as mean \pm standard error. Different letters (a and b) indicate a significant difference between groups. (*P* < .05; one-way analysis of variance and then the Tukey multiple-comparison test).



Figure 4. Effects of sesame oil on the levels of hydroxyl radical and lipid peroxidation after the onset of acetaminophen-induced hepatic injury. The rats were divided into 5 groups of 6. Group I (HC): healthy controls; Group II (SO): given oral sesame oil (8 mL/kg) at 24 hours; Group III (PEG): given oral polyethylene glycol (3 mL/kg) at 0 and 24 hours; Group IV (APAP): positive controls, given oral APAP (1,000 mg/kg) at 0 and 24 hours and Group V (ASO): first given oral APAP (1,000 mg/kg) at 0 and 24 hours. Forty-eight hours after the initial treatment, we assessed hydroxyl radical and malondialdehyde (MDA) levels in liver tissue. Data are given as mean \pm standard error. Different letters (a and b) indicate a significant difference between groups. (P < .05; one-way analysis of variance and then the Tukey multiple-comparison test).

in liver tissue increased significantly (P < .001) in the APAP group compared with those in the HC, SO, PEG, and ASO groups (all P < .001). The levels of hydroxyl radical and MDA were unaltered in the SO, PEG, and ASO groups (Figure 4).

Discussion

SO not only prevented¹⁵ but also attenuated APAPinduced acute hepatic injury in rats. SO maintained the GSH levels in the liver tissue after the onset of APAP-induced hepatic injury. GSH content has been inversely linked to APAP hepatotoxicity. GSH depletion induced either by chemicals or fasting has been shown to increase APAP toxicity.²⁰ GSH plays an important role in scavenging NAPQI, a toxic metabolite of APAP.²¹ Excessive NAPOI formation starts to deplete GSH levels and initiates hepatic injury.²² Reactive oxygen species formation starts immediately after the depletion of GSH,²³ which leads to cell death. In addition, GSH depletion increases the accumulation of superoxide anion in hepatocytes and triggers mitochondrial oxidative stress.²⁴ The mitochondrial aconitase enzyme acts as a 2-faced protein, enzyme, and iron-regulatory factor,²⁵ which can be affected by superoxide anion²⁶ during APAP-induced hepatic injury.²⁷ When superoxide anion inactivates mitochondrial aconitase activity, it may pose a significant oxidative burden because it provides equimolar amounts of hydrogen peroxide per mole of superoxide anion.²⁸ Inactive iron-sulfur centered aconitase enzyme releases free ferrous ion, which reacts with hydrogen peroxide to form potent hydroxyl radicals.²⁷ Hydroxyl radicals are highly reactive, which causes hepatic lipid peroxidation during APAP overdose.^{15,27} Hence, we hypothesize that the effect of SO is associated with maintaining GSH levels and mitochondrial aconitase activity to inhibit the generation of superoxide, hydroxyl radicals, and lipid peroxidation leading to protection against after the onset of APAPinduced acute hepatic injury.

The value of SO against after the onset of APAPinduced acute hepatic injury might reduce the incidence of liver transplantation. APAP-induced acute hepatic injury or failure requiring liver transplantation is a major concern in developing and developed countries.^{6, 29-31} It is important to provide an alternative to reduce the incidence of liver transplantation after APAP-induced acute hepatic injury or failure. SO, a natural nutrition supplement, includes various antioxidants such as tocopherol, sesamin, sesamolin, and sesaminol.^{13,32-35} SO protects the liver from APAPinduced hepatic damage without affecting the absorption of APAP.¹⁵ Thus, the protection against APAP-induced liver damage exhibited by SO suggests its high therapeutic value. However, further studies are warranted to confirm this hypothesis.

In summary, the effect of SO might be associated with maintaining GSH levels, trapping reactive oxygen species, decreasing lipid peroxidation, and maintaining the architecture of liver tissue after the onset of APAP poisoning in rats.

Acknowledgments

We thank Bill Franke for editorial assistance.

References

- 1. Thomas S. Paracetamol (acetaminophen) poisoning. J Pharmacol Exp Ther. 1993;60:91-120.
- Lesko SM, Mitchell AA. The safety of acetaminophen and ibuprofen among children younger than two years old. *Pediatrics*. 1999;104:e39.
- 3. Cranswick N, Coghlan D. Paracetamol efficacy and safety in children: the first 40 years. *Am J Ther.* 2000;7:135-141.
- Proudfoot AT, Wright N. Acute paracetamol poisoning. Br Med J. 1970;3:557-558.
- Hawton K, Fagg J, Simkin S, Bale E, Bond A. Deliberate self-harm in adolescents in Oxford, 1985-1995. J Adolesc. 2000;23:47-55.
- Litovitz TL, Klein-Schwartz W, White S, et al. 1999 annual report of the American Association of Poison Control Centers Toxic Exposure Surveillance System. Am J Emerg Med. 2000;18: 517-574.
- Farrell G. Paracetamol-induced hepatotoxicity. In: Farrell G, ed. Drug-Induced Liver Diseases. New York, NY: Churchill Livingstone; 1994:205-224.
- Genter M, Liang H, Gu J, et al. Role of CYP2A5 and 2G1 in acetaminophen metabolism and toxicity in the olfactory mucosa of the cypa2(-/-) mouse. *Biochem Pharmacol.* 1998;55:1819-1826.
- Sinclair J, Jeffery E, Wrighton S, Kostrubsky V, Szakacs J, Sinclair P. Alcohol-mediated increase in acetaminophen hepatotoxicity. *Biochem Pharmacol.* 1998;55:1557-1565.
- Zaher H, Buters J, Ward J, et al. Protection against acetaminophen toxicity in CYP1A2 and CYP2E1 double-null mice. *Toxicol Appl Pharmacol.* 1998;152:193-199.
- Shaw S, Herbert V, Colman N, Jayatilleke E. Effect of ethanolgenerated free radicals on gastric intrinsic factor and glutathione. *Alcohol.* 1990;7:153-157.
- Terneus MV, Kiningham KK, Carpenter AB, Sullivan SB, Valentovic MA. Comparison of S-Adenosyl-L-methionine and N-acetylcysteine protective effects on acetaminophen hepatic toxicity. J Pharmacol Exp Ther. 2007;320:99-107.
- Sugano M, Akimoto KA. Multifunctional gift from nature. J Chin Nutr Soc. 1993;18:1-11.
- Hsu DZ, Liu MY. Sesame oil protects against lipopolysaccharidestimulated oxidative stress in rats. *Crit Care Med.* 2004;32: 227-231.
- Chandrasekaran VR, Wan CH, Liu LL, Hsu DZ, Liu MY. Effect of sesame oil against acetaminophen-induced oxidative hepatic damage in rats. *Shock*. 2008;30:217-221.
- 16. Bancroft JD, Cook BC. *Manual of Histological Techniques*. Edinburgh, UK: Churchill Livingstone; 1984.
- Gaskill CL, Miller LM, Mattoon JS, et al. Liver histopathology and liver and serum alanine aminotransferase and alkaline phosphatase activities in epileptic dogs receiving phenobarbital. *Vet Pathol.* 2005;42:147-160.
- Yegen BC, Alican I, Yalcin AS, Oktay S. Calcium channel blockers prevent stress-induced ulcers in rats. *Agents Actions*. 1992;35:130-134.
- Frezza C, Cipolat S, Scorrano L. Organelle isolation: functional mitochondria from mouse liver, muscle and cultured filroblasts. *Nat Protoc*. 2007;2:287-295.
- Bray BJ, Rosengren RJ. Retinol potentiates acetaminopheninduced hepatotoxicity in the mouse: mechanistic studies. *Toxicol Appl Pharmacol.* 2001;173:129-136.
- Hinson JA, Reid AB, McCullough SS, James LP. Acetaminopheninduced hepatotoxicity: role of metabolic activation, reactive oxygen/nitrogen species, and mitochondrial permeability transition. *Drug Metabol Rev.* 2004;36:805-822.
- 22. Masubuchi Y, Suda C, Horie T. Involvement of mitochondrial permeability transition in acetaminophen-induced liver injury in mice. *J Hepatol.* 2005;42:110-116.

- Bajt ML, Knight TR, Lemasters JJ, Jaeschke H. Acetaminopheninduced oxidant stress and cell injury in cultured mouse hepatocytes: Protection by N-acetyl cysteine. *Toxicol Sci.* 2004;80:343-349.
- Jaeschke H, Bajt ML. Intracellular signalling mechanism of acetaminophen-induced liver cell death. *Toxicol Sci.* 2006;89:31-41.
- 25. Beinert H, Kennedy MC. Aconitase, a two-faced protein: enzyme and iron regulatory factor. *FASEB J.* 1993;7:1442-1449.
- Hausladen A, Fridovich I. Superoxide and peroxynitrite inactivate aconitases, but nitric oxide does not. J Biol Chem. 1994;47:29405-29408.
- Chandrasekaran VR, Hsu DZ, Liu MY. The protective effect of sesamol against mitochondrial oxidative stress and hepatic injury in acetaminophen-overdosed rats. *Shock*. 2009;32:89-93.
- Liochev SI, Fridovich I. The relative importance of HO* and ONOO⁻ in mediating the toxicity of O*-. Free Radic Biol Med. 1999;26:777-778.
- Kogan MD, Pappas G, Yu SM, Kotelchuck M. Over-the-counter medication use among US preschool-age children. JAMA. 1994;272:1025-1030.

- James LP, Mayeux PR, Hinson JA. Acetaminophen-induced hepatotoxicity. Drug Metab Dispos. 2003;31:1499-1506.
- Larson AM. Acetaminophen hepatotoxicity. Clin Liver Dis. 2007;11:525-548.
- 32. Simon JE, Chadwick AF, Craker LE. Herbs: An indexed bibliography, 1971-1980. In: Hamden CT, ed. The Scientific Literature on Selected Herbs and Aromatic and Medicinal Plants of the Temperate Zone. Archon Books; 1984:770. Available at: http://www.hort.purdue.edu/newcrop/med-aro/factsheets/SESAME.html. Accessed March 18, 2009.
- Chavali SR, Utsunomiya T, Forse RA. Increased survival after cecal ligation and puncture in mice consuming diets enriched with sesame seed oil. *Crit Care Med.* 2001;29:140-143.
- Fukuda Y. Food chemical studies on the antioxidants in sesame seed. Nippon Shokuhin Kogyo Gakkaishi. 1990;37:484-492.
- Kang MH, Katsuzaki H, Osawa T. Inhibition of 2,2'-azobis(2,4dimethylvaleronitrile)-induced lipid peroxidation by sesaminols. *Lipids*. 1998;33:1031-1036.