

1998 92: 4491-4508

Emerging Applications of Recombinant Human Granulocyte-Macrophage Colony-Stimulating Factor

James O. Armitage

Updated information and services can be found at: http://bloodjournal.hematologylibrary.org/content/92/12/4491.full.html

Articles on similar topics can be found in the following Blood collections Review Articles (434 articles)

Information about reproducing this article in parts or in its entirety may be found online at: http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#repub_requests

Information about ordering reprints may be found online at: http://bloodjournal.hematologylibrary.org/site/misc/rights.xhtml#reprints

Information about subscriptions and ASH membership may be found online at: http://bloodjournal.hematologylibrary.org/site/subscriptions/index.xhtml



Blood (print ISSN 0006-4971, online ISSN 1528-0020), is published weekly by the American Society of Hematology, 2021 L St, NW, Suite 900, Washington DC 20036. Copyright 2011 by The American Society of Hematology; all rights reserved.



VOL 92, NO 12

REVIEW ARTICLE

The Journal of The American Society of Hematology

DECEMBER 15, 1998

Emerging Applications of Recombinant Human Granulocyte-Macrophage Colony-Stimulating Factor

By James O. Armitage

LINICAL INDICATIONS for use of recombinant human Granulocyte-macrophage colony-stimulating factor (rHuGM-CSF) have expanded considerably since the drug first became available in the early 1990s for acceleration of myeloid engraftment in neutropenic patients. Initial clinical trials of rHuGM-CSF were based on prevailing knowledge of the biologic effects of endogenous GM-CSF at the time and therefore concentrated on the drug's myeloproliferative effects in myelosuppressed patients. As additional information accumulated from in vitro research and from results of clinical trials, it became apparent that rHuGM-CSF had diverse biologic effects and played a vital role in various functions of the immune system, including responses to inflammation and infection, as well as in hematopoiesis. Consequently, a variety of potential clinical uses for rHuGM-CSF are under investigation, such as prophylaxis or adjunctive treatment of infection in high-risk settings or immunosuppressed patient populations, use as a vaccine adjuvant, and use as immunotherapy for malignancies.

The molecular sequence of endogenous human GM-CSF was first identified in 1985; within a few years, three different synthetic human GM-CSFs were produced using recombinant DNA technology and bacterial,¹ mammalian,² and yeast expression systems.³ Sargramostim is yeast-derived rHuGM-CSF produced using Saccharomyces cerevisiae; bacterially derived rHuGM-CSF is produced using Escherichia coli and is termed molgramostim; and mammalian-derived rHuGM-CSF is produced using Chinese hamster ovary cells (CHO) and is termed regramostim. These preparations are not identical and are differentiated by their specific amino acid sequences and degree of glycosylation.¹⁻³ Sargramostim has an amino acid sequence identical to that of endogenous human GM-CSF, except that it contains leucine instead of proline at position 23 and may have a different carbohydrate moiety. Sargramostim is glycosylated to a lesser extent than regramostim, and molgramostim is not glycosylated. The degree of glycosylation of rHuGM-CSF may be an important characteristic, because it can affect pharmacokinetics, biologic activity, antigenicity, and toxicity.4-7

This review discusses current knowledge concerning the biologic effects, pharmacokinetics, and emerging clinical uses of rHuGM-CSF, with a focus on the yeast-derived rHuGM-CSF, sargramostim, the only form of synthetic rHuGM-CSF commercially available in the United States. Information on molgramostim, the form of rHuGM-CSF available in Europe, is also form of rHuGM-CSF used, the term "rHuGM-CSF" is used throughout this review to describe the drug when the expression system was not identified or when multiple studies using different forms of rHuGM-CSF reported similar findings.

PHARMACOLOGY OF SARGRAMOSTIM

provided. Because literature reports do not always indicate the

Biologic effects. GM-CSF was first identified based on its ability to stimulate the clonal proliferation of myeloid precursors in vitro.8 Endogenous GM-CSF, a heavily glycosylated polypeptide, was the first human myeloid hematopoietic growth factor to be molecularly cloned, which allowed the expression of large quantities of the protein. More than a decade of in vitro and in vivo research using murine GM-CSF and synthetic rHuGM-CSFs has shown that the name of this CSF is restrictive, because it describes only one aspect of the numerous biologic effects that have now been attributed to GM-CSF. Although GM-CSF plays a vital role in hematopoiesis by inducing the growth of several different cell lineages, it also enhances numerous functional activities of mature effector cells involved in antigen presentation and cell-mediated immunity, including neutrophils, monocytes, macrophages, and dendritic cells.9-20

The biologic effects of GM-CSF are mediated via binding to receptors expressed on the surface of target cells. The GM-CSF receptor is expressed on granulocyte, erythrocyte, megakaryocyte, and macrophage progenitor cells as well as mature neutrophils, monocytes, macrophages, dendritic cells, plasma cells, certain T lymphocytes, vascular endothelial cells, uterine cells, and myeloid leukemia cells.²¹⁻²⁷ Molecular cloning studies have shown that the GM-CSF receptor is composed of two distinct subunits, α and common β (β_c ; Fig 1).²⁸ The α -subunit binds GM-CSF with low affinity. The β_c has no detectable binding affinity for GM-CSF on its own, but forms a heterodimer with the α -subunit that has high affinity for GM-CSF.

From the Department of Internal Medicine, University of Nebraska Medical Center, Omaha, NE.

Submitted May 14, 1998; accepted September 11, 1998.

Address reprint requests to James O. Armitage, MD, University of Nebraska Medical Center, 600 S 42nd St, Omaha, NE 68198-3332.

^{© 1998} by The American Society of Hematology. 0006-4971/98/9212-0049\$3.00/0



Fig 1. Schematic representation of the GM-CSF receptor (GMR), which is composed of two distinct subunits, α and β . Binding of rHuGM-CSF to GMR leads to formation of the signaling complex and activation of a Janus kinase (JAK2). Regulation of gene expression by JAK2 activates transcription proteins STAT1, STAT3, and STAT5.

Whereas the α -subunit is unique to the GM-CSF receptor, β_c is shared with the receptors for interleukin-3 (IL-3) and IL-5.²⁹

The signal transduction pathways that occur after rHuGM-CSF binds to the GM-CSF receptor are under evaluation. There appear to be at least two distinct signaling pathways, each involving a distinct region of β_c .³⁰ The first, which leads to induction of c-*myc* and activation of DNA replication, involves activation of a Janus kinase (JAK2) that is physically associated with β_c .³¹ Regulation of gene expression by JAK2 appears to be mediated by production of a DNA-binding complex containing the signal transducer and activator of transcription (STAT) proteins STAT1, STAT3, and STAT5.^{32,33} The second pathway involves activation of *ras*³⁴ and mitogen-activated protein kinases,³⁵ with consequent induction of *c-fos* and *c-jun*, which are genes involved in regulation of hematopoietic differentiation.³¹

Pharmacokinetics. Information regarding the pharmacokinetics of rHuGM-CSF after intravenous or subcutaneous administration is available from studies in healthy adults,³⁶ adults with malignancy or myelodysplastic syndrome,^{6,37-40} and children with recurrent or refractory solid tumors.^{41,42} Because evidence exists from animal and clinical studies that the degree of glycosylation of synthetic rHuGM-CSFs influences pharmacokinetics of sargramostim and molgramostim are presented separately.

Studies have determined that the pharmacokinetics of sargramostim are similar among healthy individuals and patients.³⁹ The pharmacokinetics of sargramostim are dependent on the route of administration. Table 1 compares pharmacokinetic parameters after intravenous and subcutaneous administration of sargramostim in healthy adult males.³⁶ Peak serum concentrations are higher after intravenous administration; however, bioavailability (as determined by the area under the concentration-versus-time curve) of sargramostim is similar between administration routes. The elimination of sargramostim occurs principally by nonrenal mechanisms.³⁹ Serum concentrations are more prolonged after subcutaneous administration than after intravenous administration.^{36,42} The magnitude of the percentage of increase in absolute neutrophil count with a specific dose of sargramostim is greater after subcutaneous injection than after 2-hour intravenous infusion.³⁹

The pharmacokinetics of molgramostim (0.3 to 30 μ g/kg) also were studied after subcutaneous and intravenous administration.³⁷ Maximum serum concentrations and area under the concentration-versus-time curve increased with dose for both routes of administration, but appeared larger after intravenous administration in comparison to the same dose administered subcutaneously. However, rHuGM-CSF concentrations greater than 1 ng/mL were maintained longer after subcutaneous administration. Immunoreactive molgramostim was detected in the urine of patients, ranging from 0.001% to 0.2% of the injected dose, supporting nonrenal elimation. The half-life after intravenous administration ranged from 0.24 to 1.18 hours; the mean half life was 3.16 hours after subcutaneous administration.

USE IN ENHANCING HEMATOPOIETIC RECOVERY AFTER CANCER CHEMOTHERAPY AND BONE MARROW TRANSPLANTATION (BMT)

rHuGM-CSF is classified as a multilineage CSF because it stimulates the proliferation and differentiation of hematopoietic progenitor cells of neutrophil, eosinophil, and monocyte colonies.⁴³ Parenteral administration of rHuGM-CSF induces a dose-dependent increase in peripheral blood neutrophil

Table 1. Pharmacokinetic Parameters After Intravenous and Subcutaneous Administration of Sargramostim in Healthy Adult Males

Pharmacokinetic Parameter	Sargramostim at 250 µg/m²	
	IV	SC
C _{max} (ng/mL)	5.0-5.4	1.5
AUC (ng/mL · min)	640-677	501-549
Clearance (mL/min/m ²)	420-431	529-549
t _{1/2β} (min)	60	162

Abbreviations: C_{max} , peak plasma concentration; AUC, area under the concentration-versus-time curve; $t_{1/2\beta}$, terminal elimination half-life; IV, intravenous; SC, subcutaneous.



EMERGING APPLICATIONS OF SARGRAMOSTIM

counts.^{19,44} Sargramostim alters the kinetics of myeloid progenitor cells within the bone marrow, causing rapid entry of cells into the cell cycle and decreasing the cell-cycle time by as much as 33%.⁴⁵ The leukocyte response to rHuGM-CSF is reflected in peripheral blood principally as an increase in segmented neutrophils, but also involves an increase in monocytes and eosinophils.^{19,46-48} Leukocyte differentials generally demonstrate a shift to the left; myelocytes, promyelocytes, and myeloblasts may be present. When rHuGM-CSF is discontinued, leukocyte counts gradually decrease to pretreatment levels.^{19,43}

The myeloproliferative effects of rHuGM-CSF are also the result of its interaction with other cytokines. rHuGM-CSF functions in conjunction with erythropoietin and IL-3 to promote the proliferation and differentiation of erythroid and megakaryocytic progenitors, respectively.8,49 The addition of thrombopoietin to early acting cytokines, such as rHuGM-CSF, increases the overall in vitro megakaryocyte expansion compared with thrombopoietin alone and also generates different subpopulations of CD41+ megakaryocyte progenitors, with much less coexpression of CD42b and CD34 and slightly more coexpression of c-kit.50 In addition, the overall number of CD34⁺ cells increases approximately fivefold with the combination of thrombopoietin and early acting cytokines. A trial in sublethally irradiated nonhuman primates showed that coadministration of sargramostim and thrombopoietin augmented megakaryocyte, erythrocyte, and neutrophil recovery compared with either cytokine alone.51

Enhancing neutrophil proliferation is an important aspect of rHuGM-CSF function; however, effects of this multilineage growth factor on other cells of the immune system, including monocytes and macrophages, have been identified. Administration of rHuGM-CSF not only increases the number of circulating monocytes, but also increases the function of monocytes and macrophages, including oxidative metabolism, cytotoxicity, and Fc-dependent phagocytosis.^{19,52,53} rHuGM-CSF enhances dendritic cell maturation, proliferation, and migration.^{20,54,55} In addition, class II major histocompatibility complex (MHC) expression on macrophages and dendritic cells is increased by rHuGM-CSF, enhancing the function of antigen-presenting cells.⁵⁶

Combined, these effects of rHuGM-CSF not only increase hematopoietic cell counts, but also enhance immune function. The ability of rHuGM-CSF to accelerate myeloid recovery and to prevent infection has resulted in multiple approved indications for sargramostim and molgramostim in their respective countries. The drugs are used in patients after autologous BMT (AuBMT), peripheral blood progenitor cell (PBPC) transplantation, induction therapy for acute myelogenous leukemia (AML), engraftment delay or failure after BMT, and chemotherapyinduced neutropenia. These uses are well established and have been recently reviewed.57-61 Research has expanded in some of these settings to investigate new uses of rHuGM-CSF, including use in combination with granulocyte colony-stimulating factor (G-CSF) for PBPC mobilization, to prime leukemic cells before or during chemotherapy for AML, and as an adjunct to increase chemotherapy dose intensity.

PBPC mobilization in combination with G-CSF. There has been increasing interest in combining rHuGM-CSF with other cytokines, especially G-CSF, as a means of improving mobiliza-

tion without having to administer chemotherapy. This is especially true in the allogeneic transplant setting, where a nontoxic mobilization regimen that allows for collection of a sufficient number of cells to promote engraftment in a minimum number of leukaphereses is most critical. Lane et al⁶² evaluated the PBPC mobilization efficacy of G-CSF at 10 µg/kg/d (n = 8), sargramostim at 10 µg/kg/d (n = 5), or sargramostim plus G-CSF each at 5 µg/kg/d (n = 5) in normal donors. The median CD34⁺ cell yield with the combination regimen and with G-CSF was significantly higher than for rHuGM-CSF alone (101 × 10⁶, 119 × 10⁶, and 12.6 × 10⁶, respectively; *P* < .01 for both comparisons).

An analysis of CD34⁺ cell subsets showed some interesting differences between the different mobilization regimens. A higher proportion of cells in the combination regimen were CD34⁺CD38⁻ and CD34⁺CD38⁻ HLA-DR⁺ (Table 2). Pluripotent progenitor cells are characterized as CD34+CD38- and are likely responsible for long-term hematopoietic reconstitution after transplantation. These cells can be further subdivided according to the presence or absence of HLA-DR, with HLA-DR⁺ cells giving rise to lymphoid and myeloid precursors.⁶³ The greater percentage of this subpopulation of cells mobilized by the combination regimen translated into a higher overall number of CD34+CD38-HLA-DR+ cells in leukapheresis products than products from subjects mobilized with either G-CSF or rHuGM-CSF alone $(1.41 \times 10^6, 0.36 \times 10^6, and$ 0.12×10^6 , respectively; P < .05 for all comparisons). Moreover, the plating efficiency of colony-forming unitgranulocyte-macrophage (CFU-GM) and burst-forming uniterythroid (BFU-E) was higher in cells stimulated by rHuGM-CSF than in those stimulated by G-CSF. Whether this would correlate with more rapid engraftment is not known, although the investigators have reported that PBPCs mobilized by the combination regimen successfully engrafted after allogeneic PBPC transplantation.⁶⁴ Ali et al⁶⁵ also compared mobilization of PBPCs in normal donors using rHuGM-CSF at 5 µg/kg/d plus G-CSF at 10 μ g/kg/d (n = 15) versus G-CSF at 10 μ g/kg/d alone (n = 35). They found a statistically insignificant increase in CD34⁺ cells in the leukapheresis products from donors mobilized with the combination of cytokines; however, the number of CD3⁺ cells in the leukapheresis product was significantly lower with the combination regimen than with G-CSF alone, ie, 160 versus 328×10^6 /kg.

Investigations are ongoing to determine optimal doses and sequence of administration of the cytokines in combination.^{66,67} In a follow-up study to that reported by Lane et al,⁶² healthy volunteers received either sargramostim at 10 µg/kg/d for 3 or 4

Table 2. CD34⁺ Cell Subsets in Leukapheresis Harvests From Normal Donors Treated With Sargramostim and G-CSF⁶²

Subsets	Sargramostim (n = 3)	G-CSF (n = 4)	Sargramostim + G-CSF (n = 3)
CD34+	$0.24\pm0.22^{\star}$	1.19 ± 0.33	0.34 ± 0.18
CD34+/CD38-	4.42 ± 3.40	0.81 ± 0.22	$4.73\pm2.72^{\star}$
CD34+/HLA-DR-	20.3 ± 2.9	20.7 ± 6.9	24.0 ± 9.3
CD34+/HLA-DR-/CD38-	$1.10\pm0.22^{\star}$	0.37 ± 0.19	$1.86\pm0.34^{\star}$

Values are percentages.

*P < .05 v G-CSF.

days followed by G-CSF at 10 μ g/kg/d for 2 days.⁶⁶ In comparison to the results of single-agent G-CSF for 4 days and combination rHuGM-CSF and G-CSF for 5 days, sequential administration failed to demonstrate any differences in the extent of mobilization as measured by CD34⁺ cells. In addition, the proportion of the CD38⁻ subset, which contains the more primitive hemotopoietic cells, was higher with the combination of sargramostim and G-CSF for 5 days.

Molgramostim also has been studied in combination or in sequence with G-CSF for mobilization of PBPCs.⁶⁷ The combination of the two cytokines resulted in dramatic and sustained increases in the number of CFU-GM per kilogram collected per harvest, with administration of G-CSF to patients already receiving molgramostim increasing the hematopoietic progenitor cell content nearly 80-fold. A randomized trial comparing combination therapy versus G-CSF or molgramostim (10 µg/kg) alone is ongoing. Additional trials are required to determine the optimal scheduling of cytokine administration as well as apheresis scheduling.

Priming effect before or during chemotherapy for AML. Myeloid leukemic cells and their precursors have GM-CSF receptors, and there is in vitro evidence that the proliferation and differentiation of these cells is supported by exposure to rHuGM-CSF.68-71 Thus, recruitment of chemoresistant resting leukemic cells into sensitive phases of the cell cycle by rHuGM-CSF may enhance the antileukemic effect of chemotherapy. The rHuGM-CSF-induced increases in leukemic cells in S phase and intracellular phosphorylation of cytarabine have been shown to promote drug-induced cell kill.72 In contrast to enhancing cytarabine cytotoxicity, Lotem and Sachs73 found that the typical features of apoptosis were prevented by rHuGM-CSF and G-CSF in a murine leukemic cell line. The growth factors also inhibited apoptosis induced by cytarabine, but the overall clonogenic cell reduction was not reduced. Because contradictory laboratory data exist, it has not been possible to predict the clinical benefit of cytokine priming before results of studies in patients with AML.

In a multicenter, randomized trial of 114 patients (17 to 75 years of age) with newly diagnosed AML, Büchner et al⁷⁴ compared use of chemotherapy alone with use of chemotherapy administered in conjunction with sargramostim priming. Sargramostim at 250 µg/m² was administered once daily by subcutaneous injection starting 24 hours before chemotherapy and continuing until neutrophil recovery occurred after the induction courses, consolidation course, and first two maintenance courses. Overall, 79% of sargramostim-treated patients and 84% of controls achieved disease remission; persistent leukemia was observed in 4% and 18% of patients, respectively. In patients younger than 60 years of age, complete remissions were achieved in 82% of sargramostim-treated patients and 73% of controls, with fewer relapses in the sargramostimtreated patients during the first 6 months (3% and 22%, respectively).74

Similar studies in patients with newly diagnosed AML receiving induction chemotherapy have been conducted with molgramostim.^{47,75-77} Patients received molgramostim at 250 μ g/m² or 5 μ g/kg/d starting either on days 1 to 3 before chemotherapy or with induction chemotherapy. Although a

trend toward benefit in disease-free survival was observed, the use of rHuGM-CSF during induction therapy of AML does not appear to have a significant impact on treatment outcome. The use of rHuGM-CSF for priming remains an intriuging therapeutic approach for AML. Because negative effects on the course of AML were rarely observed, additional studies are being conducted to determine the benefits and risks of growth factors administered before or concurrently with chemotherapy regimens in the treatment of AML. However, the definitive results of an ongoing ECOG trial are awaited before use of rHuGM-CSF for priming effects can be recommended outside of a clinical trial.

Adjunctive use to increase chemotherapy dose intensity. Adjunctive use of rHuGM-CSF may allow an increase in the dose intensity of combination chemotherapy regimens including drugs with a primary toxicity of myelosuppression; however, the benefits associated with rHuGM-CSF in patients receiving dose-intensive chemotherapy may be limited to early courses of therapy, because late-cycle thrombocytopenia is not prevented.78-83 The ability of sargramostim to support a multiplecycle high-dose chemotherapy regimen was evaluated in a phase III, double-blind, randomized trial of 56 patients with lymphoma or breast cancer. Patients received submyeloablative doses of cyclophosphamide, etoposide, and cisplatin (DICEP) and were randomized to receive sargramostim (250 µg/m²) or placebo subcutaneously every 12 hours.⁸³ Sargramostim-treated patients had a significantly decreased duration of neutropenia after the first course of chemotherapy in comparison to patients who received placebo (10 v 12 days; P = .01), but the difference did not achieve statistical significance after the second or third courses. Sargramostim-treated patients experienced a statistically significant 1.5-day delay in platelet recovery during the second course. There was no difference between the groups in numbers of hospitalizations for febrile neutropenia or the incidence of bacteremia, although any potential difference might have been obscured, because prophylactic oral ciprofloxacin was administered to all patients during neutropenia. However, the duration of hospitalization for neutropenic fever was shorter for sargramostim-treated patients in the first course. Importantly, the primary endpoint of this study was duration of neutropenia during the first course of therapy. The study was stopped because of a significant difference in duration of neutropenia; therefore, the small number of patients potentially obscures other clinical benefits of sargramostim therapy.⁸³

The feasibility of EAP (etoposide, doxorubicin, cisplatin) dose escalation using molgramostim at 10 μ g/kg/d starting on day 4 and continuing unitl recovery of granulocyte count was studied by Ford et al.⁷⁸ Intolerable myelosuppression, including grade 4 neutropenia or thrombocytopenia lasting at least 7 days, occurred in 4 of 5 patients receiving escalated doses of the EAP regimen. At the lowest doses of each agent, 3 of 6 patients had intolerable myelosuppression. The investigators concluded that molgramostim did not permit dose escalation of EAP.⁷⁸ In contrast, molgramostim at 5 μ g/kg/d allowed dose escalation of 5-fluorouracil (5-FU) with leucovorin to 425 mg/m²/d, with further 5-FU dose escalation according to individual tolerance.⁷⁹

EMERGING APPLICATIONS OF SARGRAMOSTIM

* (2)100 E/T: 10/1 E/T: 20/1 (3) MTT Reduction (%) 75 (4) * (8) (5) (8) (8) 50 (8) 25 0 None DEX DEX + DEX + None DEX DEX + DEX + GM-CSF IFN GM-CSF IFN

Fig 2. (A) *Fumigatus* hyphal damage induced by elutriated human monocytes incubated with 500 nmol/L dexamethasone (DEX) alone and with either 5 ng/mL sargramostim (rHuGM-CSF) or 1.2 ng/mL interferon- γ (IFN). Vertical bars denote standard errors of means, and the number of experiments performed are shown in parentheses. **P* < .05. (Reprinted with permission.⁹⁵)

USE IN INFECTIOUS DISEASE

Modulation of host defense against bacterial and fungal infections. There are several mechanisms by which rHuGM-CSF may enhance host defense mechanisms against bacterial and fungal infection. Exposure of neutrophils to rHuGM-CSF in vitro and in vivo has been shown to enhance expression of cell surface adhesion molecules, such as β -integrins, as well as receptors for the Fc portion of IgG, and receptors for activated complement components.84,85 Other effects of rHuGM-CSF on neutrophils include enhanced chemotaxis,17 phagocytosis,10 leukotriene B4 synthesis, release of arachidonic acid,^{86,87} and superoxide anion generation.^{44,88} The upregulation of neutrophil surface antigens combined with the induction of phagocyte migration and increased phagocytic activity contribute to a role for rHuGM-CSF in host defense. Sargramostim also prolongs neutrophil survival from 96 hours to at least 216 hours by preventing apoptosis.89 Finally, sargramostim induces the expression of class II MHC molecules on neutrophils, which could potentially allow neutrophils to act as antigen-presenting cells much like B cells, macrophages, and dendritic cells.90,91

As a result of its multilineage activity, similar functional effects of rHuGM-CSF have been observed in monocytes and macrophages. Administration of rHuGM-CSF increases the level of expression of a number of receptors found on macrophages, such as CD11a, CD11b, and CD11c, that augment adhesion-dependent phenomena and FcγRII (CDw32) receptors that bind Ig during phagocytosis.⁹²⁻⁹⁴ Upregulation of these receptors would be expected to aid the phagocytic ability of macrophages. Additonally, rHuGM-CSF enhances antibody-dependent cell cytotoxic activity, respiratory burst, and superoxide anion generation by macrophages and monocytes.^{13,17,19,52} Moreover, sargramostim significantly counteracted dexamethasone-induced inhibition of superoxide anion release by monocytes, and the fungicidal activity of dexamethasone-treated monocytes against *Aspergillus fumigatus* was enhanced (Fig 2).⁹⁵

Substantial evidence exists from in vitro and in vivo studies that rHuGM-CSF activates and enhances the ability of neutrophils and macrophages to phagocytize and destroy bacteria and fungi. Enhancement of the microbicidal activity of neutrophils by rHuGM-CSF was shown in vitro against Staphylococcus aureus,^{96,97} Torulopsis glabrata,⁹⁸ and Candida albicans.^{16,88,99} Neutrophils treated with rHuGM-CSF killed 90% of intracellular C albicans in comparison to 50% of intracellular yeast cells killed by untreated neutrophils.99 Similarly, enhancement of the microbicidal activity of monocytes by rHuGM-CSF was shown in vitro against C albicans,16 A fumigatus,95,100 Histoplasma capsulatum,¹⁰¹ Cryptococcus neoformans,¹⁰² and Trypanosoma cruzi.¹⁰³ Functional studies of neutrophils and monocytes isolated from patients treated with rHuGM-CSF at 250 µg/m²/d indicate that phagocytic and cytotoxic activity against S aureus is increased.^{97,104} The percentage of S aureus phagocytosed or killed after 20 minutes significantly increased from 62% before rHuGM-CSF treatment to 72% during treatment (P = .0028).⁹⁷

Sargramostim also promotes killing of *Mycobacterium avium* complex.¹⁰⁵⁻¹⁰⁸ Significant growth inhibition of *Mycobacterium avium* complex was observed in human macrophages treated with sargramostim or tumor necrosis factor α (TNF α ; Table 3).¹⁰⁸ A similar effect was observed using a mouse model of disseminated *Mycobacterium avium* complex infection. Significantly (P = .04) lower concentrations of *Mycobacterium avium*

Table 3. Growth Inhibition of Mycobacterium Avium Complex
in Human Macrophages Treated With Sargramostim or TNF α^{108}

Cytokine (dose)	Percentage of Inhibition*
rHuGM-CSF (1 U/mL)	36 ± 6
rHuGM-CSF (10 U/mL)	58 ± 4
rHuGM-CSF (100 U/mL)	50 ± 4
TNFα (50 U/mL)	37 ± 6
TNFα (500 U/mL)	51 ± 6
TNFα (5000 U/mL)	44 ± 4
rHuGM-CSF (10 U/mL) + TNFα (500 U/mL)	41 ± 8

*Percentage of inhibition = (CFU in control well – CFU in well with each cytokine)/CFU in control well \times 100. Values are means \pm SEM.

complex were present in the liver and spleen of mice treated with sargramostim for 14 days compared with control mice.¹⁰⁵ These data also suggest that sargramostim enhances the antimy-cobacterial effect of clarithromycin, azithromycin, amikacin, and ofloxacin.^{105,107}

In other animal studies, enhancement of microbicidal activity by rHuGM-CSF has been confirmed. Survival was significantly (P < .05) improved in neonatal rats when sargramostim was administered 6 hours before inoculation of a lethal dose of *S aureus*.¹⁰⁹ Similarly, in neutropenic mice, administration of molgramostim 1 to 5 µg/d protected against lethal infections of *S aureus* and *Pseudomonas aeruginosa*; survival was significantly increased in molgramostim-treated mice infected with either *S aureus* (70% v 20%, P < .05) or *P aeruginosa* (50% v 0%, P < .01).¹¹⁰

Recombinant murine GM-CSF protected 62% of neutropenic rats from a lethal inoculum of *C albicans* and reduced lung injury. Importantly, there was no effect of murine GM-CSF on the neutrophil count, suggesting that the protective mechanism involved led to enhanced host defense mechanisms.¹¹¹

Adjunctive treatment of fungal infections. The effect of sargramostim on the incidence and severity of fungal infections was observed in randomized, double-blind studies of the drug in patients undergoing AuBMT and in patients with AML.^{112,113} Fungal infections developed in 4 AuBMT patients who received placebo in comparison to 2 AuBMT patients who received sargramostim.¹¹² Two of the infections in placebo-treated patients were disseminated aspergillosis. In the phase III ECOG trial of 99 elderly patients undergoing chemotherapy for AML, sargramostim significantly (P = .02) reduced mortality due to fungal infection.¹¹³ One of 8 patients who received sargramostim died as a result of fungal infection, whereas 9 of 12 placebo-treated patients developed fatal fungal infections.

A pilot study of molgramostim as adjuvant therapy for fungal infections was conducted in cancer patients with proven majororgan or disseminated fungal infection.114 Of 8 evaluable patients, 6 had a neutrophil response to molgramostim; 4 of these patients were completed cured of the fungal infection and the other 2 had a partial response. Several case series have reported a response to adjunctive treatment with sargramostim for fungal infections. Three human immunodeficiency virus (HIV)-infected patients with oropharyngeal candidiasis refractory to fluconazole at doses of 200 mg daily or greater for at least 14 days were treated with sargramostim at 125 μ g/m²/d.¹¹⁵ Fluconazole treatment was maintained at the same dose. All patients experienced improvement in signs and symptoms of oropharyngeal candidiasis by week 2. No significant adverse events occurred, including no upregulation of HIV-1 replication, and therapy was well tolerated. When sargramostim was discontinued, 2 of 3 patients relapsed.115

Sargramostim also has been administered to patients with rhinocerebral and disseminated mucormycosis, a rare opportunistic infection associated with a mortality rate exceeding 50%.¹¹⁶ Three of four patients with mucormycosis have been successfully treated with sargramostim (doses ranging from 250 to $500 \mu g/d$ for 14 days to 6 months) in combination with traditional surgical and medical treatment. The three patients

were disease-free at periods of 6 months, 18 months, and 3 years after surgery.

HIV infection. Initial use of sargramostim in patients with HIV infection focused on its ability to ameliorate drug-induced myelosuppression.¹¹⁷⁻¹¹⁹ In a phase I/II study in patients with Kaposi's sarcoma who became neutropenic while receiving zidovudine and interferon α , administration of sargramostim resulted in a prompt increase in absolute neutrophil count in all patients and an absolute neutrophil count greater than 1,000 cells/µL within 7 days; there was no increase in p24 antigen levels.¹¹⁸ Sargramostim (250 or 500 µg/m²/d administered by subcutaneous injection) also has been used to ameliorate chemotherapy-induced neutropenia in patients with acquired immunodeficiency syndrome (AIDS)-related Kaposi's sarcoma receiving a regimen of doxorubicin, bleomycin, and vincristine (ABV).¹¹⁷ Although both sargramostim doses allowed the chemotherapy regimen to be continued without a dose reduction, the lower sargramostim dose was better tolerated. In a recent study in 12 patients with advanced HIV infection (CD4+ cell count $\leq 200/\mu$ L) who were receiving zidovudine (300 to 1,200 mg/d), administration of sargramostim in a dosage of 50, 125, or 250 μ g/m²/d by subcutaneous injection resulted in significant increases in absolute neutrophil count and monocyte counts at all three dosage levels.119

Potential uses for the drug in HIV patients were expanded when it became evident that rHuGM-CSF activates and enhances the ability of neutrophils and macrophages to phagocytize bacteria, fungi, and intracellular parasites, which has important implications for the prophylaxis and treatment of opportunistic infections in this patient population.

Although there were initial concerns that use of rHuGM-CSF in HIV-infected patients might stimulate HIV replication and increase viral load, these concerns have not been substantiated. In vitro studies are somewhat conflicting regarding the effect of rHuGM-CSF on HIV replication. Numerous studies have demonstrated enhancement of viral replication when HIV-infected monocytes or macrophages are exposed to rHuGM-CSF¹²⁰⁻¹²⁴; however, three studies have reported suppression of HIV expression by rHuGM-CSF.¹²⁵⁻¹²⁷ An additional finding from in vitro studies is that rHuGM-CSF enhances the antiretroviral activity of some dideoxynucleoside antiretroviral agents, such as zidovudine and stavudine, possibly by increasing intracellular phosphorylation of these agents to their active metabolite.^{124,128,129}

Early clinical trials evaluating the effect of rHuGM-CSF on viral replication in HIV patients failed to show an increase in viral load as determined by serum p24 antigen levels as long as patients received concurrent zidovudine^{117,118,130-132}; however, in studies in which molgramostim was administered without an antiretroviral, some patients did experience an increase in serum p24 antigen levels.^{133,134} Subsequent trials using a more sensitive polymerase chain reaction assay for viral load determination confirmed that rHuGM-CSF does not result in an increase in viral load during or after CSF therapy in HIV patients receiving concurrent zidovudine.^{119,135} More recently, administration of sargramostim to patients on stable, highly active antiretroviral therapy including a protease inhibitor has been shown to result in no upregulation of viral load by polymerase

chain reaction.^{136,137} These HIV-positive patients receiving sargramostim have experienced a significant (P = .0372) increase in CD4 count and decrease in viral load $\ge 0.5 \log$.

Interestingly, the studies by Massari et al¹²⁶ and Matsuda et al127 that reported suppression of HIV expression by rHuGM-CSF were conducted before the role of coreceptors for HIV infection was appreciated. Indeed, the investigators examined the effects of rHuGM-CSF on CD4 expression as a potential mechanism for the reduction in HIV expression, but found CD4 expression to be unchanged. A recent study provides a possible mechanism for their findings. Exposure to sargramostim has recently been shown to downregulate expression of the C-C chemokine receptor CCR5, a β-chemokine receptor on macrophages, and to reduce the susceptibility of macrophages to infection by a macrophage-tropic strain of HIV.125,138 CCR5 has been shown to be the major coreceptor required for infection of macrophages by macrophage-tropic strains of HIV. Sargramostim-stimulated monocytes produced high levels of B-chemokines, macrophage inflammatory protein- 1α (MIP- 1α), and MIP-1 β in the medium. This medium was able to protect bystander cells from entry by JRFL, a macrophage-tropic strain of HIV.

Another possible role for sargramostim in HIV-infected patients is in the prevention or treatment of opportunistic infections. Based on results of in vitro studies demonstrating that rHuGM-CSF promotes killing of Mycobacterium aviumintracellulare^{105,107} and in vitro and murine studies indicating that rHuGM-CSF can enhance the antimycobacterial effects of some antimicrobial agents, including azithromycin, ofloxacin, and clarithromycin, 105,107 investigations of sargramostim for the adjunctive treatment of Mycobacterium avium-intracellulare were initiated. In a small study, AIDS patients with disseminated Mycobacterium avium-intracellulare were randomized to receive azithromycin with or without sargramostim for 6 weeks.¹³⁹ Mycobacteremia and monocyte function were assessed biweekly. Mean superoxide anion production was significantly increased in monocytes obtained from all 4 patients receiving sargramostim (53% to 199% relative to controls) and these patients had a 60% reduction in the number of viable intracellular Mycobacterium avium-intracellulare per milliliter at the end of treatment. Patients receiving azithromycin alone had no increase in superoxide anion production and only a 28% reduction in viable Mycobacterium avium-intracellulare per milliliter. These data indicate that sargramostim activates monocytes in AIDS patients with Mycobacterium avium-intracellulare bacteremia and deserves further study as adjunctive therapy in these patients.

A multicenter phase III randomized, double-blind, placebocontrolled trial of sargramostim in patients with advanced HIV disease is ongoing to compare the incidence and time to first opportunistic infection or death. Patients with CD4 count \leq 50 cells/µL and receiving a stable antiretroviral regimen before and during the study are eligible. Patients are randomized to receive either sargramostim 250 µg/d 3 days per week for a minimum of 24 weeks or placebo. Secondary objectives include incidence of AIDS-related opportunistic malignancies and esophageal candidiasis; survival; pharmacoeconomic and quality-of-life parameters; changes in HIV viral load or CD4⁺ lymphocyte counts; 4497

incidence, degree, and duration of neutropenia; and concurrent use of open-label cytokines.

USE AS A VACCINE ADJUVANT

rHuGM-CSF is the principal mediator of proliferation, maturation, and migration of dendritic cells, important antigenpresenting cells that play a major role in the induction of primary and secondary T-cell immune responses.14,20,140 Dendritic cells display antigens on their surface in conjunction with class II major histocompatibility complex (MHC). rHuGM-CSF also increases class II MHC expression.¹² Once presented, the antigen can be recognized by helper CD4⁺ T cells,¹⁴¹ which provide support for the development of B cells and cvtotoxic CD8⁺ T cells. By augmenting antigen presentation to lymphocytes by dendritic cells, rHuGM-CSF stimulates T-cell immune responses.12,14 rHuGM-CSF has been demonstrated to augment the primary in vitro immune response to sheep red blood cells by murine spleen cells.¹⁴² rHuGM-CSF also is important to the immune response to vaccination, because it enhances expression of costimulatory molecules such as B7 and adhesion molecules (eg, intercellular adhesion molecule [ICAM]) that are necessary for the interaction of antigen-presenting cells with T cells; it also enhances production of other cytokines such as IL-1, TNF, and IL-6, which promote expansion and differentiation of B and T lymphocytes. In addition, rHuGM-CSF primes T cells for IL-2-induced proliferation¹⁴³ and augments lymphokine-activated killer (LAK) cell generation in conjunction with IL-2.15,144 The important role of rHuGM-CSF in the maturation and function of antigen-presenting cells, such as dendritic cells and macrophages, as well as its ability to affect T-cell immunity, provides the basis for its potential evaluation as a vaccine adjuvant in new immunotherapy strategies for infectious diseases and cancer.

Local injection of rHuGM-CSF would be expected to enhance vaccine immunogenicity and would likely be well tolerated based on clinical experience in other uses. Disis et al145 evaluated the use of sargramostim as an adjuvant for protein- and peptide-based vaccines in rats. Tetanus toxoid was used as the foreign antigen system, and peptides derived from a self antigen, rat neu protein, were used as the tumor antigen system. A series of initial experiments demonstrated that intradermal injections of sargramostim every 24 hours for a total of five inoculations increased the number of class II MHC+ cells in regional lymph nodes that peaked at the fourth inoculation, whereas subcutaneous injections of sargramostim on the same schedule increased these cells with a peak after the second inoculation. This conditioning schema was then used, with tetanus toxoid administered at the beginning or end of the immunization cycle. Intradermal immunization was more effective than subcutaneous immunization in eliciting specific immunity to the tetanus toxoid antigen. In addition, intradermal injection of sargramostim as a single dose with antigen was similarly effective in eliciting specific antibody and cellular immunity as the use of Freund's adjuvant or alum (Fig 3). Inoculation with rat neu peptides and sargramostim elicited a strong delayed-type hypersensitivity response, whereas the peptides alone were nonimmunogenic. Sargramostim was as effective as Freund's adjuvant in generating rat neu-specific



delayed-type hypersensitivity responses after immunization with the peptide-based vaccine. These studies demonstrated that sargramostim was an effective adjuvant for elicitation of immunity to both antigen systems, comparing favorably with other standard adjuvants.

Results of several preliminary studies using molgramostim in conjunction with hepatitis B^{141,146} and tetravalent influenzae virus vaccine¹⁴⁷ suggest that rHuGM-CSF may have a potential role as an antiviral vaccine adjuvant; however, further evaluation is needed in this setting. Its evaluation as an adjuvant to vaccines and other immunotherapies for tumors is promising and is discussed in the subsequent section.

USE IN ANTITUMOR THERAPY

Antitumor effects. The functional effects of granulocytes, lymphocytes, and macrophages are important in patients with malignancies because of the ability of these cells to exhibit antitumor activity. In vitro, rHuGM-CSF has been shown to slightly enhance the cytotoxic activity of peripheral blood monocytes and lymphocytes and markedly increase antibody-dependent cellular cytotoxicity¹⁴⁸ and to enhance monocyte cytotoxicity against a malignant melanoma cell line.¹⁴⁹ rHuGM-CSF has also been shown to augment the cytotoxic

Fig 3. rHuGM-CSF, as an adjuvant, elicits delayedtype hypersensitivity (DTH) responses to tetanus toxoid (tt) similar to those seen in animals immunized with a standard adjuvant. Rats were injected with Freund's adjuvant (CFA) subcutaneously (sq), alum sq, rHuGM-CSF intradermally (id) or sq (5 µg), and phosphate-buffered saline (PBS) sq with tt at a concentration of 3 limit flocculation (Lf) units. Immunizations were administered on 1 day only with no repeated administration of rHuGM-CSF. Six rats were included in each experimental group. Figure represents data collected from two separate experiments. Twenty days after immunization, a DTH response was measured in the immunized animals. Antigen was applied to rat ear and responses measured at 48 hours. Ear swelling of experimental compared with control ear was measured. Results are shown as the mean and standard deviation of measurements taken from each experimental group. (Reprinted with permission¹⁴⁵)

activity of peripheral blood monocytes in antibody-dependent cellular cytotoxicity against numerous human tumor cells in the presence of various monoclonal antibodies¹⁵⁰ and to enhance IL-2-mediated LAK cell function.151,152 In tumor-infiltrating macrophages, it also increases secretion of matrix metalloelastase with subsequent production of angiostatin, which inhibits angiogenesis and suppresses the growth of lung metastases.¹⁵³ rHuGM-CSF may also enhance the immunogenicity of tumor cells through facilitation of tumor antigen presentation.⁵⁶ In a comparative study in mice, the most potent stimulator of specific antitumor immunity was tumor cells engineered to secrete GM-CSF.¹⁵⁴ Also, as previously noted, sargramostim has been shown in rats to be an excellent adjuvant for generation of immune responses to tumor antigen-derived peptides.¹⁴⁵ Thus, rHuGM-CSF might enhance functions of cells critical for immune activation against tumor cells, alone or with other cytokines or monoclonal antibodies, making it potentially useful in the biotherapy of malignant diseases.

In a phase I study in patients with cancer, administration of sargramostim enhanced monocyte antibody-dependent cellular cytotoxicity (Fig 4) and increased secretion of both TNF α and interferon.¹⁹ In another study in patients with metastatic solid tumors, sargramostim was administered once daily for 14 days

Fig 4. Antibody-dependent cellular cytotoxicity (ADCC) of monocytes after treatment with sargramostim. Monocytes were collected from patients 2 days after a bolus infection (A) and 3 (B) and 10 (C) days after the start of a continuous infusion of sargramostim. Antibody-dependent cellular cytotoxicity activity was measured against antibodycoated chicken erythrocytes by a Cr51-release assay. Experimental results were compared statistically with the average of two baseline assays. (Reprinted with permission.19)



EMERGING APPLICATIONS OF SARGRAMOSTIM

every 28 days; monocyte cytotoxicity against HT29 tumor cells was enhanced by the cytokine treatment.⁴⁶ No clinical effects on tumor regression were apparent in either study. Sargramostim is under evaluation in an open-label, phase II trial as surgical adjuvant therapy in patients with advanced melanoma at very high risk of recurrence.¹⁵⁵ Sargramostim at 125 μ g/m²/d was administered subcutaneously for 14 days every 28 days beginning within 60 days of the last evidence of tumor. Treatment is continued until recurrence or a tumor-free interval of 1 year. An interim analysis of 25 patients demonstrated a significant prolongation of disease-free survival (*P* = .04) and survival (*P* = .02) compared with 50 matched historical control patients.¹⁵⁵ These initial results are encouraging; long-term follow-up is needed.

A phase Ib trial was conducted in 20 patients with metastatic melanoma to evaluate the use of sargramostim as an adjuvant to R24, a murine monoclonal antibody that mediates complementdependent and antibody-dependent cellular cytotoxicity of melanoma tumor targets.¹⁵⁶ The rationale for this combination was the hypothesis that upregulation of monocyte and granulocyte antibody-dependent cellular cytotoxicity induced by sargramostim might enhance antitumor activity. Sargramostim (150 $\mu g/m^2/d$ administered by subcutaneous injection for 21 days) was administered alone or in conjunction with R24 (10 or 50 mg/m² administered by continuous intravenous infusion on days 8 through 15). Measurement of direct cytotoxicity and antibody-dependent cellular cytotoxicity indicated that sargramostim enhanced monocyte and granulocyte cytotoxicity by week 3 in all evaluable patients.¹⁵⁶ Of the 6 patients who received sargramostim alone, 3 had no response (2 had stable disease) and 3 had disease progression; in the 14 patients who received sargramostim plus R24, 2 had a partial response, 6 patients had no response (3 had stable disease), and 6 developed progressive disease.156

The Pediatric Oncology Group performed a phase II study to evaluate the use of sargramostim to enhance antibodydependent cellular cytotoxicity of a chimeric anti-GD2 monoclonal antibody (ch14.18) in the treatment of recurrent or refractory neuroblastoma.157 Sargramostim was administered in a dosage of 10 µg/kg daily for 14 days with 5-hour infusions of ch14.18 at 50 mg/m² daily for 4 days. Thirty-two patients who had failed to respond to 1 to 4 therapeutic regimens, including BMT in 18 patients, received 70 courses of treatment. In 27 patients evaluable for response, there were 1 complete response, 3 partial responses, 1 mixed response, and 2 stable disease. When analyzed by site of disease, in 18 patients with marrow disease, there were 4 complete responses and 1 partial response; in 21 patients with bone involvement, there were 1 complete response and 2 partial responses. Two patients with large tumor masses had greater than 60% reduction in tumor size. Among the responding patients, 4 were alive at follow-up ranging from 9 to 20 months, whereas those with progressive disease had a median survival of 3 months. All responding patients had an increase in neutrophil-mediated antibody-dependent cellular cytotoxicity to greater than 20 lytic units, whereas 9 of 12 patients with progressive disease had peak antibody-dependent cellular cytotoxicity activity less than 20 lytic units. These findings were the basis for a recommendation that a phase III trial in the setting of minimal residual disease is warranted.¹⁵⁷

Adjuvant to tumor vaccines. Based on enhancement of functional effects on monocytes, macrophages, and antigenpresenting cells (dendritic cells, macrophages),^{15,142} sargramostim has been studied for its potential to enhance the immune response to antitumor immunotherapies, including autologous tumor cell vaccines, recombinant peptide tumor vaccines, and autologous Id-KLH tumor vaccines.

Leong et al¹⁵⁸ administered sargramostim at 125 to 250 μ g as an adjuvant to a melanoma vaccine that consisted of irradiated autologous melanoma cells with Bacillus Calmette-Guérin vaccine (BCG vaccine) in 20 stage IV melanoma patients. Patients received multiple cycles that consisted of vaccine plus sargramostim on day 1, with local injection of sargramostim alone in the vaccine site on days 2 to 5; 48 hours before cycles 1, 3, and 4, cyclophosphamide at 300 mg/m² was administered. Four patients showed partial to complete responses (20%), 4 had stable disease (20%), and the remaining 12 patients had disease progression (60%). In the responding patients, regression of visceral metastases was observed. The results demonstrated the ability of patients bearing a significant tumor burden to respond specifically to their autologous melanoma.

Based on the fact that autologous tumor-derived Ig idiotype proteins (Id) have been shown to induce effective antitumor activity in experimental models and B-cell lymphoma, a vaccine containing autologous Id-KLH (keyhole limpet hemocyanin, a foreign protein used as a vaccine adjuvant) conjugates was administered to patients with multiple myeloma with either rHuGM-CSF or IL-2 as an adjuvant.¹⁵⁹ Results of skin testing with autologous and unrelated Id were used to assess the specificity of the immune response; rHuGM-CSF appeared to be a better adjuvant than IL-2 in these patients.

Immunotherapy for AML. Future directions in the treatment of AML may include immunotherapy based on the effect of rHuGM-CSF on T-lymphocyte cytotoxic functions and surface adhesion proteins. Several preliminary investigations have been conducted using molgramostim in patients with AML. The effect of molgramostim at 5 µg/kg/d on activated killer cell activity was studied in 20 patients with AML undergoing AuBMT.160 Activated killer cell function was investigated before AuBMT, during rHuGM-CSF therapy, and after withdrawal. The actuarial risk of relapse was also analyzed and compared with a historical control group of 20 patients transplanted before initiation of this study. Activated killer cell function was significantly enhanced with rHuGM-CSF (P <.001); during rHuGM-CSF treatment, median activated killer cell function increased from 1.8% before AutoBMT to 35% and remained increased after withdrawal of rHuGM-CSF (median, 20%). After a median follow-up of 24 months, the actuarial risk of relapse was 37.4% in rHuGM-CSF-treated patients compared with 49.5% in controls (P = .05). Additionally, none of the 7 patients with activated killer cell activity $\geq 20\%$ in the first 2 to 5 weeks after AutoBMT have relapsed, compared with 6 of 9 patients with activated killer cell activity less than 20% (P < .02).

Exposure of AML cells to rHuGM-CSF upregulates expression of ICAM-1 (CD54) and lymphocyte function associated molecule-3 (LFA-3; CD58), but does not increase their sensitivity to lysis by IL-2–activated natural killer cells.¹⁶¹ rHuGM-CSF induces a significantly greater upregulation of ICAM-1 on

JAMES O. ARMITAGE

leukemic CD34⁺ cells than their CD34⁻ counterparts. When AML cells are exposed to rHuGM-CSF before incubation with killer cells, their subsequent clonogenic activity is significantly reduced. These data suggest that administration of effector cell activators, such as IL-2, and target cell modulators, such as rHuGM-CSF, may have therapeutic benefit in patients with minimal residual myeloid leukemia.

OTHER POTENTIAL USES

Mucositis, stomatitis, and diarrhea. Mucositis, stomatitis, and diarrhea are frequent complications of high-dose chemoradiotherapy. Mucosal epithelial cells in the gastrointestinal tract are susceptible to direct damage from these therapies, resulting in dysphagia and decreased oral intake and potentially leading to airway compromise. Mucosal damage may be further aggravated by infections or hemorrhage related to myelosuppression. rHuGM-CSF has been shown to stimulate the migration and proliferation of endothelial cells and promote keratinocyte growth, suggesting that the growth factor has a direct effect on mucosal cells.^{162,163} In addition, by decreasing the severity and duration of neutropenia, rHuGM-CSF may reduce the severity and duration of mucositis.

Several clinical trials evaluating sargramostim for hematopoietic support have shown a coincidental benefit of the drug on mucositis. In addition to enhancing myeloprotection and permitting dose-intensification of chemotherapy, the incidence of mucositis was reduced in sarcoma patients who received sargramostim after chemotherapy.¹⁶⁴ In a phase III placebocontrolled trial of sargramostim in patients undergoing allogeneic BMT, 8% of sargramostim-treated patients compared with 29% of placebo-treated patients developed grade 3 or 4 mucositis (P = .005).¹⁶⁵

Based on these results, several prospective trials have been conducted to evaluate the effect of rHuGM-CSF on mucositis. In a nonrandomized trial, the effect of sargramostim on oral mucositis was assessed in pediatric patients undergoing stem cell transplant.¹⁶⁶ Children who received thiotepa, etoposide, and total body irradiation followed by sargramostim experienced a significantly shorter duration of mucositis than those children who did not receive sargramostim (12.2 v 20.3 days, P = .02). However, the severity of mucositis was similar between the groups. In this same study, patients treated with thiotepa, etoposide, and cyclophosphamide as the preparative regimen experienced a similar duration and severity of mucositis regardless of whether they received sargramostim. Although recovery of neutrophils was faster in sargramostim-treated patients, there was no correlation between recovery of neutrophils and resolution of mucositis.

A phase I trial of sargramostim as a mucoprotectant was conducted in 10 patients with advanced head and neck cancer who received adjuvant radiation after surgery or chemotherapy.¹⁶⁷ Four patients developed grade 3 or worse mucositis, and the remaining 6 patients had grade 1 or 2 mucositis. In comparison to 13 historical control patients, grade 3 or worse mucositis was reduced by half from 85% to 40% with sargramostim administration. In a phase I trial of colorectal cancer patients receiving escalating doses of 5-FU, sargramostim used in conjunction with leucovorin resulted in decreased rates of diarrhea relative to historical patients.¹⁶⁸

Topical application of molgramostim in the treatment or

prevention of mucositis has also been investigated.^{169,170} Of 10 BMT patients who received molgramostim (400 μ g) in 100 mL of water administered as a mouthwash and then swallowed, only grade 1 or 2 mucositis was observed; in comparison, 8 of 10 patients who did not receive the mouthwash developed grade 4 mucositis.¹⁷⁰ When the molgramostim mouthwash was used as a 2-minute rinse and not swallowed, there was no benefit with regard to mucositis; however, a positive correlation between molgramostim dose and leukocyte recovery was observed, providing evidence for systemic absorption and a hematopoietic effect.¹⁶⁹

Wound healing. As mentioned previously, rHuGM-CSF has been shown to enhance the migration and proliferation of endothelial cells and to promote keratinocyte growth.^{162,163} Animal studies have also shown that local application of rHuGM-CSF to wounds results in increased formation of granulation tissue, increased breaking strength of incisional wounds, and reversal of wound contraction in infected wounds, resulting in a faster time to wound healing.¹⁷¹⁻¹⁷³ Intradermal injections of rHuGM-CSF (molgramostim and regramostim) in humans with lepromatous leprosy resulted in enlarged keratinocytes, keratinocyte proliferation, thickening of the epidermis, accumulation of Langerhans cells, and enhanced healing.174,175 A number of case reports and small series reports have been published on the use of rHuGM-CSF as a treatment for nonhealing wounds and ulcers.¹⁷⁶⁻¹⁷⁸ Using various routes of rHuGM-CSF administration (subcutaneously around the wound, incubated with skin grafts, and as a topical application in sterile water), signs of wound healing occurred rapidly and total wound closure was achieved between 10 days and 5 weeks after treatment.

The safety and feasibility of using molgramostim to treat patients with vascular leg ulcers was evaluated by Arnold et al.¹⁷⁹ Ten patients were treated with four intradermal injections of molgramostim at 50 μ g around the perimeter of their ulcers every 2 weeks for a total of 12 weeks. No hematological abnormalities were observed and the injections were reported to be relatively painless. Although this study was not designed to determine efficacy, some patients demonstrated complete or partial healing of their ulcers.

In a double-blind, placebo-controlled study, 40 patients with chronic leg ulcers were randomized to receive either 400 µg of rHuGM-CSF or a similar volume of saline; equal-dose injections were administered into four quadrants around the wound.¹⁸⁰ The study was unblinded prematurely and data on 25 treated patients were reported. By day 8 after treatment, a significant (P < .005) difference in mean ulcer surface area reduction was observed between the two arms in favor of rHuGM-CSF. Complete healing by week 8 was observed in 8 of 16 patients treated with rHuGM-CSF and 1 of 9 placebo-treated patients. No significant side effects were reported during the trial. These findings from case reports and small studies indicate that some patients with nonhealing ulcers may benefit from rHuGM-CSF therapy. More research is needed to determine the appropriate dose, optimal dosing frequency, and efficacy of rHuGM-CSF in different types of wounds and ulcers.

Hypercholesterolemia. Elevated cholesterol concentrations result from disordered lipid metabolism. The liver is the major site of cholesterol biosynthesis and excretion; however, macrophages produce factors that activate cholesterol biosynthesis or

EMERGING APPLICATIONS OF SARGRAMOSTIM

Therapeutic Use	Preclinical Actions of rHuGM-CSF	References	Clinical Results With rHuGM-CSF	References
Fungal infections	 Increases receptor expression on macrophages. Enhances fungicidal activity against Aspergillus fumigatus, Candida albicans, Cryptococcus neoformans, Histoplasma capsulatum, Torulopsis glabrata. Counteracts dexamethasone-induced inhibition of superoxide anion release by monocytes. 	16, 88, 92-95, 98-102	 Decreases incidence of fungal infections versus placebo in AuBMT patients. Reduces mortality due to fungal infections in elderly patients with AML. As an adjunct to amphotericin B, improves recovery from <i>Candida</i> and <i>Aspergillus</i> infections. Improves oropharyngeal candidiasis refractory to fluconazole in HIV-infected patients. 	112-116
HIV infection and its complications	Suppresses HIV expression. Enhances antiretroviral activity of zidov- udine and stavudine. Downregulates expression of CCR5, reducing the susceptibility of macro- phages to HIV infection. Promotes killing of <i>Mycobacterium</i> <i>avium-intracellular</i> (MAC).	105, 107, 124-129	Increases CD4 count. Decreases viral load. As adjunctive treatment of MAC, reduces the number of viable intracellular MAC/mL.	136, 137, 139
Vaccine adjuvant	 Increases class II MHC expression and stimulates T-cell immune responses. Augments the primary in vitro immune response to sheep red blood cells by murine spleen cells. Enhances expression of costimulatory molecules and adhesion molecules and enhances production of other cytokines. Primes T cells for IL-2-induced prolifera- tion. Augments LAK cell generation in con- junction with IL-2. 	12, 14, 15, 142-145	Enhances antibody response to hepatitis B vaccine. Increases the percent of patients who seroconverted to all three strains of flu vaccine.	141, 146, 147
Antitumor therapy	Enhances monocyte cytotoxicity against human tumor cells. Enhances IL-2-mediated LAK cell func- tion. Increases secretion of matrix metal- loelastase with subsequent produc- tion of angiostatin. Facilitates tumor antigen presentation.	56, 149-153	Prolongs disease-free survival and overall survival compared with his- torical controls in patients with advanced melanoma. Of 20 stage IV melanoma patients who received sargramostim as an adjuvant to a melanoma vaccine, 4 had partial to complete responses.	155-157
Immunotherapy for AML	Enhances activated killer cell function. Upregulates expression of intercellular adhesion molecule-1 and lymphocyte function associated molecules.	160, 161	Decreases risk of relapse compared with controls (37.4% v 49.5%).	160
Mucositis, stomatitis, diarrhea	Stimulates the migration and prolifera- tion of endothelial cells and promotes keratinocyte growth.	162, 163	Reduces incidence and severity of muco- sitis in patients with sarcoma, advanced head and neck cancer, and those undergoing allogeneic BMT. Shortens the duration of mucositis in children undergoing stem cell trans- plant. Decreases rates of diarrhea in colorectal cancer patients receiving 5-FU.	164-168, 170
Wound healing	Increases formation of granulation tissue. Increases breaking strength of incisional wounds. Decreases time to wound healing.	171-173	Intradermal injections of rHuGM-CSF results in enlarged keratinocytes, kera- tinocyte proliferation, thickening of the epidermis, accumulation of Lang- erhans cells, and enhances healing. Reduces mean ulcer surface area versus saline in patients with chronic leg ulcers	174-180

Table 4. Summary of Emerging Applications of rHuGM-CSF

Abbreviations: CCR5, β-chemokine receptor on macrophages; MHC, major histocompatibility complex; LAK, lymphokine-activated killer; 5-FU, fluorouracil.

excretion, and mononuclear phagocytes play an important role in the processing and transport of cholesterol.¹⁸¹ Activated T-cell products also can affect the synthesis and accumulation of cholesterol by mononuclear phagocytes. Therefore, rHuGM- CSF could indirectly affect cholesterol levels by stimulating the activity of macrophages in the liver or phagocytic cells in the circulation or present at the site of an atherosclerotic plaque.¹⁸¹ The ability of regramostim to lower serum cholesterol

4501

. .

JAMES O. ARMITAGE

concentrations was reported almost a decade ago.¹⁸¹ Since then, efforts have focused on determining the mechanism(s) of this effect. In rabbits, the reduction in serum cholesterol is accompanied by an increase in the levels of mRNA for very low density lipoprotein (VLDL) receptors in muscle; the levels of LDL receptor mRNA in liver are unchanged. These findings suggest that the cholesterol-lowering effect of rHuGM-CSF (molgramostim) may be mediated by enhancement of macrophage functions in lipid metabolism and the increase in mRNA for VLDL receptor.¹⁸²

Treatment of pulmonary alveolar proteinosis. Pulmonary alveolar proteinosis includes a heterogenous group of diseases, both congenital and acquired, that are characterized by accumulation of large quantities of lipid- and protein-rich eosinophilic material (ie, surfactant) within the alveoli and airways.¹⁸³ Murine studies indicate that mice carrying a null allele of the GM-CSF gene develop a pulmonary abnormality that resembles alveolar proteinosis, suggesting that GM-CSF regulates the clearance or catabolism of surfactant proteins and lipids.¹⁸⁴⁻¹⁸⁶ Additionally, dysregulation of inflammatory cell activity due to the lack of GM-CSF may have detrimental effects on host defense and contribute to further lung injury.¹⁸³ An anecdotal report of rHuGM-CSF use in an adult with this disease has been published and indicates some improvement in symptoms with cytokine therapy.¹⁸⁷

SUMMARY AND CONCLUSIONS

rHuGM-CSF stimulates the proliferation and differentiation of multiple hematopoietic progenitor cells in the myeloid lineage and activates or augments many of the functional activities of mature neutrophils, monocytes/macrophages, and dendritic cells, enhancing host defenses against a broad spectrum of invading microorganisms. These properties have greatly expanded the possible therapeutic benefits of the cytokine in a wide variety of settings (Table 4), particularly those in which prevention of infection is desirable. The drug may be useful as prophylaxis or adjunctive treatment of bacterial or fungal infections in immunocompromised individuals, including cancer patients receiving myelosuppressive chemotherapy and patients with advanced HIV infection. In addition, exposure to rHuGM-CSF has recently been shown to reduce the susceptibility of macrophages to infection by HIV. Sargramostim is being evaluated as a vaccine adjuvant against infectious diseases and malignancies and as immunotherapy in the treatment of various malignancies, including melanoma and neuroblastoma.

Based on the increasing variety of biologic effects being attributed to endogenous GM-CSF, additional clinical uses for sargramostim and molgramostim are under investigation. Because rHuGM-CSF has been shown to stimulate the migration and proliferation of endothelial cells and local application of rHuGM-CSF in animal studies has shown faster wound healing times, clinical trials have evaluated rHuGM-CSF in patients susceptible to mucosal damage, such as mucositis, stomatitis, and diarrhea, and those with nonhealing wounds and ulcers. It is likely that the future will see applicaton of rHuGM-CSF in a variety of settings beyond those classically associated with myelosuppression.

REFERENCES

1. Burgess AW, Begley CG, Johnson GR, Lopez AF, Williamson DJ, Mermod JJ, Simpson RJ, Schmitz A, DeLamarter JF: Purification and properties of bacterially synthesized human granulocyte-macrophage colony stimulating factor. Blood 69:43, 1987

2. Wong GG, Witek JS, Temple PA, Wilkens KM, Leary AC, Luxenberg DP, Jones SS, Brown EL, Kay RM, Orr EC, Shoemaker C, Golde DW, Kaufman RJ, Hewick RM, Wang EA, Clark SC: Human GM-CSF: Molecular coloning of the complementary DNA and purification of the natural and recombinant proteins. Science 228:819, 1985

3. Cantrell MA, Anderson D, Cerretti DP, Price V, McKereghan K, Tushinski RJ, Mochizuki DY, Larsen A, Grabstein K, Gillis S, Cosman D: Cloning, sequence, and expression of a human granulocyte/ macrophage colony-stimulating factor. Proc Natl Acad Sci USA 82: 6250, 1985

4. Donahue RE, Wang EA, Kaufman RJ, Foutch L, Leary AC, Witek-Giannette JS, Metzger M, Hewick RM, Steinbrink DR, Shaw G, Kamen R, Clark SC: Effects of N-linked carbohydrate on the in vivo properties of human GM-CSF. Cold Spring Harb Symp Quant Biol 51:685, 1986

5. Dorr RT: Clinical properties of yeast-derived versus *Escherichia coli*-derived granulocyte-macrophage colony-stimulating factor. Clin Ther 15:19, 1993

6. Hovgaard D, Mortensen BT, Schifter S, Nissen NI: Comparative pharmacokinetics of single-dose administration of mammalian and bacterially-derived recombinant human granulocyte-macrophage colony-stimulating factor. Eur J Haematol 50:32, 1993

7. Hussein AM, Ross M, Vredenburgh J, Meisenberg B, Hars V, Gilbert C, Petros WP, Coniglio D, Kurtzberg J, Rubin P, Peters WP: Effects of granulocyte-macrophage colony stimulating factor produced in Chinese hamster ovary cells (regramostim) Escherichia coli (molgramostim) and yeast (sargramostim) on priming peripheral blood progenitor cells for use with autologous bone marrow after high-dose chemotherapy. Eur J Haematol 54:281, 1995

8. Nemunaitis J: Granulocyte-macrophage-colony-stimulating factor: A review from preclinical development to clinical application. Transfusion 33:70, 1993

9. Fabian I, Shapira E, Gadish M, Kletter Y, Nagler A, Flidel O, Slavin S: Effects of human interleukin 3, macrophage and granulocytemacrophage colony-stimulating factor on monocyte function following autologous bone marrow transplantation. Leuk Res 16:703, 1992

10. Fleischmann J, Golde DW, Weisbart RH, Gasson JC: Granulocytemacrophage colony-stimulating factor enhances phagocytosis of bacteria by human neutrophils. Blood 68:708, 1986

11. Ho AD, Haas R, Wulf G, Knauf W, Ehrhardt R, Heilig B, Körbling M, Schulz G, Hunstein W: Activation of lymphocytes induced by recombinant human granulocyte-macrophage colony-stimulating factor in patients with malignant lymphoma. Blood 75:203, 1990

12. Jones T, Stern A, Lin R: Potential roles of granulocytemacrophage colony-stimulating factor as vaccine adjuvant. Eur J Clin Microbiol Infect Dis 13:S47, 1994 (suppl 2)

13. Perkins RC, Vadhan-Raj S, Scheule RK, Hamilton R, Holian A: Effects of continuous high dose rhGM-CSF on human monocyte activity. Am J Hematol 43:279, 1993

14. Sallusto F, Lanzavecchia A: Efficient presentation of soluble antigen by cultured human dendritic cells is maintained by granulocyte/macrophage colony-stimulating factor plus interleukin 4 and downregulated by tumor necrosis factor α . J Exp Med 179:1109, 1994

15. Schiller JH, Hank JA, Khorsand M, Storer B, Borchert A, Huseby-Moore K, Burns D, Wesly O, Albertini MR, Wilding G, Sondel PM: Clinical and immunological effects of granulocyte-macrophage colony-stimulating factor coadministered with interleukin 2: A phase IB study. Clin Cancer Res 2:319, 1996

16. Smith PD, Lamerson CL, Banks SM, Saini SS, Wahl LM,

EMERGING APPLICATIONS OF SARGRAMOSTIM

Calderone RA, Wahl SM: Granulocyte-macrophage colony-stimulating factor augments human monocyte fungicidal activity for *Candida albicans*. J Infect Dis 161:999, 1990

17. Wang JM, Colella S, Allavena P, Mantovani A: Chemotactic activity of human recombinant granulocyte-macrophage colonystimulating factor. Immunology 60:439, 1987

18. Weisbart RH, Kwan L, Golde DW, Gasson JC: Human GM-CSF primes neutrophils for enhanced oxidative metabolism in response to the major physiological chemoattractants. Blood 69:18, 1987

19. Wing EJ, Magee M, Whiteside TL, Kaplan SS, Shadduck RK: Recombinant human granulocyte/macrophage colony-stimulating factor enhances monocyte cytotoxicity and secretion of tumor necrosis factor α and interferon in cancer patients. Blood 73:643, 1989

20. Young JW, Szabolcs P, Moore MAS: Identification of dendritic cell colony-forming units among normal human CD34⁺ bone marrow progenitors that are expanded by c-*kit*-ligand and yield pure dendritic cell colonies in the presence of granulocyte/macrophage colony-stimulating factor and tumor necrosis factor α . J Exp Med 182:1111, 1995

21. Colotta F, Bussolino F, Polentarutti N, Guglielmetti A, Sironi M, Bocchietto E, De Rossi M, Mantovani A: Differential expression of the common β and specific α chains of the receptors for GM-CSF, IL-3, and IL-5 in endothelial cells. Exp Cell Res 206:311, 1993

22. DiPersio JF, Hedvat C, Ford CF, Golde DW, Gasson JC: Characterization of the soluble human granulocyte-macrophage colonystimulating factor receptor complex. J Biol Chem 266:279, 1991

23. Jokhi PP, King A, Jubinsky PT, Loke YW: Demonstration of the low affinity α subunit of the granulocyte-macrophage colonystimulating factor receptor (GM-CSF-R α) on human trophoblast and uterine cells. J Reprod Immunol 26:147, 1994

24. Lanza F, Castagnari B, Rigolin G, Moretti S, Latorraca A, Ferrari L, Bardi A, Castoldi G: Flow cytometry measurement of GM-CSF receptors in acute leukemic blasts, and normal hemopoietic cells. Leukemia 11:1700, 1997

25. Park LS, Friend D, Gillis S, Urdal DL: Characterization of the cell surface receptor for human granulocyte/macrophage colony-stimulating factor. Exp Med 164:251, 1986

26. Santiago-Schwarz F, Divaris N, Kay C, Carsons SE: Mechanisms of tumor necrosis factor-granulocyte-macrophage colonystimulating factor-induced dendritic cell development. Blood 82:3019, 1993

27. Till KJ, Burthem J, Lopez A, Cawley JC: Granulocytemacrophage colony-stimulating factor receptor: Stage-specific expression and function on late B cells. Blood 88:479, 1996

28. Hayashida K, Kitamura T, Gorman DM, Arai K, Yokota T, Miyajima A: Molecular cloning of a second subunit of the receptor for human granulocyte-macrophage colony-stimulating factor (GM-CSF): Reconstitution of a high-affinity GM-CSF receptor. Proc Natl Acad Sci USA 87:9655, 1990

29. Miyajima A, Mui AL-F, Ogorochi T, Sakamaki K: Receptors for granulocyte-macrophage colony-stimulating factor, interleukin-3, and interleukin-5. Blood 82:1960, 1993

30. Sato N, Sakamaki K, Terada N, Arai K, Miyajima A: Signal transduction by the high-affinity GM-CSF receptor: Two distinct cytoplasmic regions of the common β subunit responsible for different signaling. EMBO J 12:4181, 1993

31. Quelle FW, Sato N, Witthuhn BA, Inhorn RC, Eder M, Miyajima A, Griffin JD, Ihle JN: JAK2 associates with β_c chain of the receptor for granulocyte-macrophage colony-stimulating factor, and its activation requires the membrane-proximal region. Mol Cell Biol 14:4335, 1994

32. Mui AL-F, Wakao H, O'Farrell A-M, Harada N, Miyajima A: Interleukin-3, granulocyte-macrophage colony stimulating factor and interleukin-5 transduce signals through two STAT5 homologs. EMBO J 14:1166, 1995

33. Wang Y, Morella KK, Ripperger J, Lai C-F, Gearing DP, Fey GH,

Campos SP, Baumann H: Receptors for interluekin-3 (IL-3) and growth hormone mediate an IL-6-type transcriptional induction in the presence of JAK2 or STAT3. Blood 86:1671, 1995

34. Satoh T, Nakafuka M, Miyajima A, Kaziro Y: Involvement of *ras* p21 protein in signal-transduction pathways from interkeukin 2, interleukin 3, and granulocyte/macrophage colony-stimulating factor, but not from interleukin 4. Proc Natl Acad Sci USA 88:3314, 1991

35. Okuda K, Sanghera JS, Pelech SL, Kanakura Y, Hallek M, Griffin JD, Druker BJ: Granulocyte-macrophage colony-stimulating factor, interleukin-3, and steel factor induce rapid tyrosine phosphorylation of p42 and p44 MAP kinase. Blood 79:2880, 1992

36. Leukine (sargramostim) prescribing information. Seattle, WA, Immunex, 1996

37. Cebon JS, Bury RW, Lieschke GJ, Morstyn G: The effects of dose and route of administration on the pharmacokinetics of granulocytemacrophage colony-stimulating factor. Eur J Cancer 26:1064, 1990

38. Herrmann F, Schulz G, Lindemann A, Meyenburg W, Oster W, Krumwieh D, Mertelsmann R: Hematopoietic responses in patients with advanced malignancy treated with recombinant human granulocytemacrophage colony-stimulating factor. J Clin Oncol 7:159, 1989

39. Schwinghammer TL, Shadduck RK, Waheed A, Evans C, Sulecki M, Rosenfeld CS: Pharmacokinetics of recombinant human granulocyte-macrophage colony-stimulating factor (GM-CSF) after intravenous infusion and subcutaneous injection. Pharmacotherapy 2:105, 1991 (abstr 60)

40. Shadduck RK, Waheed A, Evans C, Sulecki M, Rosenfeld CS: Serum and urinary levels of recombinant human granulocytemacrophage colony-stimulating factor: Assessment after intravenous infusion and subcutaneous injection. Exp Hematol 18:601, 1990 (abstr 201)

41. Furman WL, Fairclough DL, Huhn RD, Pratt CB, Stute N, Petros WP, Evans WE, Bowman LC, Douglass EC, Santana VM, Meyer WH, Crist WM: Therapeutic effects and pharmacokinetics of recombinant human granulocyte-macrophage colony-stimulating factor in childhood cancer patients receiving myelosuppressive chemotherapy. J Clin Oncol 9:1022, 1991

42. Stute N, Furman WL, Schell M, Evans WE: Pharmacokinetics of recombinant human granulocyte-macrophage colony-stimulating factor in children after intravenous and subcutaneous administration. J Pharm Sci 84:824, 1995

43. Metcalf D: The molecular biology and functions of the granulocyte-macrophage colony-stimulating factors. Blood 67:257, 1986

44. Kaplan SS, Basford RE, Wing EJ, Shadduck RK: The effect of recombinant human granulocyte macrophage colony-stimulating factor on neutrophil activation in patients with refractory carcinoma. Blood 73:636, 1989

45. Broxmeyer HE, Cooper S, Vadhan-Raj S: Cell cycle status of erythroid (BFU-E) progenitor cells from the bone marrows of patients on a clinical trial with purified recombinant human granulocyte-macrophage colony-stimulating factor. Exp Hematol 17:455, 1989

46. Chachoua A, Oratz R, Hoogmoed R, Caron D, Peace D, Liebes L, Blum RH, Vilcek J: Monocyte activation following systemic administration of granulocyte-macrophage colony-stimulating factor. J Immunother 15:217, 1994

47. Löwenberg B, Suciu S, Zittoun R, Ossenkoppele G, Boogaerts MA, Wijermans P, Vellenga E, Berneman Z, Dekker AW, Sonneveld P, Stryckmans P, Solbu G, Dardenne M, de Witte Th, Archimbaud E: GM-CSF during as well as after induction chemotherapy (CT) in elderly patients with acute myeloid leukemia (AML). The EORTC-HOVON phase III trial (AML 11). Blood 86:433a, 1995 (abstr, suppl 1)

48. Steis RG, VanderMolen LA, Longo DL, Clark JW, Smith JW II, Kopp WC, Ruscetti FW, Creekmore SP, Elwood LJ, Hursey J, Urba WJ: Recombinant human granulocyte-macrophage colony-stimulating factor in patients with advanced malignancy: A phase Ib trial. J Natl Cancer Inst 82:697, 1990 49. Ruef C, Coleman DL: Granulocyte-macrophage colonystimulating factor: pleiotropic cytokine with potential clinical usefulness. Rev Infect Dis 12:41, 1990

50. Birkmann J, Oez S, Smetak M, Kaiser G, Kappauf H, Gallmeier WM: Effects of recombinant human thrombopoietin alone and in combination with erythropoietin and early-acting cytokines on human mobilized purified CD34+ progenitor cells cultured in serum-depleted medium. Stem Cells 15:18, 1997

51. Neelis KJ, Hartong SCC, Egeland T, Thomas GR, Eaton DL, Wagemaker G: The efficacy of single-dose administration of thrombopoietin with coadministration of either granulocyte/macrophage or granulocyte colony-stimulating factor in myelosuppressed rhesus monkeys. Blood 90:2565, 1997.

52. Coleman DL, Chodakewitz JA, Bartiss AH, Mellors JW: Granulocyte-macrophage colony-stimulating factor enhances selective effector functions of tissue-derived macrophages. Blood 72:573, 1988

53. Wiltschke C, Krainer M, Wagner A, Linkesch W, Zielinski CC: Influence of in vivo administration of GM-CSF and G-CSF on monocyte cytotoxicity. Exp Hematol 23:402, 1995

54. Szabolcs P, Avigan D, Gezelter S, Ciocon DH, Moore MAS, Steinman RM, Young JW: Dendritic cells and macrophages can mature independently from a human bone marrow-derived, post-colony-forming unit intermediate. Blood 87:4520, 1996

55. Szabolcs P, Moore MAS, Young JW: Expansion of immunostimulatory dendritic cells among the myeloid progeny of human CD34⁺ bone marrow precursors cultured with c-*kit* ligand, granulocytemacrophage colony-stimulating factor, and TNF- α . J Immunol 154: 5851, 1995

56. Fischer H-G, Frosch S, Reske K, Reske-Kunz AB: Granulocytemacrophage colony-stimulating factor activates macrophages derived from bone marrow cultures to synthesis of MHC class II molecules and to augmented antigen presentation function. J Immunol 141:3882, 1988

57. Ganser A, Heil G: Use of hematopoietic growth factors in the treatment of acute myelogenous leukemia. Curr Opin Hematol 4:191, 1997

58. Geller RB: Use of cytokines in the treatment of acute myelocytic leukemia: A critical review. J Clin Oncol 14:1371, 1996

59. Lieschke GJ, Foote M, Morstyn G: Hematopoietic growth factors in cancer chemotherapy. Cancer Chemother Biol Response Modif 17:363, 1997

60. Lifton R, Bennett JM: Clinical use of granulocyte-macrophage colony-stimulating factor and granulocyte colony-stimulating factor in neutropenia associated with malignancy. Hematol Oncol Clin North Am 10:825, 1996

61. Montemurro F, Gallicchio M, Aglietta M: Prevention and treatment of febrile neutropenia [Italian]. Tumori 83:S15, 1997 (suppl)

62. Lane TA, Law P, Maruyama M, Young D, Burgess J, Mullen M, Mealiffe M, Terstappen LWMM, Hardwick A, Moubayed M, Oldham F, Corringham RET, Ho AD: Harvesting and enrichment of hematopoietic progenitor cells mobilized into the peripheral blood of normal donors by granulocyte-macrophage colony-stimulating factor (GM-CSF) or G-CSF: Potential role in allogeneic marrow transplantation. Blood 85:275, 1995

63. Huang S, Terstappen LWMM: Lymphoid and myeloid differentiation of single human CD34⁺, HLA-DR⁺, CD38⁻ hematopoietic stem cells. Blood 83:1515, 1994

64. Corringham RET, Ho AD: Rapid and sustained allogeneic transplantation using immunoselected CD34⁺-selected peripheral blood progenitor cells mobilized by recombinant granulocyte- and granulocyte-macrophage colony-stimulating factors. Blood 86:2052, 1995

65. Ali SM, Brown RA, Adkins DR, Todd G, Haug JS, Goodnough LT, DiPersio JF: Analysis of lymphocyte subsets and peripheral blood progenitor cells (PBPC) in apheresis products from normal donors mobilized with either G-CSF or concurrent G-CSF and GM-CSF. Blood 90:2511, 1997 (abstr, suppl 1)

66. Law P, Young D, Peterson S, Lane TA, Ho AD: Mobilization and collection of peripheral blood progenitor cells (PBPC) from normal subjects treated sequentially with GM-CSF and G-CSF. Blood 88:397a, 1996 (abstr, suppl 1)

67. Winter JN, Lazarus HM, Rademaker A, Villa M, Mangan C, Tallman M, Jahnke L, Gordon L, Newman S, Byrd K, Cooper BW, Horvath N, Crum E, Stadtmauer EA, Conklin E, Bauman A, Martin J, Goolsby C, Gerson ST, Bender J, O'Gorman M: Phase I/II study of combined granulocyte colony-stimulating factor and granulocytemacrophage colony-stimulating factor administration for the mobilization of hematopoietic progenitor cells. J Clin Oncol 14:277, 1996

68. Griffin JD, Young D, Herrmann F, Wiper D, Wagner K, Sabbath KD: Effects of recombinant human GM-CSF on proliferation of clonogenic cells in acute myeloblastic leukemia. Blood 67:1448, 1986

69. Kelleher C, Miyauchi J, Wong G, Clark S, Minden MD, McCulloch EA: Synergism between recombinant growth factors, GM-CSF and G-CSF, acting on the blast cells of acute myeloblastic leukemia. Blood 69:1498, 1987

70. Miyauchi J, Kelleher CA, Yang YC, Wong GG, Clark SC, Minden MD, Minkin S, McCulloch EA: The effect of three recombinant growth factors, IL-3, GM-CSF, and G-CSF, on the blast cells of acute myeloblastic leukemia maintained in short-term suspension culture. Blood 70:657, 1987

71. Vellenga E, Young DC, Wagner K, Wiper D, Ostapovicz D, Griffin JD: The effect of GM-CSF and G-CSF in promoting growth of clonogenic cells in acute myeloblastic leukemia. Blood 69:1771, 1987

72. Cannistra SA, Groshek P, Griffin JD: Granulocyte-macrophage colony-stimulating factor enhances the cytotoxic effects of cytosine arabinoside in acute myeloblastic leukemia and in the myeloid blast crisis phase of chronic myeloid leukemia. Leukemia 3:328, 1989

73. Lotem J, Sachs L: Hematopoietic cytokines inhibit apoptosis induced by transforming growth factor $\beta 1$ and cancer chemotherapy compounds in myeloid leukemic cells. Blood 80:1750, 1992

74. Büchner T, Hiddemann W, Wörmann B, Zühlsdorf M, Rottmann R, Innig G, Maschmeier G, Ludwig W-D, Sauerland M-C, Heinecke A: Hematopoietic growth factors in acute myeloid leukemia: Supportive and priming effects. Semin Oncol 24:124, 1997

75. Hansen PB, Johnsen HE, Jensen L, Gaarsdal E, Simonsen K, Ralfkiaer E: Priming and treatment with molgramostim (rhGM-CSF) in adult high-risk acute myeloid leukemia during induction chemotherapy: A prospective, randomized pilot study. Eur J Haematol 54:296, 1995

76. Heil G, Chadid L, Hoelzer D, Seipelt G, Mitrou P, Huber Ch, Kolbe K, Mertelsmann R, Lindemann A, Frisch J, Nicolay U, Gaus W, Heimpel H: GM-CSF in a double-blind randomized, placebo controlled trial in therapy of adult patients with *de novo* acute myeloid leukemia (AML). Leukemia 9:3, 1995

77. Zittoun R, Suciu S, Mandelli F, de Witte T, Thaler J, Stryckmans P, Hayat M, Peetermans M, Cadiou M, Solbu G, Petti MC, Willemze R: Granulocyte-macrophage colony-stimulating factor associated with induction treatment of acute myelogenous leukemia: A randomized trial by the European Organization for Research and Treatment of Cancer Leukemia Cooperative Group. J Clin Oncol 14:2150, 1996

78. Ford PA, Arbuck SG, Minniti C, Miller LL, DeMaria D, O'Dwyer PJ: Phase I trial of etoposide, doxorubicin and cisplatin (EAP) in combination with GM-CSF. Eur J Cancer 32:631, 1996

79. Grem JL, McAtee N, Murphy RF, Hamilton JM, Balis F, Steinberg S, Arbuck SG, Setser A, Jordan E, Chen A, Kohler DR, Kotite B, Allegra CJ: Phase I and pharmacokinetic study of recombinant human granulocyte-macrophage colony-stimulating factor given in combination with fluorouracil plus calcium leucovorin in metastatic gastrointestinal adenocarcinoma. J Clin Oncol 12:560, 1994

80. Lachance DH, Oette D, Schold SC Jr, Brown M, Kurtzberg J, Graham ML, Tien R, Felsberg G, Colvin OM, Moghrabi A, Browning I, Hockenberger B, Stewart E, Ferrell L, Kerby T, Duncan-Brown M, Golembe B, Fuchs H, Fredericks R, Hayes FA, Rubin AS, Bigner DD,

EMERGING APPLICATIONS OF SARGRAMOSTIM

Friedman HS: Dose escalation trial of cyclophosphamide with sargramostim in the treatment of central nervous system (CNS) neoplasms. Med Pediatr Oncol 24:241, 1995

81. Moghrabi A, Fuchs H, Brown M, Schold SC Jr, Graham M, Kurtzberg J, Tien R, Felsberg G, Lachance DH, Colvin OM, Oette D, Allegretta GJ, Hockenberger B, Stewart E, Ferrell L, Kerby T, Duncan-Brown M, Bigner DD, Friedman HS: Cyclophosphamide in combination with sargramostim for treatment of recurrent medulloblastoma. Med Pediatr Oncol 25:190, 1995

82. Neidhart JA, Mangalik A, Stidley CA, Tebich SL, Sarmiento LE, Pfile JE, Oette DH, Oldham FB: Dosing regimen of granulocytemacrophage colony-stimulating factor to support dose-intensive chemotherapy. J Clin Oncol 10:1460, 1992

83. Yau JC, Neidhart JA, Triozzi P, Verma S, Nemunaitis J, Quick DP, Mayernik DG, Oette DH, Hayes FA, Holcenberg J: Randomized placebo-controlled trial of granulocyte-macrophage colony-stimulating-factor support for dose-intensive cyclophosphamide, etoposide, and cisplatin. Am J Hematol 51:289, 1996

84. Bober LA, Grace MJ, Pugliese-Sivo C, Rojas-Triana A, Waters T, Sullivan LM, Narula SK: The effect of GM-CSF and G-CSF on human neutrophil function. Immunopharmacology 29:111, 1995

85. Lopez AF, Williamson DJ, Gamble JR, Begley CG, Harlan JM, Klebanoff SJ, Waltersdorph A, Wong G, Clark SC, Vadas MA: Recombinant human granulocyte-macrophage colony-stimulating factor stimulates in vitro mature human neutrophil and eosinophil function, surface receptor expression, and survival. J Clin Invest 78:1220, 1986.

86. DiPersio JF, Billing P, Williams R, Gasson JC: Human granulocyte-macrophage colony-stimulating factor and other cytokines prime human neutrophils for enhanced arachidonic acid release and leukotriene B_4 synthesis. J Immunol 140:4315, 1988

87. Silberstein DS, Owen WF, Gasson JC, DiPersio JF, Golde DW, Bina JC, Soberman R, Austen KF, David JR: Enhancement of human eosinophil cytotoxicity and leukotriene synthesis by biosynthetic (recombinant) granulocyte-macrophage colony-stimulating factor. J Immunol 137:3290, 1986

88. Gadish M, Kletter Y, Flidel O, Nagler A, Slavin S, Fabian I: Effects of recombinant human granulocyte and granulocyte-macrophage colony-stimulating factors on neutrophil function following autologous bone marrow transplantation. Leuk Res 15:1175, 1991

89. Brach MA, deVos S, Gruss H-J, Herrmann F: Prolongation of survival of human polymorphonuclear neutrophils by granulocytemacrophage colony-stimulating factor is caused by inhibition of programmed cell death. Blood 80:2920, 1992

90. Gosselin EJ, Wardwell K, Rigby WFC, Guyre PM: Induction of MHC class II on human polymorphonuclear neutrophils by granulocyte/ macrophage colony-stimulating factor, IFN- γ , and IL-3. J Immunol 151:1482, 1993

91. Mudzinski SP, Christian TP, Guo TL, Cirenza E, Hazlett KR, Gosselin EJ: Expression of HLA-DR (major histocompatability complex class II) on neutrophils from patients treated with granulocytemacrophage colony-stimulating factor for mobilization of stem cells. Blood 86:2452, 1995

92. Phillips N, Jacobs S, Stoller R, Earle M, Przepiorka D, Shadduck RK: Effect of recombinant human granulocyte-macrophage colonystimulating factor on myelopoiesis in patients with refractory metastatic carcinoma. Blood 74:26, 1989

93. Rossman MD, Ruiz P, Comber P, Gomez F, Rottem M, Schreiber AD: Modulation of macrophage Fc_{γ} receptors by rGM-CSF. Exp Hematol 21:177, 1993

94. Williams MA, Kelsey SM, Collins PW, Gutteridge CN, Newland AC: Administration of rHuGM-CSF activates monocyte reactive oxygen species secretion and adhesion molecule expression *in vivo* in patients following high-dose chemotherapy. Br J Haematol 90:31, 1995

95. Roilides E, Blake C, Holmes A, Pizzo PA, Walsh TJ: Granulocytemacrophage colony-stimulating factor and interferon-γ prevent dexamethasone-induced immunosuppression of antifungal monocyte activity against *Aspergillus fumigatus* hyphae. J Med Vet Mycol 34:63, 1996

96. Roilides E, Mertins S, Eddy J, Walsh TJ, Pizzo PA, Rubin M: Impairment of neutrophil chemotactic and bactericidal function in children infected with human immunodeficiency virus type 1 and partial reversal after in vitro exposure to granulocyte-macrophage colonystimulating factor. J Pediatr 117:531, 1990

97. Verhoef G, Boogaerts M: *In vivo* administration of granulocytemacrophage colony stimulating factor enhances neutrophil function in patients with myelodysplastic syndromes. Br J Haematol 79:177, 1991

98. Kowanko IC, Ferrante A, Harvey DP, Carman KL: Granulocytemacrophage colony-stimulating factor augments neutrophil killing of *Torulopsis glabrata* and stimulates neutrophil respiratory burst and degranulation. Clin Exp Immunol 83:225, 1991

99. Richardson MD, Brownlie CED, Shankland GS: Enhanced phagocytosis and intracellular killing of *Candida albicans* by GM-CSF-activated human neutrophils. J Med Vet Mycol 30:433, 1992

100. Roilides E, Holmes A, Blake C, Venzon D, Pizzo PA, Walsh TJ: Antifungal activity of elutriated human monocytes against *Aspergillus fumigatus* hyphae: enhancement by granulocyte-macrophage colonystimulating factor and interferon-γ. J Infect Dis 170:894, 1994

101. Newman SL, Gootee L: Colony-stimulating factors activate human macrophages to inhibit intracellular growth of *Histoplasma capsulatum* yeasts. Infect Immunity 60:4593, 1992

102. Collins HL, Bancroft GJ: Cytokine enhancement of complementdependent phagocytosis by macrophages: Synergy of tumor necrosis factor- α and granulocyte-macrophage colony-stimulating factor for phagocytosis of *Cryptococcus neoformans*. Eur J Immunol 22:1447, 1992b

103. Reed SG, Nathan CF, Pihl DL, Rodricks P, Shanebeck K, Conlon PJ, Grabstein KH: Recombinant granulocyte/macrophage colony-stimulating factor activates macrophages to inhibit *Trypanosoma cruzi* and release hydrogen peroxide. J Exp Med 166:1734, 1987

104. Nemunaitis J, Gordon A, Cox J, Kerr R, Hanson T, Courtney A, Mever W: Phase II pilot trial comparing neutrophil and monocyte function by microbicidal assay in oncology patients receiving rhG-CSF, rhGM-CSF or no cytokine after cytotoxic chemotherapy. Blood 84:135, 1994 (abstr, suppl 1)

105. Bermudez LE, Martinelli J, Petrofsky M, Kolonoski P, Young LS: Recombinant granulocyte-macrophage colony-stimulating factor enhances the effects of antibiotics against *Mycobacterium avium* complex infection in the beige mouse model. J Infect Dis 169:575, 1994

106. Bermudez LEM, Young LS: Recombinant granulocytemacrophage colony-stimulating factor activates human macrophages to inhibit growth or kill *Mycobacterium avium* complex. J Leukoc Biol 48:67, 1990

107. Onyeji CO, Nightingale CH, Tessier PR, Nicolau DP, Bow LM: Activities of clarithromycin, azithromycin, and ofloxacin in combination with liposomal or unencapsulated granulocyte-macrophage colonystimulating factor against intramacrophage *Mycobacterium avium*-*Mycobacterium intracellulare*. J Infect Dis 172:810, 1995

108. Suzuki K, Lee WJ, Hashimoto T, Tanaka E, Murayama T, Amitani R, Yamamoto K, Kuze F: Recombinant granulocytemacrophage colony-stimulating factor (GM-CSF) or tumour necrosis factor-alpha (TNF- α) activate human alveolar macrophages to inhibit growth of *Mycobacterium avium* complex. Clin Exp Immunol 98:169, 1994

109. Frenck RW, Sarman G, Harper TE, Buescher ES: The ability of recombinant murine granulocyte-macrophage colony-stimulating factor to protect neonatal rats from septic death due to *Staphylococcus aureus*. J Infect Dis 162:109, 1990

110. Mayer P, Schütze E, Lam C, Kricek F, Liehl E: Recombinant murine granulocyte-macrophage colony-stimulating factor augments neutrophil recovery and enhances resistance to infections in myelosuppressed mice. J Infect Dis 163:584, 1991

111. Lechner AJ, Lamprech KE, Potthoff LH, Tredway TL, Matuschak GM: Recombinant GM-CSF reduces lung injury and mortality during neutropenic *Candida* sepsis. Am J Physiol 266:L561, 1994

4506

112. Nemunaitis J, Rabinowe SN, Singer JW, Bierman PJ, Vose JM, Freedman AS, Onetto N, Gillis S, Oette D, Gold M, Buckner CD, Hansen JA, Ritz J, Appelbaum FR, Armitage JO, Nadler LM: Recombinant granulocyte-macrophage colony-stimulating factor after autologous bone marrow transplantation for lymphoid cancer. N Engl J Med 324:1773, 1991

113. Rowe JM, Rubin A, Mazza JJ, Bennett JM, Paietta E, Anderson JW, Ghalie R, Wiernick PH: Incidence of infections in adult patients (>55 years) with acute myeloid leukemia treated with yeast-derived GM-CSF (sargramostim): Results of a double-blind prospective study by the Eastern Cooperative Oncology Group, in Hiddemann W, Buchner T, Wormann B, Schellong L, Ritter J, Creutzig U (eds): Acute Leukemias V: Experimental Approaches and Management of Refractory Disease. Berlin, Germany, Springer-Verlag, 1996, p 178

114. Bodey GP, Anaissie E, Gutterman J, Vadhan-Raj S: Role of granulocyte-macrophage colony-stimulating factor as adjuvant therapy of fungal infection in patients with cancer. Clin Infect Dis 17:705, 1993

115. Swindells S, Kleinschmidt DR, Hayes FA. Pilot study of adjunctive Gm-CSF (yeast-derived) for fluconazole-resistant oral candidiasis in HIV-1 infection. Infect Dis Clin Prac 6:278, 1997

116. Palau LA, Pankey GA: Resolution of rhinocerebral and disseminated mucormycosis with adjuvant administration of subcutaneous granulocyte-macrophage colony-stimulating factor (GM-CSF). Proceedings of the 37th Interscience Conference on Antimicrobial Agents and Chemotherapy, Toronto, Ontario, Canada. Washington, DC, American Society for Microbiology, 1997, p 374 (abstr LM-55)

117. Gill PS, Bernstein-Singer M, Espina BM, Rarick M, Magy F, Montgomery T, Berry MS, Levine A: Adriamycin, bleomycin and vincristine chemotherapy with recombinant granulocyte-macrophage colony-stimulating factor in the treatment of AIDS-related Kaposi's sarcoma. AIDS 6:1477, 1992

118. Scadden DT, Bering HA, Levine JD, Bresnahan J, Evans L, Epstein C, Groopman JE: Granulocyte-macrophage colony-stimulating factor mitigates the neutropenia of combined interferon alfa and zidovudine treatment of acquired immune deficiency syndrome-associated Kaposi's sarcoma. J Clin Oncol 9:802, 1991

119. Scadden DT, Pickus O, Hammer SM, Stretcher B, Bresnahan J, Gere J, McGrath J, Agosti JM: Lack of *in vivo* effect of granulocytemacrophage colony-stimulating factor on human immunodeficiency virus type 1. AIDS Res Human Retrovir 12:1151, 1996

120. Folks TM, Justement J, Kinter A, Dinarello CA, Fauci AS: Cytokine-induced expressin of HIV-1 in a chronically infected promonocyte cell line. Science 238:800, 1987

121. Hammer SM, Gillis JM, Pinkson P, Rose RM: Effect of zidovudine and granulocyte-macrophage colony-stimulating factor on human immunodeficiency virus replication in alveolar macrophages. Blood 75:1215, 1990

122. Kitano K, Abboud CN, Ryan DH, Quan SG, Baldwin GC, Golde DW: Macrophage-active colony-stimulating factors enhance human immunodeficiency virus type 1 infection in bone marrow stem cells. Blood 77:1699, 1991

123. Koyanagi Y, O'Brien WA, Zhao JQ, Golde DW, Gasson JC, Chen ISY: Cytokines alter production of HIV-1 from primary mononuclear phagocytes. Science 241:1673, 1988

124. Perno C-F, Cooney DA, Gao W-Y, Hao Z, Johns DG, Foli A, Hartman NR, Caliò R, Broder S, Yarchoan R: Effects of bone marrow stimulatory cytokines on human immunodeficiency virus replication and the antiviral activity of dideoxynucleosides in cultures of monocyte/ macrophages. Blood 80:995, 1992

125. DiMarzio P, Mariani R, Tse J, Thomas EK, Landau NR: GM-CSF or CD40L suppresses chemokine receptor expression and HIV-1 entry in human monocytes and macrophages. Program and

Abstracts of the 5th Conference on Retroviruses and Opportunistic Infections. Chicago, IL. Alexandria, VA, Foundation of Retrovirology and Human Health, 1998, p 86 (abstr 37)

126. Massari F, Poli G, Fauci AS: GM-CSF inhibition of susceptibility of M-CSF treated human monocytes to in vitro infection with HIV. Proceedings of the Fifth International Conference on AIDS. Montreal, Quebec, Canada. Ottawa, Ontario, Canada, International Development Research Center, 1989, p 610 (abstr WCP 109)

127. Matsuda S, Akagawa K, Honda M, Yokota Y, Takebe Y, Takemori T: Suppression of HIV replication in human monocytederived macrophages induced by granulocyte/macrophasge colonystimulating factor. AIDS Res Hum Retroviruses 11:1031, 1995

128. Hammer SM, Gillis JM: Synergistic activity of granulocytemacrophage colony-stimulating factor and 3'-azido-3'-deoxythymidine against human immunodeficiency virus in vitro. Antimicrob Agents Chemother 31:1046, 1987

129. Perno C-F, Yarchoan R, Cooney DA, Hartman NR, Webb DS, Hao Z, Mitsuya H, Johns DG, Broder S: Replication of human immunodeficiency virus in monocytes. Granulocyte/macrophage colonystimulating factor (GM-CSF) potentiates viral production yet enhances the antiviral effect mediated by 3'-azido-2'3'-dideoxythymidine (AZT) and other dideoxynucleoside congeners of thymidine. J Exp Med 169:933, 1989

130. Bakker PJM, Danner SA, ten Napel CHH, Kroon FP, Sprenger HG, van Leusen R, Meenhorst PL, Muusers A, Veenhof CHN: Treatment of poor prognosis epidemic Kaposi's sarcoma with doxorubicin, bleomycin, vindesine and recombinant human granulocytemacrophage colony stimulating factor (rh GM-CSF). Eur J Cancer 31A:188, 1995

131. Hardy WD: Combined ganciclovir and recombinant human granulocyte-macrophage colony-stimulating factor in the treatment of cytomegalovirus retinitis in AIDS patients. J AIDS 4:S22, 1991 (suppl 1)

132. Krown SE, Paredes J, Bundow D, Polsky B, Gold JWM, Flomenberg N: Interferon- α , zidovudine, and granulocyte-macrophage colony-stimulating factor: A phase I AIDS Clinical Trials Group study in patients with Kaposi's sarcoma associated with AIDS. J Clin Oncol 10:1344, 1992

133. Kaplan LD, Kahn JO, Crowe S, Northfelt D, Neville P, Grossberg H, Abrams DI, Tracey J, Mills J, Volberding PA: Clinical and virologic effects of recombinant human granulocyte-macrophage colony-stimulating factor in patients receiving chemotherapy for human immunodeficiency virus-associated non-Hodgkin's lymphoma: Results of a randomized trial. J Clin Oncol 9:929, 1991

134. Pluda JM, Yarchoan R, Smith PD, McAtee N, Shay LE, Oette D, Maha M, Wahl SM, Myers CE, Broder S: Subcutaneous recombinant granulocyte-macrophage colony-stimulating factor used as a single agent and in an alternating regimen with azidothymidine in leukopenic patients with severe human immunodeficiency virus infection. Blood 76:463, 1990

135. Davison FD, Kaczmarski RS, Pozniak A, Mufti GJ, Sutherland S: Quantification of HIV by PCR in monocytes and lymphocytes in patients receiving antiviral treatment and low dose recombinant human granulocyte macrophage colony stimulating factor. J Clin Pathol 47:855, 1994

136. Bernstein AP, Brooks S, Hayes FA, Gould M, Jacob S, Tomasi TB: A pilot study in the use of GM-CSF in human immunodeficiency virus (HIV) infected individuals. Blood 90:133a, 1997 (abstr, suppl 1)

137. Skowron G, Stein D, Drusano G, Melbourne K, Mongillo A, Whitmore J, Echols R, Gilbert M: Safety and anti-HIV effect of GM-CSF in patients on highly active anti-retroviral therapy. Program and Abstracts of the 5th Conference on Retroviruses and Opportunistic Infections. Chicago, IL. Alexandria, VA, Foundation of Retrovirology and Human Health, 1998, p 267 (abstr 615)

138. DiMarzio P, Tse J, Landau NR: Chemokine receptor regulation

EMERGING APPLICATIONS OF SARGRAMOSTIM

and HIV type 1 tropism in monocyte-macrophages. AIDS Res Hum Retrovirus 14:129, 1998

139. Kemper C, Bermudez L, Agosti J, Deresinski S: Immunomodulatory therapy of *Mycobacterium avium* (MAC) bacteremia in AIDS with rhGM-CSF. Thirty-fifth Annual Meeting of the Interscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, CA. Washington, DC, American Society of Microbiology, 1995, p 177 (abstr G109)

140. Caux C, Dezutter-Dambuyant C, Schmitt D, Banchereau J: GM-CSF and TNF- α cooperate in the generation of dendritic Langerhans cells. Nature 360:258, 1992

141. Tarr PE, Lin R, Mueller EA, Kovarik JM, Guillaume M, Jones TC: Evaluation of tolerability and antibody response after recombinant human granulocyte-macrophage colony-stimulating factor (rhGM-CSF) and a single dose of recombinant hepatitis B vaccine. Vaccine 14:1199, 1996

142. Morrissey PJ, Bressler L, Park LS, Alpert A, Gillis S: Granulocyte-macrophage colony-stimulating factor augments the primary antibody response by enhancing the function of antigen-presenting cells. J Immunol 139:1113, 1987

143. Al-Aoukaty A, Giaid A, Sinoff C, Ho AD, Maghazachi AA: Priming effects of granulocyte-macrophage colony-stimulating factor are coupled to cholera toxin-sensitive guanine nucleotide binding protein in human T lymphocytes. Blood 83:1299, 1994

144. Stewart-Akers AM, Cairns JS, Tweardy DJ, McCarthy SA: Effect of granulocyte-macrophage colony-stimulating factor on lymphokine-activated killer cell induction. Blood 81:2671, 1993

145. Disis ML, Bernhard H, Shiota FM, Hand SL, Gralow JR, Husaby ES, Gillis S, Cheever MA: Granulocyte-macrophage colonystimulating factor: An effective adjuvant for protein and peptide-based vaccines. Blood 88:202, 1996

146. Hess G, Kreiter F, Kösters W, Deusch K: The effect of granulocyte-macrophage colony-stimulating factor (GM-CSF) on hepatitis B vaccination in haemodialysis patients. J Viral Hepatitis 3:149, 1996

147. Taglietti M, Rouzier-Panis R, Aymard M, Garaud JJ: A doubleblind, placebo-controlled study to assess the immune response to flu vaccine following a single dose of rhGM-CSF in elderly people. Abstracts of the 34th Interscience Conference on Antimicrobial Agents and Chemotherapy. Orlando, FL. Washington, DC, American Society for Microbiology, 1994, p 266 (abstr H76)

148. Masucci G, Wersäll P, Ragnhammar P, Mellstedt H: Granulocytemonocyte-colony-stimulating factor augments the cytotoxic capacity of lymphocytes and monocytes in antibody dependent cellular cytotoxicity. Cancer Immunol Immunother 29:288, 1989

149. Grabstein KH, Urdal DL, Tushinski RJ, Mochizuki DY, Price VL, Cantrell MA, Gillis S, Conlon PJ: Induction of macrophage tumoricidal activity by granuloctye-macrophage colony-stimulating factors. Science 232:506, 1986

150. Ragnhammar P, Frödin J-E, Trotta PP, Mellstedt H: Cytotoxicity of white blood cells activated by granulocyte-colony-stimulating factor, granulocyte/macrophage-colony-stimulating factor and macrophage-colony-stimulating factor against tumor cells in the presence of various monoclonal antibodies. Cancer Immunol Immunother 39:254, 1994

151. Baxevanis CN, Dedoussis GVZ, Papadopoulos NG, Missitzis I, Beroukas C, Stathopoulos GP, Papamichail M: Enhanced human lymphokine-activated killer cell function after brief exposure to granulocyte-macrophage-colony stimulating factor. Cancer 76:1253, 1995

152. Epling-Burnette PK, Wei S, Blanchard DK, Spranzi E, Djeu JY: Coinduction of granulocyte-macrophage colony-stimulating factor release and lymphokine-activated killer cell susceptibility in monocytes by interleukin-2 via interleukin-2 receptor β . Blood 81:3130, 1993

153. Dong Z, Kumar R, Yang X, Fidler IJ: Macrophage-derived

metalloelastase is responsible for the generation of angiostatin in Lewis lung carcinoma. Cell 88:801, 1997

154. Dranoff G, Jaffee E, Lazenby A, Golumbek P, Levistsky H, Brose K, Jackson V, Hamada H, Pardoll D, Mulligan RC: Vaccination with irradiated tumor cells engineered to secrete murine granulocytemacrophage colony-stimulating factor stimulates potent, specific, and long-lasting anti-tumor immunity. Proc Natl Acad Sci USA 90:3539, 1993

155. Spitler LE, Grossbard ML, Ernstoff MS, Silver G, Jacobs M, Hayes FA, Soong SJ: Adjuvant therapy of stage III and IV malignant melanoma using yeast derived, GM-CSF. Melanoma Res 7:160, 1997

156. Chachoua A, Oratz R, Liebes L, Alter RS, Felice A, Peace D, Vilcek J, Blum RH: Phase Ib trial of granulocyte-macrophage colonystimulating factor combined with murine monoclonal antibody R24 in patients with metastatic melanoma. J Immunother 16:132, 1994

157. Yu AL, Batova A, Alvarado C, Rao VJ, Castleberry RP: Usefulness of a chimeric anti-GD2 (ch14.18) and GM-CSF for refractory neuroblastoma: A POG phase II study. Proc Am Soc Clin Oncol 16:513a, 1997 (abstr)

158. Leong SPL, Enders-Zohr P, Zhou YM, Allen RE Jr, Sagebiel RW, Glassberg AB, Hayes FA: Active specific immunotherapy with GM-CSF as an adjuvant to autologous melanoma (AM) vaccine in metastatic melanoma. Proc Am Soc Clin Oncol 15:437, 1996 (abstr 1360)

159. Massaia M, Battaglio S, Beggiato E, Bianchi A, Borrione P, Mariani S, Napoli P, Peola S, Boccadoro M, Pileri A: Vaccination with Id/KLH and local cytokines (IL2/GM-CSF) in advanced multiple myeloma patients. Proceedings of the Keystone Symposia. Silver-thorne, CO, Keystone Symposia, 1997, p 59 (abstr 517)

160. Richard C, Baro J, Bello-Fernandez C, Hermida G, Calavia J, Olalla I, Alsar MJ, Loyola I, Cuadrado MA, Iriondo A, Conde E, Zubizarreta A: Recombinant human granulocyte-macrophage colony stimulating factor (rhGM-CSF) administration after autologous bone marrow transplantation for acute myeloblastic leukemia enhances activated killer cell function and may diminish leukemic relapse. Bone Marrow Transplant 15:721, 1995

161. Bendall LJ, Kortlepel K, Gottlieb DJ: GM-CSF enhances IL-2-activated natural killer cell lysis of clonogenic AML cells by upregulating target cell expression of ICAM-1. Leukemia 9:677, 1995

162. Bussolino F, Wang JM, Defilippi P, Turrini F, Sanavio F, Edgell C-JS, Aglietta M, Arese P, Mantovani A: Granulocyte- and granulocytemacrophage-colony stimulating factors induce human endothelial cells to migrate and proliferate. Nature 337:471, 1989

163. Hancock GE, Kaplan G, Cohn ZA: Keratinocyte growth regulation by the products of immune cells. J Exp Med 168:1395, 1988

164. Vadhan-Raj S, Broxmeyer HE, Hittelman WN, Papadopoulos NE, Chawla SP, Fenoglio C, Cooper S, Buescher ES, Frenck RW Jr, Holian A, Perkins RC, Scheule RK, Gutterman JU, Salem P, Benjamin RS: Abrogating chemotherapy-induced myelosuppression by recombinant granulocyte-macrophage colony-stimulating factor in patients with sarcoma: Protection at the progenitor cell level. J Clin Oncol 10:1266, 1992

165. Nemunaitis J, Rosenfeld CS, Ash R, Freedman MH, Deeg HJ, Appelbaum F, Singer JW, Flomenberg N, Dalton W, Elfenbein GJ, Rifkin R, Rubin A, Agosti J, Hayes FA, Holcenberg J, Shadduck RK: Phase III randomized, double-blind placebo-controlled trial of rhGM-CSF following allogeneic bone marrow transplantation. Bone Marrow Transplant 15:949, 1995

166. Gordon B, Spadinger A, Hodges E, Ruby E, Stanley R, Coccia P: Effect of granulocyte-macrophage colony-stimulating factor on oral mucositis after hematopoietic stem-cell transplantation. J Clin Oncol 12:1917, 1994

167. Dunphy F, Kim H, Dunleavy T, Harrison B, Boyd J, Petruska P: Granulocyte monocyte colony stimulating factor (GM-CSF) ameliorates radiation mucositis. Blood 90:184b, 1997 (abstr 3552, suppl 1) 168. Meropol NJ, Petrelli NJ, Rustum YM, Rodriguez-Bigas M, Proefrock A, Frank C, Creaven PJ: Granulocyte-macrophage colonystimulating factor (GM-CSF) as a diarrhea protectant in patients treated with 5-fluorouracil (FU) and leucovorin (LV). Proc Am Soc Clin Oncol 14:263, 1995 (abstr 724)

169. Cartee L, Petros WP, Rosner Gl, Gilbert C, Moore S, Affronti ML, Hoke JA, Hussein AM, Ross M, Rubin P, Vredenburgh JJ, Peters WP: Evaluation of GM-CSF mouthwash for prevention of chemotherapyinduced mucositis: A randomized, double-blind, dose-ranging study. Cytokine 7:471, 1995

170. Ovilla-Martinez R, Rubio ME, Borbolla JR, Gonzale-Llaven JE: GM-CSF mouthwashes as treatment for mucositis in BMT patients. Blood 84:717a, 1994 (abstr 2853, suppl 1)

171. Jyung RW, Wu L, Pierce GF, MustoeTA: Granulocytemacrophage colony-stimulating factor and granulocyte colonystimulating factor: Differential action on incisional wound healing. Surgery 115:325, 1994

172. Kucukcelebi A, Carp SS, Hayward PG, Hui P-S, Cowan WT, Ko F, Cooper DM, Robson MC: Granulocyte-macrophage colony stimulating factor reverses the inhibition of wound contraction caused by bacterial contamination. Wounds 4:241, 1992

173. Vyalov S, Desmoulière A, Gabbiani G: GM-CSF-induced granulation tissue formation: Relationships between macrophage and myofibroblast accumulation. Arch B Cell Pathol 63:231, 1993

174. Braunstein S, Kaplan G, Gottlieb AB, Schwartz M, Walsh G, Abalos RM, Fajardo TT, Guido LS, Krueger JG: GM-CSF activates regenerative epidermal growth and stimulates keratinocyte proliferation in human skin *in vivo*. J Invest Dermatol 103:601, 1994

175. Kaplan G, Walsh G, Guido LS, Meyn P, Burkhardt RA, Abalos RM, Barker J, Frindt PA, Fajardo TT, Celona R, Cohn ZA: Novel responses of human skin to intradermal recombinant granuloctye/ macrophage-colony-stimulating factor: Langerhans cell recruitment, keratinocyte growth, and enhanced wound healing. J Exp Med 175: 1717, 1992

176. Marques da Costa R, Aniceto C, Jesus FM, Mendes M: Quick healing of leg ulcers after molgramostim. Lancet 344:481, 1994

177. Pojda Z, Struzyna J: Treatment of non-healing ulcers with rhGM-CSF and skin grafts. Lancet 343:1100, 1994

178. Raderer M, Kornek G, Hejna M, Koperna K, Scheithauer W, Base W: Topical granulocyte-macrophage colony-stimulating factor in patients with cancer and impaired wound healing. J Natl Cancer Instit 89:263, 1997

179. Arnold F, O'Brien J, Cherry G: Granulocyte monocyte-colony stimulating factor as an agent for wound healing. J Wound Care 4:400, 1995

180. Marques da Costa R, Jesus FM, Aniceto C, Mendes M: Double-blind randomized placebo-controlled trial of the use of granulocyte-macrophage colony-stimulating factor in chronic leg ulcers. Am J Surg 173:165, 1997

181. Nimer SD, Champlin RE, Golde DW: Serum cholesterollowering activity of granulocyte-macrophage colony-stimulating factor. JAMA 260:3297, 1988

182. Ishibashi T, Yokoyama K, Shindo J, Hamazaki Y, Endo Y, Sato T, Takahashi S, Kawarabayasi Y, Shiomi M, Yamamoto T, Maruyama Y: Potent cholesterol-lowering effect by human granulocyte-macrophage colony-stimulating factor in rabbits. Possible implications of enhancement of macrophage functions and an increase in mRNA for VLDL receptor. Arterioscler Thromb 14:1534, 1994

183. Hallman M, Merritt TA: Lack of GM-CSF as a cause of pulmonary alveolar proteinosis. J Clin Invest 97:589, 1996 (editorial)

184. Dranoff G, Crawford AD, Sadelain M, Ream B, Rashid A, Bronson RT, Dickersin GR, Bachurski CJ, Mark EL, Whitsett JA, Mulligan RC: Involvement of granulocyte-macrophage colony-stimulating factor in pulmonary homeostasis. Science 264:713, 1994

185. Huffman JA, Hull WM, Dranoff G, Mulligan RC, Whitsett JA: Pulmonary epithelial cell expression of GM-CSF corrects the alveolar proteinosis in GM-CSF-deficient mice. J Clin Invest 97:649, 1996

186. Ikegami M, Ueda T, Hull W, Whitsett JA, Mulligan RC, Dranoff G, Jobe AH: Surfactant metabolism in transgenic mice after granulocyte macrophage-colony stimulating factor ablation. Am J Physiol 270: L650, 1996

187. Seymour JF, Dunn AR, Vincent JM, Presneill JJ, Pain MC: Efficacy of granulocyte-macrophage colony-stimulating factor in acquired alveolar proteinosis. N Engl J Med 335:1924, 1996 (letter)