Nrf2, a multi-organ protector?

Jong-Min Lee,^{*,‡} Jiang Li,[†] Delinda A. Johnson,[†] Thor D. Stein,[§] Andrew D. Kraft,[†] Marcus J. Calkins,[‡] Rebekah J. Jakel,[§] and Jeffrey A. Johnson^{†,‡,§,II,††,1}

*Molecular Neurogenetics Unit, Massachusetts General Hospital, Harvard Medical School, Charlestown, Massachusetts, USA; and [†]School of Pharmacy, [‡]Molecular and Environmental Toxicology Center, [§]Neuroscience Training Program, ^{II}Waisman Center, and ^{††}Center for Neuroscience, University of Wisconsin, Madison, Wisconsin, USA

ABSTRACT NF-E2-related factor 2 (Nrf2) is a basic leucine zipper transcription factor that binds to the promoter sequence "antioxidant responsive element (ARE)" leading to coordinated up-regulation of AREdriven detoxification and antioxidant genes. Since the expression of a wide array of antioxidant and detoxification genes are positively regulated by the ARE sequence, Nrf2 may serve as a master regulator of the ARE-driven cellular defense system against oxidative stress. In support of this, numerous studies have shown that Nrf2 protects many cell types and organ systems from a broad spectrum of toxic insults and disease pathogenesis. This Nrf2-conferred, multi-organ protection phenomenon raises an interesting question about how a single protein can protect many different organs from various toxic insults. A possible molecular mechanism explaining this phenomenon is that Nrf2 protects many different cell types by coordinately up-regulating classic ARE-driven genes as well as cell type-specific target genes that are required for the defense system of each cell type in its unique environment. This hypothesis is supported by microarray data indicating the protective role of Nrf2 is conveyed through both known ARE-driven genes and novel cell type-specific genes. The widespread nature of Nrf2 may have an important therapeutic potential, allowing prevention of carcinogenesis and neurodegenerative diseases.-Lee, J.-M., Li, J., Johnson, D. A., Stein, T. D., Kraft, A. D., Calkins, M. J., Jakel, R. J., Johnson, J. A. Nrf2, a multi-organ protector? FASEB J. 19, 1061-1066 (2005)

Key Words: Nrf2 · ARE · multi-organ protection

THE HYPOTHESIS

MANY STUDIES have reported that a basic leucine zipper transcription factor, NF-E2-related factor 2 (Nrf2), plays a critical role in protecting a variety of tissues (lung, liver, kidney, stomach, small intestine, central nervous system, splenocytes, macrophages, erythrocytes, and retinal epithelia) from a wide array of toxic insults (carcinogens, electrophiles, reactive oxygen species, diesel exhaust, inflammation, calcium disturbance, UV light, and cigarette smoke). To explain this "single protein-conferred multi-organ protection phenomenon," we suggest a hypothesis that Nrf2 protects various cell types by coordinately up-regulating not only classic ARE-driven genes but also cell type-specific protective genes essential for the basic defense system of each cell type.

ARE AND Nrf2

Reactive oxygen species (ROS) and electrophiles cause cellular damage leading to many diseases including cancer, autoimmune disease, and neurodegenerative disease; such toxic insults are normally detoxified by phase II detoxification enzymes and antioxidant proteins. Therefore, the *cis*-acting regulatory element "antioxidant responsive element (ARE)" that transcriptionally regulates genes encoding detoxification enzymes and antioxidant proteins plays an important role in cellular defense system. Enhancer sequences originally were described in the rat glutathione S-transferase (GST) -P (1), rat GST Ya (2), mouse GST Ya (3), rat NAD(P)H:quinone oxidoreductase-1 (NQO1) (4), and human NQO1 genes (5). Collectively, these studies have identified a common pathway for ARE-driven phase II detoxification gene induction; the underlying ARE activation mechanism was further clarified by investigation of Nrf2. Initially, Venugopal and Jaiswal (6) and Itoh et al (7) implicated Nrf2 function in the ARE-driven gene expression mechanism; subsequent studies have revealed the molecular mechanism by which Nrf2 is activated and participates in ARE-driven gene expression. Briefly, Nrf2 is sequestered in the cytoplasm by Keap1, and ARE activation signals (i.e., protein kinase pathways and electrophiles) disrupt the Nrf2-Keap1 complex leading to nuclear translocation of Nrf2. Upon activation, Nrf2 binds to ARE sites in the promoter regions of many detoxification and antioxidant genes, leading to the coordinate up-regulation of downstream targets that boost cellular detoxification processes and antioxidant potential (**Fig. 1**) (8–16).

¹ Correspondence: School of Pharmacy, University of Wisconsin, 6125 Rennebohm Hall, 777 Highland Ave., Madison, WI 53705, USA. E-mail: jajohnson@pharmacy.wisc.edu doi: 10.1096/fj.04-2591hyp

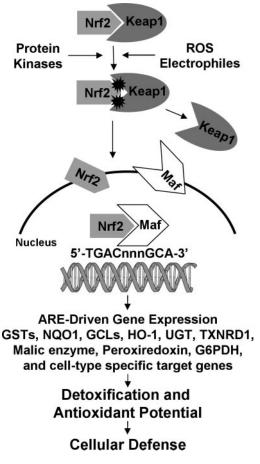


Figure 1. Cellular protection mechanism conferred by Nrf2-ARE pathway.

MULTI-ORGAN PROTECTION BY THE Nrf2 PATHWAY

The function of Nrf2 and its downstream target genes suggests that Nrf2 plays a central role in protecting cells from ROS and electrophiles. In agreement with this idea, numerous studies have shown that Nrf2 protects multiple organs from various toxic insults.

Lung and liver

Nrf2 protects lung from butylated hydroxytolueneinduced acute respiratory distress syndrome (17), hyperoxic injury (18), and bleomycin-mediated pulmonary fibrosis (19) by increasing detoxification pathways and antioxidant potentials. Recent study further demonstrated that genetic ablation of Nrf2 enhanced sensitivity to cigarette smoke-induced emphysema and identified Nrf2-dependent antioxidant and cytoprotective genes using microarray technology (20). Nrf2 plays a role in protecting liver, as evidenced by increased sensitivity to acetaminophen-induced centrilobular hepatocellular necrosis and hepatotoxicity (21, 22) as well as increased levels of lipid peroxidation and DNA damage in Nrf2^{-/-} livers (23).

GI tract

Some studies have shown that Nrf2 protects GI tract from carcinogenesis, implying a role for Nrf2 in cell cycle regulation and cancer prevention. For example, $Nrf2^{-/-}$ mice showed an increased burden of gastric neoplasia after benzo[a]pyrene treatment compared with Nrf2^{+/+} mice; oltipraz significantly decreased gastric tumors only in Nrf2^{+/+} mice, implying that the chemopreventive activity of oltipraz is dependent on the Nrf2-ARE pathway (24). Further support for this anti-carcinogenic effect is evidenced by an increased DNA adduct formation in forestomach after benzo-[a] pyrene administration in $Nrf2^{-/-}$ mice (25). Finally, an ARE-inducer, sulforaphane, is a potent bacteriostatic agent against Helicobacter pylori and blocks benzo[a]pyrene-evoked forestomach tumors (26). This chemopreventive effect of sulforaphane is not observed in $Nrf2^{-/-}$ mice, suggesting Nrf2 dependency. Thus, constitutive and inducible expression of phase II enzymes through the Nrf2-ARE pathway can modify the susceptibility of GI tract to carcinogenesis.

Nervous system

The Nrf2-ARE pathway appears to contribute to neuroprotection. First, activation of the Nrf2-ARE pathway by tert-butylhydroquinone (tBHQ) protected neuroblastoma cells from oxidative glutamate toxicity (27) and H₂O₂-induced apoptosis (28). Primary astrocytes and neurons derived from Nrf2^{-/-} mice were more sensitive to oxidative damage, calcium disturbance, and mitochondrial toxins than were wild-type cells (29, 30). Overexpression of Nrf2 and small molecule-mediated activation of the Nrf2-ARE pathway in astrocytes increased resistance of neurons to non-excitotoxic glutamate toxicity (31). Similarly, dominant negative-Nrf2 stable cells were more sensitive to NO-induced apoptosis, and siRNA-mediated knockdown of Nrf2-sensitized neuroblastoma cells to NO-induced apoptosis (32). $Nrf2^{-/-}$ mice are more susceptible to lesions produced by the mitochondrial complex II inhibitors, and transplantation of Nrf2-overexpressing astrocytes into the striatum protects from malonate-induced lesions (33). These data reinforce the pivotal role of the Nrf2-ARE pathway in protecting nervous system.

Others

The protective effects of Nrf2 have proved indispensable in many other cell types as well. Phase II gene induction by sulforaphane protected retinal pigment epithelial cells from photooxidative damage (34), and Nrf2 was important in protecting macrophages and epithelial cells from diesel exhaust chemicals (35). In addition, Nrf2 has been identified as a putative Lith 1 gallstone gene contributing to gallstone resistance (36) and plays a role in wound healing/keratinocyte differentiation (37, 38). Finally, Nrf2^{-/-} mice develop lupuslike autoimmune nephritis (39), systemic autoimmune disease (23), hemolytic anemia (40), and splenomegaly (23, 40).

Taken together, the studies outlined indicate that Nrf2 protects many different types of organs and cells, and begs an interesting question as to how a single protein can protect multiple organs from an array of different toxic insults.

PROTECTION MECHANISMS

Our hypothesis for Nrf2-conferred multi-organ protection phenomenon is that Nrf2 protects multiple tissues by coordinately up-regulating classic ARE-driven detoxification and antioxidant genes as well as cell typespecific targets that are required for basic defense in each unique environment. Although the observed protective effects are primarily mediated by classic AREdriven genes, Nrf2-mediated, cell type-specific pathways (other than classic detoxification and antioxidant genes) may substantially contribute to protection of each cell type. This hypothesis is supported by a series of gene expression profiling experiments using oligonucleotide microarrays for various tissues/cell types (**Table 1**).

First, Thimmullappa et al. showed that many xenobiotic metabolizing enzymes, detoxification, and antioxidant proteins were increased by Nrf2 in the small intestine and suggested a pivotal role for Nrf2 in modulating cellular defense against carcinogens and toxins (41). Small intestine-selective Nrf2 target genes identified by microarray experiments (epoxide hydrolase, aflatoxin aldehyde reductase, UGT, GSTs, and multidrug resistance protein) indicate that Nrf2 protects the small intestine by coordinately regulating drug metabolism enzymes in the GI tract, where xenobiotics are absorbed primarily and processed. Considering the basic function of the small intestine, these small intestine-selective genes play a role in maintaining cellular integrity of GI tract against xenobiotics.

Kwak et al. identified Nrf2-dependent, D3T (3H-1,2dithiol-3-thione) -inducible genes in liver (42). D3T induced many chaperones and ubiquitin-proteasome pathway genes in a Keap1-Nrf2-dependent manner. Based on liver-selective Nrf2 target genes, the authors expanded the role of the Nrf2-ARE pathway beyond its primary functions into secondary protective effects such as recognition and repair/removal of damaged proteins (42). This finding indicates that Nrf2 plays an important role in protecting hepatocytes and maintaining hepatocyte function by up-regulating specific target genes involved in recognition, detoxification, repair, and removal of damaged proteins. Since these processes might constantly occur during the xenobiotic metabolism in liver, the Nrf2-ARE pathway is important in supporting specialized hepatocyte functions.

DNA chip experiments have revealed that Nrf2 basally up-regulates many ARE-driven genes in primary astrocytes and neurons. In addition to classical AREdriven genes (Table 1), these studies revealed that Nrf2

TABLE 1. Identification of classic Nrf2 target genes by microarray analysis^a

Gene Species	Fold change							
	Mouse	Mouse	Mouse	Rat	Rat	Mouse	Mouse	Mouse
Cell type/gender	Small intestine	Primary astrocyte	Primary neuronal	Primary glial	Primary neuronal	Liver/ male	Liver/ female	Spleen/ female
Reference	(41)	(30)	(29)	(43)	(43)	(23)	(23)	(23)
Nrf2	_	↑ 35.6	$\uparrow 22.7$	_	_	$\uparrow 40.6$	$\uparrow 18.8$	$\uparrow 21.0$
NQO1	$\uparrow 3.7$	$\uparrow 1.9$	$\uparrow 4.9$	$\uparrow 4.3$	$\uparrow 5.4$	^ 4.3	↑ 7.5	_
GST A4 (Ya)	· _	$\uparrow 1.5$	$\uparrow 4.7$	$\uparrow 21.7$	$\uparrow 4.7$	$\uparrow 2.0$	$\uparrow 1.4$	_
GST A2 (Yc2)	_	_	·	$\uparrow 17.6$	∱ 8.8	<u>↑</u> 2.6	_	_
GST P1	_	$\uparrow 1.3$	-	_	_	_	_	_
GST P2	_	-	-	$\uparrow 3.4$	$\uparrow 2.4$	-	-	_
GST mu 1 (8.7)	$\uparrow 2.7$	$\uparrow 1.9$	$\uparrow 2.6$	-	-	$\uparrow 2.7$	$\uparrow 1.7$	_
GST mu 2	$\uparrow 1.9$	$\uparrow 1.3$	-	-	-	$\uparrow 2.2$	_	_
GST mu 3 (9.3)	$\uparrow 6.1$	$\uparrow 1.6$	$\uparrow 1.7$	-	-	$\uparrow 3.6$	$\uparrow 2.4$	$\uparrow 1.2$
GCLM	_	$\uparrow 1.3$	-	$\uparrow 13.0$	$\uparrow 5.1$	-	_	_
GCLC	-	$\uparrow 1.9$	$\uparrow 1.3$	-	-	$\uparrow 2.0$	_	_
HO-1	_	$\uparrow 1.5$	-	$\uparrow 2.8$	$\uparrow 21.3$	-	-	_
Malic enzyme	$\uparrow 3.4$	$\uparrow 1.5$	$\uparrow 1.6$	$\uparrow 8.3$	$\uparrow 10.1$	$\uparrow 2.1$	$\uparrow 2.1$	-
TXNRD1	_	$\uparrow 1.7$	-	$\uparrow 2.6$	$\uparrow 3.4$	$\uparrow 1.4$	_	-
G6PDH	$\uparrow 1.7$	$\uparrow 1.3$	_	_	_	_	$\uparrow 2.0$	_

^{*a*} Primarily, RNA from Nrf2^{-/-} and wild-type control mice (small intestine, primary astrocytes, primary neuronal, liver, and spleen) was used for gene expression profiling. Rat glial and neuronal cells were used for overexpression of Nrf2 and identification of Nrf2 target genes. Many classic Nrf2 target genes were identified by gene expression profiling experiments using oligonucleotide microarrays. Fold changes of Nrf2 target genes were obtained from the comparisons of *1*) Nrf2^{+/+} cells vs. Nrf2^{-/-} cells (23, 29, 30, 41) and 2) Nrf2-overexpressed cell vs. vector control (43). GCLM, glutamate-cysteine ligase modulatory subunit; GCLC, glutamate-cysteine ligase catalytic subunit; HO-1, heme oxygenase-1; TXNRD1, thioredoxin reductase 1; and G6PD, glucose-6-phosphate dehydrogenase. –, Either not available or no change.

TABLE 2. Examples of nervous system-selective Nrf2-dependent genes^a

Cell type	Category	Gene		
Primary Astrocytes	Reducing potential	Glucose-6-phosphate dehydrogenase Transaldolase		
	Immune/inflammation	Transketolase PAF acetylhydrolase Prostaglandin-endoperoxide synthase 2 Dithiolethione-inducible gene		
Primary Neuronal	Calcium homeostasis	Tachykinin 2 Calbindin-28K Synaptotagmin-1 Hippocalcin		
	Growth factor	S100 calcium binding protein A1 Nerve growth factor-γ Fibroblast growth factor-13		
	Signaling	Fibroblast growth factor-14 Brain-derived neurotrophic factor Neuronal GEF Protein kinase C-β		
	Receptor/channel	G-protein-γ3 Adrenomedullin Corticotropin-releasing hormone Chloride channel GABA-A receptor-1 GABA-A receptor, gamma 3		
		GABA-B receptor-1		

^{*a*} In addition to known classic Nrf2-target genes, oligonucleotide microarray experiments further identified novel cell type-specific Nrf2-dependent genes from many cell types. Some examples of novel Nrf2-dependent genes identified from primary astrocytes and neurons by microarray analysis (29, 30, 31, 43) are listed.

regulates genes involved in the reducing potential and immune/inflammation in astrocytes as well as calcium homeostasis, growth factors, signaling molecules, and receptors/channels in neuronal cultures (**Table 2**) (29, 30). Nrf2-regulated, astrocyte-specific genes may explain the observed antioxidant/reducing potential and anti-inflammatory effects of Nrf2 (18, 37), insinuating important functional roles for Nrf2-activated astrocytes in supporting neurons. Furthermore, neuronal functions (i.e., signaling and receptors) and defense activities of neurons (calcium buffering capacity) are greatly enhanced in the presence of Nrf2, suggesting toxic by-products occurring during normal neuronal activities is mitigated by Nrf2-ARE pathway.

Using a cell sorting technology, Kraft et al. isolated an astrocyte-specific (detoxification and antioxidant) and neuron-specific gene cluster (cell adhesion, synaptic transmission, calcium mobilization) (31), supporting the notion that Nrf2 regulates target gene expression depending on a cell's function and the toxic by-products generated in the cell's microenvironment during routine activities.

Finally, Li et al. showed that $Nrf2^{-/-}$ mice develop an autoimmune disease with multiple organ pathology that closely resembles human systemic lupus erythmatosus (SLE). These data indicate that the lack of Nrf2 can induce pathologies in multiple organs, and SLE might be evoked by oxidative tissue damage. In support of this, the authors identified Nrf2 target genes in liver and spleen. While Nrf2 regulates many classical AREdriven genes in liver, it also regulates many cytokines as

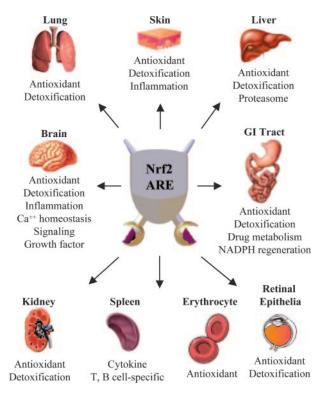


Figure 2. A multi-organ protector, Nrf2. Nrf2 protects various cell types (lung, liver, kidney, stomach, small intestine, central nervous system, splenocytes, macrophages, erythrocytes, and retinal epithelia) by coordinately increasing classic ARE-driven detoxification and antioxidant genes. Nrf2 contributes to cellular protection by enhancing cellular resistance to potential harmful insults that occur during cells' normal activities.

well as B cell and T cell-specific genes, which are important for immune and phagocytosis function of splenocytes (23).

Overall, these observations show that Nrf2-dependent, ARE-driven detoxification and antioxidant genes commonly play an important role in cellular protection (Table 1, Fig. 1). Furthermore, there are cell typedependent Nrf2 target genes that further contribute to cellular protection. Although some of these genes have been shown to be regulated through the Nrf2-ARE pathway, many have not been verified as being under direct control of Nrf2. It is quite possible that some of the changed genes could be secondary to an Nrf2 controlled gene; such information has yet to be investigated.

The basic defense system of each cell type has evolved to deal with those most likely toxic insults that each cell could encounter in its specific environment (**Fig. 2**). It would also appear that cell utilizes the Nrf2-ARE pathway as a common mechanism to regulate/activate its defense system.

CONCLUSIONS

In summary, we have proposed and presented evidence supporting the hypothesis that Nrf2 has the capability to confer and is central to a multi-organ protection phenomenon. Expansion of these known characteristics of the Nrf2-ARE pathway are important in identifying novel pathways necessary to prevent or cure toxic effects involved in multiple disease states. As numerous studies have demonstrated the great potential for the Nrf2-ARE pathway as a therapeutic target in preventing cancer, autoimmune disease, and neurodegenerative disease, it is important to identify ways to modulate cell-specific Nrf2 activity so as to facilitate the development of novel therapeutic strategies for treatment of these diseases.

REFERENCES

- Sakai, M., Okuda, A., and Muramatsu, M. (1988) Multiple regulatory elements and phorbol 12-O-tetradecanoate 13-acetate responsiveness of the rat placental glutathione transferase gene. *Proc. Natl. Acad. Sci. USA* 85, 9456–9460
- Rushmore, T. H., and Pickett, C. B. (1990) Transcriptional regulation of the rat glutathione S-transferase Ya subunit gene. Characterization of a xenobiotic-responsive element controlling inducible expression by phenolic antioxidants. *J. Biol. Chem.* 265, 14648–14653
- Friling, R. S., Bensimon, A., Tichauer, Y., and Daniel, V. (1990) Xenobiotic-inducible expression of murine glutathione S-transferase Ya subunit gene is controlled by an electrophile-responsive element. *Proc. Natl. Acad. Sci. USA* 87, 6258–6262
- 4. Favreau, L. V., and Pickett, C. B. (1991) Transcriptional regulation of the rat NAD(P)H:quinone reductase gene. Identification of regulatory elements controlling basal level expression and inducible expression by planar aromatic compounds and phenolic antioxidants. *J. Biol. Chem.* **266**, 4556–4561
- 5. Li, Y., and Jaiswal, A. K. (1992) Regulation of human NAD(P)H: quinone oxidoreductase gene. Role of AP1 binding site con-

- Venugopal, R., and Jaiswal, A. K. (1996) Nrfl and Nrf2 positively and c-Fos and Fra1 negatively regulate the human antioxidant response element-mediated expression of NAD(P)H:quinone oxidoreductase1 gene. *Proc. Natl. Acad. Sci. USA* 93, 14960– 14965
- Itoh, K., Chiba, T., Takahashi, S., Ishii, T., Igarashi, K., Katoh, Y., Oyake, T., Hayashi, N., Satoh, K., Hatayama, I., et al. (1997) An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem. Biophys. Res. Commun.* 236, 313–322
- Itoh, K., Wakabayashi, N., Katoh, Y., Ishii, T., Igarashi, K., Engel, J. D., and Yamamoto, M. (1999) Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. *Genes Dev.* 13, 76–86
- Huang, H. C., Nguyen, T., and Pickett, C. B. (2000) Regulation of the antioxidant response element by protein kinase Cmediated phosphorylation of NF-E2-related factor 2.[erratum appears in. *Proc. Natl. Acad. Sci. USA* 97, 12475–12480
- Yu, R., Chen, C., Mo, Y. Y., Hebbar, V., Owuor, E. D., Tan, T. H., and Kong, A. N. (2000) Activation of mitogen-activated protein kinase pathways induces antioxidant response element-mediated gene expression via a Nrf2-dependent mechanism. *J. Biol. Chem.* 275, 39907–39913
- Dinkova-Kostova, A. T., Holtzclaw, W. D., Cole, R. N., Itoh, K., Wakabayashi, N., Katoh, Y., Yamamoto, M., and Talalay, P. (2002) Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of phase 2 enzymes that protect against carcinogens and oxidants. *Proc. Natl. Acad. Sci. USA* 99, 11908–11913
- Dhakshinamoorthy, S., and Jaiswal, A. K. (2001) Functional characterization and role of INrf2 in antioxidant response element-mediated expression and antioxidant induction of NAD(P)H:quinone oxidoreductase1 gene. *Oncogene* 20, 3906– 3917
- Kang, K. W., Lee, S. J., Park, J. W., and Kim, S. G. (2002) Phosphatidylinositol 3-kinase regulates nuclear translocation of NF-E2-related factor 2 through actin rearrangement in response to oxidative stress. *Mol. Pharmacol.* 62, 1001–1010
- 14. Bloom, D. A., and Jaiswal, A. K. (2003) Phosphorylation of Nrf2 at Ser40 by protein kinase C in response to antioxidants leads to the release of Nrf2 from INrf2, but is not required for Nrf2 stabilization/accumulation in the nucleus and transcriptional activation of antioxidant response element-mediated NAD (P)H: quinone oxidoreductase-1 gene expression. *J. Biol. Chem.* 278, 44675–44682
- Wakabayashi, N., Dinkova-Kostova, A. T., Holtzclaw, W. D., Kang, M. I., Kobayashi, A., Yamamoto, M., Kensler, T. W., and Talalay, P. (2004) Protection against electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keapl sensor modified by inducers. *Proc. Natl. Acad. Sci. USA* 101, 2040–2045
- Kobayashi, A., Ohta, T., and Yamamoto, M. (2004) Unique function of the Nrf2-Keap1 pathway in the inducible expression of antioxidant and detoxifying enzymes. *Methods Enzymol.* 378, 273–286
- Chan, K., and Kan, Y. W. (1999) Nrf2 is essential for protection against acute pulmonary injury in mice. *Proc. Natl. Acad. Sci. USA* 96, 12731–12736
- Cho, H. Y., Jedlicka, A. E., Reddy, S. P., Kensler, T. W., Yamamoto, M., Zhang, L. Y., and Kleeberger, S. R. (2002) Role of NRF2 in protection against hyperoxic lung injury in mice. *Am. J. Respir. Cell Mol. Biol.* 26, 175–182
- Cho, H. Y., Reddy, S. P., Yamamoto, M., and Kleeberger, S. R. (2004) The transcription factor NRF2 protects against pulmonary fibrosis. *FASEB J.* 18, 1258–1260
- Rangasamy, T., Cho, C. Y., Thimmulappa, R. K., Zhen, L., Srisuma, S. S., Kensler, T. W., Yamamoto, M., Petrache, I., Tuder, R. M., and Biswal, S. (2004) Genetic ablation of Nrf2 enhances susceptibility to cigarette smoke-induced emphysema in mice. J. Clin. Invest. 114, 1248–1259
- Enomoto, A., Itoh, K., Nagayoshi, E., Haruta, J., Kimura, T., O'Connor, T., Harada, T., and Yamamoto, M. (2001) High sensitivity of Nrf2 knockout mice to acetaminophen hepatotoxicity associated with decreased expression of ARE-regulated

drug metabolizing enzymes and antioxidant genes. *Toxicol. Sci.* 59, 169–177

- Chan, K., Han, X. D., and Kan, Y. W. (2001) An important function of Nrf2 in combating oxidative stress: detoxification of acetaminophen. *Proc. Natl. Acad. Sci. USA* 98, 4611–4616
- Li, J., Stein, T. D., and Johnson, J. A. (2004) Genetic dissection of systemic autoimmune disease in Nrf2-deficient mice. *Physiol. Genomics* 18, 261–272
- Ramos-Gomez, M., Kwak, M. K., Dolan, P. M., Itoh, K., Yamamoto, M., Talalay, P., and Kensler, T. W. (2001) Sensitivity to carcinogenesis is increased and chemoprotective efficacy of enzyme inducers is lost in nrf2 transcription factor-deficient mice. *Proc. Natl. Acad. Sci. USA* 98, 3410–3415
- Ramos-Gomez, M., Dolan, P. M., Itoh, K., Yamamoto, M., and Kensler, T. W. (2003) Interactive effects of nrf2 genotype and oltipraz on benzo[a]pyrene-DNA adducts and tumor yield in mice. *Carcinogenesis* 24, 461–467
- Fahey, J. W., Haristoy, X., Dolan, P. M., Kensler, T. W., Scholtus, I., Stephenson, K. K., Talalay, P., and Lozniewski, A. (2002) Sulforaphane inhibits extracellular, intracellular, and antibioticresistant strains of Helicobacter pylori and prevents benzo-[a]pyrene-induced stomach tumors. *Proc. Natl. Acad. Sci. USA* 99, 7610–7615
- Murphy, T. H., De Long, M. J., and Coyle, J. T. (1991) Enhanced NAD(P)H:quinone reductase activity prevents glutamate toxicity produced by oxidative stress. *J. Neurochem.* 56, 990–995
- Li, J., Lee, J. M., and Johnson, J. A. (2002) Microarray analysis reveals an antioxidant responsive element-driven gene set involved in conferring protection from an oxidative stress-induced apoptosis in IMR-32 cells. *J. Biol. Chem.* 277, 388–394
- Lee, J. M., Shih, A. Y., Murphy, T. H., and Johnson, J. A. (2003) NF-E2-related factor-2 mediates neuroprotection against mitochondrial complex I inhibitors and increased concentrations of intracellular calcium in primary cortical neurons. *J. Biol. Chem.* 278, 37948–37956
- Lee, J. M., Calkins, M. J., Chan, K., Kan, Y. W., and Johnson, J. A. (2003) Identification of the NF-E2-related factor-2-dependent genes conferring protection against oxidative stress in primary cortical astrocytes using oligonucleotide microarray analysis. *J. Biol. Chem.* 278, 12029–12038
- 31. Kraft, A. D., Johnson, D. A., and Johnson, J. A. (2004) Nuclear factor E2-related factor 2-dependent antioxidant response element activation by tert-butylhydroquinone and sulforaphane occurring preferentially in astrocytes conditions neurons against oxidative insult. *J. Neurosci.* 24, 1101–1112
- Dhakshinamoorthy, S., and Porter, A. G. (2004) Nitric oxideinduced transcriptional up-regulation of protective genes by Nrf2 via the antioxidant response element counteracts apoptosis of neuroblastoma cells. *J. Biol. Chem.* 279, 20096–20107
- Calkins, M. J., Jakel, R. J., Johnson, D. A., Chan, K., and Kan, Y. W. (2004) Protection from mitochondrial complex II inhibi-

tion in vitro and in vivo by Nrf2-mediated transcription. Proc. Natl. Acad. Sci. USA 102, 244-249

- Gao, X., and Talalay, P. (2004) Induction of phase 2 genes by sulforaphane protects retinal pigment epithelial cells against photooxidative damage. *Proc. Natl. Acad. Sci. USA* 101, 10446– 10451
- 35. Li, N., Alam, J., Venkatesan, M. I., Eiguren-Fernandez, A., Schmitz, D., Di Stefano, E., Slaughter, N., Killeen, E., Wang, X., Huang, A., et al. (2004) Nrf2 is a key transcription factor that regulates antioxidant defense in macrophages and epithelial cells: protecting against the proinflammatory and oxidizing effects of diesel exhaust chemicals. J. Immunol. 173, 3467–3481
- Dyck, P. A., Hoda, F., Osmer, E. S., and Green, R. M. (2003) Microarray analysis of hepatic gene expression in gallstonesusceptible and gallstone-resistant mice. *Mamm. Genome* 14, 601–610
- 37. Braun, S., Hanselmann, C., Gassmann, M. G., auf dem Keller, U., Born-Berclaz, C., Chan, K., Kan, Y. W., and Werner, S. (2002) Nrf2 transcription factor, a novel target of keratinocyte growth factor action which regulates gene expression and inflammation in the healing skin wound. *Mol. Cell. Biol.* 22, 5492–5505
- Motohashi, H., Katsuoka, F., Engel, J. D., and Yamamoto, M. (2004) Small Maf proteins serve as transcriptional cofactors for keratinocyte differentiation in the Keap1-Nrf2 regulatory pathway. *Proc. Natl. Acad. Sci. USA* 101, 6379–6384
- Yoh, K., Itoh, K., Enomoto, A., Hirayama, A., Yamaguchi, N., Kobayashi, M., Morito, N., Koyama, A., Yamamoto, M., and Takahashi, S. (2001) Nrf2-deficient female mice develop lupuslike autoimmune nephritis. *Kidney Int.* 60, 1343–1353
- Lee, J. M., Chan, K., Kan, Y. W., and Johnson, J. A. (2004) Targeted disruption of Nrf2 causes regenerative immunemediated hemolytic anemia. *Proc. Natl. Acad. Sci. USA* 101, 9751–9756
- Thimmulappa, R. K., Mai, K. H., Srisuma, S., Kensler, T. W., Yamamoto, M., and Biswal, S. (2002) Identification of Nrf2regulated genes induced by the chemopreventive agent sulforaphane by oligonucleotide microarray. *Cancer Res.* 62, 5196–5203
- 42. Kwak, M. K., Wakabayashi, N., Itoh, K., Motohashi, H., Yamamoto, M., and Kensler, T. W. (2003) Modulation of gene expression by cancer chemopreventive dithiolethiones through the Keap1-Nrf2 pathway. Identification of novel gene clusters for cell survival. *J. Biol. Chem.* **278**, 8135–8145
- Shih, A. Y., Johnson, D. A., Wong, G., Kraft, A. D., Jiang, L., Erb, H., Johnson, J. A., and Murphy, T. H. (2003) Coordinate regulation of glutathione biosynthesis and release by Nrf2expressing glia potently protects neurons from oxidative stress. *J. Neurosci.* 23, 3394–3406

Received for publication December 14, 2004. Accepted for publication March 3, 2005.