

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/7015287>

A Review of the bioactivity and potential health benefits of peppermint tea (*Mentha piperita* L)

Article *in* *Phytotherapy Research* · August 2006

Impact Factor: 2.66 · DOI: 10.1002/ptr.1936 · Source: PubMed

CITATIONS

156

READS

1,913

2 authors:



[Diane McKay](#)

Tufts University

26 PUBLICATIONS 1,342 CITATIONS

[SEE PROFILE](#)



[Jeffrey B. Blumberg](#)

Tufts University

289 PUBLICATIONS 11,615 CITATIONS

[SEE PROFILE](#)

REVIEW ARTICLE

A Review of the Bioactivity and Potential Health Benefits of Peppermint Tea (*Mentha piperita* L.)

Diane L. McKay* and Jeffrey B. Blumberg

USDA Human Nutrition Research Center on Aging at Tufts University, 711 Washington St., Boston, MA 02111, USA

Peppermint (*Mentha piperita* L.) is one of the most widely consumed single ingredient herbal teas, or tisanes. Peppermint tea, brewed from the plant leaves, and the essential oil of peppermint are used in traditional medicines. Evidence-based research regarding the bioactivity of this herb is reviewed. The phenolic constituents of the leaves include rosmarinic acid and several flavonoids, primarily eriocitrin, luteolin and hesperidin. The main volatile components of the essential oil are menthol and menthone. *In vitro*, peppermint has significant antimicrobial and antiviral activities, strong antioxidant and antitumor actions, and some antiallergenic potential. Animal model studies demonstrate a relaxation effect on gastrointestinal (GI) tissue, analgesic and anesthetic effects in the central and peripheral nervous system, immunomodulating actions and chemopreventive potential. Human studies on the GI, respiratory tract and analgesic effects of peppermint oil and its constituents have been reported. Several clinical trials examining the effects of peppermint oil on irritable bowel syndrome (IBS) symptoms have been conducted. However, human studies of peppermint leaf are limited and clinical trials of peppermint tea are absent. Adverse reactions to peppermint tea have not been reported, although caution has been urged for peppermint oil therapy in patients with GI reflux, hiatal hernia or kidney stones. Copyright © 2006 John Wiley & Sons, Ltd.

Keywords: *Mentha piperita*; peppermint oil; menthol; herbal tea; tisane; dyspepsia.

INTRODUCTION

About 80% of the world population currently relies on indigenous or traditional medicines for their primary health needs, and most of this therapy involves the use of plant extracts, often in aqueous solutions (Zhang, 2002). Of the plant-based foods used as medicines, none have received more attention as a group than herbal remedies (Dubick, 1996). The use of herbal preparations, typically prepared by steeping or heating crude plant material, has prevailed for centuries and healthcare providers in Europe and Asia today often prescribe herbal teas. However, such practices are largely based on folklore and schools of traditional medicine rather than evidence-based research.

Peppermint (*Mentha piperita* L.) is among the most popular single ingredient herbal teas. The list of purported benefits and uses of peppermint as a folk remedy or in complementary and alternative medical therapy include: biliary disorders, dyspepsia, enteritis, flatulence, gastritis, intestinal colic, and spasms of the bile duct, gallbladder and gastrointestinal (GI) tract. We review here the available scientific literature related closely or directly to the bioactivity and potential health benefits of infusions, or tisanes, prepared with peppermint leaves, and the effects of the essential oil and other components contained therein. Information regarding the phytochemical and nutrient content, *in vitro* experiments, animal models, and human studies available in the recent scientific literature is presented.

NOMENCLATURE

Peppermint (*Mentha piperita* L.) is a perennial herb native to Europe, naturalized in the northern USA and Canada, and cultivated in many parts of the world. A hybrid of spearmint (*M. spicata* L.) and water mint (*M. aquatica* L.), peppermint grows particularly well in areas with high water-holding capacity soil. Best known for its flavoring and fragrance properties, peppermint leaves (fresh and dried) and the essential oil extracted from the leaves are used in many food, cosmetic and pharmaceutical products.

* Correspondence: Diane L. McKay, Antioxidants Research Laboratory, Jean Mayer USDA Human Nutrition Research Center on Aging at Tufts University, 711 Washington Street, Boston, MA 02111, USA.
E-mail: diane.mckay@tufts.edu

The contents of this publication do not necessarily reflect the views or policies of the USDA nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. government.
Contract/grant sponsor: U.S. Department of Agriculture (USDA) Agricultural Research Service; contract/grant number: 58-1950-001
Contract/grant sponsor: Hain Celestial Group (Boulder, CO)

PHYTOCHEMICAL AND NUTRIENT CONTENT

The chemical components of peppermint leaves and oil vary with plant maturity, variety, geographical region and processing conditions (Clark and Menary, 1981; Maffei and Scannerini, 1992; Rohloff, 1999; Gherman *et al.*, 2000; Blanco *et al.*, 2002; Pino *et al.*, 2002; Ruiz del Castillo *et al.*, 2003; Xu *et al.*, 2003). The fatty acid composition of the non-polar lipid fraction of peppermint leaves is dominated by palmitic (16:0), linoleic (18:2) and linolenic (18:3) acids (Maffei and Scannerini, 1992). The main volatile components identified in the essential oil of peppermint are menthol (33–60%), menthone (15–32%), isomenthone (2–8%), 1,8-cineole (eucalyptol) (5–13%), menthyl acetate (2–11%), menthofuran (1–10%), limonene (1–7%), β -myrcene (0.1–1.7%), β -caryophyllene (2–4%), pulegone (0.5–1.6%) and carvone (1%) (Clark and Menary, 1981; Sang, 1982; Pittler and Ernst, 1998; Dimandja *et al.*, 2000; Gherman *et al.*, 2000). The leaves contain 1.2–3.9% (v/w) essential oil (Picuric-Jovanovic *et al.*, 1997; Blumenthal *et al.*, 1998) (0.38% yield from fresh leaves) (Kaul *et al.*, 2001), while an infusion of dried leaves is reported to contain 21% of the original oil (25 mg/L). Proportions of the individual components found in oil were both higher and lower than those found in the infused tea (Duband *et al.*, 1992).

Studies regarding the mineral content of peppermint leaves are more comprehensive than those pertaining to the vitamin content. Fresh *M. piperita* leaves from Brazil were found to contain 940–1016 retinol equivalents (RE)/100 g β -carotene (de Almeida-Muradian *et al.*, 1998). The presence of other carotenoids and chlorophylls (Pilipenko *et al.*, 1998), as well as α - and γ -tocopherols (Blumenthal *et al.*, 1998) and ascorbic acid (Capecka *et al.*, 2005), has also been reported. The major minerals in dried peppermint leaves (as g/kg) include K (33), Ca (15.3), Mg (5.8) and lower amounts of Na, along with smaller amounts (as mg/kg) of Fe (239), Mn (188), Zn (51) and Cu (12). Trace amounts (as μ g/g) of Cr (941), I (325) and Se (147) are also present (Zimna and Piekos, 1988; Lozak *et al.*, 2002). Concentrations of these minerals found in an infusion of dried leaves (prepared at 95 °C, 15 min) were approximately 8–60% of the amounts present in the leaves, i.e. Ca (2.9 g/kg), Mg (2.2 g/kg), Fe (20 mg/kg), Mn (27 mg/kg), Zn (6 mg/kg), Cu (3 mg/kg), Cr (390 μ g/g), I (206 μ g/g) and Se (87 μ g/g) (Lozak *et al.*, 2002). According to Lozak *et al.* (2002), the most readily eluted elements of nutritional importance from the leaves are Se and I with Fe as the least eluted mineral. Muller *et al.* (1997) reported finding 477 μ g/g Al in dried mint leaves, approximately half the amount present in black tea (899 μ g/g); however, the transfer of Al into a peppermint tea infusion was very low (5%) compared with black tea (30%).

The total polyphenolic content of peppermint leaves is approximately 19–23% (total flavonoids 12%), which includes 59–67% eriocitrin and rosmarinic acid (combined), 7–12% luteolin 7-O-rutinoside, 6–10% hesperidin, and smaller quantities of 5,6-dihydroxy-7,8,3',4'-tetramethoxyflavone, pebrellin, gardenin B and apigenin (Hoffmann and Lunder, 1984; Zakharov *et al.*, 1990; Samejima *et al.*, 1995; Areias *et al.*, 2001; Zheng and Wang, 2001). About 75% of the polyphenolic

compounds present in the leaves are extracted in an infusion (750 mg/L) (Duband *et al.*, 1992).

The salicylic acid content of peppermint candies and tea was reportedly very high (7.7–75.8 mg/kg) in an early study by Swain *et al.* (1985); however, a more recent analysis using a more sensitive assay method by Venema *et al.* (1996) revealed <0.2 mg/kg.

IN VITRO STUDIES

Antioxidant capacity. The antioxidant capacity of peppermint has been determined using a number of different assay methods. The oxygen radical absorbance capacity (ORAC) value for an aqueous solution of previously frozen fresh *M. piperita* leaves (supernatant of 2.0 g homogenized in 15 mL buffer) was among the highest found in an analysis of popular medicinal herbs by Zheng and Wang (2001). At 15.84 ± 0.42 μ mol Trolox equivalents (TE)/g fresh weight, the ORAC value for *M. piperita* was similar to *Hypericum perforatum* (16.77 ± 0.22 μ mol TE/g) and *Valerian officinalis* (15.69 ± 0.37 μ mol TE/g), slightly higher than *Salvia officinalis* (13.28 ± 0.40 μ mol TE/g) and lower than *Thymus vulgaris* (19.49 ± 0.21 μ mol TE/g). The ORAC values for the related *M. aquatica* and *M. spicata* were 19.80 ± 0.43 and 8.10 ± 0.26 μ mol TE/g, respectively. In a study using the ferric reducing ability of plasma (FRAP) assay, Dragland *et al.* (2003) found the relative antioxidant value of dried *M. piperita* (78.5 mmol/g) to be lower than *S. officinalis* (91.2 mmol/g) and higher than *T. vulgaris* (74.6 mmol/g). FRAP values >75 mmol/g are indicative of high antioxidant concentrations. As with other compounds present in *M. piperita*, seasonal variations with regard to antioxidant activity have been observed (range 59.8–96.1 mmol/g) (Dragland *et al.*, 2003).

According to Mimica-Dukic *et al.* (2003), the free radical scavenging capacity of *M. piperita* oil was higher than that of either *M. aquatica* or *M. longifolia*. In their experiment, *M. piperita* reduced the radical generator 2,2-diphenyl-1-picrylhydrazyl (DPPH) by 50% ($IC_{50} = 2.53$ μ g/mL) and inhibited the generation of the OH radical in the Fenton reaction by 24%. In an assay based upon the oxidation of homovanillic acid (HVA) to its fluorescent biphenyl dimer in the presence of H_2O_2 and peroxidase, the antioxidant capacities of aqueous solutions of peppermint (0.1, 0.5 and 1.0%), prepared with boiling water and incubated 10 min at 95 °C, were among the highest of the tea infusions tested by Pazdzioch-Czochra and Widenska (2002). The percent of fluorescence inhibition exhibited by a 0.5% peppermint infusion (closest approximation to an amount typically used) was ~67%; lower than comparable amounts of black (78%) and green (81%) teas, but higher than other herb teas including hibiscus (56%) and rooibos (52%). When the results were expressed as Trolox equivalents, these teas were ranked similarly, i.e. black tea (0.32 ± 0.05), green tea (0.31 ± 0.03), peppermint tea (0.27 ± 0.02), hibiscus tea (0.20 ± 0.02) and rooibos (0.17 ± 0.01). One limitation of this particular method includes the ability to estimate only the H_2O_2 scavenging ability of the tested herbs, and not the scavenging of other free radicals. In other studies, ethanol extracts of dried *M. piperata* were shown to stabilize

the auto-oxidation of kinetically pure triacylglycerols of sunflower oil (Yanishlieva and Marinova, 1995), and natural sunflower oil (Marinova and Yanishlieva, 1997). Essential oil derived from peppermint exhibited an even greater antioxidant effect against sunflower oil peroxidation than butylated hydroxytoluene (BHT) (Gurdip *et al.*, 1998).

Antitumor activity. Ohara and Matsuhisa (2002) screened 120 edible plants for antitumor promoting activities against the non-12-O-tetradecanoylphorbol-13-acetate (TPA)-type promoter, okadaic acid (OA), which promotes tumor formation by inhibiting protein phosphatase-2A. Peppermint was one of only eight plants that showed strong activity (86–100%) in suppressing the effect of OA. Menthol derived from *M. piperita* appears to affect cytosolic arylamine N-acetyltransferase (NAT) activity in the human liver tumor cell line J5 differentially dependent on dose (Lin *et al.*, 2001); higher doses (32 and 3.2 mM) inhibited NAT, a more moderate dose (0.32 mM) had no effect, and lower doses (0.032 and 0.0032 mM) promoted NAT relative to controls.

Peppermint oil showed a genotoxic effect in human lymphocytes in a study by Lazutka *et al.* (2001). The frequency of chromosomal aberrations was highest ($16.0 \pm 2.3\%$) with $0.20 \mu\text{L/mL}$ peppermint oil (control = $2.0 \pm 0.6\%$) but was reduced at concentrations of 0.25 and $0.30 \mu\text{L/mL}$ (9.0 ± 1.6 and $6.0 \pm 1.4\%$, respectively). In this same experiment, peppermint oil showed weak sister chromatid exchange (SCE) activity, although the effect was not dose-dependent, and the oil inhibited mitotic activity at the lowest tested concentration ($0.10 \mu\text{L/mL}$). In contrast, using a Chinese hamster fibroblast cell chromosome aberration assay, the effects of peppermint oil were equivocal and, using a mouse lymphoma mutagenesis assay and the Ames test (using *Salmonella typhimurium*), no mutagenic activity was detected with peppermint oil (Andersen and Jensen, 1984; Anonymous, 2001); individual components of peppermint leaves and peppermint oil were also absent for evidence of genotoxicity. No significant chromosomal aberrations or changes in the frequency of SCE were observed in phytohemagglutinin-stimulated human lymphocyte cultures grown in the presence of 0.1, 1.0 or 10.0 mM menthol when compared with control samples (Murthy *et al.*, 1991). Similarly, menthol (200–700 $\mu\text{g/plate}$) and 1,8-cineole (1500–2500 $\mu\text{g/plate}$) did not increase the number of revertant colonies observed with and without addition of S9 rat hepatic microsomal enzymes in the *S. typhimurium* assay (Gomes-Carneiro *et al.*, 1998). An aqueous extract of peppermint also strongly suppressed the mutagenicity of 3-amino-1-methyl-5H-pyrido[4,3-b]indole (Trp-P-2), a human carcinogen formed in cooked meat, when evaluated in the *S. typhimurium* assay (Natake *et al.*, 1989). This effect was most likely due to the presence of the flavonoid luteolin (Samejima *et al.*, 1995).

Kim *et al.* (2002) tested the effects of the methanol extracts of ten herbs on L1210 cancer cells, and found that all of the herbs, including *M. piperita*, were cytotoxic. The addition of the herbs augmented the generation of superoxide ion and increased activities of superoxide dismutase and glutathione peroxidase, suggesting the cytotoxic mechanism may involve reactive oxygen species. Astrocytes, the most abundant glial cell

type in the brain, may pathologically affect neuronal activities following exposure to environmental stress. Koo *et al.* (2001) heat-shock induced apoptosis in rat and human and astrocytes pretreated with peppermint oil and inhibited DNA fragmentation and condensation of nuclear chromatin. In the human cells, peppermint oil also inhibited caspase-3 activation and poly-ADP-ribose polymerase fragmentation.

In human liver microsomes, Dresser *et al.* (2002) determined that peppermint oil and two of its components were moderately effective, reversible, and partially mixed inhibitors of nifedipine metabolism. Metabolism of this calcium channel blocker is mediated by the cytochrome P450 isoform CYP3A4. The effects of peppermint oil, menthol and menthyl acetate (K_i 35.9, 87.0, $124.0 \mu\text{mol/L}$, respectively) were less potent than two other partially mixed reversible CYP3A4 inhibitors, buspirone (K_i $20.2 \mu\text{mol/L}$) and propafenone (K_i $38.9 \mu\text{mol/L}$), and non-significant when compared with irreversible, or mechanism-based inhibitors. These findings suggest that peppermint may affect the bioavailability of certain drugs that require CYP3A4 for their effective metabolism. Using a different assay method, Unger and Frank (2004) reported that peppermint oil moderately inhibited all CYP enzymes tested except for 3A4, which was only 20% inhibited at $500 \mu\text{g/mL}$.

Antiallergenic activity. In rat peritoneal mast cells, Inoue *et al.* (2002) observed an antiallergenic activity among the flavonoid glycosides derived from *M. piperita*, including eriocitrin, narirutin, hesperidin, luteolin-7-O-rutinoside, isorhoifolin, diosmin, rosmarinic acid and 5, 7-dihydroxycromone-7-O-rutinoside. Of the compounds tested, only luteolin-7-O-rutinoside showed a potent inhibitory effect on histamine release induced by compound 48/80 and an antigen-antibody reaction. However, beyond the flavonoid activity in this assay, using lipopolysaccharide (LPS)-stimulated monocytes from healthy human subjects, Juergens *et al.* (1998a) found that menthol ($0.1 \mu\text{g/mL}$) significantly suppressed the production of the inflammatory mediating compounds leukotriene (LT) B₄ (64.4%), prostaglandin (PG) E₂ (56.6%) and interleukin (IL)- β 2 (64.2%). Mint oil ($0.1 \mu\text{g/mL}$) had a similar effect on LTB₄, PGE₂ and IL- β 2, but at lower concentrations ($<0.01 \mu\text{g/mL}$) there was a paradoxical increase in PGE₂ production. Another peppermint constituent, 1,8-cineole, significantly inhibited the production of tumor necrosis factor (TNF)- α , IL-1 β , LTB₄ and thromboxane B₂ in the same *in vitro* model (Juergens *et al.*, 1998b). In canine airway epithelial cells, menthol increased cytosolic calcium (Takeuchi *et al.*, 1994) and stimulated the secretion of Cl⁻ through a calcium-dependent mechanism (Chiyotani *et al.*, 1994). Using Caco-2 cells, Satsu *et al.* (2004) observed the increased secretion of IL-8 with an ethanol extract of peppermint, most likely attributable to the presence of the monocyclic sesquiterpene α -humulene.

Antiviral activity. Herrmann and Kucera (1967) found significant antiviral activity in aqueous extracts of peppermint leaves towards Influenza A, Newcastle disease virus, Herpes simplex virus (HSV) and Vaccinia virus in egg and cell-culture systems. An alcohol extract of *M. piperita* in combination with four other herbs (*Thymus serpyllum*, *Viscum album*, *Salvia officinalis* and

Glycyrrhiza glabra) inhibited the reproduction of influenza viruses A/Gabrovo (H1N1), A/Hong Kong (H3N2) and A/PR/8 (H1N1) in tissue cultures and embryonated eggs, reducing their infectious titers by 3.5, 3.0 and 2.0 log₁₀ inhibitory dose (ID)₅₀/mL, respectively (Manolova *et al.*, 1995). Yamasaki *et al.* (1998) tested an aqueous extract of *M. piperita* and found potent anti-human immunodeficiency virus-1 (HIV)-1 activity at an effective dose of 16 µg/mL in MT-4 cells. Water-soluble polar substances in the extract also showed inhibitory activity against HIV-reverse transcriptase.

Minami *et al.* (2003) found peppermint essential oil (1%) suppressed the replicative ability of HSV-1 in Vero cells incubated at 4 °C for 24 h. As no viral activity was observed in Vero cells treated with essential oil either before or after viral adsorption, the investigators suggested a direct interaction between the oil and the HSV-1 virion. Similarly, Schuhmacher *et al.* (2003) found both HSV-1 and HSV-2 were significantly inhibited when the viruses were treated with peppermint oil prior to adsorption, but not after penetration into the host cell. The 50% inhibitory concentration (IC₅₀) of peppermint oil was 0.002% for HSV-1 and 0.0008% for HSV-2 in RC-37 cells using a plaque reduction assay. Peppermint oil also exhibited virucidal activity in viral suspension tests. Viral titers of HSV-1 were reduced by 82%, HSV-2 by 92% and plaque formation of an acyclovir resistant strain (HSV-1-ACV[res]) was reduced by 99%.

Antibacterial activity. Many studies have assessed the antibacterial activity of peppermint (Piccaglia *et al.*, 1993; Shapiro *et al.*, 1994; Larsen *et al.*, 1996; Pattnaik *et al.*, 1996; Nelson, 1997; Carvalho *et al.*, 1999; Tkachenko *et al.*, 1999; Furuhashi *et al.*, 2000; Tassou *et al.*, 2000; Akin *et al.*, 2001; Imai *et al.*, 2001; Inouye *et al.*, 2001; Marino *et al.*, 2001; Montes Belmont and Flores Moctezuma, 2001; Aridogan *et al.*, 2002; Bonyadian and Karim, 2002; Iscan *et al.*, 2002; Azuma *et al.*, 2003) and antifungal (Sarbhoy *et al.*, 1978; Rai and Upadhyay, 1988; Pattnaik *et al.*, 1996; Zambonelli *et al.*, 1996; Carvalho *et al.*, 1999; Blaszczyk *et al.*, 2000; Ezzat, 2001; Karanika *et al.*, 2001; Giamperi *et al.*, 2002; Edris and Farrag, 2003; Mimica-Dukic *et al.*, 2003). For example, Iscan *et al.* (2002) tested peppermint oil and its components menthol and menthone against 21 human and plant pathogens and found moderate inhibitory activity against the human pathogens. *Staphylococcus aureus* was inhibited by 0.63 mg/mL (minimum inhibitory concentration (MIC) of oil, *Listeria monocytogenes* by 0.16–63 mg/mL, and *Staphylococcus epidermidis* by 0.63–2.5 mg/mL. The oil showed stronger inhibition (0.07–1.25 mg/mL) against the *Pseudomonas* and *Xanthomonas* strains of plant pathogens. Pattnaik *et al.* (1996) found that peppermint oil was effective against 22 different bacterial strains, including Gram-positive cocci and rods and Gram-negative rods, with an MIC of 0.16–20 µL/mL. According to Tassou *et al.* (2000), the addition of 0.4–1.2% of peppermint oil to nutrient broth, either with or without glucose, reduced the total viable count of *Staphylococcus aureus* by 6–7 logs colony forming units (cfu), while 0.1–1.0% reduced *Salmonella enteritidis* by 3 log cfu. At a concentration of 0.1% (v/v), peppermint oil was also able to inhibit the production of *S. aureus* toxin by a factor of 100 000. In drug resistant *S. aureus* and *Enterococcus faecium*, Nelson (1997) determined the

effective bacteriostatic and bactericidal dose of peppermint oil was 0.5–2.0%. Mimica-Dukic *et al.* (2003) found *M. piperita* oil was more effective against a multiresistant strain of *Shigella sonnei* and *Micrococcus flavus* than oils from other *Mentha* species.

Inouye *et al.* (2001) reported the major respiratory tract pathogens, including *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Streptococcus pyogenes* and *Staphylococcus aureus*, were susceptible to peppermint oil and its components menthol and menthone, but not 1,8-cineole. Menthol was the most effective of the peppermint components with a MIC range of 0.04–0.08% (w/v), followed by peppermint oil at 0.08–0.32%. Another respiratory tract pathogen, *Legionella pneumophila*, was also found to be susceptible to peppermint (Furuhashi *et al.*, 2000).

Azuma *et al.* (2003) demonstrated the effectiveness of menthol against the gastrointestinal bacteria *Helicobacter pylori* at 0.5 mM (MIC), but found no inhibition with 1,8-cineole tested at concentrations ≤4 mM. Mahady *et al.* (2005) reported a methanol extract of peppermint as weakly active against 15 strains of *H. pylori* with an MIC range of 25–100 µg/mL. The reported effects of peppermint oil on *Escherichia coli* are mixed, possibly reflecting a differential susceptibility of various strains used and/or testing conditions (Pattnaik *et al.*, 1995; Arakawa and Osawa, 2000; Inouye *et al.*, 2001; Aridogan *et al.*, 2002; Mimica-Dukic *et al.*, 2003).

Fungicidal and antimicrobial activity. The fungicidal activity of peppermint oil was demonstrated in 11 of 12 fungi tested by Pattnaik *et al.* (1996), including *Candida albicans*, *Trichophyton mentagrophytes*, *Aspergillus fumigatus* and *Cryptococcus neoformans*, at an MIC range of 0.25–10 µL/mL. Peppermint extracts were shown to have a moderate effect against these and other pathologically relevant fungi in other studies as well (Guerin and Reveillere, 1985; Rai and Upadhyay, 1988; Blaszczyk *et al.*, 2000; Ezzat, 2001; Duarte *et al.*, 2005; Tampieri *et al.*, 2005).

Peppermint oil has also been shown to be an effective antimicrobial and pest control agent in food crops and foodstuffs (Mabrouk and El-Shayeb, 1980; Tassou *et al.*, 1995; Damayanti *et al.*, 1996; Montes-Belmont and Carvajal, 1998; Hirazawa *et al.*, 2000; Karanika *et al.*, 2001; Montes Belmont and Flores Moctezuma, 2001; Lee *et al.*, 2002; Al-Abbadi and Nazer, 2003; Araujo *et al.*, 2003; Choi *et al.*, 2003; Guynot *et al.*, 2003).

ANIMAL MODEL STUDIES

Gastrointestinal actions. The effects of peppermint on the muscular actions and secretory processes of the gastrointestinal (GI) tract have been examined in many different animal models. Aqueous extracts of *M. piperita* showed a significant, dose-dependent relaxation effect on isolated rabbit duodenum in a study by Mahmood *et al.* (2003). The effect of dried leaf extract was greater than fresh leaf extract and a decrease in spontaneous activity was also observed. Acetylcholine-induced contraction of the muscle was only slightly modified in the presence of peppermint extract suggesting the mechanism of relaxation was most likely not due to cholinergic

antagonism. Further, the occurrence of an extract-induced relaxation after the addition of barium chloride (to increase spontaneous activity of the duodenum) suggests the relaxation was not due to adrenergic agonism.

Experiments by Hawthorn *et al.* (1988) using other isolated muscle preparations, including guinea-pig ileal smooth muscle, found that both menthol and peppermint oil at 78 µg/mL competitively inhibited the binding of the labeled calcium channel blockers ³H-nitrendipine and ³H-PN 200-110. The mechanism of action on GI smooth muscle relaxation appears to involve calcium channel antagonism. In a set of patch clamp experiments performed on rabbit jejunum and guinea-pig colon by Hills and Aaronson (1991), a reduction in calcium influx was also observed with peppermint oil. The calcium channel blocking action of peppermint and its components may influence the transport and secretory activity of enterocytes lining the intestinal lumen according to Beesley *et al.* (1996). In their experiment using intestinal sheets of Wistar rat jejunum, peppermint oil (1 and 5 mg/mL) applied to the mucosal side significantly inhibited active sodium-dependent glucose absorption and active transport of the amino acid glycine, while serosal application (1 mg/mL) inhibited acetylcholine induced secretion.

Peppermint oil and menthol have been shown to effectively stimulate choleric activity (bile flow) in rats at doses of 25–50 mg/kg administered i.v. (Trabace *et al.*, 1992; Trabace *et al.*, 1994). Vo *et al.* (2003) observed a significant increase of bile flow in rats treated with 830 µL/kg by gavage, but not at lower doses (83 or 8.3 µL/kg). One possible mechanism may involve the ability of menthol to inhibit the binding of β-D-glucuronide, a cholestatic compound, to rat liver plasma membranes (Takacs and Vore, 1987).

In the large intestine of pigs, the production of volatile sulfur compounds by the metabolism of intestinal bacteria was significantly reduced with peppermint (Ushid *et al.*, 2002). The digesta of pigs supplemented with peppermint extract and L-methionine (to enhance methanethiol (MeSH) production) were sampled at 24 h. While the number of volatile sulfur-producing bacteria remained similar in each group, peppermint significantly decreased hydrogen sulfide, MeSH and ammonia production, but had no effect on the production of short chain fatty acids. In a study by Ando *et al.* (2003) of peppermint-fed Holstein steers, the ammonia nitrogen concentrations were also lowered and the total numbers of protozoa, including *Entodinium*, *Isotrica* and *Diplodium*, in the rumen were reduced. The digestibility of nutrients also tended to be higher in the steers given peppermint than in controls. In a feeding study of 72 New Zealand White rabbits, Ibrahim *et al.* (2000) compared the effects of different herbs, including peppermint, sweet basil, oregano, thyme and catnip, on growth and metabolic changes. Changes in weight gain, feed intake and biochemical parameters (red blood cell count, hemoglobin, packed cell volume, blood glucose, total protein, liver aspartate aminotransferase and alanine aminotransferase, urea, creatinine and total lipids) were only slightly affected by peppermint compared with all of the other herbs tested. The concentration of the antioxidant compound 6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (HTCA) was, however, significantly higher (2.39 mol/L) in the

milk of Holstein cows fed 1 kg peppermint per day for 14 days compared with cows fed either basil (2.21 mol/L), lemongrass (2.05 mol/L) or a basal diet without the addition of herbs (1.40 mol/L) (Uegaki *et al.*, 2001). In a study of 48 rats, Akdogan *et al.* (2004a) found that replacing their drinking water with 20 g/L of *M. piperita* tea for 30 days inhibited iron absorption, significantly reduced serum iron and ferritin levels, and increased unsaturated iron-binding capacity.

Hepatic and renal actions. A study of female Wistar rats by Maliakal and Wanwimolruk (2001) demonstrated the modulatory effects of peppermint tea on selected hepatic phase I metabolizing enzymes. Pretreatment with a 2% tea solution for 4 weeks ($n = 5$) significantly decreased the activities of cytochrome P450 isoforms CYP1A2 (24%) and CYP2E (48%) compared with an equal sized control group given free access to water. Similarly, peppermint oil (50–100 mg/kg) was also shown to inhibit CYP3A after 24 h and consequently improve the oral bioavailability of cyclosporine in rats (Wacher *et al.*, 2002). Rats given daily oral doses of 83 µL/kg peppermint oil for 28 days had significantly increased alkaline phosphatase (ALP) levels (262 ± 21 U/L, $n = 6$) compared with controls (181 ± 18 U/L, $n = 5$), but no increase in bilirubin, glutamyl transpeptidase (GGT) or alanine aminotransferase (ALT) and no changes in liver histology (Vo *et al.*, 2003). In a study comparing the effects of *M. piperita* and *M. spicata* teas, Akdogan *et al.* (2003) found no evidence of nephrotoxicity from *M. piperita* in male Wistar albino rats. In this experiment, rats ($n = 48$) were given water, 20 g/L *M. piperita* tea, 20 or 40 g/L *M. spicata* tea as their sole source of drinking water for 30 days. *M. spicata* tea significantly increased plasma urea, creatinine and TBARS and decreased the activities of superoxide dismutase, glutathione peroxidase and catalase compared with water; in contrast, none of these parameters were significantly altered in rats administered *M. piperita*. Although histopathological changes were observed in groups given either type of tea, i.e. hydropic degeneration of tubular epithelial cells, epithelial cells with picnotic nuclei and eosinophilic cytoplasm, tubular dilation and enlargements in Bowman capsules, these changes in the *M. piperita* group were slight compared with the *M. spicata* group. Similarly, a later experiment showed only minimal hepatocyte degeneration with *M. piperita* compared with *M. spicata* (Akdogan *et al.*, 2004b).

Chemopreventive potential. Peppermint appears to prevent or reduce carcinogenesis induced by various agents in some animal models. A powdered tobacco mixture (15 g) without or with 15 g peppermint leaves was painted onto cheek pouches of Syrian golden hamsters 3 times/week for 20 weeks by Samman *et al.* (1998). At week 30, the number of animals showing morphological changes and frank tumors was recorded. Compared with animals treated with tobacco plus peppermint, the non-mint containing tobacco mixture increased mucosal thickening ($n = 20/22$ in non-mint group vs $9/15$ in mint-containing group), leukoplakia ($20/22$ vs $3/15$) and frank tumors ($19/22$ vs $0/15$) in the oral cavity. Overall 86.3% of the animals treated with non-mint tobacco were tumor-bearing at week 30, while those treated with the mint-containing tobacco mix exhibited no tumors (0%). The effects of an aqueous

peppermint suspension on papillomagenesis of the skin induced by 7,12-dimethylbenz (a) anthracene (DMBA) in mice were studied by Sameena and Ashok (2001). Mice treated with peppermint topically for 2 weeks prior to and 2 weeks after DMBA application (just before application of 1% Croton oil) and mice fed peppermint beginning 2 weeks after DMBA (at the time of Croton oil application) showed a significant reduction in the cumulative number of papillomas compared with mice either topically treated with peppermint during the 2 weeks between DMBA and Croton oil application or untreated mice. These results suggest that the chemopreventive effect of peppermint on skin papillomas is most active during the promotional stage of carcinogenesis.

Samarth *et al.* (2001a; 2001b; 2002a; 2002b; 2004) and Samarth and Kumar (2003) explored the modifying effects of peppermint extracts against sublethal and lethal doses of gamma radiation in Swiss albino mice in several studies. Pretreatment with an aqueous extract of peppermint prior to whole body gamma irradiation at 4, 6, 8 and 10 Gy significantly increased the spleen weight and the number of endogenous spleen colonies compared with irradiated mice without pretreatment (Samarth *et al.*, 2001a). A daily oral dose of 1 g/kg administered for 3 days prior to irradiation at 8 Gy significantly increased hematological parameters (erythrocytes, leukocytes, hemoglobin, hematocrit) and improved the survival rate compared with irradiated control animals at 10 days post-irradiation (Samarth *et al.*, 2001b). The same protocol using peppermint extract (1 g/kg) and peppermint oil (40 µL/animal) decreased serum acid phosphatase and increased serum alkaline phosphatase compared with controls after irradiation, but the levels returned to normal within 5 d (Samarth *et al.*, 2001a; 2002b). Significant alterations in the intestinal mucosa of mice treated daily with 1 g/kg peppermint extract were observed within 20 days post-irradiation at 8 Gy (Samarth *et al.*, 2002b). Compared with controls, peppermint pretreatment increased villus height, total number of cells and mitotic cells, and decreased the number of goblet and dead cells. A regression analysis of the survival data in irradiated mice revealed that mice pretreated with peppermint were able to withstand a 1.78-fold higher dose of radiation than untreated mice (Samarth and Kumar, 2003).

Antiallergenic and antiinflammatory actions. A 50% ethanol extract of *M. piperita* leaves and stems administered orally to rats with nasal symptoms (induced by antigen challenge in actively sensitized animals) significantly inhibited sneezing at a dose of 300 mg/kg and nasal rubbing at 1000 mg/kg (Kamei *et al.*, 2000; Inoue *et al.*, 2001). In a study of flavonoid glycosides derived from *M. piperita*, only the fraction containing luteolin-7-O-rutinoside caused a dose-related inhibition of an antigen-induced nasal response at 100 and 300 mg/kg (Inoue *et al.*, 2002). These results suggest that peppermint may be helpful in alleviating the nasal symptoms of allergic rhinitis.

The immunomodulating effects of *M. piperita* and its constituents have also been examined in animal models. Intraperitoneal administration of peppermint oil, 1-menthol and 1,8-cineole to guinea-pigs suppressed homologous passive cutaneous anaphylaxis mediated by IgE antibodies (Arakawa *et al.*, 1992; Arakawa and

Osawa, 2000). At doses of 200 and 400 mg/kg, an ethanol extract of dried *M. piperita* leaves injected into male Swiss mice ($n = 5/\text{group}$) 30 min prior to topical xylene-induced ear edema significantly inhibited acute inflammation by 49–50% (Atta and Alkofahi, 1998). At 400 mg/kg, the extract also significantly reduced the weight of cotton granuloma in rats ($n = 6$) indicating a potential effect of peppermint on chronic inflammatory processes as well.

Laude *et al.* (1994) demonstrated the antitussive effect of menthol in guinea-pigs. The frequency of coughing in animals ($n = 13$) treated with 10 and 30 µg/L menthol vapors 5 min prior to an aerosolized citric acid challenge was reduced 28 and 56%, respectively, when compared with air (placebo). Similarly, Wright *et al.* (1997; 1998) showed that menthol had a direct action on guinea-pig bronchial smooth muscle ($n = 13$) where the bronchoconstrictors capsaicin and neurokinin A (NKA) were used to increase airway resistance *in vivo* in the presence of either air or menthol vapor (7.5 µg/L); after the removal of bronchial rings and pre-contraction with KCl or acetylcholine, relaxation was measured. Menthol significantly reduced both capsaicin- (51.3%) and NKA- (41.0%) induced airway restriction *in vivo*, and relaxed the KCl- and acetylcholine-induced bronchi constriction *ex vivo*.

Nervous system actions. Peppermint has been shown to affect both central and peripheral nervous system activity. Atta and Alkofahi (1998) examined the analgesic effects of an ethanol extract of *M. piperita* on Swiss mice ($n = 10/\text{group}$). Employing 0.7% acetic acid injected i.p. to induce pain observed by writhing, mice were first treated for 30 min with an oral dose of 200 or 400 mg/kg of the peppermint extract. The writhing in peppermint-treated animals was significantly lower by 38–44% compared with saline-injected controls. Using a hot plate test ($n = 10$) 400 mg/kg peppermint increased the latency of response to thermal stimulation, though the onset of analgesia was delayed (45–60 min) and temporary (subsided by 75 min).

The local anesthetic effect of 30–100 µg/mL menthol was demonstrated in rabbits using a conjunctival reflex test where an increased number of stimuli were necessary to provoke the reflex in a dose-dependent fashion (Galeotti *et al.*, 2001). Examining other central nervous system effects of *M. piperita*, Della Loggia *et al.* (1990) used a lyophilized infusion, initially prepared with 50 g dried leaves in 500 mL hot water for 10 min, for their series of experiments with male CD1 albino Swiss mice. A moderate increase (32% at 60 min) in the onset of barbiturate-induced sleep was seen with a dose of 300 mg/kg. The same dose diminished exploratory behavior at 60 and 90 min by 31% and 17%, respectively, and depressed motor activity during this period (after an initial excitatory period), but had no effect on motor coordination up to 3 h post treatment. In contrast, Umezu *et al.* (2001) found that 400–800 mg/kg of peppermint oil injected intraperitoneally in mice significantly increased their ambulatory activity 10–40 min after administration. The isolated peppermint constituents 1,8-cineole, menthone, isomenthone, menthol, pulegone, menthyl acetate and caryophyllene also significantly increased ambulatory behavior. Further studies examining the behavioral effects of menthol suggest that dopamine may mediate the

Table 1. Human studies examining the effects of orally ingested peppermint leaves (*Mentha piperita*)^a

Reference	Delivery method	Subjects	Dose	Duration	Outcome
Westphal <i>et al.</i> , 1996	Tablet	70 patients with chronic dyspepsia	2 tablets (containing 100 mg peppermint plus other herbs) 3 times/day or placebo	14 days	Relief of symptoms after 1 week in treatment group compared with baseline. No change in placebo group.
Madisch <i>et al.</i> , 2001	Encapsulated powder	60 patients with functional dyspepsia (25–70 years)	Daily consumption of herbal mixture containing peppermint or placebo	4 weeks	Improved GI symptom score
Uehleke <i>et al.</i> , 2002	Tablet	12 patients with idiopathic dyspepsia	3, 6, or 9 tablets (containing 100 mg peppermint plus other herbs) after a meal	1 time	3 tablets were sufficient to reduce acute GI symptoms

^a Not including studies of peppermint oil preparations.

activity-enhancing effect of peppermint (Umezu, 2002; 2003).

HUMAN STUDIES AND POTENTIAL APPLICATIONS TO HEALTH AND DISEASE

In Germany, peppermint leaf is licensed for use as a standard medicinal tea to treat dyspepsia. The German Commission E has also approved the internal use of the leaf for spastic complaints of the GI tract, gallbladder and bile ducts (Blumenthal *et al.*, 1998). Peppermint oil is approved for internal use in the event of spastic discomfort of the upper GI tract and bile ducts, irritable colon or irritable bowel syndrome (IBS), catarrhs of the respiratory tract and inflammation of the oral mucosa. Externally, the use of peppermint oil is approved for myalgia and neuralgia. With the exception of peppermint oil and IBS, studies providing evidence to either support or refute the applicability of peppermint as a treatment for many of these conditions in humans is somewhat limited (Table 1).

Effect on drug and nutrient bioavailability/metabolism.

Dresser *et al.* (2002) examined the effects of peppermint oil (660 µL) on the bioavailability of the calcium channel blocking drug felodipine in 12 healthy subjects (18–43 years). Subjects fasted for 10 h prior to testing, refrained from using alcohol or medications, and were given either grapefruit juice, peppermint oil, ascorbyl palmitate or water in a single dose, randomized, 4-way cross-over study at 1 week intervals. Peppermint oil significantly increased the plasma felodipine concentration over time (30 ± 4 nmol h L, area under the curve, AUC) and its inactive metabolite dehydrofelodipine (59 ± 6 nmol h/L), which is formed by the action of the P450 cytochrome CYP3A4. The effect of grapefruit juice on the AUC of felodipine (37 ± 4 nmol .h/L) and dehydrofelodipine (59 ± 5 nmol .h/L) was similar to that of peppermint. However, unlike grapefruit juice, peppermint oil had no effect on the ratio of dehydrofelodipine/felodipine indicating a lack of inhibition at the primary step of felodipine metabolism, i.e. CYP3A4. Thus, although peppermint oil increased the bioavailability of this felodipine, the exact mechanism may not be identical to that of grapefruit juice.

Hurrell *et al.* (1999) found that the bioavailability of non-heme iron was reduced by peppermint tea. Compared with water, all the beverages tested in this study (coffee, herbal teas, black tea, and cocoa) inhibited iron absorption from a bread meal in a dose-dependent manner as estimated by measuring the incorporation of radiolabeled Fe (⁵⁵Fe or ⁵⁹Fe) into erythrocytes of adult subjects. Inhibition by black tea was 79–94%, peppermint tea was comparable at 84% followed by pennyroyal at 73%, cocoa at 71%, vervain 59%, lime flower 52% and chamomile 47%. When concentrations of the beverages were adjusted so that each contained the same amount of total polyphenols, black tea and peppermint were equally effective and their inhibitory actions were higher than the other beverages.

Gastrointestinal actions. In a study of 12 subjects, Goerg and Spilker (2003) found that 90 mg peppermint oil (in 0.10 mL capsule) did not significantly affect gastric emptying time (assessed by ultrasonography and H₂ breath tests) compared with a placebo, but did cause a complete inhibition of gallbladder emptying and significantly increased gallbladder volume during the refilling phase. Transit time through the small intestine was also significantly delayed with peppermint oil (85.0 ± 7.8 min) compared with placebo (65.0 ± 6.1 min). In contrast, Dalvi *et al.* (1991) studied 26 subjects fed a radiolabeled test meal either with or without the addition of peppermint oil (0.2 mL in 25 mL water) and found the gastric emptying rate was accelerated after the peppermint treatment.

Tate (1997) tested the efficacy of peppermint oil inhalation at a treatment for postoperative nausea in 18 patients who underwent major gynecological surgery in a British hospital and received either no treatment (control), a placebo containing peppermint essence but no effective volatile constituents or peppermint. Although patients retained control over the frequency of administration, most inhaled the treatment only when feeling nauseous. Using a standardized descriptive ordinal scale survey to assess their degree of nausea, the reported nausea rate for the control and placebo groups was 100%, compared with 66% in the peppermint oil group, but the results were not statistically significant.

Peppermint oil has been found to reduce painful muscle spasms in patients undergoing endoscopy of the

upper and lower GI tract as well as in people subjected to barium enema. Leicester and Hunt (1982) observed that peppermint oil relieved colon spasm in 20 patients undergoing colonoscopy within 30 s of administration. In a study of 445 patients undergoing colonoscopy, Asao *et al.* (2001) found that intracolonic administration of a 0.8% peppermint oil solution reduced the spasmolytic effect inherent in this procedure by 88.5% in the treated group ($n = 409$) compared with a reduction of 33.3% in the control group ($n = 36$). In a randomized controlled trial of 100 patients undergoing endoscopy of the upper GI tract, Hiki *et al.* (2003) compared the antispasmodic effects of peppermint oil administered intraluminally with hyoscine-N-butylbromide administered intramuscularly. The opening ratio (percent change in diameter of the pyloric ring before and after treatment) was significantly higher in the peppermint oil group while the contraction ratio (percent change in diameter between the maximally and minimally opened pyloric ring) was significantly lower. In addition, the time required for the disappearance of the contraction rings in the gastric antrum was shorter in the peppermint group (97.1 ± 11.4 s) than in the hyoscine-N-butylbromide group (185.9 ± 10.1 s).

When added to a barium sulfate suspension, Sparks *et al.* (1995) found that peppermint oil eliminated residual spasm in 60% of patients ($n = 70$) undergoing a double contrast barium enema (DCBE) examination compared with 35% of patients ($n = 71$) in a control group. In a study of 383 patients subjected to DCBE, Asao *et al.* (2003) compared the effects of scopolamine ($n = 105$), an antispasmodic agent, with peppermint oil delivered either via a barium solution ($n = 91$) or enema tube ($n = 90$). The presence of spasm was evaluated on a second series of spot films. Compared with the non-treated group ($n = 97$), patients given either the drug or peppermint had a significantly higher rate of non-spasm examinations (13.4% vs 37.8–41.8%, respectively). The effects of the peppermint oil in the transverse and descending colon were comparable to the drug treatment; however, in the cecum and ascending colon, the effect of the peppermint oil was significantly stronger.

Pimentel *et al.* (2001) observed that manometry readings of lower esophageal sphincter pressures and contractile pressures of both upper and lower esophagus in eight patients with diffuse esophageal spasm, recorded before and after ingestion of a solution containing 5 drops peppermint oil in 10 mL water, were no different. However, peppermint oil completely eliminated simultaneous esophageal contractions in all patients and improved the number of multiphasic, spontaneous and missed contractions. The variability of amplitude and duration of esophageal contractions improved after peppermint oil treatment as well. Micklefield *et al.* (2000) also found a decreased number of duodenal contractions and contraction amplitudes in six patients measured with a manometer after the administration of a capsule containing 90 mg peppermint oil plus 50 mg caraway oil. In a follow up study of 24 patients, a capsule containing 90 mg of peppermint oil alone was also able to reduce significantly the frequency and duration of duodenal contractions and the duration of contractions in the gastric corpus, producing smooth-muscle relaxation in the stomach and duodenum (Micklefield *et al.*, 2003).

Abdominal pain and dyspepsia have been found to respond well to treatment with either peppermint leaves (Table 1) or oil. An herbal preparation containing peppermint leaves (in combination with extracts from bitter candy tuft, chamomile, caraway, licorice, lemon balm, angelica, celandine, and milk thistle) significantly improved the GI symptom score of 60 patients (mean age 46.8 years) with functional dyspepsia after 2 and 4 weeks of treatment in a randomized controlled trial by Madisch *et al.* (2001). This formulation was later shown to be as effective as the antispasmodic drug cisapride in relieving GI symptoms among patients whose initial symptoms were reported as moderate to severe (Rosch *et al.*, 2002). Uehleke *et al.* (2002) also reported improvement in patients with acute and chronic symptoms of dyspepsia after the administration of peppermint leaves (100 mg) in combination with other herbs (caraway, fennel, gentian). Similarly, Westphal *et al.* (1996) found that an herbal preparation (100 mL) containing 9.26 g peppermint leaves along with fennel (8.13 g), caraway (3.78 g) and wormwood (1.92 g) diluted and administered 20 min prior to meals 3 times daily for 2 weeks was more effective in alleviating symptoms than the antispasmodic drug metoclopramide. Significantly fewer patients taking the peppermint containing preparation ($n = 17$) complained of pain, nausea, heartburn, gastrospasms, retching, sensation of pressure and belching than those taking the drug ($n = 27$).

Two preparations of peppermint oil (90 or 36 mg) combined with caraway oil (50 mg or 20 mg, respectively) both reduced pain intensity and frequency over baseline measures in 213 patients with dyspepsia in an experiment by Freise and Kohler (1999). Compared with the antispasmodic cisapride, Madisch *et al.* (1999) found in a 4 week randomized controlled trial of 120 patients with functional dyspepsia, the effects of a peppermint oil (90 mg) plus caraway oil (50 mg) preparation were comparable with regard to pain score reduction and reduced frequency of pain. In another randomized clinical trial of the same formulation (90 mg peppermint oil plus 50 mg caraway oil) administered twice daily for 28 days in 96 patients with functional dyspepsia, May *et al.* (2000) observed a significant 40% reduction in pain intensity, 43% reduction in the sensation of pressure, heaviness and fullness, and 67% global improvement compared with baseline assessments. The respective changes for these measures in the placebo group were 21–22% lower than baseline values. May *et al.* (1996) also instituted a 4 week trial of 45 non-ulcer dyspepsia patients with the same preparation administered 3 times daily and found significantly improved reports on pain intensity and measures of pain frequency, medical prognosis, and severity of the disorder according to the Clinical Global Impressions Scale when compared with a placebo. The efficacy of this preparation was reportedly unaffected by *Helicobacter pylori*, which is present in approximately 50% of patients suffering from functional dyspepsia (May *et al.*, 2003). In a systematic review of studies examining the efficacy of herbal products in treating dyspepsia, Thompson Coon and Ernst (2002) concluded that the effects of peppermint and caraway were similar or greater in magnitude to conventional therapies and that the safety profile of this combination was encouraging.

Peppermint oil has also found to be efficacious in relieving symptoms attributable to irritable bowel

syndrome (IBS) in both adults and children. A summary of intervention studies in IBS patients is presented in Table 2. In their meta-analysis of eight randomized controlled trials using peppermint oil as a treatment for IBS symptoms, Pittler and Ernst (1998) found a significant positive effect compared with placebo in five of the studies. Although the earlier peppermint oil trials were criticized for design flaws and questionable statistical analyses, later studies were considered more robust.

Respiratory tract actions. Eccles *et al.* (1988) found that menthol, but neither of its isomers isomenthol nor neomenthol, had a specific pharmacological action on nasal sensory nerve endings that was not related to its peppermint aroma. In their experiment, the inhalation of menthol significantly enhanced the nasal sensation of airflow in 40 subjects. The same sensation also occurred after 5 min of exposure to menthol vapor in 31 subjects from a different experiment, but nasal airflow resistance was not decreased (Eccles and Jones, 1983). Inhalation of menthol caused a significant reduction in the sensation of respiratory discomfort during flow resistant loading and elastic loading in 11 healthy subjects, but had no effect on breathing pattern or ventilation (Nishino *et al.*, 1997). Oral administration of 11 mg menthol (lozenges) did not decrease nasal decongestion in a randomized, double-blind, placebo-controlled trial of 62 subjects diagnosed with the common cold (Eccles *et al.*, 1990), but did cause a marked change in nasal sensation of airflow with a subjective sensation of nasal decongestion. According to Naito *et al.* (1991; 1997), the effect of menthol on nasal airflow sensation is due to its stimulatory effect on the palantine nerve and the sensory nerve endings of the nasal mucosa, which do not influence airflow resistance. Compared with placebo (pine oil or air), menthol did reduce coughing induced by inhalation of 33 μ mol citric acid in 20 healthy subjects when given 5 min prior to each citric acid challenge (Morice *et al.*, 1994), suggesting its effectiveness as an antitussive agent. In contrast, Tamaoki *et al.* (1995) found no significant differences in vital capacity, forced expiratory volume or change in peak flow rate in a 4 week randomized controlled trial of 23 individuals with mild asthma given either nebulized menthol (10 mg) or placebo twice daily. However, using a different approach, Juergens *et al.* (2003) found the oral administration of 200 mg 1,8-cineole thrice daily for 12 weeks enabled 32 patients with steroid-dependent bronchial asthma to tolerate a 36% reduction in their daily prednisolone therapy (mean: 3.75 mg, range: 2.5–10.0 mg) compared with a 7% reduction (mean: 0.91 mg, range: 2.5–5.0 mg) with placebo. The same dose of 1,8-cineole also significantly inhibited the production of the arachidonic acid metabolites leukotriene 4 (40.3–57.9%) and prostaglandin E2 (31.3–42.7%) within 4 days in an *ex vivo* experiment of monocytes from both asthma patients ($n = 10$) and healthy subjects ($n = 12$) (Juergens *et al.*, 1998c). These results suggest a potential anti-inflammatory effect of peppermint oil when ingested over a period of time.

Analgesic actions. The application of peppermint oil to the skin of the forehead produced an analgesic effect in a randomized controlled trial of 32 healthy men (25 ± 2.1 y) by Gobel *et al.* (1994; 1995). Each

Table 2. Summary of studies examining the effects of peppermint oil in IBS patients

Study	Design	Subjects (n)	Intervention	Outcome
Rees <i>et al.</i> , 1979	Double-blind, placebo-controlled	18	12 \times 0.2 mL capsules, 3/day	Relieved abdominal symptoms; patients felt better
Dew <i>et al.</i> , 1984	Double-blind, crossover	29	12 \times 0.2 mL capsules, 3/day	Improved daily symptoms; no effect on bowel actions
Nash <i>et al.</i> , 1986	Double-blind, placebo-controlled	41	2 \times 0.2 mL capsules, 3/day for 4 weeks	No difference in symptoms or stool frequency
Wildgrube, 1988	Uncontrolled	40	14 day course	Prolonged intestinal transit time; improved bloating, abdominal pain
Lawson <i>et al.</i> , 1988	Double-blind, crossover	25	0.2–0.4 mL capsules, 3/day for 4 weeks	No change in global symptoms or severity; small increase in defecation frequency
Lech <i>et al.</i> , 1988	Double-blind, placebo-controlled	42	200 mg, 3/day for 4 weeks	Improved symptoms compared with placebo
Liu <i>et al.</i> , 1997	Double-blind, placebo-controlled	110	1 \times 187 mg capsule ^a , 3–4/day for 4 weeks	Improved symptoms compared with placebo
Kline <i>et al.</i> , 2001	Double-blind, placebo-controlled	42 (children)	1 or 2 \times 187 mg capsules ^a , 3/day for 2 weeks	Improved severity of symptoms, including pain

^a Capsules were manufactured by Tiliots of Switzerland under the trademark name Colpermin. Each capsule contained 187 mg or 0.2 mL peppermint oil.

subject received one of four different 3 min treatments consisting of ethanol preparations of either 10% peppermint plus 5% eucalyptus oils, 10% peppermint oil alone, 5% eucalyptus oil alone or ethanol plus oil essences (placebo) applied with a small sponge to the forehead and temples; treatment were separated by a 48 h washout period. Measures of pericranial muscle tension were significantly reduced by 30.6% with the combination of peppermint and eucalyptus oils and 28.8% with peppermint oil alone; however, only the peppermint oil preparation significantly reduced measures of pain sensitivity after thermal (40.3%) and ischemic (27.0%) stimuli to the head. The intensity of pain experienced by 41 patients (age 18–65 years) with tension-type headaches was significantly reduced with either the application of 10% peppermint oil or ingestion of 1000 mg acetaminophen in a follow-up study (Gobel *et al.*, 1996). There were no significant differences between the efficacies of these treatments, although a non-significant additive effect of simultaneous treatment was observed. It may be relevant to note here the electroencephalographic (EEG) data collected by Miki *et al.* (1997) showed significantly increased alpha and decreased beta waves in 15 healthy males (22–39 years) after peppermint inhalation, suggesting an activation of cerebral white matter. Satoh and Sugawara (2003) also reported a significant decrease in the magnitude of beta waves after peppermint inhalation.

ADVERSE REACTIONS/TOXICITY

Toxicology studies of peppermint oil and its components have been performed in animals. Histopathological changes in the white matter of the cerebellum were seen in rats ($n = 20$) given peppermint oil at doses of 40 and 100 mg/kg orally for 28 days, but no adverse effects were observed at 10 mg/kg (Thorup *et al.*, 1983a). No adverse effects were observed at 10 mg/kg. In a comparable 90 day rat study ($n = 28$), cyst-like spaces in the white matter of the cerebellum and hyaline droplets in the proximal tubules of the kidneys were observed in the highest dose group only (Spindler and Madsen, 1992). Interestingly, the extension of the cyst-like spaces was not aggravated with prolonged dosing in this study. Menthol administered to rats by gavage at 200, 400 and 800 mg/kg for 28 days significantly increased absolute and relative liver weights and the vacuolization of hepatocytes at all doses, although no sign of encephalopathy was observed (Thorup *et al.*, 1983b). At 80 and 160 mg/kg, pulegone administered for 28 days induced atonia, decreased blood creatinine levels, lowered body weight and caused histopathological changes in the liver

and white matter of the cerebellum (Thorup *et al.*, 1983b). No adverse effects were observed with 20 mg/kg pulegone. Menthone given orally to rats ($n = 20$) at 200, 400 and 800 mg/kg for 28 days decreased creatinine and increased alkaline phosphatase in a dose-dependent manner, increased bilirubin and liver and spleen weights, and also caused histopathological changes in the white matter of the cerebellum in the two highest dose groups (Madsen *et al.*, 1986). The accumulation of protein droplets containing $\alpha_2\mu$ -globulin in proximal tubular epithelial cells of rats ($n = 10$ /group) was observed after the administration of either 500–1000 mg/kg 1,8-cineole or 800–1600 mg/kg limonene for 28 days, however, no histopathological changes were observed in the brain (Kristiansen and Madsen, 1995).

A review on the use of *M. piperita* oil, leaf extract, leaf and leaf water in cosmetic formulations by Nair (2001) concluded that each are considered safe, although the concentration of pulegone in products containing these ingredients should be limited to 1%. Although the toxicity of menthol is considered to be low, it has the ability to enhance the penetration and absorption of other agents contained in some formulations, thereby increasing the effective dose of these agents at the indicated intake. A few case study reports have described contact sensitivities to peppermint oil and its components in topical and oral preparations (Dooms-Goossens *et al.*, 1977; Andersen, 1978; Morton *et al.*, 1995; Bonamonte *et al.*, 2001), but a patch test study of 4000 patients by Kanerva *et al.* (2001) found that menthol and peppermint oil provoked neither allergic nor irritant reactions.

Akdogan *et al.* (2004c) reported increased follicle-stimulating hormone and luteinizing hormone levels and decreased testosterone levels in rats given 20 g/L peppermint tea in place of their drinking water. As opposed to *M. spicata* tea, the only effect of *M. piperita* on testicular tissue was segmental maturation arrest in the seminiferous tubules.

There are no chronic toxicity studies of peppermint in humans, although the German Commission E (Blumenthal *et al.*, 1998) reports that the use of peppermint oil is contraindicated in patients with bile duct, gallbladder and liver disorders. Caution is also recommended for the use of peppermint oil capsules in patients with GI reflux, hiatal hernia or kidney stones.

Acknowledgements

Research support was provided by the U.S. Department of Agriculture (USDA) Agricultural Research Service under Cooperative Agreement No. 58-1950-001 and a grant from the Hain Celestial Group (Boulder, CO).

REFERENCES

- Akdogan M, Gultekin F, Yontem M. 2004a. Effect of *Mentha piperita* (Labiatae) and *Mentha spicata* (Labiatae) on iron absorption in rats. *Toxicol Ind Health* **20**: 119–122.
- Akdogan M, Kilinc I, Oncu M, Karaoz E, Delibas N. 2003. Investigation of biochemical and histopathological effects of *Mentha piperita* L. and *Mentha spicata* L. on kidney tissue in rats. *Hum Exp Toxicol* **22**: 213–219.
- Akdogan M, Ozguner M, Aydin G, Gokalp O. 2004b. Investigation of biochemical and histopathological effects of *Mentha piperita* L. and *Mentha spicata* L. on liver tissue in rats. *Hum Exp Toxicol* **23**: 21–28.
- Akdogan M, Ozguner M, Kocak A, Oncu M, Cicek E. 2004c. Effects of peppermint teas on plasma testosterone, follicle-stimulating hormone, and luteinizing hormone levels and testicular tissue in rats. *Urology* **64**: 394–398.

- Akin M, Aktumsek A, Okur O. 2001. Investigation of antimicrobial activities of some essential oils against some animal pathogens. *Veterinarian* **12**: 83–85.
- Al-Abbadi A, Nazer IK. 2003. Control of Varroa mite (*Varroa destructor*) on honeybees by aromatic oils and plant materials. *SQU J Sci Res Agric Sci* **8**: 15–20.
- Andersen KE. 1978. Contact allergy to toothpaste flavors. *Contact Dermatitis* **4**: 195–198.
- Andersen PH, Jensen NJ. 1984. Mutagenic investigation of peppermint oil in the Salmonella/mammalian-microsome test. *Mutat Res* **138**: 17–20.
- Ando S, Nishida T, Ishida M, Hosoda K, Bayaru E. 2003. Effect of peppermint feeding on the digestibility, ruminal fermentation and protozoa. *Livest Prod Sci* **82**: 245–248.
- Anonymous. 2001. Final report on the safety assessment of *Mentha piperita* (peppermint) oil, *Mentha piperita* (peppermint) leaf extract, *Mentha piperita* (peppermint) leaf, and *Mentha piperita* (peppermint) leaf water. *Int J Toxicol* **20**: 61–73.
- Arakawa T, Osawa K. 2000. Pharmacological study and application to food of mint flavour – antibacterial and antiallergic principles. *Aroma Res* **1**: 20–23.
- Arakawa T, Shibata M, Hosomi K *et al.* 1992. Anti-allergic effects of peppermint oil, chicle and jelutong. [Japanese]. *Shokuhin Eiseigaku Zasshi* **33**: 569–575.
- Araujo C, Sousa MJ, Ferreira MF, Leao C. 2003. Activity of essential oils from Mediterranean Lamiaceae species against food spoilage yeasts. *J Food Prot* **66**: 625–632.
- Areias FM, Valentao P, Andrade PB, Ferreres F, Seabra RM. 2001. Phenolic fingerprint of peppermint leaves. *Food Chem* **73**: 307–311.
- Aridogan BC, Baydar H, Kaya S, Demirci M, Ozbasar D, Mumcu E. 2002. Antimicrobial activity and chemical composition of some essential oils. *Arch Pharm Res* **25**: 860–864.
- Asao T, Kuwano H, Ide M *et al.* 2003. Spasmolytic effect of peppermint oil in barium during double-contrast barium enema compared with Buscopan. *Clin Radiol* **58**: 301–305.
- Asao T, Mochiki E, Suzuki H *et al.* 2001. An easy method for the intraluminal administration of peppermint oil before colonoscopy and its effectiveness in reducing colonic spasm. *Gastrointest Endosc* **53**: 172–177.
- Atta AH, Alkofahi A. 1998. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. *J Ethnopharmacol* **60**: 117–124.
- Azuma K, Ito H, Ippoushi K, Higashio H. 2003. *In vitro* antibacterial activity of extracts from four Labiatae herbs against *Helicobacter pylori* and *Streptococcus mutans*. *Bull Nat Inst Veg Tea Sci* **2**: 83–91.
- Beesley A, Hardcastle J, Hardcastle PT, Taylor CJ. 1996. Influence of peppermint oil on absorptive and secretory processes in rat small intestine. *Gut* **39**: 214–219.
- Blanco MCSG, Ming LC, Marques MOM, Bovi OA. 2002. Drying temperature effects in peppermint essential oil content and composition. *Acta Hort* **569**: 95–98.
- Blaszczuk T, Krzyzanowska J, Lamer-Zarawska E. 2000. Screening for antimycotic properties of 56 traditional Chinese drugs. *Phytother Res* **14**: 210–212.
- Blumenthal M, Busse WR, Goldberg A *et al.* (eds). 1998. *The Complete German Commission E Monographs – Therapeutic Guide to Herbal Medicines*. American Botanical Council: Austin.
- Bonamonte D, Mundo L, Daddabbo M, Foti C. 2001. Allergic contact dermatitis from *Mentha spicata* (spearmint). *Contact Dermatitis* **45**: 298.
- Bonyadian M, Karim G. 2002. Study of the effects of some volatile oils of herbs (pennyroyal, peppermint, tarragon, caraway seed and thyme) against *E. coli* and *S. aureus* in broth media. *J Fac Vet Med Univ Tehran* **57**: e81–e83.
- Capecka E, Mareczek A, Leja M. 2005. Antioxidant activity of fresh and dry herbs of some Lamiaceae species. *Food Chem* **93**: 223–226.
- Carvalho JCT, Vignoli VV, de Souza GHB, Ujikawa K, Neto JJ. 1999. Antimicrobial activity of essential oils from plants used in Brazilian popular medicine. *Acta Hort* **501**: 77–81.
- Chiyotani A, Tamaoki J, Takeuchi S, Kondo M, Isono K, Konno K. 1994. Stimulation by menthol of Cl secretion via a Ca(2+)-dependent mechanism in canine airway epithelium. *Br J Pharmacol* **112**: 571–575.
- Choi WI, Lee EH, Choi BR, Park HM, Ahn YJ. 2003. Toxicity of plant essential oils to *Trialeurodes vaporariorum* (Homoptera: Aleyrodidae). *J Econ Entomol* **96**: 1479–1484.
- Clark RJ, Menary RC. 1981. Variations in composition of peppermint oil in relation to production areas. *Econ Bot* **35**: 59–69.
- Dalvi SS, Nadkarni PM, Pardesi R, Gupta KC. 1991. Effect of peppermint oil on gastric emptying in man: a preliminary study using a radiolabelled solid test meal. *Indian J Physiol Pharmacol* **35**: 212–214.
- Damayanti M, Susheela K, Sharma GJ. 1996. Effect of plant extracts and systemic fungicide on the pineapple fruit-rotting fungus, *Ceratocystis paradoxa*. *Cytobios* **86**: 155–165.
- De Almeida-Muradian LB, Rios MD, Sasaki R. 1998. Determination of provitamin A of green leafy vegetables by high performance liquid chromatography and open column chromatography. *Boll Chim Farm* **137**: 290–294.
- Della Loggia R, Tubaro A, Lunder TL. 1990. Evaluation of some pharmacological activities of a peppermint extract. *Fitoterapia* **61**: 215–221.
- Dew H, Evans B, Rhodes J. 1984. Peppermint oil for the irritable bowel syndrome: A multicentre trial. *Br J Clin Pract* **38**: 394, 398.
- Dimandja JMD, Stanfill S, Grainger J, Patterson DG Jr. 2000. Application of comprehensive two-dimensional gas chromatography (GCxGC) to the qualitative analysis of essential oils. *J High Resolut Chromatogr* **23**: 208–214.
- Dooms-Goossens A, Degreef H, Holvoet C, Maertens M. 1977. Turpentine-induced hypersensitivity to peppermint oil. *Contact Dermatitis* **3**: 304–308.
- Dragland S, Senoo H, Wake K, Holte K, Blomhoff R. 2003. Several culinary and medicinal herbs are important sources of dietary antioxidants. *J Nutr* **133**: 128–190.
- Dresser GK, Wachter V, Wong S, Wong HT, Bailey DG. 2002. Evaluation of peppermint oil and ascorbyl palmitate as inhibitors of cytochrome P4503A4 activity *in vitro* and *in vivo*. *Clin Pharmacol Ther* **72**: 247–255.
- Duarte MCT, Figueira GM, Sartoratto A, Rehder VLG, Delarmelina C. 2005. Anti-candida activity of Brazilian medicinal plants. *J Ethnopharmacol* **97**: 305–311.
- Duband F, Carnat AP, Carnat A, Petitjean-Freytet C, Clair G, Lamaison JL. 1992. [Aromatic and polyphenolic composition of infused peppermint, *Mentha × piperita* L.]. *Ann Pharm Fr* **50**: 146–155.
- Dubick MA. 1996. Historical perspectives on the use of herbal preparations to promote health. *J Nutr* **116**: 1348–1354.
- Eccles R, Griffiths DH, Newton CG, Tolley NS. 1988. The effects of menthol isomers on nasal sensation of airflow. *Clin Otolaryngol Allied Sci* **13**: 25–29.
- Eccles R, Jawad MS, Morris S. 1990. The effects of oral administration of (–)-menthol on nasal resistance to airflow and nasal sensation of airflow in subjects suffering from nasal congestion associated with the common cold. *J Pharm Pharmacol* **42**: 652–654.
- Eccles R, Jones AS. 1983. The effect of menthol on nasal resistance to air flow. *J Laryngol Otol* **97**: 705–709.
- Edris AE, Farrag ES. 2003. Antifungal activity of peppermint and sweet basil essential oils and their major aroma constituents on some plant pathogenic fungi from the vapor phase. *Nahrung* **47**: 117–121.
- Ezzat SM. 2001. *In vitro* inhibition of *Candida albicans* growth by plant extracts and essential oils. *World J Microbiol Biotechnol* **17**: 757–759.
- Freise J, Kohler S. 1999. Peppermint oil/caraway oil-fixed combination in non-ulcer dyspepsia. Equivalent efficacy of the drug combination in an enteric coated or enteric soluble formulation. *Pharmazie* **54**: 210–215.
- Furuhata K, Dogasaki C, Hara M, Fukuyama M. 2000. Antibacterial activities of several herbs on *Legionella pneumophila*. *J Azabu Univ* **1/2**: 15–20.
- Galeotti N, Ghelardini C, di Cesare Mannelli L, Mazzanti G, Baghiroli L, Bartolini A. 2001. Local anaesthetic activity of (+)- and (–)-menthol. *Planta Med* **67**: 174–176.
- Gherman C, Culea M, Cozar O. 2000. Comparative analysis of some active principles of herb plants by GC/MS. *Talanta* **53**: 253–262.
- Giamperi L, Fraternali D, Ricci D. 2002. The *in vitro* action of essential oils on different organisms. *J Essent Oil Res* **14**: 312–318.

- Gobel H, Fresenius J, Heinze A, Dworschak M, Soyka D. 1996. [Effectiveness of *Oleum menthae piperitae* and paracetamol in therapy of headache of the tension type]. *Nervenarzt* **67**: 672–681.
- Gobel H, Schmidt G, Dworschak M, Stolze H, Heuss D. 1995. Essential plant oils and headache mechanisms. *Phyto-medicine* **2**: 93–102.
- Gobel H, Schmidt G, Soyka D. 1994. Effect of peppermint and eucalyptus oil preparations on neurophysiological and experimental algometric headache parameters. *Cephalalgia* **14**: 228–234.
- Goerg KJ, Spilker T. 2003. Effect of peppermint oil and caraway oil on gastrointestinal motility in healthy volunteers: a pharmacodynamic study using simultaneous determination. *Aliment Pharmacol Ther* **17**: 445–452.
- Gomes-Carneiro MR, Felzenszwalb I, Paumgarten FJR. 1998. Mutagenicity testing of (\pm)-camphor, 1,8-cineole, citral, citronellal, (–)-menthol and terpineol with the Salmonella/microsome assay. *Mutat Res* **416**: 129–136.
- Guerin JC, Reveillere HP. 1985. Antifungal activity of plant extracts used in therapy. II. Study of 40 plant extracts against 9 fungal species. [French]. *Ann Pharm Fr* **43**: 77–81.
- Gurdip S, Kapoor IPS, Pandey SK. 1998. Studies on essential oils – part thirteen: natural antioxidant for sunflower oil. *J Sci Ind Res* **57**: 139–142.
- Guynot ME, Ramos AJ, Seto L, Purroy P, Sanchis V, Marin S. 2003. Antifungal activity of volatile compounds generated by essential oils against fungi commonly causing deterioration of bakery products. *J Appl Microbiol* **94**: 893–899.
- Hawthorn M, Ferrante J, Luchowski E, Rutledge A, Wei XY, Triggler DJ. 1988. The actions of peppermint oil and menthol on calcium channel dependent processes in intestinal, neuronal and cardiac preparations. *Aliment Pharmacol Ther* **2**: 101–118.
- Herrmann EC Jr, Kucera LS. 1967. Antiviral substances in plants of the mint family (Labiatae). 3. Peppermint (*Mentha piperita*) and other mint plants. *Proc Soc Exp Biol Med* **124**: 874–878.
- Hiki N, Kurosaka H, Tatsutomi Y *et al.* 2003. Peppermint oil reduces gastric spasm during upper endoscopy: a randomized, double-blind, double-dummy controlled trial. *Gastrointest Endosc* **57**: 475–482.
- Hills JM, Aaronson PI. 1991. The mechanism of action of peppermint oil on gastrointestinal smooth muscle: An analysis using patch clamp electrophysiology and isolated tissue pharmacology in rabbit and guinea pig. *Gastroenterology* **101**: 55–65.
- Hirazawa N, Ohtaka T, Hata K. 2000. Challenge trials on the anthelmintic effect of drugs and natural agents against the monogenean *Heterobothrium okamotoi* in the tiger puffer *Takifugu rubripes*. *Aquaculture* **188**: 1–13.
- Hoffmann BG, Lunder LT. 1984. Flavonoids from *Mentha piperita* leaves. *Planta Med* **50**: 361.
- Hurrell RF, Reddy M, Cook JD. 1999. Inhibition of non-haem iron absorption in man by polyphenolic-containing beverages. *Br J Nutr* **81**: 289–295.
- Ibrahim SAM, El-Ghamry AA, El-Mallah GM. 2000. Effect of some medicinal plants of Labiatae family as feed additives on growth and metabolic changes of rabbits. *Egypt J Rabbit Sci* **10**: 105–120.
- Imai H, Osawa K, Yasuda H, Hamashima H, Arai T, Sasatsu M. 2001. Inhibition by the essential oils of peppermint and spearmint of the growth of pathogenic bacteria. *Microbios* **106**: 31–39.
- Inoue T, Sugimoto Y, Masuda H, Kamei C. 2001. Effects of peppermint (*Mentha piperita* L.) extracts on experimental allergic rhinitis in rats. *Biol Pharm Bull* **24**: 92–95.
- Inoue T, Sugimoto Y, Masuda H, Kamei C. 2002. Antiallergic effect of flavonoid glycosides obtained from *Mentha piperita* L. *Biol Pharm Bull* **25**: 256–259.
- Inouye S, Yamaguchi H, Takizawa T. 2001. Screening of the antibacterial effects of a variety of essential oils on respiratory tract pathogens, using a modified dilution assay method. *J Infect Chemother* **7**: 251–254.
- Iskan G, Kirimer N, Kurkcuoğlu M, Husnu Can Baser K, Demirci F. 2002. Antimicrobial screening of *Mentha piperita* essential oils. *J Agric Food Chem* **50**: 3943–3946.
- Juergens UR, Dethlefsen U, Steinkamp G, Gillissen A, Repges R, Vetter H. 2003. Antiinflammatory activity of 1.8-cineole (eucalyptol) in bronchial asthma: a double-blind placebo-controlled trial. *Respir Med* **97**: 250–256.
- Juergens UR, Stober M, Schmidt-Schilling L, Kleuver T, Vetter H. 1998a. The anti-inflammatory activity of L-menthol compared to mint oil in human monocytes *in vitro*: a novel perspective for its therapeutic use in inflammatory diseases. *Eur J Med Res* **3**: 539–545.
- Juergens UR, Stober M, Vetter H. 1998b. Inhibition of cytokine production and arachidonic acid metabolism by eucalyptol (1.8-cineole) in human blood monocytes *in vitro*. *Eur J Med Res* **3**: 508–510.
- Juergens UR, Stober M, Vetter H. 1998c. Antiinflammatory effects of eucalyptol (1.8-cineole) in bronchial asthma: inhibition of arachidonic acid metabolism in human blood monocytes *ex vivo*. *Eur J Med Res* **3**: 407–412.
- Kamei C, Inoue T, Sugimoto Y, Masuda H. 2000. Effects of peppermint extracts on experimental allergic rhinitis in rats. *Aroma Res* **1**: 61–66.
- Kanerva L, Rantanen T, Aalto-Korte K *et al.* 2001. A multicenter study of patch test reactions with dental screening series. *Am J Contact Dermat* **12**: 83–87.
- Karanika MS, Komaitis M, Aggelis G. 2001. Effect of aqueous extracts of some plants of Lamiaceae family on the growth of *Yarrowia lipolytica*. *Int J Food Microbiol* **64**: 175–181.
- Kaul PN, Bhattacharya AK, Singh K, Rao BRR, Mallavarapu GR, Ramsh S. 2001. Chemical investigation of peppermint (*Mentha piperita* L.) oil. *J Essent Oil Bear Plants* **4**: 55–58.
- Kim SM, Cho YS, Park S. 2002. Cytotoxicity of methanol extracts of edible herbs against L1210 cells with the changes of antioxidant enzymes activities. [Korean]. *Korea J Pharmacog* **33**: 376–383.
- Kline R, Kline J, Di Palma J, Barbero G. 2001. Enteric-coated, pH-dependent peppermint oil capsules for the treatment of irritable bowel syndrome in children. *J Pediatr* **138**: 125–128.
- Koo HN, Jeong HJ, Kim CH *et al.* 2001. Inhibition of heat shock-induced apoptosis by peppermint oil in astrocytes. *J Mol Neurosci* **17**: 391–396.
- Kristiansen E, Madsen C. 1995. Induction of protein droplet (alpha 2 mu-globulin) nephropathy in male rats after short-term dosage with 1,8-cineole and l-limonene. *Toxicol Lett* **80**: 147–152.
- Larsen T, Fiehn N, Ostergaard E. 1996. The susceptibility of dental plaque bacteria to the herbs included in Longo Vital(R). *Microb Ecol Health Dis* **9**: 91–95.
- Laude EA, Morice AH, Grattan TJ. 1994. The antitussive effects of menthol, camphor and cineole in conscious guinea-pigs. *Pulm Pharmacol* **7**: 179–184.
- Lawson M, Knight R, Tran K, Walker G, Roberts-Thompson I. 1988. Failure of enteric-coated peppermint oil in the irritable bowel syndrome: A prospective randomized trial. *J Gastroenterol Hepatol* **3**: 235–238.
- Lazutka JR, Mierauskiene J, Slapsyte G, Dedonyte V. 2001. Genotoxicity of dill (*Anethum graveolens* L.), peppermint (*Mentha x piperita* L.) and pine (*Pinus sylvestris* L.) essential oils in human lymphocytes and *Drosophila melanogaster*. *Food Chem Toxicol* **39**: 485–492.
- Lech Y, Olesen KM, Hey H, Rask-Pedersen E, Vilien M, Ostergaard O. 1988. [Treatment of irritable bowel syndrome with peppermint oil. A double-blind study with a placebo]. *Ugeskr Laeger* **150**: 2388–2389.
- Lee B, Lee S, Annis PC, Pratt SJ, Park B, Tumaalii F. 2002. Fumigant toxicity of essential oils and monoterpenes against the red flour beetle, *Tribolium castaneum* Herbst. *J Asia Pac Entomol* **5**: 237–240.
- Leicester RJ, Hunt RH. 1982. Peppermint oil to reduce colonic spasm during endoscopy. *Lancet* **2**: 989.
- Lin JP, Li YC, Lin WC, Hsieh CL, Chung JG. 2001. Effects of (–)-menthol on arylamine N-acetyltransferase activity in human liver tumor cells. *Am J Chin Med* **29**: 321–329.
- Liu JH, Chen GH, Yeh HZ, Huang CK, Poon SK. 1997. Enteric-coated peppermint-oil capsules in the treatment of irritable bowel syndrome: a prospective, randomized trial. *J Gastroenterol* **32**: 765–768.
- Lozak A, Soltyk K, Ostapczuk P, Fijalek Z. 2002. Determination of selected trace elements in herbs and their infusions. *Sci Total Environ* **289**: 33–40.
- Mabrouk SS, El-Shayeb NM. 1980. Inhibition of aflatoxin formation by some spices. *Z Lebensm Unters Forsch* **171**: 344–347.

- Madisch A, Heydenreich CJ, Wieland V, Hufnagel R, Hotz J. 1999. Treatment of functional dyspepsia with a fixed peppermint oil and caraway oil combination preparation as compared to cisapride: a multicenter, reference-controlled double-blind equivalence study. *Arzneimittelforschung* **49**: 925–932.
- Madisch A, Melderis H, Mayr G, Sassin I, Hotz J. 2001. Commercially available herbal preparation and its modified dispense in patients with functional dyspepsia. Results of a double-blind, placebo-controlled, randomized multicenter trial. *Z Gastroenterol* **39**: 511–517.
- Madsen C, Wurtzen G, Carstensen J. 1986. Short-term toxicity study in rats dosed with menthone. *Toxicol Lett* **32**: 147–152.
- Maffei M, Scannerini S. 1992. Seasonal variations in fatty acids from non-polar lipids of developing peppermint leaves. *Phytochemistry* **31**: 479–484.
- Mahady GB, Pendland SL, Stoia A *et al.* 2005. *In vitro* susceptibility of *Helicobacter pylori* to botanical extracts used traditionally for the treatment of gastrointestinal disorders. *Phytother Res* **19**: 988–991.
- Mahmood SA, Abbas NA, Rojas RL. 2003. Effects of aqueous extracts of peppermint, fennel, dill and cumin on isolated rabbit duodenum. *U Aden J Nat Appl Sci* **7**: 377–383.
- Maliakal PP, Wanwimolruk S. 2001. Effect of herbal teas on hepatic drug metabolizing enzymes in rats. *J Pharm Pharmacol* **53**: 1323–1329.
- Manolova N, Serkedjieva J, Ivanova V. 1995. Antiinfluenza activity of the plant preparation 'Broncho Pam'. *Fitoterapia* **66**: 223–226.
- Marino M, Bersani C, Comi G. 2001. Impedance measurements to study the antimicrobial activity of essential oils from Lamiaceae and Compositae. *Int J Food Microbiol* **67**: 187–195.
- Marinova EM, Yanishlieva NV. 1997. Antioxidative activity of extracts from selected species of the family Lamiaceae in sunflower oil. *Food Chem* **58**: 245–248.
- May B, Funk P, Schneider B. 2003. Peppermint oil and caraway oil in functional dyspepsia – efficacy unaffected by *H. pylori*. *Aliment Pharmacol Ther* **17**: 975–976.
- May B, Kohler S, Schneider B. 2000. Efficacy and tolerability of a fixed combination of peppermint oil and caraway oil in patients suffering from functional dyspepsia. *Aliment Pharmacol Ther* **14**: 1671–1677.
- May B, Kuntz HD, Kieser M, Kohler S. 1996. Efficacy of a fixed peppermint oil/caraway oil combination in non-ulcer dyspepsia. *Arzneimittelforschung* **46**: 1149–1153.
- Micklefield GH, Greving I, May B. 2000. Effects of peppermint oil and caraway oil on gastroduodenal motility. *Phytother Res* **14**: 20–23.
- Micklefield G, Jung O, Greving I, May B. 2003. Effects of intraduodenal application of peppermint oil (WS((R)) 1340) and caraway oil (WS((R)) 1520) on gastroduodenal motility in healthy volunteers. *Phytother Res* **17**: 135–140.
- Miki S, Kinogiri M, Izaki Y, Okura M, Ikuta T. 1997. The effect of odours of lavender and peppermint on the human SEP (Somatosensory Evoked Potential) and EEG. [Japanese]. *Shikoku Acta Med* **53**: 248–257.
- Mimica-Dukic N, Bozin B, Sokovic M, Mihajlovic B, Matavulj M. 2003. Antimicrobial and antioxidant activities of three *Mentha* species essential oils. *Planta Med* **69**: 413–419.
- Minami M, Kita M, Nakaya T, Yamamoto T, Kuriyama H, Imanishi J. 2003. The inhibitory effect of essential oils on herpes simplex virus type-1 replication *in vitro*. *Microbiol Immunol* **47**: 681–684.
- Montes-Belmont R, Carvajal M. 1998. Control of *Aspergillus flavus* in maize with plant essential oils and their components. *J Food Prot* **61**: 616–619.
- Montes Belmont R, Flores Moctezuma HE. 2001. Sorghum seeds treated with natural products for the control of *Fusarium thapsinum* and *Claviceps africana*. [Spanish]. *Manejo Integrado de Plagas* **61**: 23–30.
- Morice AH, Marshall AE, Higgins KS, Grattan TJ. 1994. Effect of inhaled menthol on citric acid induced cough in normal subjects. *Thorax* **49**: 1024–1026.
- Morton CA, Garioch J, Todd P, Lamey PJ, Forsyth A. 1995. Contact sensitivity to menthol and peppermint in patients with intra-oral symptoms. *Contact Dermatitis* **32**: 281–284.
- Muller M, Anke M, Illing-Gunther H. 1997. Availability of aluminium from tea and coffee. *Z Lebensm Unters Forsch* **205**: 170–173.
- Murthy PBK, Ahmed MM, Regu K. 1991. Lack of genotoxicity of menthol in chromosome aberration and sister chromatid exchange assays using human lymphocytes *in vitro*. *Toxicol In Vitro* **5**: 337–340.
- Nair B. 2001. Final report on the safety assessment of *Mentha piperita* (peppermint) oil, *Mentha piperita* (peppermint) leaf extract, *Mentha piperita* (peppermint) leaf, and *Mentha piperita* (peppermint) leaf water. *Int J Toxicol* **20**: 61–73.
- Naito K, Komori M, Kondo Y, Takeuchi M, Iwata S. 1997. The effect of L-menthol stimulation of the major palatine nerve on subjective and objective nasal patency. *Auris Nasus Larynx* **24**: 159–162.
- Naito K, Ohoka E, Kato R, Kondo Y, Iwata S. 1991. The effect of L-menthol stimulation of the major palatine nerve on nasal patency. *Auris Nasus Larynx* **18**: 221–226.
- Nash P, Gould S, Barnardo D. 1986. Peppermint oil does not relieve the pain of irritable bowel syndrome. *Br J Clin Pract* **40**: 292–293.
- Natake M, Kanazawa K, Mizuno M *et al.* 1989. Herb water-extracts markedly suppress the mutagenicity of Trp-P-2. *Agric Biol Chem* **53**: 1423–1425.
- Nelson RRS. 1997. In-vitro activities of five plant essential oils against methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant *Enterococcus faecium*. *J Antimicrob Chemother* **40**: 305–306.
- Nishino T, Tagaito Y, Sakurai Y. 1997. Nasal inhalation of L-menthol reduces respiratory discomfort associated with loaded breathing. *Am J Respir Crit Care Med* **156**: 309–313.
- Ohara A, Matsuhisa T. 2002. Anti-tumor promoting activities of edible plants against okadaic acid. *Food Sci Technol Res* **8**: 158–161.
- Pattnaik S, Subramanyam VR, Kole C. 1996. Antibacterial and antifungal activity of ten essential oils *in vitro*. *Microbios* **86**: 237–246.
- Pattnaik S, Subramanyam VR, Rath CC. 1995. Effect of essential oils on the viability and morphology of *Escherichia coli* (SP-11). *Microbios* **84**: 195–199.
- Pazdziuch-Czochra M, Widencka A. 2002. Spectrofluorimetric determination of hydrogen peroxide scavenging activity. *Anal Chim Acta* **452**: 177–184.
- Piccaglia R, Marotti M, Giovanelli E, Deans SG, Eaglesham E. 1993. Antibacterial and antioxidant properties of Mediterranean aromatic plants. *Ind Crop Prod* **2**: 47–50.
- Picuric-Jovanovic K, Milovanovic M, Poludnyonny LV. 1997. Chemical composition of essential oils of several wild-growing species of *Mentha piperita* L. *Zbornik Radova Poljop Fak* **42**: 243–248.
- Pilipenko LN, Oleinik LB, Kozhukhar VV. 1998. Liposoluble pigments of food plants and their extracts. *Chem Nat Compd* **34**: 269–271.
- Pimentel M, Bonorris GG, Chow EJ, Lin HC. 2001. Peppermint oil improves the manometric findings in diffuse oesophageal spasm. *J Clin Gastroenterol* **33**: 27–31.
- Pino JA, Borges P, Martinez MA *et al.* 2002. Essential oil of *Mentha piperita* L. grown in Jalisco. *J Essent Oil Res* **14**: 189–190.
- Pittler MH, Ernst E. 1998. Peppermint oil for irritable bowel syndrome: a critical review and metaanalysis. *Am J Gastroenterol* **93**: 1131–1135.
- Rai MK, Upadhyay S. 1988. Laboratory evaluation of essential oil of *Mentha piperita* Linn. against *Trichophyton mentagrophytes*. *Hindustan Antibiot Bull* **30**: 82–84.
- Rees W, Evans B, Rhodes J. 1979. Treating irritable bowel syndrome with peppermint oil. *Br Med J* **6**: 835–836.
- Rohloff J. 1999. Monoterpene composition of essential oil from peppermint (*Mentha × piperita* L.) with regard to leaf position using solid-phase microextraction and gas chromatography/mass spectrometry analysis. *J Agric Food Chem* **47**: 3782–3786.
- Rosch W, Vinson B, Sassin I. 2002. A randomised clinical trial comparing the efficacy of a herbal preparation STW 5 with the prokinetic drug cisapride in patients with dysmotility type of functional dyspepsia. *Z Gastroenterol* **40**: 401–408.
- Ruiz del Castillo ML, Santa-Maria G, Herraiz M, Blanch GP. 2003. A comparative study of the ability of different techniques to extract menthol from *Mentha piperita*. *J Chromatogr Sci* **41**: 385–389.

- Samarth RM, Goyal PK, Ashok K. 2001a. Radioprotective effects of *Mentha piperita*. *J Med Aromatic Plant Sci* **22**: 91–97.
- Samarth RM, Goyal PK, Ashok K. 2004. Protection of Swiss albino mice against whole-body gamma irradiation by *Mentha piperita* (Linn.). *Phytother Res* **18**: 546–550.
- Samarth RM, Goyal PK, Kumar A. 2001b. Modulatory effect of *Mentha piperita* (Linn.) on serum phosphatases activity in Swiss albino mice against gamma irradiation. *Indian J Exp Biol* **39**: 479–482.
- Samarth RM, Goyal PK, Kumar A. 2002a. Modulation of serum phosphatases activity in Swiss albino mice against gamma irradiation by *Mentha piperita* Linn. *Phytother Res* **16**: 586–589.
- Samarth RM, Kumar A. 2003. Radioprotection of Swiss albino mice by plant extract *Mentha piperita* (Linn.). *J Radiat Res* **44**: 101–109.
- Samarth RM, Saini MR, Maharwal J, Dhaka A, Kumar A. 2002b. *Mentha piperita* (Linn) leaf extract provides protection against radiation induced alterations in intestinal mucosa of Swiss albino mice. *Indian J Exp Biol* **40**: 1245–1249.
- Sameena Y, Ashok K. 2001. Evaluation of chemoprevention of skin papilloma by *Mentha piperita*. *J Med Aromatic Plant Sci* **22**: 84–88.
- Samejima K, Kanazawa K, Ashida H, Danno G. 1995. Luteolin: a strong antimutagen against dietary carcinogen, Trp-P-2, in peppermint, sage, and thyme. *J Agric Food Chem* **43**: 410–414.
- Samman MA, Bowen ID, Taiba K, Antonius J, Hannan MA. 1998. Mint prevents shamma-induced carcinogenesis in hamster cheek pouch. *Carcinogenesis* **19**: 1795–1801.
- Sang JP. 1982. Estimation of menthone, menthofuran, menthyl acetate and menthol in peppermint oil by capillary gas chromatography. *J Chromatogr* **253**: 109–112.
- Sarbhoy AK, Varshney JL, Maheshwari ML, Saxena DB. 1978. Efficacy of some essential oils and their constituents on few ubiquitous molds. *Zentralbl Bakteriell Naturwiss* **133**: 723–725.
- Satoh T, Sugawara Y. 2003. Effects on humans elicited by inhaling the fragrance of essential oils: sensory test, multi-channel thermometric study and forehead surface potential wave measurement on basil and peppermint. *Anal Sci* **19**: 139–146.
- Satsu H, Matsuda T, Toshimitsu T *et al.* 2004. Regulation of interleukin-8 secretion in human intestinal epithelial Caco-2 cells. *Biofactors* **21**: 137–139.
- Schuhmacher A, Reichling J, Schnitzler P. 2003. Virucidal effect of peppermint oil on the enveloped viruses herpes simplex virus type 1 and type 2 *in vitro*. *Phytomedicine* **10**: 504–510.
- Shapiro S, Meier A, Guggenheim B. 1994. The antimicrobial activity of essential oils and essential oil components towards oral bacteria. *Oral Microbiol Immunol* **9**: 202–208.
- Sparks MJ, O'Sullivan P, Herrington AA, Morcos SK. 1995. Does peppermint oil relieve spasm during barium enema? *Br J Radiol* **68**: 841–843.
- Spindler P, Madsen C. 1992. Subchronic toxicity study of peppermint [*Mentha piperita* and *Mentha arvensis*] oil in rats. *Toxicol Lett* **62**: 215–220.
- Swain AR, Dutton SP, Truswell AS. 1985. Salicylates in foods. *J Am Diet Assoc* **85**: 950–960.
- Takacs AL, Vore M. 1987. Binding of 3H-estradiol-17 beta-(beta-D-glucuronide), a cholestatic organic anion, to rat liver plasma membranes. Evidence consonant with identification of organic anion carriers. *Mol Pharmacol* **32**: 511–518.
- Takeuchi S, Tamaoki J, Kondo M, Konno K. 1994. Effect of menthol on cytosolic Ca²⁺ levels in canine airway epithelium in culture. *Biochem Biophys Res Commun* **201**: 1333–1338.
- Tamaoki J, Chiyotani A, Sakai A, Takemura H, Konno K. 1995. Effect of menthol vapour on airway hyperresponsiveness in patients with mild asthma. *Respir Med* **89**: 503–504.
- Tampieri MP, Galuppi R, Macchioni F *et al.* 2005. The inhibition of *Candida albicans* by selected essential oils and their major components. *Mycopathologia* **159**: 339–345.
- Tassou CC, Drosinos EH, Nychas GJ. 1995. Effects of essential oil from mint (*Mentha piperita*) on *Salmonella enteritidis* and *Listeria monocytogenes* in model food systems at 4 degrees and 10 degrees C. *J Appl Bacteriol* **78**: 593–600.
- Tassou C, Koutsoumanis K, Nychas GJE. 2000. Inhibition of *Salmonella enteritidis* and *Staphylococcus aureus* in nutrient broth by mint essential oil. *Food Res Intern* **33**: 273–280.
- Tate S. 1997. Peppermint oil: a treatment for postoperative nausea. *J Adv Nurs* **26**: 543–549.
- Thompson Coon J, Ernst E. 2002. Systematic review: herbal medicinal products for non-ulcer dyspepsia. *Aliment Pharmacol Ther* **16**: 1689–1699.
- Thorup I, Wurtzen G, Carstensen J, Olsen P. 1983a. Short term toxicity study in rats dosed with peppermint oil. *Toxicol Lett* **19**: 211–215.
- Thorup I, Wurtzen G, Carstensen J, Olsen P. 1983b. Short term toxicity study in rats dosed with pulegone and menthol. *Toxicol Lett* **19**: 207–210.
- Tkachenko KG, Kazarinova NV, Muzychenko LM, Shurgaya AM, Pavlova OV, Safonova NG. 1999. Sanitation properties of essential oils of some plant species. *Rastitel'Nye Resursy* **35**: 11–24.
- Trabace L, Avato P, Mazzoccoli M, Siro-Brigiani G. 1992. Choleric activity of some typical components of essential oils. *Planta Med* **58**: A650–A651.
- Trabace L, Avato P, Mazzoccoli M, Siro-Brigiani G. 1994. Choleric activity of Thapsia Chem I, II, and III in rats: comparison with terpenoid constituents and peppermint oil. *Phytother Res* **8**: 305–307.
- Uegaki R, Ando S, Ishida M, Takada O, Shinokura K, Kohchi Y. 2001. Antioxidative activity of milk from cows fed herbs. *Nippon Nogeikagaku Kaishi* **75**: 669–671.
- Uehleke B, Silberhorn H, Wohling H. 2002. A plant cocktail soothes upset stomachs. *MMW Fortschr Med* **144**: 695.
- Umezu T. 2002. Pharmacological effects of plant-derived essential oils on the central nervous system. *Aroma Res* **3**: 376–382.
- Umezu T. 2003. Evidence for involvement of dopamine in the ambulation promoted by menthol in mice. *Aroma Res* **4**: 27–34.
- Umezu T, Sakata A, Ito H. 2001. Ambulation-promoting effect of peppermint oil and identification of its active constituents. *Pharmacol Biochem Behav* **69**: 383–390.
- Unger M, Frank A. 2004. Simultaneous determination of the inhibitory potency of herbal extracts on the activity of six major cytochrome P450 enzymes using liquid chromatography/mass spectrometry and automated online extraction. *Rapid Commun Mass Spectrom* **18**: 2273–2281.
- Ushid K, Maekawa M, Arakawa T. 2002. Influence of dietary supplementation of herb extracts on volatile sulfur production in pig large intestine. *J Nutr Sci Vitaminol* **48**: 18–23.
- Venema DP, Hollman PCH, Janssen KPLTM, Katan MB. 1996. Determination of acetylsalicylic acid and salicylic acid in foods, using HPLC with fluorescence detection. *J Agric Food Chem* **44**: 1762–1767.
- Vo LT, Chan D, King RG. 2003. Investigation of the effects of peppermint oil and valerian on rat liver and cultured human liver cells. *Clin Exp Pharmacol Physiol* **30**: 799–804.
- Wacher VJ, Wong S, Wong HT. 2002. Peppermint oil enhances cyclosporine oral bioavailability in rats: comparison with D-alpha-tocopheryl poly(ethylene glycol 1000) succinate (TPGS) and ketoconazole. *J Pharm Sci* **91**: 77–90.
- Westphal J, Horning M, Leonhardt K. 1996. Phytotherapy in functional upper abdominal complaints. Results of a clinical study with a preparation of several plants. *Phytomedicine* **2**: 285–291.
- Wildgrube HJ. 1988. Untersuchungen zur Wirksamkeit von Pfefferminzöl auf Beschwerdebild und funktionelle Parameter bei Patienten mit Reizdarm-Syndrom (Studie). *Naturheilpraxis* **41**: 591–596.
- Wright CE, Bowen WP, Grattan TJ, Morice AH. 1998. Identification of the L-menthol binding site in guinea-pig lung membranes. *Br J Pharmacol* **123**: 481–486.
- Wright CE, Laude EA, Grattan TJ, Morice AH. 1997. Capsaicin and neurokinin A-induced bronchoconstriction in the anaesthetised guinea-pig: evidence for a direct action of menthol on isolated bronchial smooth muscle. *Br J Pharmacol* **121**: 1645–1650.
- Xu P, Jia W, Bi L, Liu X, Zhao Y. 2003. Studies on components and quality of essential oil from *Mentha piperita* L. produced in Xinjiang, China. *Chem Ind Forest Prod* **23**: 43–45.
- Yamasaki K, Nakano M, Kawahata T *et al.* 1998. Anti-HIV-1 activity of herbs in Labiatae. *Biol Pharm Bull* **21**: 829–833.

- Yanishlieva NV, Marinova EM. 1995. Antioxidant activity of selected species of the family Lamiaceae grown in Bulgaria. *Nahrung* **39**: 458–463.
- Zakharov AM, Zakharova OI, Smirnova LP. 1990. Flavonoids of *Mentha piperita*, variety Krasnodarskaya 2. *Chem Nat Compd* **26**: 96.
- Zambonelli A, Zechini D'Aulerio A, Bianchi A, Albasini A. 1996. Effects of essential oils on phytopathogenic fungi *in vitro*. *J Phytopathol* **144**: 491–494.
- Zhang X. 2002. *WHO Traditional Medicine Strategy 2002–2005*. World Health Organization: Geneva, Switzerland.
- Zheng W, Wang SY. 2001. Antioxidant activity and phenolic compounds in selected herbs. *J Agric Food Chem* **49**: 5165–5170.
- Zimna D, Piekos R. 1988. Extraction of eight essential elements from the leaves of peppermint, *Mentha piperita* (L.) Huds. *Herba Hungar* **27**: 65–75.