

links between biomarker structure and biochemical function.

Similarly, geologists have much to offer evolutionary biology by helping constrain the time period and physical context of the appearance of new life forms.

References

1. Snow, C.P. (1959). The Two Cultures and the Scientific Revolution. *Encounter* 12, 17–24.
2. Canfield, D.E. (2005). The early history of atmospheric oxygen: Homage to Robert A. Garrels. *Annu. Rev. Earth Planet. Sci.* 33, 1–36.
3. Tice, M.M., and Lowe, D.R. (2006). Hydrogen-based carbon fixation in the earliest known photosynthetic organisms. *Geology* 34, 37–40.
4. Tian, F., Toon, O.B., Pavlov, A.A., and De Sterck, H. (2005). A hydrogen-rich early Earth atmosphere. *Science* 308, 1014–1017.
5. Des Marais, D.J., Strauss, H., Summons, R.E., and Hayes, J.M. (1992). Carbon isotope evidence for the stepwise oxidation of the Proterozoic environment. *Nature* 359, 605–609.
6. Anbar, A.D., and Knoll, A.H. (2002). Proterozoic ocean chemistry and evolution: a bioinorganic bridge? *Science* 297, 1137–1142.
7. Summons, R.E., Jahnke, L.L., Hope, J.M., and Logan, G.A. (1999). 2-Methylhopanoids as biomarkers for cyanobacterial oxygenic photosynthesis. *Nature* 400, 554–557.
8. Brocks, J.J., Logan, G.A., Buick, R., and Summons, R.E. (1999). Archean molecular fossils and the early rise of eukaryotes. *Science* 285, 1033–1036.
9. Brocks, J.J., Love, G.D., Summons, R.E., Knoll, A.H., Logan, G.A., and Bowden, S.A. (2005). Biomarker evidence for green and purple sulphur bacteria in a stratified Palaeoproterozoic sea. *Nature* 437, 866–870.
10. Rohmer, M., Bouvier, P., and Ourisson, G. (1979). Molecular evolution of biomembranes: structural equivalents and phylogenetic precursors of sterols. *Proc. Natl. Acad. Sci. USA* 76, 847–851.
11. Bloch, K. (1987). Summing-Up. *Annu. Rev. Biochem.* 56, 1–19.
12. Pike, L.J. (2004). Lipid rafts: heterogeneity on the high seas. *Biochem. J.* 378, 281–292.
13. Lindmark, D.G., and Muller, M. (1973). Hydrogenosome, a cytoplasmic organelle of the anaerobic flagellate *Trichomonas foetus*, and its role in pyruvate metabolism. *J. Biol. Chem.* 248, 7724–7728.
14. Boxma, B., de Graaf, R.M., van der Staay, G.W., van Alen, T.A., Ricard, G., Gabaldon, T., van Hoek, A.H., Moon-van der Staay, S.Y., Koopman, W.J., van Hellemond, J.J., et al. (2005). An anaerobic mitochondrion that produces hydrogen. *Nature* 434, 74–79.
15. Martin, W., and Muller, M. (1998). The hydrogen hypothesis for the first eukaryote. *Nature* 392, 37–41.
16. Javaux, E.J., Knoll, A.H., and Walter, M.R. (2001). Morphological and ecological complexity in early eukaryotic ecosystems. *Nature* 412, 66–69.

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Revisiting Neandertal diversity with a 100,000 year old mtDNA sequence

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The cohabitation of Neandertals and modern humans in Europe about 35,000 years ago has stimulated considerable debate regarding hypothetical admixture. Recently, sequences of the hypervariable region-1 (HVR-1) of mitochondrial DNA (mtDNA) from 9 Neandertal specimens dated between 29,000 and 42,000 years ago from dispersed locations have revealed the genetic diversity of Neandertals around the time of the cohabitation [1–4]. The genetic signatures before and after contact with modern humans were found to be similar. They fall outside the range of modern human genetic diversity and show no specific affinity with modern or Paleolithic Europeans [5]. Such observations are generally taken as strong evidence for the ‘Rapid replacement’ model for the origin of modern humans [4,6], though further evidence is needed to completely exclude admixture [7].

The first presence of modern humans in Europe before 35,000 years ago as well as the survival of Neandertals beyond 30,000 years ago are still controversial issues [9]. Our goal was to recover a Neandertal sequence that unambiguously predates the cohabitation period. A comparison of this sequence with published Neandertal sequences might reveal either the long-time stability of the Neandertal mtDNA-pool or drastic modifications around the time of cohabitation. We, therefore, retrieved 123 bp of the mtDNA HVR-1 from a 100,000 year old Neandertal tooth from the Scladina cave (Meuse Basin,

Belgium), which represents the most ancient Neandertal sample analyzed at the DNA level.

The experiments were conducted in a specific laboratory respecting the current authentication standards [10]. The extract was treated with uracil DNA-glycosylase (UDG) to excise deaminated cytosines formed after death, because they lead to artefactual GC→AT polymorphisms during PCR [11,12] and have already been shown to be present in sequences from Scladina fossils [13–15]. We took advantage of previously reported Neandertal sequences to design primers that favor the amplification of Neandertal DNA. PCR was never successful when fragments larger than 173 bp were targeted (Supplemental Data). We amplified four fragments spanning in total 221 bp of the HVR-1. Each PCR product was cloned and the final sequence was deduced from the consensus of 61 clones. Each position was found in at least two amplification products, except for the first 39 and last 59 nucleotides for which PCR replication was not possible. These nucleotides were consequently excluded from the sequence analyses. The remaining 123 bp (Figure 1) fulfilled all standards to guarantee the absence of DNA-damage-induced errors [10]. In addition, we are confident that the conditions in the Scladina cave favour DNA preservation, because an atomic C:N ratio typical of well-preserved collagen was found on the maxillary from the Scladina Neandertal [8], cave bear bones from the same excavation layer have already yielded authentic ancient DNA sequences [13,14] and 60,000–70,000 thousand year old nuclear DNA sequences were successfully amplified from woolly rhinoceroses from Scladina [15].

The Scladina Neandertal sequence has not been found among the 7161 human HVR-1 sequences present in the HvrBase++ [16]. It appears more distantly related to the human than to the already reported Neandertal sequences (Figure 1). Of the 123 nucleotides considered, only one polymorphic site (at position 16258) has already been

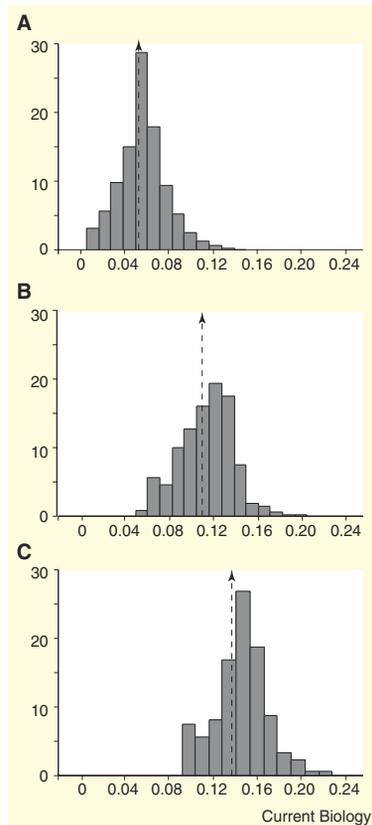


Figure 2. Pairwise distance distribution between modern humans and Neanderthals.

The distributions of pairwise distances expressed in terms of number of substitutions per site are estimated from the best-fitting model, within the *Homo sapiens* sequences (A, $n = 14535$), between the *Homo sapiens* and the first six Neanderthal sequences of Figure 1 except Scladina (B, $n = 855$), between the *Homo sapiens* and the Neanderthal sequence from Scladina (C, $n = 171$). The y-axis shows the percentage of the pairwise counts. The arrows show the means of each distribution. The means of (B) and (C) (0.108 and 0.136, respectively) are significantly different ($p < 0.001$). The distributions in (A) and (B) overlap extensively: 95% (and 99%) of this distribution depicted in (A) overlap with 21% (and 69%) of the distribution in (B). The extent of this overlap is drastically reduced to 7% (and 21%) of the distribution in (C). Actually, 94% of the distribution in (A) does not overlap with (C).

contribution to early modern humans. *PLoS Biol.* 2, E57.

5. Caramelli, D., Lalueza-Fox, C., Vernesi, C., Lari, M., Casoli, A., Mallegni, F., Chiarelli, B., et al. (2003). Evidence for a genetic discontinuity between Neanderthals and 24,000-year-old anatomically modern Europeans. *Proc. Natl. Acad. Sci. USA* 100, 6593–6597.
6. Currat, M., and Excoffier, L. (2004). Modern humans did not admix with Neanderthals during their range expansion into Europe. *PLoS Biol.* 2, E421.

7. Gutierrez, G., Sanchez, D., and Marin, A. (2002). A reanalysis of the ancient mitochondrial DNA sequences recovered from Neanderthal bones. *Mol. Biol. Evol.* 19, 1359–1366.
8. Bocherens, H., Billiou, D., and Mariotti, A. (1999). Palaeoenvironmental and palaeodietary implications of isotopic biochemistry of last interglacial Neanderthal and mammal bones in Scladina cave (Belgium). *J. Arc. Sci.* 26, 599–607.
9. Higham, T., Ramsey, C.B., Karavanic, I., Smith, F.H., and Trinkaus, E. (2006). Revised direct radiocarbon dating of the Vindija G1 Upper Paleolithic Neanderthals. *Proc. Natl. Acad. Sci. USA* 103, 553–557.
10. Gilbert, M.T.P., Bandelt, H.J., Hofreiter, M., and Barnes, I. (2005). Assessing ancient DNA studies. *Trends Ecol. Evol.* 20, 541–544.
11. Hofreiter, M., Jaenicke, V., Serre, D., von Haeseler, A., and Pääbo, S. (2001). DNA sequences from multiple amplifications reveal artifacts induced by cytosine deamination in ancient DNA. *Nucleic Acids Res.* 29, 4793–4799.
12. Gilbert, M.T., Hansen, A.J., Willerslev, E., Rudbeck, L., Barnes, I., Lynnerup, N., and Cooper, A. (2003). Characterization of genetic miscoding lesions caused by postmortem damage. *Am. J. Hum. Genet.* 72, 48–61.
13. Orlando, L., Bonjean, D., Bocherens, H., Thénot, A., Argant, A., Otte, M., and Hänni, C. (2002). Ancient DNA and the population genetics of cave bears (*Ursus spelaeus*) through space and time. *Mol. Biol. Evol.* 19, 1920–1933.
14. Loreille, O., Orlando, L., Patou-Mathis M., Philippe, M., Taberlet, P., and Hänni, C. (2001). Ancient DNA analysis reveals divergence of the cave bear, *Ursus spelaeus*, and brown bear, *Ursus arctos*, lineages. *Curr. Biol.* 11, 200–203.
15. Orlando, L., Leonard, J., Thénot, A., Laudet, V., Guérin, C., and Hänni, C. (2003). Ancient DNA analysis reveals woolly rhino evolutionary relationships. *Mol. Phylogenet. Evol.* 28, 485–499.
16. Handt, O., Meyer, S., and von Haeseler, A. (1998). Compilation of human mtDNA control region sequences. *Nucleic Acids Res.* 26, 126–129.
17. Bower, M.A., Spencer, M., Matsumura, S., Nisbet, R.E., and Howe, C. (2005). How many clones need to be sequenced from a single forensic or ancient DNA sample in order to determine a reliable consensus sequences? *Nucleic Acids Res.* 33, 2549–2556.

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Do angry men get noticed?

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In humans, the physical differences between the sexes are readily apparent, but possible cognitive and perceptual differences are less obvious. As social animals, humans have specialized mechanisms for recognizing facial expressions, but the extent to which these mechanisms are tuned to differences between male and female faces remains unclear. We measured the efficiency with which emotional expressions conveyed by male and female faces are detected by male and female observers. Angry male faces were detected significantly more rapidly by male than female observers. Moreover, detection of angry male faces by either male or female observers was scarcely affected by the addition of neutral distractor faces to the search display. Our findings are consistent with the notion of a perceptual system in both males and females that has evolved to rapidly detect aggression in males.

In humans, evolution has resulted in marked differentiation between males and females [1,2], including differences in the structural and functional organization of the brain. These differences are reflected in patterns of cognitive and behavioural abilities [3]. For example, females tend to perform better than males at fine motor and perceptual discrimination tasks, whereas males are better at route-finding tasks [3]. Males are also physically larger and more aggressive than females, and so more likely to pose a physical threat [4]. Such physical differences between the sexes may in turn have shaped the cognitive processes involved in detecting threatening behaviour in others. Early detection of an angry facial expression, for example, might reduce the