Induction of apoptosis and cell cycle arrest by a chalcone panduratin A isolated from *Kaempferia pandurata* in androgen-independent human prostate cancer cells PC3 and DU145

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Because of unsatisfactory treatment options for prostate cancer (CaP) there is a need to develop novel preventive approaches for this malignancy. One such strategy is through chemoprevention by the use of non-toxic dietary substances and botanical products. We have shown previously that panduratin A isolated from the extract of *Kaempferia pandurata* (Zingiberaceae) is a strong inhibitor of cyclooxygenase-2 in RAW264.7 cells and induces apoptosis in HT-29 cells. In the present study, we provide evidence that panduratin A treatment to androgen-independent human CaP cells PC3 and DU145 result in a time and dose-dependent inhibition of cell growth with an IC50 of 13.5–14 μM and no to little effect on normal human prostate epithelial cells. To define the mechanism of these anti-proliferative effects of panduratin A, we determined its effect on critical molecular events known to regulate the cell cycle and the apoptotic machinery. Annexin V/propidium iodide staining provided the evidence for the induction of apoptosis which was further confirmed by the observation of cleavage of poly (ADP-ribose) polymerase and degradation of acinus. Panduratin A treatment to cells was found to result in inhibition of procaspases 9, 8, 6 and 3 with significant increase in the mitochondrial-dependent apoptotic pathway. Panduratin A-mediated apoptosis was accompanied with upregulation of Fas death receptor and TNF-related apoptosis-inducing ligand (TRAIL). Furthermore, cell cycle analysis using flow cytometry showed that panduratin A treatment of cells resulted in a G2/M arrest in a dose-dependent manner. The immunoblot analysis data revealed that in both cell lines panduratin A treatment resulted in a dose-dependent (i) induction of p21WAF1/Cip1 and p27Kip1, (ii) downregulation of cdks 2, 4 and 6 and (iii) decrease in cyclins D1 and E. These findings suggest that panduratin A may be an effective chemopreventive or therapeutic agent against CaP.

Introduction

Prostate cancer (CaP) is the second most prevalent malignancy and the second leading cause of cancer-related deaths among men in the USA with similar trend in many western countries (1). Clinically, CaP is usually diagnosed in men over 50 years of age; with increasing life expectancy the incidence of CaP is likely to increase worldwide (1–2). Thus, there is a promising opportunity for its intervention using cancer chemopreventive compounds that can prevent or slow the progression of this disease (3–4). For a variety of reasons naturally occurring botanicals and dietary substances are gaining increasing attention as cancer chemopreventive agents. Important for CaP chemoprevention is the fact that in recent years, the use of dietary substances and botanical products is showing sustained increase by CaP patients. In CaP a fine balance between cell proliferation and apoptotic death is lost which contributes to increase in cellular mass and tumor progression. In this regard, for CaP chemoprevention at the present time there is considerable emphasis in identifying novel botanicals that selectively induces apoptosis and growth arrest of CaP cells without producing cytotoxic effects on normal cells.

The rhizome of *Kaempferia pandurata* Roxb. (Zingiberaceae), a herb cultivated in some tropical countries including Indonesia and Thailand, has been used as a condiment and folk medicine for treatment of various ailments, including colic disorder, fungal infections, dry cough, rheumatism and muscular pains (5–6). Since chalcones are natural products which have been reported to possess a variety of biological properties, including anti-inflammatory, analgesic and antioxidant activities (7). We recently isolated panduratin A (Figure 1), a cyclohexenyl chalcone derivative, from the methanolic extract of *K.pandurata* and reported that it exhibits strong COX-2 inhibitory activity in mouse peritoneal macrophages (8) and induces apoptosis in human colon cancer HT-29 cells (9). In the present study, we demonstrate antiproliferative and proapoptotic effect of panduratin A in human androgen-independent CaP cells, PC3 and DU145, and delineate the mechanism of this effect.

Abbreviations: CaP, prostate cancer; cdc2, cell division cycle 2 kinase; cdk, cyclin dependent kinase; cdkI, cdk inhibitor; FADD, fas-associated death domain; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; PARP, poly (ADP-ribose) polymerase; PC3 and DU145, androgen-independent human prostate carcinoma cells; PI, propidium iodide; pEC, normal human prostate epithelial cell line; TNF, tumor necrosis factor; TRAIL, TNF-related apoptosis-inducing ligand.

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MTT solution was removed from the wells by aspiration and the formazan vehicle-treated cells were taken as 100% viable. IC50 values were determined as described previously (8). Panduratin A (Figure 1) was identified by comparison of the spectral (496-NMR, 13C-NMR and FAB-MS) properties with published data (6,7,10). Copies of the original spectra can be obtained on request from the corresponding author. Optical rotation was measured with a Perkin-Elmer 241 polarimeter as $\alpha$ D $\geq$ 0.0066 ($\epsilon$ = 0.1, CHCl3). The purity of panduratin A used was >99%. All other chemicals, unless otherwise stated, were obtained from Sigma (St Louis, MO).

Materials and methods

Plant material and chemicals

Anti-caspases, anti-FADD, anti-Fas, anti-FasL, anti-TRAIL, anti-Bid and anti-p27Kip1 antibodies were procured from Santa Cruz Biotechnology, (Santa Cruz, CA). Anti-cyclins, anti-Bcl-2, anti-Bax, anti-cdc2, anti-p-cdc2, anti-cdc25C and anti-p27 (Santa Cruz) antibodies were procured from Cell Signaling Technology (Beverly, MA). Anti-PARP116 and anti-cdk2 antibodies were procured from Upstate Biotechnology (Lake Placid, NY). Anti-PARP p85 antibody was procured from Promega (Madison, WI).

The androgen-independent human prostate carcinoma cells, PC3 and DU145, were cultured in RPMI and Minimum Essential Eagle Medium (ATCC; Manassa, VA) containing 10% fetal bovine serum, 100 U/ml penicillin and 100 µg/ml streptomycin (Gibco BRL, USA). The PC3 cells were cultured in prostate epithelial basal medium (PHEM; Cambrex, USA) supplemented with growth factors bovine pituitary extract (BPE), hydrocortisone, human epidermal growth factor (hEGF), epinephrine, insulin, triiodothyronine, transferrin, gentamicine, amphotericin-B and retinoic acid; Cambrex Bioscience, Walkersville, MD, USA). All cells were maintained at 37°C in a humidified atmosphere containing 5% CO2. Panduratin A was dissolved in dimethyl sulfoxide (DMSO) and the final concentration of DMSO used in cell culture was 0.1% (v/v).

MTT assay for cellular viability

Cell viability was determined using the MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide; Sigma) assay (11). The cells were plated at 1 x 10^5 cells per well in 200 µl of complete culture medium and treated with designed concentrations of panduratin A in 96 well microtiter plates for 24 h. Each concentration of panduratin A was repeated in 10 wells. After incubation for specified times at 37°C in a humidified incubator, MTT reagent (50 µl, 2 mg/ml in PBS) was added to each well and incubated for 2 h. The microtiter plate containing the cells was centrifuged at 1800 rpm for 5 min at 4°C. The MTT solution was removed from the wells by aspiration and the formazan crystals were dissolved in DMSO (150 µl). Absorbance was recorded on a microplate reader at 570 nm wavelength. The effect of panduratin A on growth inhibition was assessed as the percentage of inhibition in cell growth where vehicle-treated cells were taken as 100% viable. IC50 values were determined from three independent experiments.

Apoptosis assessment by annexin-V/propidium iodide staining

The annexin-V-Fluos staining kit was used for the detection of apoptotic and necrotic cells according to vendor’s protocol. This kit uses a dual-staining protocol in which the apoptotic cells are stained with annexin-V (green fluorescence), and the necrotic cells are stained with propidium iodide (PI) (red fluorescence). Cells were grown to ~70% confluency and treated with panduratin A (2.5, 5, 10 and 20 µM concentrations) for 24 h. The fluorescence was measured by a Zeiss 410 confocal microscope (Thornwood, NY). Confocal images of green annexin-FITC fluorescence were collected using 488 nm excitation light form an argon/krypton laser, a 560 nm dichroic mirror and a 514–540 nm bandpass barrier filter. Images of red PI fluorescence were collected using a 568 nm excitation light form the argon/krypton laser, a 560 nm dichroic mirror and a 590 nm-long pass filter.

DNA cell cycle analysis by flow cytometry

The effect of panduratin A on PC3 and DU145 cell cycle phase distribution was assessed using flow cytometry. Briefly, after treatment of cells with panduratin A (2.5, 5, 10 and 20 µM concentrations) for 24 h, floating cells were discarded by aspiration and the attached cells were trypsinized and then washed twice with cold PBS, and centrifuged. The pellet was resuspended in 50 µl cold PBS and 450 µl cold methanol for 1 h at 4°C. The cells were centrifuged at 110 g for 5 min. pellet washed twice with cold PBS, suspended in 500 µl PBS and incubated with 5 µl RNAase (20 µg/ml final concentration) at 37°C for 30 min. The cells were then chilled over ice for 10 min and stained with PI (50 µg/ml final concentration) for 1 h for analysis by flow cytometry. Flow cytometry was performed with a FACSscan (Becton Dickinson, Germany).

A minimum of 10 000 cells/sample were collected, and the DNA histograms were further analyzed using ModFitLT software (Verity Software House, Topsham, ME, USA) for cell cycle analysis.

Protein extraction and western blot analysis

Following treatment of cells with panduratin A, the medium was aspirated and the cells were washed with cold PBS (10 mmol/l) at 4°C. The cells were then incubated in ice-cold lysis buffer [50 mmol/l Tris–HCl, 150 mmol/l NaCl, 1 mmol/l EDTA, 20 mmol/l NaF, 100 mmol/l Na3VO4, 0.5% NP40, 1% Triton X-100 and 1 mmol/l phenylmethylsulfonyl fluoride (pH 7.4)] with freshly added protease inhibitor cocktail (protease inhibitor cocktail set III, Calbiochem, La Jolla, CA) over ice for 20 min. The cells were scraped and the lysate was collected in a microfuge tube and passed through a 21.5-gauge needle to break up the cell aggregates. The lysate was centrifuged at 13000×g for 15 min at 4°C, and the supernatant (total cell lysate) was collected, aliquoted and used on the day of preparation or immediately stored at −80°C for use at a later time. For western blot analysis, 40 µg of the protein was resolved over 12% polyacrylamide gels and transferred onto a nitrocellulose membrane. The non-specific sites on blots were blocked by incubating in blocking buffer [5% non-fat dry milk, 1% Tween-20 in 20 mmol/l TBS, pH 7.6] for 1 h at room temperature, incubated with appropriately monoclonal or polyclonal primary antibody in blocking buffer for 90 min to overnight at 4°C, followed by incubation with anti-mouse or anti-rabbit secondary antibody horseradish peroxidase conjugate and detected by chemiluminescence and autoradiography using Hyperfilm obtained from Amershams Biosciences. Densitometric measurement of the bands in western blot analysis were done using digitalized scientific software program UN-SCAN-IT purchased from Silk Science Corporation (Orem, UT).

Reverse-transcription-polymerase chain reaction (RT–PCR)

Total RNA was isolated from the cells using a commercial RNeasy kit (Qiagen, Valencia, CA), and RNA concentration was measured spectrophotometrically at 260 nm. Total RNA (1 µg) was subjected to RT–PCR using one-step RT–PCR premix kit (Promega) containing all components required to synthesize cDNA and to perform the PCR, and specific primers for p27kip1 and GAPDH as a loading control: p27kip1 (size of PCR product: 523 bp); sense 5'-GAA CCG GCA TTT GGC GAA CC-3', antisense 5'-TAA CCC GGC GGT TGG AGA AG-3'; GAPDH (size of PCR product: 735 bp); sense 5'-ATT CCA TGG CAC CGT CAA GG-3', antisense 5'-GTC GTT GAG GAC AAT GCC AG-3'. The first cDNA synthesis was performed following the manufacturer’s instructions. PCR was performed after a 4 min denaturation at 94°C, and the cycles of 94, 55 and 72°C each for 40 s; the number of cycles was specific for each primer set. PCR products were electrophoresed in a 1.5% agarose gel containing ethidium bromide.

Statistical analysis

Each experiment was performed at least three times. Results are expressed as the means ± SD. Statistical analysis was performed using Student’s t-test and statistical significance is expressed as $^a, P < 0.05$, $^{**}, P < 0.01$. 1455

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Results

Inhibition of PC3 and DU145 cell growth by panduratin A

The effect of panduratin A on cell viability was determined employing a MTT assay. As shown in Figure 2, panduratin A treatment to PrEC (Figure 2A) cells produced little to no cytotoxicity in 24 h at which time it produced significant growth inhibition in PC3 and DU145 cells. At 48 and 72 h treatment this differential cytotoxic response of panduratin A persisted. At 24 h treatment with panduratin A IC_{50} values of 13.5 and 14 \mu M in PC3 (Figure 2B) and DU145 (Figure 2C) cells, respectively were obtained. For further mechanistic studies a dose of 2.5, 5, 10 and 20 \mu M panduratin A and a treatment time of 24 h was selected.

Induction of apoptosis by panduratin A in PC3 and DU145 cells

Annexin-V/PI staining was performed to determine early, late apoptotic and necrotic cells following panduratin A treatment to PC3 and DU145 cells. Annexin-V specially binds to phosphatidylserine and has been employed for determination of apoptotic cells. When PC3 and DU145 cells were stained with annexin-V/PI and examined under a fluorescence microscope, early and late apoptosis (annexin V-stained) cells were found to be increased in panduratin A treated cells in a dose-dependent manner. Data obtained with 10 and 20 \mu M concentration of panduratin A is depicted in Figure 3A. The cleavage of poly (ADP-ribose) polymerase (PARP) and acinus are regarded as hallmarks for induction of apoptotic response. The cytosplasmic levels of the native PARP (116 kDa) and its cleaved product (85 kDa) and degradation of acinus were determined by immunoblot analysis in panduratin A treated cells. As shown in Figure 3B, the native 116 kDa PARP protein was found to be cleaved into its characteristic 85 kDa fragment upon treatment with panduratin A in both cells. Also, treatment of panduratin A was found to cause degradation of acinus in both cell types (Figure 3B). Taken together, these data indicate that panduratin A-induced cell death is due to the induction of apoptosis in both cell types.

Effect of panduratin A on the expression of apoptosis-related procaspases in PC3 and DU145 cells

Caspases are responsible for many of the biochemical and morphological changes that occur during apoptosis. In next series of experiments, we determined whether treatment of PC3 and DU145 cells to panduratin A leads to activation of caspase. Immunoblot analysis depicted in Figure 4 indicates that panduratin A treatment to both cell types led to activation of initiator caspases 9 and 8 as well as procaspases 6 and 3.

Panduratin A modulates Bcl-2 family proteins in PC3 and DU145 cells

In next series of experiments, we determined the effect of panduratin A treatment to PC3 and DU145 cells on levels of proteins of the Bcl-2 family members. The immunoblot analysis data showed that the expression of Bax (proapoptosis protein) was significantly increased dose-dependently and time-dependently after treatment of panduratin A in both cells (Figure 5), whereas the expression of Bcl-2 (antiapoptosis protein) was significantly decreased in a dose-dependent and a time-dependent manner. Thus, panduratin A treatment was found to result in alteration in Bax:Bcl-2 ratio in favor of apoptosis that was ~6-fold higher (at 20 \mu M) in PC3 cells and ~15-fold higher (at 20 \mu M) in DU145 cells, compared with their respective controls (Figure 5). The levels of native Bid were also significantly decreased dose-dependently in panduratin A treated cell.

Panduratin A induces Fas death receptor-mediated apoptosis in PC3 and DU145 cells

Because panduratin A induced the cleavage of initiator caspase 8, we considered that its proapoptotic response, at least in part,
Fig. 3. Panduratin A induces apoptosis in PC3 and DU145 cells as assessed by (A) fluorescence microscopy and (B) by immunoblot analysis. (A) The figures contain the representative micrographs of PC3 and DU145 cells undergoing apoptosis induced by treatment with panduratin A as assessed by fluorescence microscopy. Cells were treated with vehicle alone or specified concentration of panduratin A for 24 h. Apoptosis and necrosis was detected by a Zeiss Axiovert 100 microscope as described in Material and methods. The samples were excited at 330–380 nm, and the image was observed and photographed under a combination of a 400 nm dichoric mirror and the 420 nm high pass filter. Data from a typical experiment repeated effects are shown; Magnification ×200. (B) Cells were treated with specified concentrations of panduratin A for 24 h, harvested and total cell lysates were prepared. PARP cleavage and the expression of acinus were determined by western blot analysis. Equal loading was confirmed by stripping immunoblots and reprobing them for β-actin. A typical immunoblot from three separate experiments with similar results is shown here. The details are described under Materials and methods.

Fig. 4. Dose-dependent effect of panduratin A on the expression of apoptosis-related procaspases in PC3 and DU145 cells. Cells were treated with specified concentrations of panduratin A for 24 h, harvested and total cell lysates were prepared. The expression of procaspases 3, 6, 8 and 9 were determined by western blot analysis. Equal loading was confirmed by stripping immunoblots and reprobing them for β-actin. A typical immunoblot from three separate experiments with similar results is shown here. The details are described under Materials and methods.
may be mediated via the death receptor-signaling pathway. As shown in Figure 6, in panduratin A treated PC3 and DU145 cells, a significant increase in the expression of fas-associated death domain (FADD) protein was observed. Panduratin A treatment was found to cause a dose-dependent upregulation of the Fas receptor protein without a significant change in the expression of Fas-L in both cells. In addition, a significant change in the expression of TNF-related apoptosis-inducing ligand (TRAIL) was also observed in both cell lines (Figure 6). These data suggest that panduratin A treatment also induces apoptosis through extrinsic pathway involving the death receptors.

**Panduratin A treatment caused G2/M phase arrest in PC3 and DU145 cells**

To investigate whether panduratin A treatment has an effect on the cell cycle regulation, we determined its effect on cell cycle distribution by flow cytometry after staining with PI. As shown in Figure 7, concomitant with growth inhibitory effects, panduratin A treatment induced a strong G2/M phase arrest in
a dose-dependent manner. In PC3 cells G2/M phase distribution was 12.4, 14.9, 21.0 and 38.6% at 2.5, 5, 10 and 20 μM concentrations of panduratin A, respectively (Figure 7A). In DU145 cells G2/M phase distribution was 17.0, 25.9, 22.0 and 38.6% at 2.5, 5, 10 and 20 μM concentrations of panduratin A, respectively (Figure 7B). Interestingly, the increase in cell population in G1 phase was observed at a concentration of 2.5 μM panduratin A treatment in both the cells.

**Effect of panduratin A on the expression of cell cycle regulators in PC3 and DU145 cells**

Since panduratin A was observed to cause an arrest of cells in G2/M phase, we next assessed its effect on G2/M cell cycle regulators including cdc25C, p-cdc2, cdc2 and cyclin B1. As shown by immunoblot analysis in Figure 8A, panduratin A treatment of cells caused a dose-dependent decrease in the levels of cdc25C, cdc2 and cyclin B1 proteins. However, the phosphorylation of cdc2 was found to be decreased only
Fig. 8. Effect of panduratin A treatment to PC3 and DU145 cells on change in G2/M cell cycle regulator, other cell cycle regulator and cyclin-dependent kinase inhibitor (cdki). Cells were treated with specified concentrations of panduratin A for 24 h, harvested and total cell lysates were prepared. The expression of (A) Cyclin B1, cdc2, p-cdc2 and cdc25C, (B) cyclin D1, cdk6, cdk4, cyclin E and cdk2 and (C) cdkI (p27Kip1 and p21WAF1/Cip1) were determined by western blot analysis. Equal loading was confirmed by stripping immunoblots and reprobing them for β-actin. A typical immunoblot from three separate experiments with similar results is shown here. The details are described under Materials and methods.
Apoptosis represents a universal and exquisitely efficient suicide pathway, considered as an ideal way for elimination of damaged cells (15). Recently, the apoptosis signaling systems have been shown to provide promising targets for the development of novel anticancer agents (16). Several plant-derived bioactive agents are known to be chemopreventive agents inducing apoptosis in a number of experimental models of carcinogenesis (17–19). Thus induction of apoptosis is considered as a possible mechanism of chemopreventive agents. Panduratin A (Figure 1) is a cyclohexenyl chalcone derivative isolated from K. pandurata. Chalcones are natural products, which have been reported to possess a variety of biological effects, including anti-inflammatory, analgesic and anti-oxidant properties (7). Recently we have shown that panduratin A possesses anti-inflammatory effects in RAW264.7 cells (8). However, little is known about the effect of panduratin A on experimental carcinogenesis. As a step towards evaluating an anti-tumorigenic potential of this compound, we have recently shown that it induces apoptosis in HT-29 cells (9). Interestingly, we found that in colon cancer cells panduratin A was a more potent inhibitor of growth than selective COX-2 inhibitor Celecoxib and antitumor drugs (5-flurouracil and cisplatin). This prompted us to explore the effect of panduratin A on human CaP cells. In the present study, we investigated the anti-proliferative activity of panduratin A and the underlying mechanism in androgen-independent human prostate cancer PC3 and DU145 cells.

Caspases plays a central role in many forms of apoptosis (20,21). These enzymes are synthesized as inactive zymogens that must be cleaved after conserved aspartate residues to be activated. Both the intrinsic and extrinsic pathways have been shown to trigger caspase activation in cells undergoing apoptosis (22,23). In the mitochondrial pathway (intrinsic pathway), the death signals are relayed to mitochondria where release of cytochrome c is induced. Cytochrome c binds to Apaf-1, partipating in formation of the apoptosome-dATP-dependent complex, which activates caspase 9. Active caspase 9 can then cleave downstream effector caspases (24–26). The extrinsic pathway for cell death involves plasma membrane death receptors (3,22). These receptors trimerize and recruit the adaptor molecule FADD, which, in turn, activates caspase 8. This, in turn, also leads to the activation of downstream execution caspases (27–30). In both pathways, activation of effector caspases leads to a series of morphological changes characteristics of apoptosis (15).
In this study, panduratin A treatment to CaP cells led to a remarkable induction of apoptotic cells. Panduratin A treatment also led to activation of initiator caspase 8 and 9 as well as of downstream effector caspases 3 and 6. The activation of effector caspases 3 and 6 in response to panduratin A treatment also resulted in cleavage of PARP and acinus in both cell types. The ratio of Bax:Bcl-2 is critical to cell survival such that an increase in Bax levels can shift the process in favor of apoptosis (31). Panduratin A-induced apoptosis and cell growth inhibition was accompanied with decrease in Bcl-2 with concomitant increase in Bax in both cell types. This altered expression of Bcl-2 family members by panduratin A treatment may trigger the activation of initiator caspases 8 and 9 followed by activation of effector caspases 3 and 6. This, in turn, also may lead to cleavage of PARP and acinus.

The Fas receptor (CD95 or Apo1) is the most completely characterized death receptor. Studies have demonstrated that apoptosis induced by anticancer therapy involves the CD95 system (32). TRAIL is a member of the TNF family and can also induce apoptotic cell death in a variety of cell types including CaP cells (32,33). Stimulation of death receptors of the TNF receptor superfamily such as CD95 (APO-1/Fas) or TRAIL receptors results in receptor aggregation and recruitment of the adapter molecule FADD (28,34,35). Upon recruitment, caspase 8, becomes activated and initiates apoptosis by direct cleavage of downstream effector caspases (35). Activation of caspase 8 then leads to a series of downstream apoptotic events (29,30). Studies have shown that the death receptor-induced apoptosis is linked to the mitochondrial machinery through Bid (36,37). Furthermore, the death signal form the Fas receptor may be linked to mitochondria by Bid, a BH3 domain containing protein of the Bcl-2 family (38). Bid then translocates to the mitochondria and mediates the release of cytochrome c that leads to apoptotic changes (38). In this study, panduratin A induced expression of Fas and TRAIL in PC3 and DU145 cells which may, in turn, lead to aggregation and recruitment of the adaptor protein FADD, which results in the activation of caspase 8. These sequence of events may also leads to the activation of downstream execution caspases. Several lines of evidence suggest that both TRAIL and FAS signaling pathways are involved in chemotherapeutic-induced apoptosis, either by activating the initiator caspase 8 or at the level of downstream effector caspase (caspases 3, 6 and 7) activation (39,40). Thus, it is likely that panduratin A-induced apoptosis of CaP cells follows this pathway.

Induction of apoptosis and/or inhibition of cell proliferation are highly correlated with the activation of a variety of intracellular signaling pathways leading to arrest the cell cycle in the G1, S, or G2/M phase of the cell cycle (15) Cell cycle regulation and its modulation by various plant-derived agents are gaining widespread attention in recent years. A large number of phytochemicals have been shown to inhibit cell cycle progression of various cancer cells (41). Especially, G2/M transition provides an effective checkpoint in cell cycle progression that is regulated by the sequential activation and deactivation of cdc family proteins and cyclin complexes. cdc2 interacts with cyclin B1, and activation of the cyclin B–cdc2 complex is required for transition from G2 to M phase of the cell cycle (42–44). Moreover, cdc25C functions as a mitotic activator by dephosphorylating cdc2 that forms a complex with cyclin B1 and drives the cell from G2→M phase. The activation of cdc25C is controlled via its inhibitory phosphorylation by upstream kinase Chk1/2, in response to DNA damage, and thus inhibits mitosis (45–49). In the present study, we found that panduratin A treatment resulted in G2/M phase arrest in a dose-dependent manner in both PC3 and DU145 cells. We further investigated the effect of panduratin A on the expression of G2/M regulatory proteins. Panduratin A treatment was found to result in a remarkable decrease in the protein levels of cyclin B1, cdc25C and cdc2 in both cell types. These results suggest that reduced expression of cyclin B1 and cdc2 and cdc25C may be involved in panduratin A-induced G2/M phase arrest, leading to cell growth inhibition and possible apoptotic death. The effect of panduratin A on the expression of other cyclins and their corresponding protein partners were also investigated. We found that treatment of cells to panduratin A resulted in a dose-dependent decrease in the protein expression of cyclins D1 and E1 with a concomitant decrease in cdks 2, 4 and 6 in both cell types. These data indicate that panduratin A modulates multiple regulatory molecules important in the cell cycle regulation.

Studies have demonstrated that cell cycle arrest at G2/M transition by DNA damaging agent is also tightly associated with the induction of p21WAF/Cip1 (50–53). The p27Kip1 is another member of cdkI, which could bind and inhibit a broader range of cdks (54–57). We found that p27Kip1 and/or p21WAF/Cip1 were upregulated in PC3 and DU145 cells treated with panduratin A. Therefore, The upregulation of p21WAF/Cip1 and/or p27Kip1 by panduratin A is, at least in part, probably responsible for the downregulation of cyclins and cdks expression. Moreover, the upregulation of p21WAF/Cip1 and p27Kip1 and the downregulation of cdks may be one of the molecular mechanisms by which panduratin A inhibits CaP cell growth and induces cell-cycle arrest.

Taken together, this study provided first evidence that panduratin A inhibited cell proliferation of CaP cells by apoptosis which was associated with upregulation of the Fas death receptor and G2/M phase arrest resulting in upregulation of the cdk inhibitor p21WAF/Cip1 and p27Kip1 and inhibition of cdks, cyclins and cdc2/cdc25C. These abilities of panduratin A to induce apoptosis and arrest of cell cycle implies its potential as chemotherapeutic agent because many anticancer drugs are known to achieve their antitumor function by inducing apoptosis and cell cycle arrest in tumor cells. Although the precise molecular mechanism by which apoptosis is induced by panduratin A remains unclear, it might be a potent useful antitumor agent against CaP.

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Efficacy of panduratin A against prostate cancer cells


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