Pretreatment of cromolyn sodium prior to reperfusion attenuates early reperfusion injury after the small intestine ischemia in rats

Zi-Qing Hei, Xiao-Liang Gan, Gang-Jian Luo, Shang-Rong Li, Jun Cai

Abstract

AIM: To investigate the effects of Cromolyn Sodium (CS) pretreated prior to reperfusion on the activity of intestinal mucosal mast cells (IMMC) and mucous membrane of the small intestine in ischemia-reperfusion (IR) injury of rats.

METHODS: Thirty-two Sprague-Dawley (SD) rats were randomly divided into four groups: sham group (group S), model group (group M), high and low dosage of CS groups, (treated with CS 50 mg/kg or 25 mg/kg, group C1 and C2). Intestinal IR damage was induced by clamping the superior mesenteric artery for 45 min followed by reperfusion for 60 min. CS was intravenously administrated 15 min before reperfusion. Ultrastructure and counts of IMMC, intestinal structure, the expression of tryptase, levels of malondialdehyde (MDA), TNF-α, histamine and superoxide dismutase (SOD) activity of the small intestine were detected at the end of experiment.

RESULTS: The degranulation of IMMC was seen in group M and was attenuated by CS treatment. Chiu’s score of group M was higher than the other groups. CS could attenuate the up-regulation of the Chiu’s score, the levels of MDA, TNF-α, and expression of tryptase and the down-regulation of SOD activity and histamine concentration. The Chiu’s score and MDA content were negatively correlated, while SOD activity was positively correlated to the histamine concentration respectively in the IR groups.

CONCLUSION: Pretreated of CS prior to reperfusion protects the small intestine mucous from ischemia-reperfusion damage, the mechanism is inhibited IMMC from degranulation.
The method of CS pretreatment according to Cordeiro et al.[11] was done and CS (50 mg/kg or 25 mg/kg, the dosage and sham group). In another two groups, the same operation was performed for ventilation. The right femoral vein was cut from 5 cm to terminal ileum andleased and reperfusion of the splanchnic region was main-
fined and clamped for 45 min. Then the clamp was re-
was opened and its superior mesenteric artery (SMA) was
operative arterioles and largely prevented pulmonary injury.

The studies proved that IMMC are associated with the small intestine injury after ischemia-reperfu-
sion, and MC membrane stabilizer pretreatment prior to ischemia can protects against the injury, such as CS and MAR-99. While the studies about the intestinal mucosal injury with CS pretreatment after the small intestine ischemia before reperfusion were few. Oxidative stress is one of the mechanism about the small intestine ischemia-reperfusion injury has been generally acknowledged. We hypothesized that CS have an influence on the oxidative stress during the small intestine ischemia-reperfusion, and the purpose of our present study was to investigate whether CS pretreatment prior to reperfusion could protect against early intestinal mucosal damage induced by ischemia-reperfusion through inhibition of IMMC degranulation or oxidative stress. To test our hypothesis, we showed in a rat IR gut injury model: (1) the ultrastructure and counts of IMMC in the early reperfusion; (2) mucosal damage with CS pre-
treatment; (3) expression of tryptase in the IMMC; and (4) the levels of malondialdehyde (MDA), TNF-α, histamine and superoxide dismutase (SOD) activity of the small intes-
tine in rats.

MATERIALS AND METHODS

Acute ischemia-reperfusion injury of the intestinal mucosa in rats

Thirty-two healthy Sprague-Dawley rats (200-250 g, provided by Animal Center of Sun Yat-Sen University and approved by the University Animal Study Committee) were randomly divided into four groups each of which contained 8 rats. Laboratory temperature was kept at 25°C-27°C. Surgery was conducted under general anesthesia with intra-peritoneal sodium pentobarbital (45 mg/kg) after they were fasted for 18 h. Tracheotomy was performed for ventilation. The right femoral vein was cannulated for fluid infusion and drugs. The rat abdomen was opened and its superior mesenteric artery (SMA) was found and clamped for 45 min. Then the clamp was released and reperfusion of the splanchic region was maintained for 60 min (in M group). In control group, SMA was found but not clamped and i.v. saline solution via the right femoral vein at 30th min after the start of experiment (sham group). In another two groups, the same operation was done and CS (50 mg/kg or 25 mg/kg). The dosage and method of CS pretreatment according to Cordeiro et al.[8] for Cromolyn sodium is poorly absorbed by oral.) was given via right femoral vein 15 min before the opening of the clamp (C1 and C2 groups).

Preparation of specimens and measurements

After ischemia-reperfusion, the rats were killed and paunched rapidly. A segment of 0.5-1.0 cm intestine was cut from 5 cm to terminal ileum and fixed in 4% formal-
dehyde polymerisatum, then embedded in paraffin for sec-
tion. Another segment of small intestine was washed with frozen saline and dried with suction paper and at -70°C.

The segment of small intestine was stained with hematoxylin-eosin. The damages of intestinal mucosa were evaluated by two different pathologist according to the cri-
teria of Chiu's method[22]. Criteria of Chiu grading system consists from 5 subdivisions according to the changes of villus and gland of intestinal mucosa: grade 0, normal mu-
cosa; grade 1, development of subepithelial Gruenhagen's space at the tip of villus; grade 2, extension of the space with moderate epithelial lifting; grade 3, massive epithe-
\[\text{Transmission electron microscopy}\]

Intestines were immersed and fixed in 2.5% glutaraldehyde overnight at 4°C and washed three times in PBS. Then they were postfixed in aqueous 1% OsO₄ and 1% KFe(CN)₆ for 1 h. After three times of PBS washes, the tissue was dehydrated through a graded series of 30% to 100% ethanol and then infiltrated in 1:1 mixture of propylene oxide and Polybed 812 epoxy resin for 1 h. The infiltration solution was changed to 100% res-
in. After 24 h of infiltration, the tissue was embedded in molds and cured at 37°C overnight, followed by additional hardening at 65°C for 2 d. Ultrathin (70 nm) sections were collected on 200-mesh copper grids and stained with 2% uranyl acetate in 50% methanol for 10 min, followed by 1% lead citrate for 7 min. Sections were photographed using a Hitachi H-600 transmission electron microscope (TOSHI-
\[\text{Detection of concentration of protein in intestine}\]

Intestinal tissues were homogenized with normal sa-
ilne. Intestinal protein quantitation was by the Bradford method[19] with a BSA standard using kits were provided by Shenerg Biocolor BioScience & Technology Company, Shanghai, China.

\[\text{Detection of content of MDA in the intestine}\]

Intestinal tissues were homogenized with normal saline. MDA content was determined by the TBA method (Jian
ccheng Bioengineering Ltd, Nanjing, China). Homoge-
eenate (0.1 mL) was taken to detect MDA content. Briefly, 0.1 mL 8.1% SDS, 0.8 mL acetic acid buffer, 0.8 mL 0.8% TBA and 0.2 mL distilled water were added into the sample tubes and one standard tube (containing 0.1 mL tetrae-
thoxyp propane). All the tubes were then incubated at 100°C for 1 h. After cooled at -20°C for 5 min, 2 mL of n-butyl alcohol was added into the sample, which was then vibrat-
ed for 1 minute and centrifuged for 10 min at 3000 r/min. The supernatant of the samples were assayed to detect absorbance at 532 nm; and the results were expressed as nmol/mL. The content of MDA in intestine was calcu-
\[\text{Detection of activity of SOD in the intestine}\]

Intestinal tissues were made into a homogenate with nor-
mal saline. After centrifugation for 5 min and centrifuged for 15 min at 4000 r/min. Supernatants were transfered into fresh tubes for evaluation of SOD activity. SOD activity was evaluated with an SOD detection kit according to the

www.wjgnet.com
manufacturer’s instructions (Jiancheng Bioengineering Ltd, Nanjing, China). Results were expressed as nmol/mL. The activity of SOD in the intestine was calculated as U per milligram of protein.

**Detection of the concentration of TNF-α in the intestine**

Intestinal tissues were made into a homogenate with normal saline, frozen at -20°C for 5 min and centrifuged for 15 min at 4000 r/min. Supernatants were transferred into fresh tubes for evaluation of concentration of TNF-α (Biosource, USA) using a commercially available ELISA kit in accordance with the manufacturer’s instructions, results were expressed as pg/mL. The concentration of TNF-α in the intestine was calculated as picogram per milligram of protein.

**Detection of the concentration of histamine in the intestine**

Intestinal tissues were made into a homogenate with normal saline, frozen at -20°C for 5 min and centrifuged for 15 min at 4000 r/min. Supernatants were transferred into fresh tubes for evaluation of concentration of histamine (RapidBio Lab, USA) using a commercially available ELISA kit in accordance with the manufacturer’s instructions, results were expressed as ng/mL. The concentration of histamine in the intestine was calculated as nanogram per milligram of protein.

**Immunohistochemical detection of tryptase in intestine**

Five µm thick sections were prepared from paraffin-embedded tissue. After deparaffinization, endogenous peroxidase was quenched with 3% H2O2 in deionised water for 10 min. Nonspecific binding sites were blocked by incubating the sections in 10% normal rabbit serum for 1 h. The sections were then incubated with polyclonal rat anti-mast cell tryptase (dilution 1: 50) for 30 min at 37°C, followed by incubation with biotinylated goat-anti-rat IgG at room temperature for 10-15 min. After 3 × 5 min PBS rinses, the horseradish-peroxidase-conjugated streptavidin solution was added and incubated at room temperature for 10-15 min. The antibody binding sites were visualized by incubation with a diaminobenzidine-H2O2 solution. The sections incubated with PBS instead of the primary antibody were used as negative controls. Brown-yellow granules in the cytoplasm were recognized as positive staining for tryptase. We calculated the tryptase positive mast cells and their intensity in 5 representative areas at × 400 magnification by Image-Pro Plus 5.0 (USA).

**Statistical analysis**

Data were expressed as mean ± SD and analysis of variance was performed using SPSS 11.0 software. One-way analysis of variance was used for multiple comparison, least significant difference test (LSD-t) was used for intra-group comparison or Tamhane’s T2 test was used if equal variances was not assumed. Pearson analysis was used for the correlation in the ischemia and reperfusion groups. Differences were considered significant when P was < 0.05.

**RESULTS**

**Changes of intestinal mucosa under light microscope**

The villus and glands were normal and no inflammatory cell infiltration was observed in mucosal epithelial layer in sham group. Multiple erosions and bleeding were observed in model group. Light edema of mucosa villus and infiltration of few necrotic epithelial inflammatory cells neutrophil leukomonocyte were found in mucosa epithelial layer in C2 and C1 groups (Figure 1).
Chiu’s score of small intestinal structure
The Chiu’s score in sham group was the lowest, while in the model group it was the highest in the four groups \((P < 0.05)\). The Chiu’s score in C1 group was significantly lower than in C2 group after treated with CS \((P < 0.05)\) (Table 1).

Changes of ultrastructure of small intestinal
The ultrastructure of small intestinal was normal in group S. There was seen the karyopyknosis of epithelial cell of small intestine in group M, the nuclear membrane was more irregularity, and the swelling microvillus became shorter and thicker, most of the microvillus were shedding. The nucleus of epithelial cell of small intestine in group C1 and C2 was deflated, the nuclear membrane was irregularity, and the light swelling microvillus became shorter (Figure 2).

Changes of ultrastructure of IMMC
The ultrastructure of IMMC was normal in sham group. There were abundant vacuolus with a reduction granulation in their endochylema in model group. There were few swollen granules with a reduction in IMMC homogeneity in C1 and C2 group (Figure 3).

Changes of MDA in small intestine
The content of MDA in intestine of model group was the highest in all experimental groups, it decreased significantly compared with the model group after treated with CS \((P < 0.05)\) and there was no significant difference compared with the sham group \((P > 0.05)\) (Table 2).

Changes of activity of SOD in small intestine
The activity of intestinal SOD decreased significantly in ischemia-reperfusion injury groups compared with the sham group \((P < 0.05)\), treated with CS it increased significantly compared with the model group \((P < 0.05)\), and there was no significant difference between group C1 and C2 \((P > 0.05)\) (Table 2).

Changes of TNF-\(\alpha\) in small intestine
The concentration of TNF-\(\alpha\) of intestine in model group rats was higher than the other three groups \((P < 0.05)\). There were no significant difference in sham group, C1 group and C2 group \((P > 0.05)\) (Table 2). There was a positive correlation between the Chiu’s score and the concentration of TNF-\(\alpha\) in the ischemia and reperfusion groups \((r = 0.734, P < 0.05)\).

Changes of histamine in small intestine
The histamine concentration of intestine in the model and C2 groups decreased significantly compared with the sham group \((P < 0.05)\), it increased significantly after pretreated with CS compared with the model group \((P < 0.05)\).
were no significant difference between C1 and C2 groups ($P > 0.05$) (Table 2). The Chiu's score and MDA content were negatively correlated to the histamine concentration respectively ($r = -0.676$, $P < 0.05$ or $r = -0.452$, $P < 0.05$), while the SOD activity was positively correlated to the concentration of histamine in the ischemia and reperfusion groups ($r = 0.579$, $P < 0.05$).

Counts and expression of tryptase of IMMC
Expression of tryptase in sham group was the lowest, while in the model group it was the highest in the four expression groups ($r = 0.813$, $P < 0.05$). Tryptase is one of the specific markers of IMMC [16]. Boros [15] proved that intestinal ischemia induced the release of a variety of IMMC-derived inflammatory compounds and resulted in a spectrum of injury ranging from reversible permeability changes to structural mucosal damage.

DISCUSSION
IMMC are located in close proximity to submucosal collecting venules, which are primary targets of leukocyte-endothelial interactions during ischemia-reperfusion injury. IMMC are particularly frequent in close proximity to epithelial surfaces where they are strategically located for optimal interaction with the environment and for their putative functions for host defense. They sense the foreign material invading the mucosa in an appropriate inflammatory response, and were considered as one of components of the fourth level of mucosal defense [16]. Acute inflammation could lead to increase of IMMC counts and release of a multi-faceted spectrum of proinflammatory mediators by IMMC such as cytokines and chemokines, and MC have the capacity to coordinate trafficking of leukocytes [15]. Boros [15] proved that intestinal ischemia induced the release of a variety of IMMC-derived inflammatory compounds and resulted in a spectrum of injury ranging from reversible permeability changes to structural mucosal damage.

The ischemia time of small intestine rats’ model is from 30 min to 60 min [17-19], here we used the median time (45 min). Cizova reported that the concentration of thiobarbituric acid reactive substances was increased at the end of the ischemia lasting from 30 to 90 min [20], and CS plasma life in vivo is very short. Thus the reperfusion time in our study was watched in 60 min, it was the early reperfusion according to Hamar et al [21]. All of previous studies were focused on the MC membrane stabilizer pretreatment prior to ischemia, and had proved that IMMC were associated with the damage to intestinal mucosal after the small intestine ischemia-reperfusion. While the main purpose of our study was to see whether pretreatment with CS prior to reperfusion also have the protective effects during early reperfusion after the small intestine ischemia.

Tryptase is one of the specificity markers of IMMC [22]. We counted the IMMC counts through the expression of tryptase using immunohistochemical methods which is more accuracy than oluidine blue staining. Our study found that the expression of tryptase and IMMC counts increased significantly in 60 min reperfusion injury in model group. There were abundant vacuolus in IMMCs in the model group after they were degranulated by electron microscope. IMMC is the main source of histamine in intestine. The levels of intestinal histamine includes the concentration of histamine intra- and extro-IMMC. The level of intestinal histamine is mainly represent of the concentration of histamine intra-IMMC as the extracellular...

**Table 2** Changes of TNF-α, MDA content, SOD activity and histamine concentration in small intestine in various groups (mean ± SD)

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>TNF-α (pg/mg protein)</th>
<th>MDA (nmol/mg protein)</th>
<th>SOD (U/mg protein)</th>
<th>Histamine (ng/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S</td>
<td>8</td>
<td>3.7 ± 0.4</td>
<td>0.44 ± 0.09</td>
<td>179.2 ± 15.3</td>
<td>5.7 ± 0.5</td>
</tr>
<tr>
<td>M</td>
<td>8</td>
<td>4.7 ± 0.4</td>
<td>0.66 ± 0.07</td>
<td>130.6 ± 10.6</td>
<td>4.2 ± 0.4</td>
</tr>
<tr>
<td>C1</td>
<td>8</td>
<td>3.8 ± 0.4</td>
<td>0.45 ± 0.06</td>
<td>147.9 ± 12.4</td>
<td>5.3 ± 0.6</td>
</tr>
<tr>
<td>C2</td>
<td>8</td>
<td>3.9 ± 0.4</td>
<td>0.47 ± 0.07</td>
<td>145.3 ± 15.7</td>
<td>4.8 ± 0.6</td>
</tr>
</tbody>
</table>

$^aP < 0.05$, $^bP < 0.01$ vs Group S; $^cP < 0.05$, $^dP < 0.01$ vs Group M.

![Figure 3](image_url) Ultrastructure of intestinal mucosal mast cells of rats in each group. There are abundant vacuolus with a reduction of granulation in their endochylema in model group; there are filled with granulation endochylema and there is no vacuolus in their endochylema in sham group, these changes of ultrastructure are ameliorated by treatment with Cromolyn Sodium (group C1 and C2).
histamine released from IMMC in the gastrointestinal tract is rapidly cleared and degraded\(^23\) and the more IMMC degranulate, the lower concentration of histamine is found in intestine\(^24\). Our study found the histamine level decreased significantly in model group while IMMC counts increased, suggesting that IMMC may degranulate and release histamine in 60 min reperfusion.

Histamine has many pathophysiological roles, and it is an important messenger in the gut\(^25\). Akerstrom et al\(^26\) reported that anti-histaminergic pretreatment could decrease the trauma-induced leakage of albumin by mechanisms which may involve readjustments of pressures and flows in capillaries as well as a prevention of histamine effects on capillary permeability on a model of mechanical intra-abdominal trauma in rats. Our results found that the intestinal histamine concentration decreased after early ischemic-reperfusion, and there was a negative correlation between the small intestinal Chiu's score and the level of histamine in the intestine. This result suggested that histamine took part in the ischemia-reperfusion intestinal mucosal damage.

TNF-\(\alpha\) is an inflammatory cytokine that may be an important mediator in the development of reperfusion-induced tissue injury and lethality\(^27\). Grewal\(^28\) demonstrated that treatment of rats with anti-TNF antibodies could prevent neutrophil influx, tissue injury. Our study found that the intestinal TNF-\(\alpha\) concentration increased after ischemic-reperfusion; and there was a positive correlation between the small intestinal Chiu's score and the level of TNF-\(\alpha\) in the intestine. This result suggested that TNF-\(\alpha\) also took part in the ischemia-reperfusion intestinal mucosal damage. Although the most intestinal TNF-\(\alpha\) is considered by some one from the mast cells\(^29\), it has been proved that many sorts of cells also release the TNF-\(\alpha\) besides the mast cells. We believe the increase of the TNF-\(\alpha\) is contributed by many factors.

CS is a stabilizing agent of mast cell which prevents histamine and TNF-\(\alpha\) released from IMMC\(^30\). Szabo et al\(^31\) reported that 30 min segmental ischemia and 120 min reperfusion induced significant tissue injury, elevated the segmental vascular resistance, and decreased intramucosal pH (pHi), and CS pretreatments prior to ischemia significantly inhibited the permeability changes, but did not influence the pHi and morphological alterations induced by ischemia-reperfusion, they conclude that intestinal mast cells and mast cell-induced reactions contribute to the mucosal permeability alterations during reperfusion, but play only a minor role in ischemia-reperfusion-induced structural injury. Pretreatment with CS protecting against degranulation, caused a significant impairment of plasma exudation at 30 min of inflammation corresponding to a significantly decreased level of histamine, one of the most potent vasoactive factors released from activated mast cells\(^32\).

The results in our study showed that the injury of small intestinal villus and microvillus was alleviated after CS pretreatment prior to reperfusion and that the ultrastructure of IMMC was basically normal. The expression of trypase and TNF-\(\alpha\) concentration were also alleviated by CS pretreatment prior to reperfusion, and the concentration of histamine in intestine was increased compared with the model group after CS pretreatment prior to reperfusion. The results indicated that CS decreased ischemia-reperfusion injury by prevention of IMMC degranulation, thus it decreased the release of histamine and TNF-\(\alpha\). This protection may be dose-dependent as high dose of CS with
more powerful effect.

There were many reports about intestinal ischemia and reperfusion resulted in the increase of MDA and decrease of SOD activity, toxic-free oxygen radicals are produced in the ischemic tissue[33,34]. Our study also demonstrated that ischemia-reperfusion injury elevated the oxygen radicals and lipid radicals. Frossi[35] reported that oxidative stress could induce a pro-type 2 inflammatory response and degranulation of mast cells. Fukushii[36] found the compound 48/80, a typical histamine liberator elicited superoxide anion generation in mast cells in a dose-dependent fashion. These studies indicated that degranulation of mast cells was able to induce oxidative stress injury and oxygen radicals could make mast cells to degranulate. In this study we found there were correlations among the MDA, SOD activity and the concentration of histamine, the other findings of our study were that MDA content increased and SOD activity decreased remarkably in the model group, while pretreatment by cromolyn sodium prior to reperfusion could attenuate the up-regulation of MDA content and the down-regulation of SOD activity. The results were indicating that IMMC degranulation and oxidative stress can affect each other, and the less IMMC degranulation can make less oxidative stress. Future studies are need to focus on the relationships in vitro.

In conclusion, pretreatment of Cromolyn Sodium prior to reperfusion could attenuate early reperfusion injury after the small intestine ischemia in rats. The mechanisms includes: inhibited IMMC from degranulation, decreased the release of histamine and TNF-α from IMMC, and decreased oxidative stress.

REFERENCES


Peer review
This paper reports an experimental study very well designed and performed and very elegant results and discussion. The final conclusions are nicely shown. Their English is of good quality. The purpose of this paper was to determine whether cromolyn sodium reduces or prevents injury of the small intestine of rats following ischemia-reperfusion. To this end the authors perform a number of biochemical measurements (MDA, TNFα, histamine, SOD) as well as ultrastructural studies of mast cells and microscopical investigations of the small intestine.

COMMENTS

Background
Intestinal mucosal mast cells (IMMCs) is associated with the mucosal damage. The aim of this study was to investigate the effects of Cromolyn Sodium (CS) pretreated prior to reperfusion on the activity of IMMC and mucous membrane of the small intestine in ischemia-reperfusion (IR) injury of rats.

Research frontiers
Previous studies proved that IMMC are associated with the small intestine injury after ischemia-reperfusion, and MC membrane stabilizer pretreatment prior to ischemia can protect against the injury, such as CS and MAR-99.

Innovations and breakthroughs
While the studies about the intestinal mucosal injury with CS pretreatment after the small intestine ischemia before reperfusion were few. Oxidative stress is one of the mechanism about the small intestine ischemia-reperfusion injury has been generally acknowledged. We hypothesized that CS have an influence on the oxidative stress during the small intestine ischemia-reperfusion, and the purpose of our present study was to investigate whether CS pretreatment prior to reperfusion could protect against early intestinal mucosal damage induced by ischemia-reperfusion through inhibition of IMMC degranulation or oxidative stress.

Applications
Pretreated of CS prior to reperfusion protects the small intestine mucous from ischemia-reperfusion damage, the mechanism is inhibited IMMC from degranulation.

Terminology
Restoration of blood supply to tissue which is ischemic due to decrease in normal blood supply. The decrease may result from any source including atherosclerotic obstruction, narrowing of the artery, or surgical clamping. It is primarily a procedure for treating infarction or other ischemia, by enabling viable ischemic tissue to recover, thus limiting further necrosis. However, it is thought that reperfusion can itself further damage the ischemic tissue, causing REPERFUSION INJURY.
T. Role of complement activation and mast cell degranulation in the pathogenesis of rapid intestinal ischemia/reperfusion injury in rats. Digestion 2001; 63 Suppl 1: 103-107


S- Editor Liu Y  L- Editor Li M  E- Editor Li JL