

RESEARCH ARTICLE

Chemopreventive Activity of Turmeric Essential Oil and Possible Mechanisms of Action

Vijayasteltar Belsamma Liju, Kottarapat Jeena, Ramadasan Kuttan*

Abstract

This study aimed to evaluate the antimutagenic and anticarcinogenic activity of turmeric essential oil as well as to establish biochemical mechanisms of action. Antimutagenicity testing was accomplished using strains and known mutagens with and without microsomal activation. Anticarcinogenic activity was assessed by topical application of 7, 12 – dimethylbenz[a]anthracene (DMBA) as initiator and 1% croton oil as promoter for the induction of skin papillomas in mice. Inhibition of p450 enzymes by TEO was studied using various resorufins and aminopyrene as substrate. Turmeric essential oil (TEO) showed significant antimutagenic activity ($p < 0.001$) against direct acting mutagens such as sodium azide (NaN_3), 4-nitro-O-phenylenediamine (NPD) and N-methyl-N-nitro N'-nitrosoguanine (MNNG). TEO was found to have significant antimutagenic effect ($>90\%$) against mutagen needing metabolic activation such as 2-acetamidoflourene (2-AAF). The study also revealed that TEO significantly inhibited ($p < 0.001$) the mutagenicity induced by tobacco extract to *Salmonella* TA 102 strain. DMBA and croton oil induced papilloma development in mice was found to be delayed and prevented significantly by TEO application. Moreover TEO significantly ($P < 0.001$) inhibited isoforms of cytochrome p450 (CYP1A1, CYP1A2, CYP2B1/2, CYP2A, CYP2B and CYP3A) enzymes *in vitro*, which are involved in the activation of carcinogens. Results indicated that TEO is antimutagenic and anticarcinogenic and inhibition of enzymes (p450) involved in the activation of carcinogen is one of its mechanisms of action.

Keywords: Antimutagenicity - turmeric essential oil - dimethylbenz[a]anthracene - cytochrome p450

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Introduction

Many human cancers are caused by chemical carcinogens, such as polycyclic aromatic hydrocarbons, heterocyclic amines, aromatic amines etc present in our environment. Continued exposure of these substances to human cells leads to genomic instability, including repair deficiency and accumulation of genetic alteration (Ames, 1983). Mutation is a major factor in carcinogenesis and incidence of cancer may be reduced by decreasing the rate of mutations induced by various chemical mutagens. Many carcinogens are activated through Phase I (cytochrome p450) enzymes present in endoplasmic reticulum of the liver cells. Induction of Phase II detoxification enzymes, such as glutathione S-transferase and UDP-glucuronyl transferase, is another mechanisms of protection against carcinogenesis (Piengchai et al., 2011). Modulating these enzymes may reduce cancer incidence.

Spices which are widely used as food ingredient exhibits different pharmacological properties. Essential oils from spices are volatile compounds produced as secondary metabolites. The essential oil from *Curcuma longa* rhizome is a complex mixture obtained by steam distillation. A total number of 12 components of the essential oil are identified by GC-MS analysis and the principal compounds include ar-turmerone (61%),

curlone (11.2%), and cucumene (5.5%) (Liju et al., 2011). Ar-turmerone, the major compound present in TEO has been shown to exhibit anticancer properties (Baik et al., 1993). Biological properties of TEO such as its antimicrobial, anti-inflammatory, antioxidant and antinociceptive activities have been reported (Negi et al., 1999; Liju et al., 2011). Studies have revealed that TEO has chemopreventive efficacy against submucous fibrosis in human (Hastak et al., 1997; Deepa et al., 2010). TEO also showed antiproliferative activity against both human oral epidermal carcinoma (KB) and murine leukemia (P388) cell lines (Manosroi et al., 2006). Nevertheless few studies have been carried out to evaluate its antimutagenic and anticarcinogenic activities.

This study examines whether TEO possess any antimutagenic ability against direct acting mutagens such as sodium azide (NaN_3), 4-nitro-o-phenylenediamine (NPD), MNNG as well as mutagens needing mammalian liver microsomal activation, such as 2-acetamidoflourene (2-AAF) using *Salmonella* strains TA 98, TA 100, TA 1535 and TA 102. Effect of TEO on tobacco induced mutagenicity was also evaluated. Moreover chemopreventive efficacy of TEO against skin papilloma development in mice induced by DMBA as initiator and croton oil as promoter is also being reported in this study. In order to determine the possible mechanism of action

Department of Biochemistry, Amala Cancer Research Centre, Kerala, India *For correspondence: amalacancerresearch@gmail.com, amalaresearch@hotmail.com

of TEO we have evaluated the inhibition of different cytochrome p450 enzymes by TEO *in vitro*.

Materials and Methods

Turmeric essential oil

The essential oil isolated by steam distillation from the rhizome of *Curcuma longa* was purchased from Kancore Ingredients Limited., Angamali, Kerala, India. TEO was dissolved in DMSO for *in vitro* studies. The colour and appearance of TEO was pale yellow liquid (Sample No. TONE-00321). Essential oil was stored at 4°C away from direct light. For oral administration, TEO was dissolved in paraffin oil. Composition of TEO has been reported earlier (Liju et al., 2011).

Chemicals

Nicotinamide adenine dinucleotide phosphate reduced (NADPH), Agar agar, L- histidine, D-biotin, NADP, glucose-6-phosphate, dimethyl sulphoxide and sodium azide (NaN₃) were obtained from Sisco Research Laboratories, Mumbai, India. Nutrient broth was purchased from Hi-media laboratories, Mumbai. N-Methyl-N'-nitro-N-nitrosoguanidine (MNNG), 4-nitro-O-phenylenediamine (NPD), 2-acetamidoflourene (2-AAF), 7-ethoxy resorufin (ER), 7-pentoxo resorufin (PR), 7-methoxy resorufin (MR) and 7, 12-dimethylbenz (a) anthracene (DMBA) were purchased from Sigma chemicals (St. Louis, MO, USA). Phenobarbitone (Gardenal[®] 60, Batch No.B03007) was purchased from Nicholas-Piramel India Ltd, Gujarat, India. Dimethyl sulfoxide (DMSO) and liquid paraffin were purchased from Merck Specialties Private Ltd., Mumbai, India. Tobacco was purchased from local market. All other chemicals were of analytical grade. Croton oil was prepared from the seeds of *Croton tiglium L.* by light ether petroleum extraction. All other reagents were of analytical grade

Animals

Male Wistar rats (8-10 weeks aged) weighing 150±20 g and Balb/C mice weighing 20-25g were purchased from Small Animal Breeding Station, Kerala Veterinary and Animal Sciences University, Mannuthy, Kerala, India. The animals were maintained under standardized environmental conditions (22-28°C, 60-70% relative humidity, 12h dark/light cycle). They were housed in well ventilated cages and fed with pelleted diet (Sai Durga Agency, Bangalore, India) and water ad libitum. All the animal experiments were done as per the instructions prescribed by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Environment and Forest, Government of India, and implemented through the Institutional Animal Ethical Committee of the Amala Cancer Research Centre.

Evaluation of antimutagenicity of TEO

Bacterial strains: auxotrophic *Salmonella typhimurium* strains TA 98, TA 100 and TA 1535 were purchased from Gene Bank & MTCC, Chandigarh, India. TA 102 mutant strain was kindly provided by Dr. Padma Ambalam,

Sourashtra University, Gujarat, India. For antimutagenicity study *S. typhimurium* strains were prepared from frozen permanent by inoculating the strains in nutrient broth (5 ml) and incubating for 12h at 37°C. Genotype of the *S. typhimurium* strains TA 98, TA 100 and TA 1535 were checked using crystal violet sensitivity, histidine requirement, ampicillin and tetracycline resistance. TA 102 strain was checked by UV-sensitivity (Maron and Ames, 1983).

Mutagens: the mutagens, N-Methyl-N'-nitro-N-nitrosoguanidine (MNNG) (1 µg/plate), 2-acetamidoflourene (2-AAF) (20 µg/ plate), sodium azide (NaN₃) (2.5 µg/plate) were dissolved in water and 4-Nitro-O-phenylenediamine (NPD) (20 µg /plate) was dissolved in dimethyl-sulfoxide (DMSO). For antimutagenicity studies tobacco extract was prepared by cutting 100 g of tobacco into small pieces and boiling in 500 ml of distilled water. This was evaporated to dryness and 50 mg of the extract was incorporated into each plate, which can produce mutagenic response.

Evaluation of antimutagenic potential of TEO

Antimutagenicity of TEO was tested against mutagens such as sodium azide, NPD and MNNG using TA 100, TA 1535, TA 98 and TA 102 strains *Salmonella typhimurium* by the method of Maron and Ames, (1983) and modified by Kaur et al. (1998) in triplicate. Concentrations of TEO used for evaluating the antimutagenicity were 0.1, 0.5 and 1 mg/ plate. TEO was added to 2 ml of top agar at 45°C (0.5% NaCl and 0.6% agar) containing 0.5 mM histidine-biotin, bacterial culture of 1-2×10⁹ cells/ml (0.1 ml) and direct acting mutagens at concentrations mentioned above. It was mixed well and poured into minimal agar plates. After incubation at 37°C, the number of histidine independent revertant colonies were counted using colony counter. The plates with mutagen alone acted as positive control and plates without test sample and mutagen were considered as negative controls or spontaneous revertants.

In the case of mutagen needing activation, 2-acetamidoflourene (2-AAF) (20 µg/plate), *Salmonella* strains TA 98 or TA 100, different concentrations of TEO, 0.1 ml bacteria (1-2×10⁹ cells/ml), 0.5 ml S9 mix containing 0.2 M sodium phosphate buffer (pH 7.4), NADP (0.1 M), 1 M glucose 6 phosphate, 10 µl MgCl₂-KCl and 2-AAF were incubated for 45 min at 37°C. This mixture was then added to 2 ml of melted top agar, gently mixed and overlaid onto the minimal glucose agar plates. After solidification, the plates were inverted and incubated for 48 h at 37°C. The number of revertant colonies were counted using colony counter. All the plates were prepared in triplicate. The percentage inhibition of mutagenicity was calculated using the formula: **Percentage inhibition**=[(C-SR)×(T-SR)÷(C-SR)]×100, where C is the number of revertants in the presence of mutagen alone, T is the number of revertants in the presence of TEO with mutagens and SR is the spontaneous revertants. No antimutagenic effect was considered if the percentage of inhibition was smaller than 25%, a moderate effect if value is between 25% and 40% and a strong antimutagenic activity if value is greater than 40%. Untreated plates (without mutagens) and plates treated

with DMSO (Vehicle control) were also kept. The plates with diagnostic mutagen acted as positive control and plates without test sample and mutagen were considered as negative controls or spontaneous revertants.

Determination of the effect of TEO on DMBA and croton oil induced papilloma formation in mice

Male Balb/c mice were used for the DMBA induced skin papilloma study. DMBA, croton oil and TEO was applied on the shaved dorsal side (2 cm diameter) of the mice at least two days before the application of chemicals. Animals having no hair growth after 2 days were selected for the experiment. Balb/C mice were divided in to eight groups and each group having 8 animals.

Single dose of DMBA (470 nmol/mouse dissolved in 200 μ l acetone) was used for the study. All groups were applied with 1% croton oil in acetone (200 μ l/ animal) twice weekly for six weeks except group VI and VIII. TEO dissolved in paraffin oil was applied twice weekly for 6 weeks 30 minutes before each croton oil application. The animals in all groups were watched for food intake as well as any apparent toxicity such as weight loss or mortality during the entire period of the study. Skin papilloma formation was recorded weekly and the tumor growth greater than 1 mm in diameter was included in the cumulative total if they persisted for 2 weeks or more. Formation of the onset papilloma and number of papilloma per cage in various groups were recorded every week.

Effect of TEO on the inhibition of various isoforms of cytochrome p450 enzymes

Rats were administered with phenobarbitone continuously for 4 days (60 mg/kg body weight, once daily) and sacrificed 24 h after the last dose of phenobarbitone. The livers of all the rats were excised quickly, washed thoroughly in ice-cold saline and kept at -70°C . Liver homogenate (25%) was made in cold phosphate buffer (pH 7.4, 0.1 M). Homogenate was initially centrifuged at 14000g for 20 min in a cold centrifuge (Remi) and supernatant was then further centrifuged at 10, 5000 g for 1 h in an ultracentrifuge (Sorvall). The microsomes obtained from liver homogenate were washed and resuspended in cold phosphate buffer (pH 7.4, 0.1 M) and used for determination of the effect of TEO on the dealkylation of methoxy resorufin by 7-methoxyresorufin-O-demethylase (MROD), CYP1A2, pentoxy resorufin by 7-pentoxyresorufin-O-depentylase (PROD), CYP2B1/2 and ethoxy resorufin by 7-ethoxyresorufin-O-deethylase (EROD), CYP1A1 (Pohl and Fouts, 1980; Nerurkar et al.,

1993).

Reaction mixture containing sodium phosphate buffer (0.1 M, pH 7.4), 6.25 mM MgSO_4 , 60 μM EDTA, 5 μM (ER, MR or PR), 100 μg microsomal protein, and various concentrations of essential oil (50-200 μg) in a final volume of 1 ml was incubated for 5 min at 37°C . The reaction was stopped by the addition of 2 ml of chilled methanol. The precipitated protein was centrifuged and supernatant was used for the estimation of enzyme activity by measuring resorufin formed using a fluorescent spectrophotometer (Nanodrop ND-3300, Thermo Scientific, USA.) at the excitation wavelength of 550 nm and the emission wavelength of 585 nm. Blanks were set without addition of NADPH. The percentage inhibition was calculated by the following formula: $\text{C-T}/\text{C}\times 100$. Where C is the optical density of control without essential oil, T is the optical density with essential oil.

Aminopyrene-N-demethylase activity (an indicator of CYP 1A, 2A, 2B, 2D and 3A activity) was measured at various concentration of TEO (50, 100 and 200 μg) by the method of Mazel, (1971). Aminopyrene was dealkylated by microsomal enzymes to form 4-amino antipyrine and formaldehyde. Formaldehyde so formed was measured by condensation with Nash reagent. The colour development and absorbance was measured at 412 nm. The percentage of inhibition was calculated.

Statistical analysis

The values are expressed as mean \pm SD. The statistical significance was compared between control and tested groups by one way analysis of variance (ANOVA) followed by appropriate post hoc test (Dunnet multiple comparison test) using Graph pad in Stat software.

Results

Evaluation of antimutagenic potential of TEO

Earlier studies indicated that TEO did not show any mutagenicity and cytotoxicity up to a concentration of 3 mg/plate (Liju et al., 2013) against *S. typhimurium* strains.

Table 1. Groups, Treatment and Number of Animals

Groups	Treatment	No. of animals
SI	DMBA+croton oil	8
II	DMBA+croton oil+Paraffin oil	8
III	DMBA+croton oil+10% TEO	8
IV	DMBA+croton oil+25% TEO	8
V	DMBA+croton oil+50% TEO	8
VI	DMBA alone	6
VII	Croton oil alone	6
VIII	DMBA+TEO	8

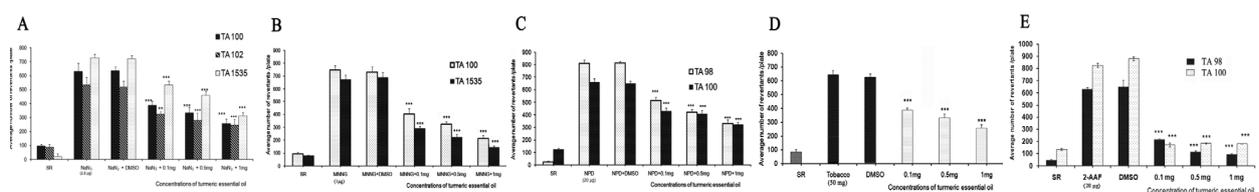


Figure 1. Antimutagenic Activity of Turmeric Essential Oil on Mutagenicity Induced by NaN_3 , MNNG, NPD, Tobacco and 2-AAF in *Salmonella* Strains. (A) NaN_3 , Sodium azide. DMSO, dimethyl-sulfoxide (Vehicle control); (B) MNNG, N-Methyl-N'-nitro-N-nitrosoguanidine; (C) NPD, 4-Nitro-O-phenylenediamine; (D) tobacco; (E) 2-AAF, 2-acetamidofluorene; Data shown are mean values with bars indicating the standard deviation of the mean (n=3). *** $p < 0.001$, ** $p < 0.01$ when compared with positive control

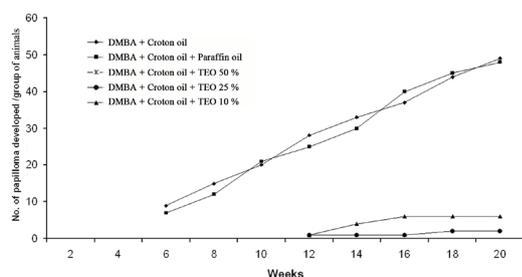


Figure 2. Chemopreventive Effect of Turmeric Essential Oil (TEO) on the Onset of Skin Papilloma Formation after Topical Application of 7, 12-dimethylbenz (a) Anthracene (DMBA) (470 nmol/mouse, dissolved in 200 µl acetone) and 1% Croton Oil in mice

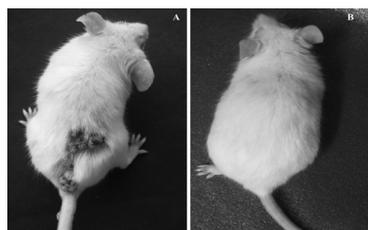


Figure 3. Effect of Turmeric Essential Oil (TEO) on Skin Papilloma Induction Initiated by Topical Application of 7, 12-dimethylbenz (a) Anthracene (DMBA) (470 nmol/mouse, dissolved in 200 µl acetone) and Promoted by 1% Croton Oil in mice. (A) DMBA and croton oil treated mice with skin papilloma. (B) DMBA + Croton oil+50% TEO treated mice with no papilloma

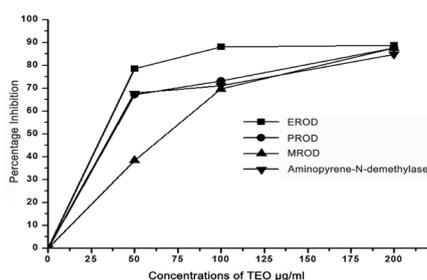


Figure 4. Effect of Turmeric Essential Oil (TEO) on Inhibition of Cytochrome p450 Enzymes *in vitro*

Table 2. Inhibition of Cytochrome p450 Enzymes by TEO (*in vitro*)

Cytochrome p450 enzymes	IC ₅₀ Values of TEO
CYP1A2 (MROD)	66 µg
CYP2B 1/2 (PROD)	36 µg
CYP1A1 (EROD)	31 µg
CYP 1A, 2A, 2B, 2D and 3A (Aminopyrene-N-demethylase)	45 µg

*MROD, 7-methoxyresorufin-O-demethylase; PROD, pentoxy resorufin by 7- pentoxyresorufin O-depentyase; EROD, ethoxy resorufin by 7-ethoxyresorufin O-deethylase

We selected three different concentrations of TEO - 0.1, 0.5 and 1 mg/plate for evaluating antimutagenicity. All the strains of *S. typhimurium* (TA 98, TA 100, TA 102 and TA 1535) produced several fold revertant colonies compared to spontaneous revertant after treatment with mutagens. TEO showed significant antimutagenic activity against mutagens such as NaN₃, MNNG, NPD, 2-AAF and tobacco on all the 4 *Salmonella* strains studied with or without activations in a concentration dependent manner. DMSO alone (vehicle control) did not produce any antimutagenic activity.

TEO inhibited NaN₃-induced mutagenicity by 70.3% (TA100), 65.8% (TA102) and 59.2% (TA 1535) at a concentration of 1 mg/plate, (Figure 1A). MNNG induced mutagenicity was inhibited by 81.6% (TA100) and 89.1% (TA1535) (Figure 1B) and NPD-induced mutagenicity by 62.6% (TA98) and 63.1% (TA100) by TEO at a concentration of 1 mg/plate (Figure 1C). TEO also significantly exhibited antimutagenicity (81.7%) against tobacco extract in *Salmonella* strain TA 102 at the concentration of 1mg/plate (Figure 1D).

TEO exhibited strong (P<0.001) antimutagenic activity against 2-AAF after metabolic activation. TEO inhibited mutagenicity by 92.6 % and 94.5% at the concentration of 1mg/ plate in *Salmonella* TA 98 and TA 100 strains respectively. The percentage inhibition of antimutagenic activity of TEO against 2AAF is shown in Figure 1E.

Determination of the effect of TEO on DMBA and croton oil induced papilloma development in mice

Papilloma development in mice was started 6th week in control and vehicle control group. Onset of papilloma was found to be significantly delayed or prevented by TEO treatment. In TEO (10 and 25%) treated groups, formation of papillomas were started in the 12th week after DMBA application. DMBA alone, croton oil alone and DMBA+croton oil+50% TEO treated groups did not produce any papilloma on mice. At the end of 20th week the average number of papilloma per mouse was 7±1.06 and 6.86±1.04 in control and vehicle control group respectively and this number was reduced to 1.2±0.5 and 1±0.3 by topical application of TEO 10% and 25% respectively (Figure 2). Inhibition of development of papilloma was 82.9, 85.7 and 100% in 10, 25 and 50% TEO treated groups respectively (Figure 3).

Effect of TEO on the inhibition of various isoforms of cytochrome p450 enzymes

Administration of phenobarbitone increased the level of cytochrome p450 enzymes EROD, MROD and PROD in rats, when compared with untreated group. Oral gavage of TEO significantly inhibited EROD activity. Percentage inhibition of EROD at various concentration of TEO such as 50, 100 and 200 µg/ml was 78.7%, 80.6% and 83.8% respectively (Figure 4). TEO significantly inhibited MROD activity (P<0.001) and 80% inhibition was shown at 200 µg/ml. Similarly activity of PROD was also inhibited by TEO at various concentrations which were 63.5%, 66.4% and 81.1%. Aminopyrene-N-demethylase activity is an indicator of CYP 1A, 2A, 2B, 2D and 3A. TEO inhibited the activity of aminopyrene-N-demethylase significantly (P<0.001) at various concentration (50, 100 and 200 µg/ml) (Figure 4). IC₅₀ values of MROD, PROD, EROD and aminopyrene-N-demethylase by TEO is given in Table 2.

Discussion

Cancer is one of the major health problems in the present world. The main reason for the cause of cancer is genetic mutation especially influenced by industrial and environmental pollutants. Mutagenicity indicates

perpetual transmissible changes in the structure of genetic material in the cells of living organisms. Prolonged exposure to chemical mutagens present in agricultural, food, medicine and industry causes DNA mutations that can lead to carcinogenesis. Mutagenic potential of substances can be detected through Ames assay which is a very fast and economical test (Ames et al., 1975). It is assessed that 90% of the mutagens are carcinogens. Secondary metabolites from medicinal plants and spices have significant role in preventing cancer by acting as chemopreventive agents. Chemopreventive agents can be targeted for intervention at any of stage of carcinogenesis stage i.e., initiation, promotion or progression (Sulaiman et al., 2012).

Curcuma longa is a well known traditional spice with more than 1700 research articles published about this spice. TEO isolated from rhizome of *Curcuma longa* by steam distillation, has significant pharmacological and medicinal properties (Funk et al., 2010). In the present study, we have evaluated the antimutagenic and anticarcinogenic potential of TEO. We have also studied the effect of TEO in the regulation of cytochrome p450 enzymes *in vitro*.

NaN_3 is a potent mutagen in plants, bacterial species and a mammalian cell as well as it has high specificity in *Salmonella* and *E. coli* species. MNNG can produce DNA adduct formation and may cause gastrointestinal tract related cancer (Matsuyama et al., 1970; Lucchesi et al., 1986). It is present in detergent, cosmetic items and cigarettes smoke etc. NPD is mutagen present in cosmetics and induced frame shift mutation in *Salmonella typhimurium* (TA98 and TA100). Tobacco contains different compounds; most of them are identified as cancer causing. Moreover 2-Acetylaminofluorene (2-AAF) is a potent arylamine carcinogen and activation is done by cytochrome p450 enzymes. Antimutagenic studies using different auxotrophic *Salmonella* strains indicated that the TEO significantly inhibited the mutagenicity produced by all the direct acting mutagens, sodium azide, NPD, MNNG and tobacco and also inhibited the activation of 2-AAF by rat liver S9 fraction in a dose dependent manner. The antimutagenic property of essential oil may be due to inactivation of mutagens by inhibition of free radicals or activation of cellular antioxidant enzymes, inhibition of cytochrome p450 enzymes or activation of detoxification of mutagens (Shankel et al., 1993; Water et al., 1996; Ipek et al., 2005). TEO is also known to possess significant *in vivo* and *in vitro* antioxidant activity (Liju et al., 2011). Significant antimutagenic activity of TEO may be linked to anticarcinogenic potential.

The pro-carcinogens are known to under go bioactivation by cytochrome p450 enzymes and converted to active carcinogens. Cytochrome p450 is a superfamily most of which are present in liver and plays an important role in the metabolism of drugs as well as endogenous and exogenous chemicals. Well-known pro-carcinogenic compounds such as aflatoxin, 3-methylcholanthrene (3-MC), 2-AAF, DMBA are metabolized by the action of isoforms of cytochrome p450 enzymes and converted into potent carcinogenic compounds. Activation of pro-mutagen 2-AAF is done by the presence of enzymes

CYP1A2 in liver (Firdous et al., 2010; Yueh et al., 2001). CYP1A2 is the most efficient cytochrome p450 enzyme catalysing the N-hydroxylation of the dietary heterocyclic amines. Moreover Cytochrome p450 enzymes such as CYP1A1 and 1B1 usually metabolize polycyclic aromatic hydrocarbons (3-MC) into carcinogen (Thorgeirsson and Nebert, 1977). Present study revealed that level of cytochrome p450 enzymes such as CYP1A2, CYP2B 1/2, CYP1A1, CYP2A, CYP2D and CYP3A are significantly reduced after treated with TEO in a concentration dependent manner. Reactive oxygen species plays an important role as initiators as well as promoters of mutagenesis and carcinogenesis and our previous study indicated that TEO significantly scavenge reactive oxygen species *in vitro* and *in vivo* (Arya and Kumar, 2011; Liju et al., 2011). Antioxidants are generally acted as antimutagenic and anticarcinogenic agents due to their free radical scavenging properties (Ames et al., 1993; Clark, 2002).

DMBA is a polycyclic aromatic hydrocarbon and mostly used for studies in carcinogenic animal model experiment (Hamizah et al., 2012). DMBA is metabolized by CYP1A1 and CYP1B1, which convert DMBA to the ultimate carcinogen 1, 2-epoxide-3, 4-diol DMBA (Anqus et al., 1999; Buters et al., 1999; Kawajiri and Ikuta, 1999; Sharma et al., 2012), which form adducts with DNA and it leads to carcinogenesis (Cheng et al., 1988). Suppression of CYP1A1 enzyme activity by TEO can reduce the effect of DMBA induced carcinogenesis. In present study we have showed that the incidence and number of skin papillomas formation after treated with 7, 12 dimethylbenz[a]anthracene and promoted by croton oil was significantly decreased (10 and 25% TEO; $p < 0.001$) and absolutely prevented (50% TEO treated group) by treatment with TEO in mice. This implies that TEO may have either inhibited the metabolism of DMBA to its active form, or delayed the promotion phase of carcinogenesis (Agrawal and Sonam, 2009).

Detoxification and elimination of DMBA are done by phase II enzymes such as glutathione-s-transferase (GST) and UDP-glucuronyl transferase. Previous study showed that TEO administration enhanced phase II enzymes. Inhibition of cytochrome p450 enzymes and promotion of phase II enzymes in liver would inhibit the metabolic activation of DMBA and prevent cancer formation. These results suggest the role of TEO as chemopreventive and anticancer agent. It is already reported that TEO did not show any toxicity and genotoxicity in rats up to concentration of 500 mg/kg body weight (Liju et al., 2013). FDA has accepted that TEO can be used as food additive and is recommended as GRAS.

Previous study indicated that TEO did not produce any mutagenicity to *Salmonella* strains with and without activation of S9 mixture (Liju et al., 2013). In this study we have shown TEO has strong antimutagenic potential against know chemical mutagens such as NaN_3 , MNNG, NPD and tobacco as well as 2-AAF which needs metabolic activation. Moreover TEO significantly delayed and prevented the skin papilloma development in DMBA and croton oil treated mice. Antimutagenicity and anticarcinogenic activity of TEO may be due to its

antioxidant activity as well as suppression of mutagen inducing metabolizing cytochrome p450 enzymes such as CYP1A1, CYP1A2, CYP2B, CYP2A, CYP2D and CYP3A.

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