Response to Therapy in Breast Cancer

Norbert Avril1, Stefanie Sassen2, and Rebecca Roylance3

1Department of Nuclear Medicine, Barts and The London School of Medicine, Queen Mary, University of London, London, United Kingdom; 2Department of Pathology, Technische Universität München, Munich, Germany; and 3Centre for Molecular Oncology and Imaging, Institute of Cancer, Barts and the London School of Medicine and Dentistry, London, United Kingdom

Increasing numbers of patients with newly diagnosed breast cancer receive primary systemic therapy followed by surgery. Histopathology provides an accurate assessment of treatment efficacy on the basis of the extent of residual tumor and regressive changes within tumor tissue. However, only approximately 20% of breast cancer patients achieve a pathologic complete response, a fact that necessitates methods for monitoring therapeutic effectiveness early during therapy.18F-FDG PET and 18F-FDG PET/CT provide essential information regarding a response to primary chemotherapy. Patients with low tumor metabolic activity on pretreatment 18F-FDG PET are not likely to achieve a histopathologic response. The degree of changes in 18F-FDG uptake after the initiation of therapy is correlated with the histopathologic response after the completion of therapy. Thus, tumor metabolic changes assessed early during therapy predict therapeutic effectiveness in individual patients. Early identification of ineffective therapy also might be helpful in patients with metastatic breast cancer because many palliative treatment options are available. Changes in metabolic activity generally occur earlier than changes in tumor size, which is the current standard for the assessment of a response. Although treatment stratification based on a metabolic response is an exciting potential application of PET, specific PET response assessment criteria still need to be developed and validated on the basis of patient outcomes before changes in treatment regimens can be implemented. There is increasing clinical evidence for metastatic breast cancer and other tumors that 18F-FDG PET/CT is the most accurate imaging procedure for assessment of the response at the end of treatment when both CT information and tumor metabolic activity are considered. Importantly, in the setting of primary chemotherapy, neither PET/CT nor conventional imaging procedures can assess the extent of residual breast cancer as accurately as histopathology. Observation of changes in tumor blood flow or tumor cell proliferation is an additional encouraging approach for predicting a response. Ultimately, the prediction of therapeutic effectiveness by PET and PET/CT could help to individualize treatment and to avoid ineffective chemotherapies, with their associated toxicities.

Key Words: breast cancer; treatment monitoring; histopathology;18F-FDG;15O-water; PET/CT

DOI: 10.2967/jnumed.108.057240

The treatment of newly diagnosed breast cancer is primarily determined by the extent of disease and generally includes surgery—either breast-conserving surgery, usually combined with postoperative radiation therapy, or total mastectomy. Axillary lymph node dissection has now been replaced by sentinel lymph node biopsy in many patients presenting without clinically apparent axillary tumor involvement. Adjuvant systemic therapy after surgery is frequently recommended to reduce the risk of tumor recurrence, especially in patients with lymph nodes showing positive findings, tumors larger than 1 cm, or other unfavorable prognostic features.

Primary systemic chemotherapy was first introduced for managing inoperable and large or locally advanced breast tumors (1). It is also now increasingly being used for tumors that, although operable, would require mastectomy; therefore, it is used to try to reduce tumor size and therefore permit breast-conserving surgery. In this situation, the term “neoadjuvant chemotherapy” is often used. The primary aim of treatment is a reduction in tumor volume (2), but disease-free survival, overall survival, and locoregional control are similar to those achieved with initial surgery followed by conventional adjuvant chemotherapy (3).

Primary systemic chemotherapy followed by surgery allows for assessment of the histopathologic response. Although approximately 70% of patients demonstrate a clinical response either on physical examination or on anatomic imaging, only 3%—27% achieve a complete histopathologic response (2,4–6). The degree of the response provides important prognostic information, and patients achieving a complete pathologic response show significantly longer disease-free overall and overall survival than nonresponders (7,8).

A recent review of data presented at the National Cancer Institute State of the Science Conference on preoperative therapy in invasive breast cancer indicated that preoperative systemic therapy in operable breast cancer is effective and can improve breast conservation rates (9). A variety of clinical, imaging, and pathologic measurements can be used to assess the response of a tumor to treatment; in particular, a pathologic complete response may be useful as a surrogate endpoint in the evaluation of new therapies. However, at
present, there are no data to suggest that systemic treatment should be tailored on the basis of the initial tumor response or the extent of residual disease.

For patients with metastatic breast cancer, several therapeutic options are available (10,11). The choice of treatment depends on histopathologic tumor features—that is, whether tumors are estrogen receptor positive or human epidermal growth factor receptor 2 (HER2) positive—and the extent of disease. Three groups of tumors are generally treated with different regimens: triple-negative breast cancer, which is defined as estrogen receptor–negative, progesterone receptor–negative, and HER2-negative tumors; HER2-positive tumors; and estrogen receptor–positive tumors and a small volume of metastatic disease are considered for treatment with endocrine agents (11).

**ASSESSMENT OF RESPONSE TO TREATMENT**

Histopathology is often used as the reference standard for assessing the response to primary chemotherapy in breast cancer. However, there is no single definition of a histopathologic response, and response criteria vary among studies. Most commonly, a pathologic complete response is defined by the absence of residual invasive tumor (2,5,6). A modified classification was suggested by Honkoop et al., who found no difference in survival between patients with scattered microscopic foci of residual tumor cells and patients who achieved a pathologic complete response (14). These 2 groups of responses were combined into a response category referred to as “minimal residual disease,” and all other responses were classified as “gross residual disease.”

Other response classifications include changes in tumor cellularity (15–17) and the presence of regressive changes in residual tumor tissue (18,19). However, regressive morphologic features, such as hyaline fibrosis, inflammatory cell infiltrates, iron-storing macrophages, or calcifications, are observed in approximately 50% of cases after chemotherapy (6,20) and have limited sensitivity (7%–67%) and specificity (48%–99%) compared with nontreated tumor specimens (6,21). Although the extent of residual viable tumor is the most important predictor of long-term outcome (2,8,15,22–25), histopathologic response criteria have distinct limitations. The rate of complete pathologic response is relatively low, ranging from 3% to 27% (2,4–6). Between 13% and 25% of patients will experience a systemic recurrence during 5 y of follow-up (8,23,26,27). Furthermore, a subgroup of patients classified as not showing a histopathologic response will remain disease free but have not been recognized by current histopathologic criteria.

In the absence of tissue specimens, generally the case in the setting of metastatic disease, the most frequently used surrogate endpoint for a therapeutic effect is a change in tumor size. The Response Evaluation Criteria in Solid Tumors (RECIST) define a response as a decrease in the maximum tumor diameter of at least 30% (28). Anatomic imaging, predominantly CT, MRI, and ultrasound, is used to obtain measurements of reference tumor lesions before and after treatment for response assessment and follow-up.

Variable results have been found for the agreement between anatomic imaging and pathologic tumor size (29). A large retrospective review found no evidence that mammography or ultrasound performed better than physical examination in measuring residual disease after chemotherapy (29). Breast MRI has been found to be more accurate for estimating the extent of residual disease (30). However, MRI cannot rule out residual tumor tissue with sufficient accuracy (30–34). Another approach currently under investigation is dynamic contrast-enhanced MRI. More than a 65% reduction in tumor volume and a reduction in the early enhancement ratio after 2 cycles of chemotherapy were associated with a histopathologic response (35). Combining both criteria yielded a diagnostic accuracy of 93% for identifying tumors showing a pathologic complete response (35).

The use of RECIST is established for response assessment in the setting of metastatic disease. However, several cycles of treatment are needed before a change in tumor size can be assessed by anatomic imaging. Moreover, changes in the sizes of metastases in bone and liver, which are the most common sites of breast cancer metastases, are difficult to evaluate.

**MONITORING RESPONSE TO TREATMENT WITH 18F-FDG PET AND PET/CT**

Metabolic Response to Primary Systemic Therapy of Breast Cancer

The prediction of a response to treatment refers to the early identification of treatment effectiveness through a comparison of the levels of radiotracer uptake before and after 1 or 2 cycles of systemic therapy. The concept of using 18F-FDG PET for predicting a therapeutic response is based on early changes in tumor glucose use and the close correlation of changes in 18F-FDG uptake with the effectiveness of treatment (36,37). At least 2 sequential 18F-FDG PET scans are currently necessary for the prediction of a treatment response; one is obtained before treatment to serve as a baseline, and one is obtained after the initiation of chemotherapy, for example, after the first or second cycle.

Fifteen years ago, Wahl et al. (36) reported on changes in tumor metabolic activity in a series of 11 women who had locally advanced primary breast cancers and who had received a combination of primary chemotherapy and hormone therapy. Tumor 18F-FDG uptake promptly decreased in 8 patients, with subsequent partial or complete pathologic responses, whereas tumors in 3 nonresponding patients did not show a significant decrease in 18F-FDG uptake. Later studies confirmed a more pronounced decrease in 18F-FDG uptake (standardized uptake value
Table 1 summarizes trials reporting on the response. A decrease in $^{18}$F-FDG uptake (SUV) of 40% alterations in tumor size for monitoring a treatment re-
of a treatment failure might be clinically more important.

The accuracies for the prediction of a histopathologic response were 88% after the first cycle of chemotherapy and 91% after the second cycle. In another study, 30 breast cancer patients received 8 cycles of primary chemotherapy, and the mean reduction in $^{18}$F-FDG uptake after the first cycle was significantly higher in lesions with a partial, complete macroscopic or complete microscopic response (39). In a recent report, 64 breast cancer patients underwent $^{18}$F-FDG PET after the first, second, and third cycles of chemotherapy (40). Relative changes in tumor $^{18}$F-FDG uptake were superior to alterations in tumor size for monitoring a treatment response. A decrease in $^{18}$F-FDG uptake (SUV) of 40% predicted a histopathologic response with accuracies of 77% after the first cycle of chemotherapy and 87% after the second cycle. Table 1 summarizes trials reporting on the prediction of a histopathologic response by $^{18}$F-FDG PET.

In comparisons of those studies, the definition of a treatment response as a reference for a metabolic response is critical. Importantly, the histopathologic response criteria used in those PET studies varied widely, with response rates ranging from 16.3% to 55.6% (38–41). Translation of the observations described earlier into clinical practice is currently difficult because there is no agreement about the optimal timing of PET, that is, whether it should be performed after the first, second, or third cycle of chemotherapy. In addition, different thresholds for the relative changes in tumor metabolic activity (SUV) used to predict a response were used in those PET studies. Although it is essential to investigate the ability of $^{18}$F-FDG PET to identify patients responding to systemic therapy, the detection of a treatment failure might be clinically more important.

A recent prospective multicenter trial in which 272 $^{18}$F-FDG PET scans were performed for 104 patients confirmed that the greater the reduction in tumor metabolic activity early in the course of therapy, the more likely that patients would achieve a histopathologic response (42). In patients who showed a histopathologic response, the SUV decreased by 50.5% ± 18.4% (mean ± SD) after the first cycle of primary chemotherapy; in comparison, the SUV decreased by 36.5% ± 20.9% in nonresponders. Patients who did not show a histopathologic response were identified with a negative predictive value of 89.5% after the first cycle of therapy when a relative decrease in the SUV of less than 45% was used as a cutoff. Correspondingly, the negative predictive value after the second cycle of therapy was 88.9% when the cutoff was a 55% decrease in the SUV. Interestingly, a metabolic response after one cycle of therapy predicted outcomes in patients regardless of whether they continued to receive a combination of epirubicin and paclitaxel or received a planned switch to epirubicin followed by paclitaxel. A high negative predictive value is essential for clinical decision making to ensure the continuation of treatment in all patients potentially responding to therapy.

An important observation of this multicenter trial (42) is that $^{18}$F-FDG PET identified patients with low tumor metabolic activity before treatment as not achieving histopathologic response. Twenty-four of 104 breast carcinomas (23%) had a baseline SUV of less than 3.0, and none responded to chemotherapy. These tumors were more likely to be well differentiated and estrogen receptor positive. These findings mean that breast cancers with low metabolic activity identified by $^{18}$F-FDG PET before treatment are not likely to respond to primary chemotherapy. Therefore, for patients with such cancers, it is possible that initial management should be different; for example, if the cancers are operable, patients should undergo surgery immediately or perhaps should have primary hormone therapy. These findings are supported by the results of smaller series additionally reporting that it is difficult to measure changes in $^{18}$F-FDG uptake in tumors with low pretherapy $^{18}$F-FDG uptake (40,41,43). In the multicenter trial (43), a positive correlation was also found between the histopathologic response and the level of $^{18}$F-FDG uptake after the first and second cycles of chemotherapy, but no defined SUV threshold was clearly superior for the optimal separation of responders and nonresponders during chemotherapy.

The main rationale for primary chemotherapy is to test for chemosensitivity, allowing for subsequent changes in the chemotherapy regimen, with the aim of designing a more individualized treatment plan. In the only trials in which early changes in primary chemotherapy were implemented, a clinical response was used to guide treatment decisions (16,44,45). It is still unclear which patients would benefit most from an early change in treatment, such as a switch to a non–cross-resistant or second-line chemotherapy regimen, because there was no advantage of a switch for most nonresponders (45). Nevertheless, patients showing a clinical response to primary chemotherapy benefited from either the addition of a non–cross-resistant chemotherapeutic agent (such as a taxane) or prolonged treatment (4,46). For the establishment of therapy modifications based on response assessments during chemotherapy, a tool more suitable than the clinical response is desirable. Metabolic PET could serve as such a method and could provide a clinically useful surrogate endpoint. There is clearly a need for prospective randomized trials to evaluate and validate PET-guided treatment stratification for identifying subsets of patients who would benefit most from early treatment modifications.

### Metabolic Response to Therapy of Metastatic Breast Cancer

There are significant differences between the goals of primary systemic therapy of newly diagnosed breast cancer and the goals of treatment of metastatic disease. Generally,
A few chemotherapeutic regimens are used for primary systemic therapy, but many palliative treatment options are available for metastatic disease. Histopathology is used as a surrogate endpoint for the validation of primary chemotherapy, whereas almost always no tissue specimens are available from metastatic disease for the evaluation of a response.

Only a few studies have reported on the clinical utility of sequential 18F-FDG PET in patients with metastatic disease. A decrease in tumor 18F-FDG uptake was observed in 8 of 12 patients who had metastatic disease and who responded after the first cycle of chemotherapy (47). In another study, a rapid decrease in tumor glucose metabolism was observed after the first cycle of therapy in 6 of 9 responding patients, but there was no substantial decrease in nonresponding patients (48). Specht et al. (49) retrospectively reviewed 28 patients who underwent serial 18F-FDG PET during systemic therapy for bone-dominant metastatic breast cancer. The treatments were heterogeneous, including endocrine therapy, chemotherapy, biologic therapy, and bisphosphonates. Smaller relative decreases in the SUV (or increases in the SUV) were associated with a shorter time to progression. A patient with no change in the SUV was twice as likely to progress as a patient with a 42% median decrease in the SUV (49).

Table 2 summarizes trials reporting on the prediction of a response in metastatic breast cancer by 18F-FDG PET.

The results of a recent study confirmed previous observations on the predictive value of information about early changes in glucose metabolism for metastatic breast cancer (50). Compared with the baseline PET data, the 18F-FDG uptake in responding metastatic lesions decreased to 72% ± 21% after the first cycle of chemotherapy and to 54% ± 16% after the second cycle. In contrast, the 18F-FDG uptake in metastases not responding to chemotherapy declined only to 94% ± 19% after the first cycle of chemotherapy and to 79% ± 9% after the second cycle. 18F-FDG PET correctly predicted the responses in all patients after the first cycle of chemotherapy and was more accurate than conventional imaging procedures after the third cycle of chemotherapy.
potential benefit most from the early identification of ineffective treatments, particularly in cases of metastatic breast cancer, because various alternative treatment options are available; in addition, patients could be spared the toxicities of ineffective treatments.

Weber et al. proposed defining the metabolic response as a decrease in the SUV larger than 2 times the SD (20%) of spontaneous changes in 18F-FDG uptake (51, 52). In a study of breast cancer, a decrease in 18F-FDG uptake of more than 20% compared with the baseline correctly identified 5 of 7 nonresponding lesions (71.2%) and 12 of 14 responding lesions (85.7%) after the first cycle of chemotherapy (50). A specific strength of 18F-FDG PET appears to be the identification of nonresponders, who are characterized by virtually no change in 18F-FDG uptake after the initiation of chemotherapy. The overall survival of patients not showing a metabolic response was 8.8 ± 6.7 mo, and that of patients showing a metabolic response was 19.2 ± 13.6 mo (50). An important finding of that study (50) was that 14 cycles of chemotherapy could be avoided in 5 nonresponding patients.

Importantly, the pattern of changes in glucose metabolism also depends on the type of therapeutic agent. For example, a transient increase in glucose metabolism followed by a decrease in 18F-FDG uptake has been observed for tumors responding to hormone therapy with tamoxifen. Dehdashti et al. (53) studied 11 women with 18F-FDG PET before and approximately 1 wk after the initiation of tamoxifen therapy. Seven patients who responded on the basis of clinical and radiologic follow-up showed an initial increase in 18F-FDG uptake 1 wk after the initiation of therapy. An explanation for this “metabolic flare” phenomenon is that tamoxifen has initial agonist effects before antagonist effects dominate. Clinically, this effect occurs within the first 1–2 wk of treatment and is characterized by erythema of soft-tissue lesions and increased pain in osseous metastases. The same group of investigators confirmed their findings in a larger series (54). In patients responding to tamoxifen, tumor 18F-FDG uptake increased by 28.4%, and only 5 of these patients showed evidence of a clinical metabolic flare reaction. In nonresponding patients, no significant change in tumor 18F-FDG uptake compared with the baseline was observed. Importantly, so far there have been no reports of a metabolic flare in chemotherapeutic regimens, nor is the response known to occur with the newer endocrine agents (such as the aromatase inhibitors), which are now the treatment of choice for postmenopausal women.

In a recent comparison of 18F-FDG PET and 16α-[18F]fluorooestradiol-17β (18F-FES) PET, both the metabolic flare after estradiol challenge and the level of 18F-FES uptake before endocrine therapy predicted the response to therapy (55). 18F-FES uptake in breast cancer reflects estrogen receptor status, and an SUV of ≥2.0 is considered positive for estrogen receptor expression. Fifty-one postmenopausal women with advanced estrogen receptor–positive breast cancer underwent 18F-FES PET and 18F-FDG PET scans at baseline and a second 18F-FDG PET

<table>
<thead>
<tr>
<th>Tumor type or stage</th>
<th>Authors</th>
<th>Year</th>
<th>No. of patients</th>
<th>PET timing</th>
<th>PET criterion</th>
<th>Outcome measure</th>
<th>Design</th>
<th>PET performance</th>
<th>Response rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bone-dominant metastatic breast cancer</td>
<td>Specht et al. (49)</td>
<td>2007</td>
<td>28</td>
<td>During treatment</td>
<td>No change in SUV (vs. 42% decrease)</td>
<td>Disease progression</td>
<td>Retrospective</td>
<td>Hazard ratio, 2.2 ($P = 0.006$)</td>
<td>13/28 (46%)*</td>
</tr>
<tr>
<td>Metastatic breast cancer</td>
<td>Dose Schwarz et al. (50)</td>
<td>2005</td>
<td>11</td>
<td>After first cycle</td>
<td>Visual analysis</td>
<td>Clinical response</td>
<td>Prospective</td>
<td>Accuracy, 100%</td>
<td>6/11 (55%)†</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>After first cycle</td>
<td>Relative decrease in SUV</td>
<td>Clinical response</td>
<td></td>
<td>28% vs. 6% ($P = 0.02$)§</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>After second cycle</td>
<td>Relative decrease in SUV</td>
<td>Clinical response</td>
<td></td>
<td>46% vs. 21% ($P = 0.003$)§</td>
<td></td>
</tr>
<tr>
<td>Metastatic breast cancer</td>
<td>Gennari et al. (48)</td>
<td>2000</td>
<td>9</td>
<td>After first cycle</td>
<td>Absolute decrease in SUV</td>
<td>Clinical or radiographic response</td>
<td>Prospective</td>
<td>SUV decrease from 7.7 to 5.7‡</td>
<td>6/9 (67%)†</td>
</tr>
</tbody>
</table>

*Response was defined as no progression of bone metastases during median follow-up time of 17.5 mo.
†Clinical response based on changes in tumor size assessed by conventional imaging.
‡Relative decrease in SUV in patients showing clinical response vs. nonresponders.
§Absolute decrease in SUV in patients showing clinical response vs. no change in nonresponders.

TABLE 2. Prediction of Clinical Response of Metastatic Breast Cancer by Relative or Absolute Changes in 18F-FDG Uptake on 18F-FDG PET
scan after the administration of 30 mg of estradiol. A ≥12% increase in the SUV for 18F-FDG represented a metabolic flare and was therefore considered to indicate a response. After the estradiol challenge, only the responders showed a metabolic flare, with a significantly higher percentage change in the SUV for 18F-FDG in responders (20.9% ± 24.2%) than in nonresponders (−4.3% ± 11.0%). On 18F-FES PET, significantly higher uptake was noted in responders (SUV, 3.5 ± 2.5) than in nonresponders (SUV, 2.1 ± 1.8). 18F-FES PET appears to be a promising approach for noninvasively determining the estrogen receptor status of metastatic breast cancer and predicting the response to endocrine therapy. Further trials are needed to clarify whether patients with low 18F-FES uptake would benefit from more aggressive chemotherapy regimens.

**TREATMENT-INDUCED CHANGES IN TUMOR BLOOD FLOW**

The Seattle group has extensively studied 15O-water PET perfusion measurements in breast cancer and compared these measurements with glucose metabolism measured as the rate of 18F-FDG metabolism (56–59). In 31 patients who underwent 15O-water perfusion and 18F-FDG PET before the initiation of primary chemotherapy, high glucose metabolism normalized to blood flow was associated with a significantly poorer response to therapy (56).

Subsequently assessed changes in the rate of 18F-FDG metabolism during primary chemotherapy revealed no significant correlation with histopathologic responses at the completion of treatment (57). In comparisons of relative changes in metabolic rates at baseline and midtherapy (2 mo after the initiation of chemotherapy), patients showing a histopathologic response had an average decrease in the metabolic rate of 65%; for patients showing a partial response and patients not showing a response, the average decreases were 49% and 40%, respectively. These findings are in line with those of previous studies demonstrating a good correlation between a decrease in tumor glucose metabolism measured as the SUV and a histopathologic response (38–40). On the other hand, tumor blood flow decreased by 53% in patients showing a histopathologic response and by 12% in patients showing a partial response but increased, on average, by 45% in patients not showing a response. An important finding of that study (57) was that patients whose tumors failed to show a decrease in tumor blood flow after 2 mo of therapy had shorter disease-free survival and overall survival.

A series of 53 patients with locally advanced breast cancer were evaluated for changes in the rate of 18F-FDG metabolism, 18F-FDG blood-to-tissue transport (18F-FDG $K_t$), and tumor blood flow before and 2 mo after the initiation of primary chemotherapy (59). Patients who achieved a complete pathologic response had average decreases of 84% in the rate of 18F-FDG metabolism, 79% in 18F-FDG $K_t$, and 76% in tumor blood flow. In contrast, the average changes in nonresponding patients were 62% (rate of 18F-FDG metabolism), 19% (18F-FDG $K_t$), and 14% (tumor blood flow). Patients who had no significant decrease in tumor blood flow or 18F-FDG $K_t$ had a higher risk of recurrence and a higher mortality rate. Changes in tumor perfusion predicted disease-free survival and overall survival. The primary additional finding of that study (59) was the correlation of response and outcome with perfusion measurements, even more so than with changes in metabolism. The relationship between metabolism and blood flow was also shown to be predictive. The ability to predict responses on the basis of changes in tumor blood flow is encouraging, but further validation, including the definition of thresholds for changes in tumor perfusion, is needed to prove accurate identification of responses to therapy.

**MONITORING RESPONSE BY IMAGING CHANGES IN TUMOR CELL PROLIFERATION**

Imaging changes in tumor cell proliferation is an interesting new approach for monitoring therapeutic effects. Of the currently available PET proliferation tracers, 18F-fluorothymidine (18F-FLT) has been the most extensively evaluated (60). In one of the first reports about 18F-FLT PET in breast cancer, its use for diagnostic purposes was assessed (61). Increased 18F-FLT uptake was noted in primary and metastatic breast cancer; however, the level of uptake was lower than that of 18F-FDG. A subsequent study revealed increased 18F-FLT uptake predominately in larger primary tumors and axillary lymph node metastases (62).

18F-FLT PET for treatment monitoring in metastatic breast cancer detected changes in breast cancer proliferation 1 wk after the initiation of combination chemotherapy with 5-fluorouracil, epirubicin, and cyclophosphamide (63). Eight patients were scanned twice before chemotherapy, and 18F-FLT uptake in 9 discrete tumor lesions was found to be reproducible, with a high test–retest correlation coefficient. Six patients had a significant reduction in 18F-FLT uptake at 1 wk and a partial or complete clinical response later during treatment. In a study of 14 patients with newly diagnosed primary or metastatic breast cancer, 18F-FDG PET and 18F-FLT PET were performed at baseline, 2 wk after the first cycle of a new treatment regimen, and after completion of the regimen (64). Changes in 18F-FLT uptake in primary and metastatic tumors after the first cycle were significantly correlated with the overall response. An interesting observation was that in most cases, the maximum tumor 18F-FLT uptake assessed by dynamic data acquisition was reached within 10 min after tracer injection. The authors concluded that 18F-FLT PET performed 2 wk after the end of the first cycle of chemotherapy is useful for predicting the longer-term efficacy of chemotherapy in breast cancer.
To define the role of $^{18}$F-FLT PET in treatment monitoring in breast cancer, further validation with more patients is needed. A potential advantage of the use of $^{18}$F-FLT is the early identification of therapeutic effectiveness, particularly in the evaluation of new biologic agents.

**ASSESSMENT OF RESPONSE TO TREATMENT WITH $^{18}$F-FDG PET AND PET/CT**

There are limited data on the role of $^{18}$F-FDG PET and PET/CT in the assessment of a response after the completion of therapy in breast cancer. However, comparisons with conventional imaging procedures used for staging (CT, ultrasound, and MRI) revealed that metabolic whole-body PET has distinct advantages (Fig. 1). A recent meta-analysis of the diagnostic performance of $^{18}$F-FDG PET in metastatic breast cancer included 18 articles published from January 1995 to June 2004 (65). In the studies with patient-based data, the median sensitivity was 92.7%, and the median specificity was 81.6%. The pooled sensitivity was 90%, and the pooled false positivity rate was 11% after the exclusion of outliers. The maximum combined sensitivity and specificity was 88%.

A distinct advantage of $^{18}$F-FDG PET is the ability to assess tumor viability after previous treatments. Bony metastases are particularly difficult to evaluate because radiologic appearances often persist despite successful treatment. In a recent study, 146 bone metastases in 25 patients with recurrent breast cancer identified by $^{18}$F-FDG PET/CT were monitored (66). After treatment, 58 osteolytic lesions (80.5%) became $^{18}$F-FDG negative and osteoblastic on CT, with only 14 relatively large lesions (19.5%) remaining $^{18}$F-FDG avid. Of the 25 $^{18}$F-FDG–avid osteoblastic lesions, 13 (52%) became $^{18}$F-FDG negative, but 12 (48%) remained $^{18}$F-FDG avid and increased in size on CT. Five of the mixed-pattern lesions remained $^{18}$F-FDG avid after treatment. All 17 CT-negative lesions became $^{18}$F-FDG negative; however, 9 of them became osteoblastic. None of the initially $^{18}$F-FDG–negative lesions showed increased metabolic activity during follow-up. The findings of that study (66) indicated that it is likely that $^{18}$F-FDG uptake reflects the immediate tumor activity of bone metastases, whereas radiographically revealed morphologic changes vary greatly with time in patients. Another group of investigators recently compared morphologic and metabolic changes in bone metastases in response to systemic therapy in 102 patients (67). The progression of sclerotic changes after treatment was identified in 49 patients (48%). Increases in attenuation on CT were significantly correlated with decreases in $^{18}$F-FDG uptake, and a decrease in the SUV after treatment was an independent predictor of a response.

With the increasing availability of PET/CT, the decision to use this imaging modality is becoming easier. There is increasing clinical evidence for breast cancer and other tumors that $^{18}$F-FDG PET/CT after treatment is the most accurate procedure for the assessment of a treatment response when both CT information and tumor metabolic activity are considered.

**CONCLUSION**

In the United States, health insurance providers reimburse for monitoring of the response to treatment in breast cancer as an adjunct to standard imaging modalities for women with locally advanced and metastatic breast cancer when a change in therapy is anticipated. However, specific $^{18}$F-FDG PET criteria for determining a response or ineffective therapy have yet to be established. Generally, it appears to be possible to predict a response after the first cycle of chemotherapy. Such timing would have a notable clinical impact because patients receiving ineffective treatment could be switched to alternative therapies and responding patients could receive more aggressive chemotherapy.

There are several potential clinical applications for the PET-based prediction of a response to treatment with primary chemotherapy. First, metabolic imaging could identify breast cancer with low tumor metabolic activity before treatment; patients with such cancer might be more suitable candidates for surgery or hormone therapy. Second, patients for whom PET predicts a poor response after the first cycle of chemotherapy could be switched to alternative therapies earlier. The Aberdeen neoadjuvant chemotherapy trial (16) demonstrated improved overall survival with the addition of taxanes. In particular, patients showing clinical partial and complete responses benefited, but those not showing clinical responses did not greatly benefit. The use of metabolic imaging might offer better stratification of patients for optimal treatment and an earlier switch from ineffective chemotherapy.

**FIGURE 1.** (Top row) Soft-tissue mass showing intense increase in $^{18}$F-FDG uptake in left parasternal location in 59-year-old woman with prior left mastectomy. (Bottom row) Mass decreased in size after completion of chemotherapy, and no abnormal metabolic activity was visualized—results consistent with good response to treatment. Panels from left to right represent $^{18}$F-FDG PET, CT, and fused $^{18}$F-FDG PET/CT images.
Importantly, the prediction of a clinically meaningful response to a particular chemotherapy regimen or drug remains a challenge. The concept of a response to treatment and therapy monitoring is complicated by the lack of an unequivocal definition of a response. For example, there is a fundamental difference between overall treatment efficacy and a complete pathologic response or a response assessed by changes in tumor size on the basis of RECIST. In a neoadjuvant chemotherapy trial (National Surgical Adjuvant Breast and Bowel Project Protocol B-27) (4), a 2-fold increase in the rate of a pathologic complete response was observed when a taxane was added to a standard anthracycline-based chemotherapy regimen. However, this increase did not translate into an increase in overall survival. This finding raises the question of which response criteria should be used in assessments of novel drugs, new treatment regimens, or novel techniques, such as metabolic imaging. The response to chemotherapy is a continuous variable rather than a dichotomized one. Therefore, it may be important to establish response criteria that relate tumor response to patient outcome to guide subsequent treatment decisions. Therapy-induced changes in tumor glucose metabolism could play such a role.

The critical issue in the use of 18F-FDG PET for monitoring and prediction of a response to treatment in the setting of metastatic disease is the accurate identification of ineffective treatment. For example, if a threshold of a 20% decrease in the SUV after the first cycle of therapy is used to identify ineffective therapy, then serial 18F-FDG PET will not identify all nonresponding patients because some tumors might exhibit an initial decrease in 18F-FDG uptake. On the other hand, clinically applicable thresholds need to be set so that treatment is not switched in patients who may be responding to first-line therapy. Therefore, cutoff thresholds for relative changes in 18F-FDG uptake (SUV) must be carefully chosen and validated in prospective trials with a sufficient number of patients before chemotherapy can be discontinued or a treatment regimen can be changed. Finally, more information about PET treatment monitoring of endocrine and new biologic agents, such as trastuzumab, is also needed.

The current use of 18F-FDG PET and PET/CT for treatment monitoring in breast cancer varies among institutions and reflects different practice in various countries. Important requirements include sufficient experience and dedication of the PET team to ensure the reproducible quantitation of tumor 18F-FDG uptake. Generally, baseline 18F-FDG PET before systemic therapy provides an accurate assessment of the extent of disease, and 18F-FDG PET approximately 6 wk after the completion of therapy accurately reflects the response to treatment. Prediction of the therapeutic response by 18F-FDG PET after the first or second cycle of therapy as well as early changes to alternative therapies in patients not showing a metabolic response should be done only in the context of clinical trials.

REFERENCES


