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Ras Is both Necessary and Sufficient for the Inhibition of Myeloid Differentiation Mediated by Ras

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Hyperactivation of Ras is one of the most common abnormalities in acute myeloid leukemia. In experimental models, Ras inhibits myeloid differentiation, which is characteristic of leukemia; however, the mechanism through which it disrupts hematopoiesis is poorly understood. In multipotent FDCP-mix cells, Ras inhibits terminal neutrophil differentiation, thereby indefinitely extending their proliferative potential. Ras also strongly promotes the sensitivity of these cells to granulocyte-macrophage colony-stimulating factor (GM-CSF). Using this model, we have dissected the signaling elements downstream of Ras to determine their relative contribution to the dysregulation of hematopoiesis. Cells expressing Ras mutants selectively activating Raf (Ras*T35S) or phosphatidylinositol 3-kinase (Ras*Y40C) did not significantly affect differentiation or proliferative capacity, whereas Ras*E37G (which selectively activates RafGEFs) perpetuated proliferation and blocked neutrophil development in a manner similar to that of Ras. Correspondingly, expression of constitutively active versions of these effectors confirmed the overriding importance of Ras guanine nucleotide exchange factors. Cells expressing Ras demonstrated hyperactivation of Ral, which itself was able to exactly mimic the phenotype of Ras, including hypersensitivity to GM-CSF. Conversely, dominant negative Ral promoted spontaneous neutrophil development. Ras, in turn, appears to influence differentiation through multiple effectors. These data show, for the first time, the importance of Ras in regulating differentiation and self-renewal in hematopoietic cells.

The 21-kDa guanine nucleotide-binding Ras proteins function as membrane-bound molecular switches linking the transduction of extracellular signals, via receptor and nonreceptor tyrosine kinases, to downstream effectors regulating diverse processes such as differentiation, proliferation, and cell cycle progression (7). Direct activation of Ras via point mutation is among the most common molecular abnormalities in hematopoietic malignancy, with an incidence of around 30% in myeloid leukemia and preleukemia (9, 25, 40, 58). Hyperactivation of Ras in leukemia also arises as a result of constitutive upstream signals arising from other common abnormalities such as those involving the Flt3 receptor (22, 46) and the Bcr-Abl fusion (24, 90). Conversely, Ras also becomes constitutively active as a result of loss of GTPase-activating protein (GAP) activity as seen in neurofibromatosis patients, who lack Nf1 Ras-GAP expression. These patients are at high risk of developing juvenile myelomonocytic leukemia (JMML) (65). Ras mutations also arise at high frequency in the preleukemic condition known as myelodysplastic syndrome (68), characterized by disorders of development resulting in hyperplastic marrow and peripheral blood cytopenia. The association of hyperactive Ras with both leukemia and preleukemia suggests that Ras signaling plays an important role during hematopoietic dysregulation and leukemic transformation.

Activated Ras genes are most commonly associated with promotion of proliferation of a variety of cell types. However, in the context of myeloid leukemia it has long been established that there are no differences in the proliferative capacity of leukemic cells compared to their normal counterparts. The accumulation of malignant cells is instead due to the failure of cells to undergo terminal differentiation and apoptosis (5). There is evidence from both in vitro and in vivo models that Ras influences the development of monocytic, erythroid, and granulocytic cells. This is substantiated by observations of primary mouse and human cells, where Ras appears to promote monocytic differentiation (10, 21), as well as to oppose the differentiation of erythroid progenitors (17, 88). In addition, neutrophilic leukocytosis has been observed with K-Ras transgenic mice (12), implying that Ras also affects the development of granulocytic cells.

Another commonly observed effect of Ras on myeloid hematopoietic cells is the capacity to reduce the requirements for exogenous growth factors (55, 73). This may arise in part from the fact that many hematopoietic cytokine receptors act through Ras, as previously reported for granulocyte-colony-stimulating factor (G-CSF), granulocyte-macrophage CSF (GM-CSF), and interleukin 3 (IL-3) receptors (18, 39, 72). Cytokines are known to influence the development of hematopoietic cells (37), with dysregulated cytokine signaling identified as a common feature of acute myeloid leukemia, ranging from mutational activation of growth factor receptor, as exemplified by the Flt3 receptor (28, 59, 89), to hyperresponsiveness to cytokine, such as the enhanced response to GM-CSF seen with JMML (23).

We have previously studied the effects of Ras on the multipotent factor-dependent cell line, FDCP-mix. These cells possess a nonleukemic phenotype, have a normal karyotype, are factor dependent, and can be directed to undergo multilinage development in response to cytokines (77). By changing growth conditions (serum and cytokines), it is possible to grow
this cell line under self-renewing conditions, such that they retain their multipotent state; alternatively, they may be directed to differentiate to form erythroid or myeloid cells. In these cells, Ras selectively inhibits granulocytic differentiation of neutrophils, giving rise to continued proliferation of these partly differentiated cells. In addition, Ras also selectively increases the cells’ sensitivity to GM-CSF, suggesting a link between the heightened response to this cytokine and an inhibition of differentiation (16). This phenotype therefore closely resembles that seen with JMML, which also commonly arises from hyperactivation of Ras (58).

As described above, the effects of Ras on myeloid development have been well documented; however, the mechanism by which it elicits these changes is poorly understood. Ras interacts with multiple downstream effectors via a highly conserved amino-terminal loop domain (switch I), encompassing amino acid residues 32 to 40 (83). The three best-established effector families are the Raf serine-threonine kinases, phosphatidylinositol 3-kinases (PI3-kinases), and the Ras guanine nucleotide exchange factor (RalGEF) family (54). In the context of myelopoesis, activation of the mitogen-activated protein kinase (MAPK) cascade via Ras has been implicated in both disruption of differentiation and changes in cytokine dependency (20, 21, 83). PI3-kinase activity has been linked with both differentiation and inhibition of apoptosis (14, 20, 50) with demonstration of the synergistic requirement for Raf-mediated activation in leukemogenesis (6). Comparatively little has been reported regarding the role of RalGEFs in hematopoiesis, although they have been associated with both Jak/Stat signaling and the promotion of survival following growth factor withdrawal (33, 55). This family of effectors acts as GEFs for the Ras GTPases. Two highly similar Ras proteins, RalA and RalB, associate with a variety of downstream effectors (for a review, see reference 26): Sec5 and Exo84 during formation of the Exocyst complex, which participates in delivering vesicles to the plasma membrane (60, 61); the actin-binding protein, filamin (67); the ZO-1-associated nucleic acid-binding protein (designated ZONAB) (29), and Ral-binding protein 1 (RalBP1), which mediates GAP activity towards the Rho family GTPases Rac and Cdc42 (27).

Here, we show for the first time that Ras is a critical developmental regulator in hematopoietic cells. Ras promotes the activity of this GTPase through RalGEF family members. Further, activation of Ras is both necessary and sufficient to mimic the inhibition of neutrophil development imposed by Ras and is also responsible for promoting the sensitivity of these cells to GM-CSF. The data demonstrate an as-yet-unidentified role for Ras in controlling hematopoietic differentiation.

**MATERIALS AND METHODS**

**Cell culture.** Culturing of FDCP mix (clone A4) cells (77) under self-renewing conditions was carried out in Iseove’s modified Dulbecco’s medium (Sigma) supplemented with 20% batch-tested horse serum (GibcoBRL) and 10% WEHI-3B conditioned medium and maintained at 37°C with 5% CO2. Under these conditions, >95% of cells exhibited an immature blast cell morphology. Analysis of the effect of ectopically expressed genes on differentiation was carried out as follows. Mid-logarithmic-phase cells were harvested and washed twice in 20 ml Hanks buffered saline solution containing 25 mM HEPES buffer (GibcoBRL), and resuspended to 2 × 108 cells/ml in 5 ml of differentiation medium: Iseove’s modified Dulbecco’s medium containing 20% batch-tested fetal calf serum, 1,000-U/ml human G-CSF, and 50-U/ml mouse GM-CSF (R&D Systems). Cells were subcultured if necessary in the same medium. Automated May-Grunwald-Giemsa staining (Bayer) of cytospins (20,000 cells) was carried out at day 7 (or as indicated), and morphological assessments were made under blind conditions. Cell proliferation was determined by viable cell counts as assessed by trypan blue exclusion, as previously described (4).

**Sensitivity to cytokines and inhibitors.** To assess cytokine sensitivity, mid-logarithmic-phase cells were harvested and washed twice in 20 ml Hanks buffered saline solution. These were resuspended to 2 × 108 cells/ml in 5 ml differentiation medium (as above but excluding cytokines). Recombinant mouse IL-3 (R&D Systems) at a final specific activity of 0 to 1,000 U/ml, human G-CSF (0 to 10,000 U/ml), or mouse GM-CSF (0 to 500 U/ml) was added. GM-CSF hyperresponsiveness was assessed in the presence of constant G-CSF (50 U/ml), since GM-CSF alone did not support the viability of these cells. Cumulative proliferative capacity was determined by cell counts at day 5. The effect of the Rac inhibitor (nic23766; Calbiochem) was assessed by addition at the start of the 7-day differentiation assay. The effect of PI3-kinase and MEK inhibitors was studied on FDCP-mix Ras* cells preincubated for 7 days under differentiating conditions. These were exposed to LY294002 or PD98059 (Sigma) or U0126 (Cell Signalling Technology). Morphological and proliferative end points were scored 3 days later as described above.

**cDNA constructs.** Ras effector loop domain mutants and PI3-kinase* (K227E) (both provided by Julian Downward, London, United Kingdom) were expressed in plXSN. The c-Raf1 catalytic domain was expressed in the pG5 retroviral vector (56). RalGEFs (Michael A. White, Dallas, Tex., and Johannes L. Bos, The Netherlands) and constitutively active and inhibitory RalA constructs (Chris J. Marshall, London and David Wynford-Thomas, Cardiff, Wales) were all subcloned into the retroviral expression vector pHABE- puro. RalB, RabA Q72L, RabA Q72L DON, and the RalA effector loop domain mutant (Rala A Q72L, DON), and RalA Q72L, AN1 constructs, all in pBABE-puro, were kindly provided by Christopher Counter (Duke University Medical Center).

**Retrovirus infection.** FDCP-mix-Ras* and neomycin control lines were created as previously described (16). The remaining lines were created via the generation of replication-defective amphotropic retrovirus by transient transfection of the Phoenix-Ampho packaging line (American Type Culture Collection). This was subsequently used to infect the parental FDCP-mix line as described below. Non-tissue-culture 24-well plates were treated with 15 μg of RetroNectin (Takara Biochemicals) per well and incubated at room temperature for 2 h. RetroNectin was replaced by 1% bovine serum albumin, and incubation was carried out for 30 min for blocking. The blocking solution was replaced by 400 μl of retroviral supernatant together with 10⁵ cells at semilogarithmic growth phase in 1.5 ml of culture medium. This was incubated overnight at 37°C with 5% CO2, after which the infection procedure was repeated using fresh retroviral supernatant. Following infection, antibiotic selection was carried out in the presence of puromycin (2 μg/ml) or neomycin (800 μg/ml).

**Activation assays and Western blotting.** Cells were lysed in 50 mM Tris (pH 7.4), 1% NP-40, 15% glycerol, 200 mM NaCl, 5 mM MgCl2, 5 mM NaF, 1 μM leupeptin, 0.1 μM aprotinin, and 1 mM phenylmethylsulfonyl fluoride. Detergent-insoluble material was removed by centrifugation (16,000 × g at 4°C for 20 min). Activation sensitive pull-down assays were carried out using the binding domain of RalBP1 (amino acids 397 to 518) and PKD (amino acids 1 to 232) (both were kindly provided by Chris J. Marshall, London, United Kingdom), which specifically interact with active GTP-bound forms of Ras and Rac/Cdc42 proteins, respectively, essentially as described previously (71, 86). Glutathione S-transferase-binding domain–coupled beads were prepared by mixing at 4°C for 1 h, while the pull-down was carried out by mixing with the whole-cell lysate for 45 min. GTP proteins were eluted using 2× reducing sample buffer, for 5 min at 95°C, and assessed by Western blotting (12% sodium dodecyl sulfate–polyacrylamide gel electrophoresis). Comparative total protein content was determined by parallel Western blotting of the whole-cell lysates. Immunoprecipitation was carried out using the following antibodies: Ras G12V-sensitive monoclonal (Calbiochem), pMek1/2 (#2354), pErk1/2 (#4695), and pAkt/PKB (#9271), all from Cell Signaling Technology and RabA (BD Transduction Laboratories), Rac1 (Upstate Biotechnology), and Cdc42 (Santa Cruz Biotechnology).

**RESULTS**

***Activation of Ras guanine nucleotide dissociation stimulator (RalGDS) mediates Ras-induced inhibition of differentiation.*** FDCP-mix cells undergo either self-renewal or differentiation, depending on the presence of hematopoietic growth factors. In the presence of G-CSF plus GM-CSF, these cells undergo predominantly neutrophil development, culminating in terminal differentiation and growth arrest. Expression of mutant...
Ras (Ras*) strongly inhibits the capacity of these cells to undergo terminal neutrophil development under these conditions (Fig. 1A and E). To determine which downstream targets of Ras influenced the development of these cells, we repeated these experiments using cells expressing effector mutants of Ras. Selective activation of Raf (Ras*T35S) or PI3-kinase (Ras*Y40C) did not significantly affect differentiation capacity; these cells consequently underwent terminal neutrophil development and growth arrest. In contrast, Ras*E37G (selectively activating RalGEFs-phospholipase C [PLC]/H9280) inhibited neutrophil development (Fig. 1A), with concomitant increase in proliferation capacity (Fig. 1B). This observation was not due to the relative overexpression of Ras*E37G, as demonstrated by Western blotting (Fig. 2A); indeed, normalized expression of Ras*E37G was marginally lower than that of Ras*T35S or Ras*Y40C. Therefore, the Ras*E37G mutant is sufficient to replicate the antidifferentiation phenotype resulting from activation of Ras.

The Ras*E37G effector loop domain mutant has been previously shown to interact with a group of guanine nucleotide exchange factors for the Ral GTPase (e.g., RalGDS and Rlf) (84), as well as the phosphoinositide-specific PLCε (15, 45, 75). However, PLCε RNA is undetectable in FDCP-mix cells, as assessed by reverse transcription-PCR (data not shown), suggesting that RalGEFs were the principal target in this context.

FIG. 1. Differentiation and proliferation of FDCP-mix cells in the presence of Ras loss of function and constitutively active effector mutants. Morphological scoring and proliferative expansion of FDCP-mix cells in the presence of Ras loss-of-function mutants (A and B) and constitutively active mutants (C and D). Cells were seeded at $2 \times 10^4$/ml in medium containing G-CSF plus GM-CSF, and differentiation was assessed following May-Grunwald-Giemsa staining. Significant differences versus FDCP-mix-Neo cells were defined as follows: *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.005$; ****, $P < 0.001$. Error bars represent standard deviations from the mean of at least three independent experiments. (E) Morphology (magnification, $\times 400$) after 7 days under differentiating conditions. Control cultures (Neo) are dominated by large segmented mature neutrophils (n). Cells expressing Ras* either fail to differentiate (b) or give rise to immature neutrophils with doughnut-shaped nuclei characteristic of mouse metamyelocytes (m); these cells retain proliferative capacity (16). Macrophage differentiation is unaffected (mac).
Active Ral is necessary and sufficient for dysregulated myelopoietic development. The above data indicate a correlation between active Ral and dysregulated hematopoiesis in FDCP-mix cells. To substantiate whether Ral is necessary and/or sufficient to inhibit neutrophil differentiation, we examined the effect of activated and dominant negative versions of RalA. The constitutively active Ral mutants used were that which is insensitive to Ral-GAP activity (mutated at Q72L) and the “fast-cycling” mutation (F39L) (52, 53, 70). Both RalA mutants examined mimicked the blocked development and increased proliferative capacity demonstrated by FDCP-mix-Ras* (Fig. 3A). This was also associated with increased proliferative capacity (Fig. 3B). The effects of dominant negative Ral constructs on the parental FDCP-mix cells substantiated these observations. Expression of dominant negative Ral (RalBP1-RalBD) in conjunction with Ras* abrogated the capacity of Ras* to inhibit differentiation and promote differentiation (Fig. 3A and B). Expression of dominant negative Ral constructs alone (RalBP1-RalBD or RalA S28N) promoted spontaneous differentiation of FDCP-mix cells grown under self-renewing conditions (IL-3) compared to the empty vector control line (Fig. 3C and D). Interestingly, these constructs also promoted monocyte differentiation. Therefore, constitutive inhibitory RalA is able to compromise the self-renewing capacity of FDCP-mix cells, resulting in spontaneous development. Recent work has demonstrated functional differences between the roles of RalA and RalB in the transformation of human cells (51). We therefore examined the possibility that there may also be differences in the capacity of these proteins to affect hematopoietic differentiation by repeating these experiments with the corresponding constitutively active RalB mutants. In this context, however, we found RalB to be just as effective as RalA (Fig. 3E and F).

Together, these data suggest that activation of RalA (or that of RalB) directly inhibits neutrophil differentiation and that its activity is necessary to maintain FDCP-mix cells in a self-renewing state.

Like Ras*, both RalGDS-CAAX and RalA F39L enhance GM-CSF sensitivity. Previously, it was shown that the block in neutrophil development in this model was associated with an increased sensitivity to GM-CSF and that the block in development was dependent upon the continued presence of this cytokine (16). To establish the possible role of the downstream effectors of Ras in augmenting cytokine responsiveness, the growth of the aforementioned cell lines was examined by dose-response assays. We observed that expression of RalGDS-CAAX promoted hyperresponsiveness to GM-CSF in a manner similar to that seen with cells expressing Ras* (P < 0.01) (Fig. 4A). Similarly, cells expressing constitutively active Ral (RalA F39L) also replicated the increase in GM-CSF sensitivity seen in Ras* (Fig. 4B). In contrast, the absolute requirement of all these cell lines for exogenous IL-3 remained, with no significant difference in the dose-response data between Ras loss-of-function mutants (data not shown) or constitutively active mutants versus control cells (Fig. 4C). The data suggest that expression of Ras* selectively increased the sensitivity to GM-CSF and that both increased sensitivity to GM-CSF and differentiation arrest are mediated via the Ral pathway.

Role of downstream effectors of Ral in the inhibition of neutrophil differentiation. Mutants of Ral which have im-

FIG. 2. Expression and activity of effector mutants of Ras and constitutively active downstream targets of Ras. (A) Mutant Ras expression was probed using a mutant H-Ras-specific antibody. (B) Raf activity was investigated using the phosphorylation-specific antibodies to MEK1/2 and ERK1/2. (C) p13-kinase activity was assessed using a pAkt/PKB antibody. (D) The effect of constructs activating RalGDS/Ral on Erk1/2 and pAkt/PKB activity was similarly determined. (E) Raf activity was determined using the GTP-sensitive probe as described in Materials and Methods. Results are representative of two independent experiments.
paired capacity to activate effectors downstream of Ral have been previously described; deletion of the N-terminal amino acids (RalAAAT) reduces phospholipase D1 binding (41), the RalAD49E mutant is defective in binding Sec5 and Exo84 (60, 61), and the RalAD49N mutant has diminished affinity for RalBP1 (11). We examined the phenotype conferred by constitutively active (RalAQ72L) versions of these mutants to indicate the contribution of these Ral effectors to the dysregulation of hematopoiesis. A mutant lacking the biologically essential CAAX motif acted as the negative control in these experiments. Each mutant appeared to be expressed at similar levels against the relatively high endogenous levels of RalA in these cells (Fig. 5A). Compared to RalAQ72L, we found that each of the effector mutants compromised the activity of constitutively active RalA in the inhibition of neutrophil differentiation (Fig. 5B). Correspondingly, none of the effector mutants retained any capacity to promote the proliferation of these cells under differentiating conditions (Fig. 5C). In common with RalAQ72L, these mutants did not affect the proliferative rate of these cells under self-renewing conditions (data not shown). The simplest interpretation of these data is that multiple effectors of Ral contribute towards its ability to inhibit differentiation and promote self-renewal of hematopoietic cells.

Given that activation of one of these Ral effectors, RalBP1, has been previously associated with inhibition of differentiation (32), we investigated the individual contribution of this effector to the observed inhibition of differentiation. Activation of RalBP1 functions as a GAP towards the Rho-related proteins Rac and Cdc42. Hyperactivation of Ral by Ras should therefore result in negative regulation of Rac1 and Cdc42 activity if these proteins are in fact a target of Ral in this context. We therefore examined the active-protein levels via binding-domain pull-down assays. This analysis revealed that whereas control FDCP-mix cells (Neo) expressed these Rho-related proteins in their active (GTP-bound) state, the lines expressing Ras* and RalGDS-CAAX were almost completely deficient in GTP-bound Rac1 and Cdc42 (Fig. 6A). The comparative difference in expression of these GTP-bound proteins between the Rif-CAAX and RalGDS-CAAX lines showed that the lat-

FIG. 3. Effect of RalA and RalB mutants on the development of FDCP-mix cells. (A and B) The effect of constitutively active mutants (F39L and Q72L) of RalA on the development of FDCP-mix cells and the effect of dominant negative RalA (RalBP1-RalBD) on FDCP-mix cells coexpressing Ras* are shown; conditions are as described in the legend to Fig. 1. (C) Spontaneous differentiation under self-renewing conditions of FDCP-mix cells expressing dominant negative RalA constructs alone. (D) Corresponding morphology of cells grown under self-renewing conditions is shown. Control cells (pBABE-puro) demonstrate a uniform blast cell appearance, while RalA S28N cells exhibit a variety of differentiated forms including mature neutrophils (n), macrophages (mac), and disintegrating forms of segmented neutrophils (a). Morphology was scored as described in Materials and Methods. (E and F) Effect of constitutively active mutants of RalB on FDCP-mix cells; conditions are as described in the legend to Fig. 1. Significant differences versus FDCP-mix–Neo cells are indicated as follows: *, P < 0.05; **, P < 0.01. Error bars represent standard deviations from the mean of at least three independent experiments.
ter effector more closely mimicked the effects of Ras*. Interestingly, we also observed that PI3-kinase* weakly reduced GTP-bound levels of Rac1 and Cdc42. These observations correlate with the weak inhibitory effect on neutrophil development described above (Fig. 1C) and suggest a degree of cross talk from this effector.

Recently, a highly selective cell-permeable inhibitor for Rac1, NSC23766, has become available (31); we therefore examined the influence of Rac1 inhibition on FDCP-mix development. NSC23766 strongly inhibited neutrophil differentiation in a dose-dependent manner (Fig. 6B), however, at higher doses, it was also antiproliferative (Fig. 6C). Nevertheless, the degree of inhibition at low doses was at least consistent with a role for Rac1 inactivation in mediating part of the antidifferentiation effects of Ras.

PI3-kinase activity is required to maintain the differentiation block imposed by Ras. Experiments using activated effectors and effector mutants of Rac suggested that hyperactivation of PI3-kinase was not sufficient to block neutrophil differentiation; however, the data in Fig. 6A and 1C indicated that...
PI3-kinase activity did participate in this process. We therefore examined the requirement for both PI3-kinase activity, as well as MAPK activity, to maintain the differentiation block imposed by Ras. We treated differentiation-blocked FDCP-mix–Ras* cells with inhibitors to either PI3-kinase or MEK. Inhibition of PI3-kinase with LY294002 strongly promoted differentiation (Fig. 7A) and correspondingly inhibited proliferation (Fig. 7B), indicating that the maintenance of the differentiation-inhibited phenotype was dependent on PI3-kinase activity. In contrast, inhibition of MEK with PD98059 had no significant effect on the differentiation of these cells, despite the fact that it inhibited their proliferation (Fig. 7B). The MEK1/2 inhibitor, U0126, gave similar results (data not shown). These indicate that PI3-kinase activity is required to maintain the antiderivation phenotype elicited by Ras. Correspondingly, MAPK activity does not appear to play a role in inhibiting the differentiation of these cells. These data are consistent with the lack of influence of activated Raf and the corresponding effector mutant of Ras on neutrophil development (Fig. 1). They are also consistent with the fact that Ras itself does not constitutively promote MAPK activity in this context (Fig. 2).

**DISCUSSION**

Although there is some understanding of the processes which regulate hematopoietic differentiation at a transcriptional level, little is known of the intracellular events which regulate this developmental programming. These are influenced by environmental factors such as the presence of cytokines and stromal factors (3). Since many of these environmental signals are transduced via Ras proteins, it is perhaps not surprising that in cells expressing constitutively active Ras these signals become “distorted,” leading to a dysregulation of development. Previous work from our laboratory established that mutational activation of Ras selectively inhibited neutrophil development and concomitantly increased sensitivity to GM-CSF (16). In this study, we identify the downstream targets of Ras, which mediate these changes in developmental programming and cytokine responsiveness. We examined three well-characterized Ras effector proteins (Raf, PI3-kinase, and RaLGGEFs) and their related pathways (54). Studies using effector mutants of Ras indicated that it was RaLGGEF signaling which was shown to have the predominant role in the dysregulated developmental mediated by Ras. Although the use of these effector mutants allows the dissection of signaling activity in a relatively nonmanipulative manner (in that the expression level and activity of downstream targets are not artificially altered), the use of these mutants does not provide direct evidence of the interacting proteins (including as-yet-unidentified effectors). We therefore repeated these experiments using constitutively active Raf, PI3-kinase, and RaLGGEFs. Data from these experiments replicated the phenotypes of the corresponding effector loop domain mutants, both substantiating our initial observations and definitively identifying RaLGGEFs as the key effector in mediating this phenotype. Although the involvement of these RaLGGEFs or their putative effector, Ral, during hematopoiesis has not been previously reported, their ubiquitous expression has been identified within the relevant compartments including bone marrow, spleen, and thymus (2, 85).

The involvement of the RaLGGEFs implicated Ral signaling as being important for neutrophil development in FDCP-mix cells. Use of the activation-specific probe for Ral confirmed the association of increased Ral activity with the expression of Ras and RaLGGEFs. Furthermore, we also demonstrated that promoting the activity of Ral by using constitutively active mutants was itself sufficient to block neutrophil development. Conversely, expression of dominant negative Ral promoted differentiation of FDCP-mix cells even when cultured under self-renewing conditions. These data suggest that the activity of this GTPase is essential to maintain the self-renewing potential of these cells. Given that a universal property of leukemic blasts is their capacity for self-renewal, these data suggest that Ral activity may be an important factor in maintaining the self-renewing potential of leukemic cells. This is the first demonstration of a role for Ral in hematopoietic differentiation and self-renewal; however, Ral has been previously implicated as a developmental regulator in Drosophila melanogaster eye (57) and in inhibiting differentiation of PC12 cells, as defined by neurite outgrowth (32). In the context of cellular transformation, Ral activity has a long-established role (30, 80, 84); this role may be more significant in human cells (35, 69). The function of Ral activity has been assessed in a variety of contexts: a RaLGDS knockout study has shown a role for Ral activity in inhibiting apoptotic death (34), and distinct functions for RaLa and RaLb have been identified, with RaLb being required for suppression of apoptosis while RaLa is required for anchorage-independent proliferation (13). RaLa has also been reported to have a preferential affinity for exocyst complex (74), although it is possible that in the context of overexpression of Ral, differences in affinity may not be limiting. Recent work has also identified a dominant role for RaLa in the transformation of human cells (51).

We have not so far established how Ral ultimately inhibits neutrophil development. Studies employing effector mutants...
of Ral suggest that this GTPase influences neutrophil development through a variety of effectors. Previous work has shown that all the established Ral effectors are able to influence differentiation. Increase in phospholipase D1 activity is closely associated with neutrophil differentiation (64, 66). The exocyst complex has not yet been directly linked with hematopoietic development, but it is associated with neurite outgrowth in neuronal and PC12 cells (47, 82). In both these instances, however, the increase in effector activity is associated with promoting rather than inhibiting differentiation. RalBP1 is thought to act as a GAP (or negative regulator) towards the Rho superfamily of Rac1/Cdc42 GTPases (43). These Rho family proteins have been previously associated with differentiation of epithelial cells (78), myoblasts (19, 36, 76), dendrites (49), and dorsal root ganglia (42, 81) and during PC12 neurite formation (87). It might therefore be predicted that increased RalBP1 activity would be associated with an inhibition of differentiation in these contexts; in the PC12 model, there is indeed evidence that RalGEFs inhibit differentiation by suppressing the activity of Rac and Cdc42 (32). Similarly, we present evidence that inhibition of Rac1 activity does suppress differentiation in FDCP-mix cells.

The surprisingly negligible contribution of the Raf and PI3-kinase effectors to hematopoietic dysregulation in this study may be due to the prevailing importance of the Ral pathway in this context. Alternatively, optimum activation of Raf and PI3-kinase may already be present in FDCP-mix cells; therefore, increasing their activity via Ras may be of little significance. In the context of the Raf effector pathway, it is interesting to note that, as with primary leukemias (38), the presence of activated Ras did not provoke constitutive activation of MAPK in FDCP-mix cells; even inhibition with MAPK inhibitors failed to influence the differentiation-inhibited status of FDCP-mix Ras* cells. These data, together with the lack of effect of constitutive activation of Raf, indicate that this effector is not playing a role in the regulation of neutrophil differentiation, although it may, as previously suggested (63), play a role in maintaining their proliferative capacity. In contrast to Raf, activation of PI3-kinase via the Ras*Y40C effector mutant or the constitutively active version of this kinase did provoke a modest antidifferentiation effect in these cells, implying that this effector may also participate in this process. These data were supported by the observation that inhibition of PI3-kinase activity reversed the effect of Ras on neutrophil differentiation, indicating that the maintenance of the antidifferentiated state was at least dependent on PI3-kinase activity. Since Ras has also been shown to be activated in a PI3-kinase-dependent manner in neutrophils (62), this may indicate that multiple effectors of Ras (RalGEFs and PI3-kinases) may converge on this GTPase. This possibility is also supported by the observation that expression of constitutively active PI3-kinase also suppressed Ral1 activation and, to a certain extent, Cdc42 activation. It has been suggested that PI3-kinase may participate in RalGDS activation by promoting its association with PI3-kinase-dependent kinase 1, which in turn appears to relieve autoinhibition of the catalytic domain of RalGDS (79).

A further observation of interest relates to the increased sensitivity to GM-CSF in cells expressing constructs that directly or indirectly activated Ral. We have previously shown that the influence of GM-CSF is crucial in maintaining the block in differentiation, in that its withdrawal leads to spontaneous terminal differentiation. These observations have interesting parallels with that of JMML. In JMML, 20 to 30% of cases are associated with loss of Nf1 Ras-GAP function, leading to hyperactivation of Ras. JMML-derived progenitor cells display a marked hypersensitivity towards GM-CSF, as shown by CFU-GM assays (23), with a similar phenotype demonstrated with Nf1−/− mouse cells (8, 48). While these studies strongly suggest that heightened sensitivity to GM-CSF in JMML is explicitly mediated via Ras (44), there is uncertainty as to the mechanism through which this arises. In Myb-transformed Nf1−/− murine cells, increased sensitivity appears to arise from autocrine production of GM-CSF that was dependent on Raf/MEK/ERK signaling (20). However, only around 50% of JMML patients demonstrate excessive GM-CSF production, despite the fact that they universally show increased sensitivity to this cytokine. In the model described here, we could find no evidence of autocrine production of this cytokine, also suggesting that additional and/or alternative mechanisms exist. In this context, it will be of interest to determine the activation state of Ral in Nf1−/− cells and whether their heightened response to GM-CSF is dependent on the activity of this GTPase.

Despite the variety of approaches aimed at dysregulated Ras signaling in a clinical setting (1), none has proved particularly effective so far. Therefore, the identification of alternative pharmacological targets may be of significance. In the context of leukemia, such a target may be the small GTPase Ral.

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